

A Baylis–Hillman/ozonolysis route towards (±) 4,5-dihydroxy-2,3-pentanedione (DPD) and analogues

Marine Frezza, Laurent Soullère, Yves Queneau and Alain Doutheau*

Laboratoire de Chimie Organique, UMR CNRS-UCBL 5181, Institut National des Sciences Appliquées,
20 avenue A. Einstein, 69621 Villeurbanne, France

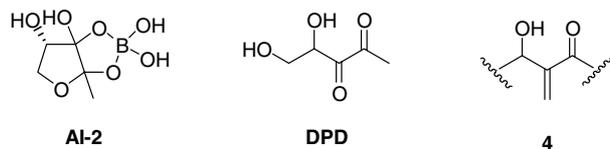
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Abstract—The Baylis–Hillman reaction between 2-(*tert*-butyldimethylsilyloxy)ethanal and 3-buten-2-one followed by desilylation gave rise to the corresponding α -methylene- β,γ -dihydroxy ketone further converted by reductive ozonolysis of the carbon–carbon double bond into racemic 4,5-dihydroxy-2,3-pentanedione (DPD), a significant molecule in bacterial cell–cell communication systems. The same sequence applied to other substrates allowed the preparation of chain elongated analogues and 5-*O*-acylated derivatives of DPD.

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Many bacterial species are able to sense their own population size through a cell–cell communication system, named quorum sensing (QS), based on the production and exchange of diffusible extracellular signaling molecules called autoinducers.¹ Since a link between this process and virulence factor production² or biofilms formation³ has been established for a number of pathogenic bacteria, the design of QS inhibitors has recently attracted considerable interest for the development of anti-infection therapies.⁴ Among the various autoinducers which have been identified so far, a borate, known as autoinducer-2 (AI-2), formed from the metabolic product (*S*) 4,5-dihydroxy-2,3-pentanedione (DPD) (Scheme 1), is produced by a large number of both Gram-negative and Gram-positive bacteria.⁵

As part of a programme aimed at obtaining QS inhibitors,^{6,7} we recently became interested in designing a



Scheme 1.

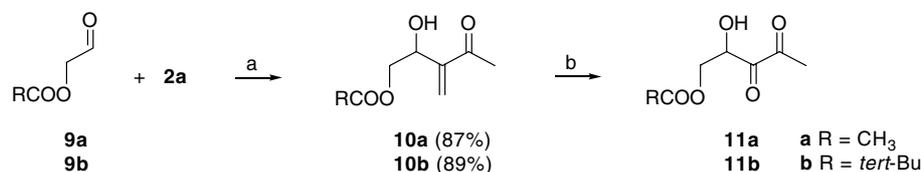
Keywords: Quorum sensing; AI-2; Baylis–Hillman reaction; Ozonolysis.

* Corresponding author. Tel.: +33 (0)4 72 43 82 21; fax: +33 (0)4 72 43 88 96; e-mail: alain.doutheau@insa-lyon.fr

new access to DPD and analogues. Despite its very simple structure, the synthesis of DPD is not an easy task owing, in particular, to its instability at high concentrations.⁸ At the time we initiated this work, only two syntheses of the (*S*) enantiomer had been reported in the literature, both involving the oxidation of a triple bond to generate the α -diketone function.^{9,10} We imagined an alternative approach in which the α -diketo- β -hydroxy moiety in DPD would be generated by the reductive ozonolysis of the carbon–carbon double bond in α -methylene- β -hydroxy ketones of type **4** resulting from a Baylis–Hillman reaction.

Thus, using dimethyl sulfide as the reducing agent, the byproducts formed during the last step will be formaldehyde and dimethylsulfoxide. The latter compounds being reputed non-toxic, at least at low concentrations, the purification of DPD or analogues would become unnecessary before biological assays. In addition, asymmetric Baylis–Hillman reaction being well documented,¹¹ the preparation of non-racemic compounds could be considered. The very recent report of Vanderleyden and co-workers¹² describing a synthesis of (*S*)-DPD by reductive ozonolysis of (*S*)-1,2-dihydroxy-4-methyl-4-penten-3-one (Scheme 2) prompts us to report our preliminary results.

When carried out in THF at 0 °C,¹³ the Baylis–Hillman reaction between 2-(*tert*-butyldimethylsilyloxy) ethanal **1a**¹⁴ and 3-buten-2-one **2a** (4 equiv) in the presence of DABCO (0.25 equiv) was completed in 21 h and



Scheme 5. Synthetic sequence to compounds **11**. Reagents and conditions: (a) THF, DABCO (0.25 equiv), 0 °C, 20 h. (b) O₃, MeOH, –78 °C, then DMS, –78 °C to rt.

Acknowledgements

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- A solution of **4a** (8 mg, 62 μmol) in CD₃OD (6 mL) was cooled to –78 °C and ozone was bubbled through the solution. The reaction mixture first turned light yellow and after about 30 min, became light green. At this time, the solution was purged with oxygen until the green colour disappeared and Me₂S (45 μL, 10 equiv) was added. The mixture was then allowed to warm at rt and stirred for 16 h. To 1 mL of the colourless reaction mixture, D₂O (1 mL) was added and the resulting solution was concentrated under vacuum to a residual volume of about 1 mL.
- ¹H NMR (500 MHz, D₂O). **5a** (20%) + **6** (40%) + **7** (40%). δ 4.35 (dd, *J* = 6.9, *J* = 5.7 Hz, H-4), 4.16 (m, 2H, H-5, H-8), 4.03 (dd, *J* = 6.0, *J* = 3.5 Hz, H-7), 3.95 (dd, *J* = 7.4, *J* = 3.5 Hz, H-1), 3.80 (m, 2H, H-3, H-9), 3.63 (dd, *J* = 11.9, *J* = 7.4 Hz, H-2), 3.56 (dd, *J* = 9.4, *J* = 5.7 Hz, H-6), 2.35 (s, CH₃), 1.42 (s, CH₃), 1.39 (s, CH₃). Assignments are based on published data and 2D-COSY experiments.
- ¹H NMR (300 MHz, D₂O). **8a** δ 8.04 (m, 1H), 7.92 (m, 1H), 7.80 (m, 2H), 5.30 (dd, *J* = 6.7, *J* = 4.5 Hz, 1H), 3.99 (dd, *J* = 12, *J* = 4.5 Hz 1H), 3.90 (dd, *J* = 12, *J* = 6.7 Hz, 1H), 2.77 (s, 3H) in agreement with published data.
- Minor signals were observed in the ¹H NMR spectra of **DPD** (three singlets in the 2.0–2.2 ppm area and a multiplet at 3.95 ppm) and of **8a** (multiplet at 7.49 and 7.70 ppm).
- The *anti* stereochemistry was temporarily assigned to the major isomer by analogy with the results reported in the literature for Baylis–Hillman reactions between α-alkoxyaldehydes and vinyl ketones or esters: Drewes, S. E.; Manickum, T.; Roos, G. H. P. *Synth. Commun.* **1988**, *18*, 1065–1070.
- In these cases, compounds **4c** are in equilibrium with small amounts (25% for **4c anti**, 5% for **4c syn**) of isomeric cyclic anomers.
- ¹H NMR (500 MHz, D₂O). **5b** δ 4.38 (dd, *J* = 7.3, *J* = 6.0 Hz, 1H), 4.16 (dd, *J* = 10.1, *J* = 6.0 Hz, 1H), 4.15 (dd, *J* = 9.5, *J* = 7.3 Hz, 1H), 4.02 (dd, *J* = 6.0, *J* = 3.2 Hz, 1H), 3.94 (dd, *J* = 7.3, *J* = 4.0 Hz, 1H), 3.79 (dd, *J* = 10.1, *J* = 3.2 Hz, 1H), 3.77 (dd, *J* = 12.0, *J* = 4.0 Hz, 1H), 3.62 (dd, *J* = 12.0, *J* = 7.3 Hz, 1H), 3.53 (dd, *J* = 9.5, *J* = 6.0 Hz, 1H) 2.77 (m, 2H), 1.75 (m, 4H), 1.03 (t, *J* = 7.1 Hz, 3H), 0.98 (t, *J* = 7.5 Hz, 3H), 0.97 (t, *J* = 7.5 Hz, 3H). (300 MHz, D₂O) **5c anti** δ 3.97 (m, 1H), 3.93 (d, *J* = 7.9 Hz, 1H), 3.76 (m, 1H), 3.56 (d, *J* = 6.0 Hz, 1H), 1.40 (s, 3H), 1.39 (s, 3H), 1.31 (d, *J* = 6.4 Hz, 3H), 1.29 (d, *J* = 6.4 Hz, 3H). **5c syn** δ 4.37 (m, 1H), 4.30 (m, 1H), 4.05 (d, *J* = 5.3 Hz, 1H), 3.86 (d, *J* = 4.5 Hz, 1H), 1.41 (s, 3H), 1.36 (s, 3H), 1.24 (d, *J* = 6.8 Hz, 3H), 1.15 (d, *J* = 6.8 Hz, 3H).
- ¹H NMR (300 MHz, D₂O). **8b** δ 8.06 (m, 1H), 7.98 (m, 1H), 7.83 (m, 2H), 5.33 (dd, *J* = 6.8, *J* = 4.9 Hz, 1H), 3.99 (dd, *J* = 11.7, *J* = 4.9 Hz, 1H), 3.93 (dd, *J* = 11.7, *J* = 6.8 Hz, 1H), 3.11 (m, 2H), 1.36 (t, *J* = 7.5 Hz, 3H). **8c anti** δ 8.10 (m, 1H), 7.98 (m, 1H), 7.83 (m, 2H), 5.06 (d, *J* = 6.8 Hz, 1H), 4.26 (m, 1H), 2.81 (s, 3H), 1.33 (d, *J* = 6.4 Hz, 3H). **8c syn** δ 8.05 (m, 1H), 7.95 (m, 1H), 7.83 (m, 2H), 5.04 (d, *J* = 5.7 Hz, 1H), 4.32 (m, 1H), 2.78 (s, 3H), 1.19 (d, *J* = 6.4 Hz, 3H).
- The aldehydes **1b** and **1c** were prepared by the reductive ozonolysis of known *tert*-butyldimethylsilyl protected 3-buten-2-ol and 2-methyl-3-buten-2-ol, respectively.
- The starting aldehyde **1c** was recovered unchanged after 30 h of stirring at 0 °C. This failure was probably due to the sterically hindered carbonyl function of the neopentyl type aldehyde **1c**.

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26. Compounds **4a–c** and **10a,b** gave ^1H , ^{13}C NMR and HRMS data in agreement with the proposed structures. Data for **4b** and **10a** are given as examples. **4b** ^1H NMR (300 MHz, CD_3OD) δ 6.29 (s, 1H), 6.14 (d, $J = 1.1$ Hz, 1H), 4.63 (m, 1H), 3.60 (dd, $J = 3.8, 11.3$ Hz, 1H), 3.36 (dd, $J = 6.8, 11.3$ Hz, 1H), 2.78 (m, 2H), 1.06 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (75 MHz, CD_3OD) δ 203.95, 149.60, 125.86, 71.31, 66.93, 32.16, 8.56; HRMS-CI MH^+ calcd for $\text{C}_7\text{H}_{13}\text{O}_3$: 145.0865; found: 145.0866. **10a** ^1H NMR (300 MHz, CDCl_3) δ 6.24 (s, 1H), 6.18 (d, $J = 1.1$ Hz, 1H), 4.75 (m, 1H), 4.24 (dd, $J = 3.4, 11.3$ Hz, 1H), 4.13 (dd, $J = 6.8, 11.3$ Hz, 1H), 3.00 (d, $J = 5.7$ Hz, 1H), 2.35 (s, 3H), 2.08 (s, 3H). ^{13}C NMR (75 MHz, CDCl_3) δ 199.64, 171.42, 146.60, 127.61, 69.33, 67.61, 26.21, 20.86. HRMS-CI MH^+ calcd for $\text{C}_8\text{H}_{13}\text{O}_4$: 173.0736; found: 173.0817.
27. **11a** ^1H NMR (300 MHz, D_2O) δ 4.32 (m, 1H), 4.13 (m, 2H), 2.38 (s, 3H), 2.09 (s, 3H); HRMS-CI MH^+ calcd for $\text{C}_7\text{H}_{11}\text{O}_5$: 175.0606; found: 175.0607. **11b** ^1H NMR (300 MHz, D_2O) δ 4.33 (m, 1H), 4.12 (m, 2H), 2.37 (s, 3H), 1.19 (s, 9H).