

Available online at www.sciencedirect.com



Tetrahedron Letters 46 (2005) 6495-6498

Tetrahedron Letters

A Baylis–Hillman/ozonolysis route towards (±) 4,5-dihydroxy-2,3-pentanedione (DPD) and analogues

Marine Frezza, Laurent Soulère, Yves Queneau and Alain Doutheau*

Laboratoire de Chimie Organique, UMR CNRS-UCBL 5181, Institut National des Sciences Appliquées, 20 avenue A. Einstein, 69621 Villeurbanne, France

> Received 7 June 2005; revised 19 July 2005; accepted 20 July 2005 Available online 9 August 2005

Abstract—The Baylis–Hillman reaction between 2-(*tert*-butyldimethylsilyloxy)ethanal and 3-buten-2-one followed by desilylation gave rise to the corresponding α -methylene- β , γ -dihydroxy ketone further converted by reductive ozonolysis of the carbon–carbon double bond into racemic 4,5-dihydroxy-2,3-pentanedione (**DPD**), a significant molecule in bacterial cell–cell communication systems. The same sequence applied to other substrates allowed the preparation of chain elongated analogues and 5-*O*-acylated derivatives of **DPD**.

© 2005 Elsevier Ltd. All rights reserved.

Many bacterial species are able to sense their own population size through a cell–cell communication system, named quorum sensing (QS), based on the production and exchange of diffusible extracellular signaling molecules called autoinducers.¹ Since a link between this process and virulence factor production² or biofilms formation³ has been established for a number of pathogenic bacteria, the design of QS inhibitors has recently attracted considerable interest for the development of anti-infection therapies.⁴ Among the various autoinducers which have been identified so far, a borate, known as autoinducer-2 (AI-2), formed from the metabolic product (S) 4,5-dihydroxy-2,3-pentanedione (**DPD**) (Scheme 1), is produced by a large number of both Gram-negative and Gram-positive bacteria.⁵

As part of a programme aimed at obtaining QS inhibitors,^{6,7} we recently became interested in designing a



Scheme 1.

Keywords: Quorum sensing; AI-2; Baylis–Hillman reaction; Ozonolysis.

0040-4039/\$ - see front matter © 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2005.07.102

new access to **DPD** and analogues. Despite its very simple structure, the synthesis of **DPD** is not an easy task owing, in particular, to its instability at high concentrations.⁸ At the time we initiated this work, only two syntheses of the (S) enantiomer had been reported in the literature, both involving the oxidation of a triple bond to generate the α -diketone function.^{9,10} We imagined an alternative approach in which the α -diketo- β -hydroxy moiety in **DPD** would be generated by the reductive ozonolysis of the carbon–carbon double bond in α -methylene- β -hydroxy ketones of type **4** resulting from a Baylis–Hillman reaction.

Thus, using dimethyl sulfide as the reducing agent, the byproducts formed during the last step will be formaldehyde and dimethylsulfoxide. The latter compounds being reputed non-toxic, at least at low concentrations, the purification of **DPD** or analogues would become unnecessary before biological assays. In addition, asymmetric Baylis–Hillman reaction being well documented,¹¹ the preparation of non-racemic compounds could be considered. The very recent report of Vander-leyden and co-workers¹² describing a synthesis of (*S*)-**DPD** by reductive ozonolysis of (*S*)-1,2-dihydroxy-4-methyl-4-penten-3-one (Scheme 2) prompts us to report our preliminary results.

When carried out in THF at 0 °C,¹³ the Baylis–Hillman reaction between 2-(*tert*-butyldimethylsilyloxy) ethanal $1a^{14}$ and 3-buten-2-one 2a (4 equiv) in the presence of DABCO (0.25 equiv) was completed in 21 h and

^{*} Corresponding author. Tel.: +33 (0)4 72 43 82 21; fax: +33 (0)4 72 43 88 96; e-mail: alain.doutheau@insa-lyon.fr



Scheme 2.

afforded the expected enone **3a** in 74% yield (Scheme 3 and Table 1). Removal of the silvl protecting group led to the corresponding diol **4a** which was subjected to ozonolysis. When conducted on a diluted ($\approx 10 \text{ mM}$) CD₃OD solution,¹⁵ the reaction gave rise to **5a** (**DPD**).

As already reported,^{9,10,12} the ¹H NMR spectrum of **DPD** is rather complicated due to the presence of three isomers in equilibrium: the open diketone, and its two cyclic anomers **6** and **7**, in about a 0.5:1:1 ratio.¹⁶ In order to get structure verification, this mixture was submitted to the action of 1,2-phenylene-diamine (3 equiv)^{9,10,12} and we observed that it was totally converted into the quinoxaline **8a** (Scheme 4).^{17,18}

We then extended this approach to the preparation of some chain elongated analogues of **DPD** by varying the nature of both the vinyl ketone and the silyloxyaldehyde. A similar sequence was carried out from aldehyde **1a** and 4-penten-3-one **2b** (Scheme 3 and Table 1) to give diketone **5b**. The Baylis–Hillman reaction of α -methyl substituted aldehyde **1b** with **2a** gave **3c** as a mixture ($\approx 65:35$) of *anti/syn* diastereoisomers,¹⁹ which were incompletely separated by column chromatography. The deprotection step was carried out on each pure stereoisomer to give the corresponding *anti* or *syn* diol **4c**,²⁰ which were submitted to ozonolysis and led to expected dicarbonyl compounds **5c**.²¹ As seen above in the case of **5a**, when **5b** and **5c** were reacted with excess 1,2-phenylene-diamine, they were quantitatively transformed into the corresponding quinoxaline **8b** and **8c**.²² Conversely, when carried out from α -dimethyl substituted aldehydes **1c**²³ and **2a**, the Baylis–Hillman reaction did not take place.²⁴

The method was also successfully applied for the preparation of derivatives bearing 5-OH enzymolabile masking groups, which could be used as in vivo precursors of **DPD**. Thus, the Baylis–Hillman reaction between acetoxyacetaldehyde $9a^{13}$ or pivaloxyacetaldehyde $9b^{25}$ and **2a** led to the corresponding adducts **10a** and **10b** in very good yields.²⁶ Further, ozonolysis gave the diketonic compounds **11a** and **11b** (Scheme 5).²⁷

In conclusion, we developed a short reaction sequence leading to racemic **DPD**. The sequence was successfully applied for the preparation of elongated chain analogues or 5-*O*-acylated derivatives. We are presently exploring the possibilities of our method to obtain more elaborated analogues of **DPD** as well as non-racemic compounds.



Scheme 3. Synthetic sequence to compounds 5. Reagents and conditions: (a) THF, DABCO (0.25 equiv), 0 °C. (b) TBAF (1 equiv), THF, rt. (c) O_3 , MeOH, -78 °C, then DMS, -78 °C to rt.

Table 1.						
Aldehyde	Vinyl-ketone	Baylis–Hillman adduct	Reaction time (h)	Yield (%)	Deprotected product	Yield (%)
1a	2a	3a	21	74	4a	78
1a	2b	3b	30	80	4b	90
1b	2a	3c	24	70	4c anti	71
		(anti/syn 65:35)			4c syn	78



Scheme 4. Structures of DPD, its two cyclic anomers 6 and 7, and quinoxalines 8.



Scheme 5. Synthetic sequence to compounds 11. Reagents and conditions: (a) THF, DABCO (0.25 equiv), 0 °C, 20 h. (b) O₃, MeOH, -78 °C, then DMS, -78 °C to rt.

Acknowledgements

Financial support from MENESR and CNRS is gratefully acknowledged. M.F. thanks the MENESR for a scholarship.

References and notes

- 1. For a recent review, see: Lyon, G. J.; Muir, T. W. Chem. Biol. 2003, 10, 1007–1021.
- Finch, R. G.; Pritchard, D. I.; Bycroft, B. W.; Williams, P.; Stewart, G. S. J. Antimicrob. Chemother. 1998, 42, 569– 571.
- Rice, S. A.; McDougald, D.; Kumar, N.; Kjelleberg, S. Curr. Opin. Investig. Drugs 2005, 6, 178–184.
- Raffa, R. B.; Iannnuzzo, J. R.; Levine, D. R.; Saeid, K. K.; Schwartz, R. S. C.; Sucic, N. T.; Terleckyj, O. D.; Young, J. M. J. Pharmacol. Exp. Ther. 2005, 312, 417– 423.
- Chen, X.; Schauder, S.; Potier, N.; Van Dorsselaer, A.; Pelczer, I.; Bassler, B. L.; Hughson, F. M. *Nature* 2002, *415*, 545–549.
- Reverchon, S.; Chantegrel, B.; Deshayes, C.; Doutheau, A.; Cotte-Pattat, N. *Bioorg. Med. Chem. Lett.* 2002, 12, 1153–1157.
- Castang, S.; Chantegrel, B.; Deshayes, C.; Dolmazon, R.; Gouet, P.; Haser, R.; Reverchon, S.; Nasser, W.; Hugouvieux-Cotte-Pattat, N.; Doutheau, A. *Bioorg. Med. Chem. Lett.* 2004, *14*, 5145–5149.
- Hoffman, T.; Schieberle, P. J. Agric. Food Chem. 1998, 46, 235–241.
- Meijler, M. M.; Hom, L. G.; Kaufmann, G. F.; McKenzye, K. M.; Sun, C.; Moss, J. A.; Matsushita, M.; Janda, K. D. Angew. Chem., Int. Ed. 2004, 43, 2106–2108.
- Semmelhack, M. F.; Campagna, S. R.; Federle, M. J.; Bassler, B. L. Org. Lett. 2005, 7, 569–572.
- 11. Matsui, K.; Takizawa, S.; Sasai, H. *Tetrahedron Lett.* **2005**, *46*, 1943–1946, and references cited therein.
- De Keersmaecker, S. C. J.; Varszegi, C.; van Boxel, N.; Habel, L. W.; Metzger, K.; Daniels, R.; Marchal, K.; De Vos, D.; Vanderleyden, J. J. Biol. Chem. 2005, 280, 19563– 19568.
- Brzezinski, L. J.; Rafel, S.; Leahy, J. W. *Tetrahedron* 1997, 53, 16423–16434.
- Nicolaou, K. C.; Liu, J.-J.; Yang, Z.; Ueno, H.; Sorensen, E. J.; Claiborne, C. F.; Guy, R. K.; Hwang, C.-K.; Nakada, M.; Nantermet, P. G. J. Am. Chem. Soc. 1995, 117, 634–644.
- 15. A solution of 4a (8 mg, 62 μ mol) in CD₃OD (6 mL) was cooled to -78 °C and ozone was bubbled through the solution. The reaction mixture first turned light yellow and after about 30 min, became light green. At this time, the solution was purged with oxygen until the green colour disappeared and Me₂S (45 μ L, 10 equiv) was added. The mixture was then allowed to warm at rt and stirred for 16 h. To 1 mL of the colourless reaction mixture, D₂O

(1 mL) was added and the resulting solution was concentrated under vacuum to a residual volume of about 1 mL.

- 16. ¹H NMR (500 MHz, D₂O). **5a** (20%) + **6** (40%) + **7** (40%). δ 4.35 (dd, J = 6.9, J = 5.7 Hz, H-4), 4.16 (m, 2H, H-5, H-8), 4.03 (dd, J = 6.0, J = 3.5 Hz, H-7), 3.95 (dd, J = 7.4, J = 3.5 Hz, H-1), 3.80 (m, 2H, H-3, H-9), 3.63 (dd, J = 11.9, J = 7.4 Hz, H-2) 3.56 (dd, J = 9.4, J = 5.7 Hz, H-6), 2.35 (s, CH₃), 1.42 (s, CH₃), 1.39 (s, CH₃). Assignments are based on published data and 2D-COSY experiments.
- 17. ¹H NMR (300 MHz, D₂O). **8a** δ 8.04 (m, 1H), 7.92 (m, 1H), 7.80 (m, 2H), 5.30 (dd, J = 6.7, J = 4.5 Hz, 1H), 3.99 (dd, J = 12, J = 4.5 Hz 1H), 3.90 (dd, J = 12, J = 6.7 Hz, 1H), 2.77 (s, 3H) in agreement with published data.
- Minor signals were observed in the ¹H NMR spectra of DPD (three singulets in the 2.0–2.2 ppm area and a multiplet at 3.95 ppm) and of 8a (multiplet at 7.49 and 7.70 ppm).
- The *anti* stereochemistry was temporarily assigned to the major isomer by analogy with the results reported in the literature for Baylis–Hillman reactions between α-alkoxyaldehydes and vinyl ketones or esters: Drewes, S. E.; Manickum, T.; Roos, G. H. P. *Synth. Commun.* 1988, 18, 1065–1070.
- 20. In these cases, compounds **4c** are in equilibrium with small amounts (25% for **4c** *anti*, 5% for **4c** *syn*) of isomeric cyclic anomers.
- 21. ¹H NMR (500 MHz, D₂O). **5b** δ 4.38 (dd, J = 7.3, J = 6.0 Hz, 1H), 4.16 (dd, J = 10.1, J = 6.0 Hz, 1H), 4.15 (dd, J = 9.5, J = 7.3 Hz, 1H), 4.02 (dd, J = 6.0, J = 3.2 Hz, 1H), 3.94 (dd, J = 7.3, J = 4.0 Hz, 1H), 3.79 (dd, J = 10.1, J = 3.2 Hz, 1H), 3.77 (dd, J = 12.0, J = 4.0 Hz, 1H), 3.62 (dd, J = 12.0, J = 7.3 Hz, 1H), 3.53 (dd, J = 9.5, J = 6.0 Hz, 1H) 2.77 (m, 2H), 1.75 (m, 2H)4H), 1.03 (t, J = 7.1 Hz, 3H), 0.98 (t, J = 7.5 Hz, 3H), 0.97 (t, J = 7.5 Hz, 3H). (300 MHz, D₂O) **5c** anti δ 3.97 (m, 1H), 3.93 (d, J = 7.9 Hz, 1H), 3.76 (m, 1H), 3.56 (d, J = 6.0 Hz, 1H), 1.40 (s, 3H), 1.39 (s, 3H), 1.31 (d, J = 6.4 Hz, 3H), 1.29 (d, J = 6.4 Hz, 3H). 5c syn δ 4.37 (m, 1H), 4.30 (m, 1H), 4.05 (d, J = 5.3 Hz, 1H), 3.86 (d, J = 4.5 Hz, 1H), 1.41 (s, 3H), 1.36 (s, 3H), 1.24 (d, J = 6.8 Hz, 3H), 1.15 (d, J = 6.8 Hz, 3H).
- 22. ¹H NMR (300 MHz, D₂O). **8b** δ 8.06 (m, 1H), 7.98 (m, 1H), 7.83 (m, 2H), 5.33 (dd, J = 6.8, J = 4.9 Hz, 1H), 3.99 (dd, J = 11.7, J = 4.9 Hz, 1H), 3.93 (dd, J = 11.7, J = 6.8 Hz, 1H), 3.11 (m, 2H), 1.36 (t, J = 7.5 Hz, 3H). **8c** anti δ 8.10 (m, 1H), 7.98 (m, 1H), 7.83 (m, 2H), 5.06 (d, J = 6.4 Hz, 3H). **8c** syn δ 8.05 (m, 1H), 7.95 (m, 1H), 7.83 (m, 2H), 5.04 (d, J = 5.7 Hz, 1H), 4.32 (m, 1H), 2.78 (s, 3H), 1.19 (d, J = 6.4 Hz, 3H).
- 23. The aldehydes **1b** and **1c** were prepared by the reductive ozonolysis of known *tert*-butyldimethylsilyl protected 3-buten-2-ol and 2-methyl-3-buten-2-ol, respectively.
- 24. The starting aldehyde **1c** was recovered unchanged after 30 h of stirring at 0 °C. This failure was probably due to the sterically hindered carbonyl function of the neopen-tylic type aldehyde **1c**.

- Audouard, C.; Fawcett, J.; Griffiths, G. A.; Percy, J. M.; Pintat, S.; Smith, C. A. Org. Biomol. Chem. 2004, 528– 541.
- 26. Compounds **4a–c** and **10a,b** gave ¹H, ¹³C NMR and HRMS data in agreement with the proposed structures. Data for **4b** and **10a** are given as examples. **4b** ¹H NMR (300 MHz, CD₃OD) δ 6.29 (s, 1H), 6.14 (d, J = 1.1 Hz, 1H), 4.63 (m, 1H), 3.60 (dd, J = 3.8, 11.3 Hz, 1H), 3.36 (dd, J = 6.8, 11.3 Hz, 1H), 2.78 (m, 2H), 1.06 (t, J = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CD₃OD) δ 203.95, 149.60, 125.86, 71.31, 66.93, 32.16, 8.56; HRMS-CI MH⁺ calcd for C₇H₁₃O₃: 145.0865; found: 145.08666. **10a** H

NMR (300 MHz, CDCl₃) δ 6.24 (s, 1H), 6.18 (d, J = 1.1 Hz, 1H), 4.75 (m, 1H), 4.24 (dd, J = 3.4, 11.3 Hz, 1H), 4.13 (dd, J = 6.8, 11.3 Hz, 1H), 3.00 (d, J = 5.7 Hz, 1H), 2.35 (s, 3H), 2.08 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 199.64, 171.42, 146.60, 127.61, 69.33, 67.61, 26.21, 20.86. HRMS-CI MH⁺ calcd for C₈H₁₃O₄: 173.0736; found: 173.0817.

27. **11a** ¹H NMR (300 MHz, D_2O) δ 4.32 (m, 1H), 4.13 (m, 2H), 2.38 (s, 3H), 2.09 (s, 3H); HRMS-CI MH⁺ calcd for C₇H₁₁O₅: 175.0606; found: 175.0607. **11b** ¹H NMR (300 MHz, D_2O) δ 4.33 (m, 1H), 4.12 (m, 2H), 2.37 (s, 3H), 1.19 (s, 9H).