Full Paper

Subscriber access provided by LUNDS UNIV

A Scalable Total Synthesis of Halofuginone

Hua Xu, Wenhao Yin, Haoqiang Liang, Yanbo Nan, Fayang Qiu, and Yehua Jin

Org. Process Res. Dev., Just Accepted Manuscript • DOI: 10.1021/acs.oprd.9b00059 • Publication Date (Web): 06 Mar 2019

Downloaded from http://pubs.acs.org on March 6, 2019

Just Accepted

"Just Accepted" manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides "Just Accepted" as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. "Just Accepted" manuscripts appear in full in PDF format accompanied by an HTML abstract. "Just Accepted" manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). "Just Accepted" is an optional service offered to authors. Therefore, the "Just Accepted" Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the "Just Accepted" Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these "Just Accepted" manuscripts.



is published by the American Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

Published by American Chemical Society. Copyright © American Chemical Society. However, no copyright claim is made to original U.S. Government works, or works produced by employees of any Commonwealth realm Crown government in the course of their duties.

A Scalable Total Synthesis of Halofuginone

Hua Xu,[‡] Wenhao Yin,[‡] Haoqiang Liang, Yanbo Nan, Fayang, G. Qiu,* and Yehua Jin*

Launch-Pharma Technologies, Ltd., 188 Kaiyuan Boulevard, Building D, 5th Floor, the Science

Park of Guangzhou, Guangzhou, 510530, China.



Table of Contents Graphic:



ABSTRACT: A scalable total synthesis of halofuginone has been accomplished. This synthetic route features a total of 12 steps of highly efficient reactions, without any chromatographic purification. Halofuginone was obtained in 17% overall yield and over 98.5% HPLC purity. All the reaction conditions are mild and reliable. In addition, no hazardous materials were used or produced. All reagents are commercially available and inexpensive. This route is safe, robust, scalable, cost-effective and environmentally benign.

KEYWORDS: Halofuginone, Total Synthesis, Dieckmann Condensation, Isomerization,

Quinazolinone

Halofuginone **1**, a synthetic analogue of the quinazolinone alkaloid febrifugine **2** originally isolated from the Chinese herb *Dichroa febrifuga*,¹ displayed extremely rich biological activities. Halofuginone hydrobromide (racemic) has been used as an anticoccidial feed additive for broilers and turkeys for several decades under the trade name Stenorol®,² while its lactate (racemic) named Halocur® has been used for the prevention of diarrhoea caused by diagnosed *Cryptosporidium parvum* in new born calves.³ In addition to veterinary applications, halofuginone has also demonstrated great potential in the development of human medicine. For examples, it reportedly inhibits the development of proinflammatory Th17 cells, procollagen type I gene expression and extracellular matrix deposition, thus may be effective for the treatment of autoimmune disorders, fibrosis or cancer.⁴ As a result, the total synthesis of halofuginone has attracted the attention of many chemists.⁵ A detailed summary about the synthetic work of febrifugine and its analogues (including halofuginone) has been reported by Evans *et al.*^{5a, 5b} From a practical point of view, we have developed a scalable synthetic route which may provide kilograms of high purity halofuginone.



 $R_1 = R_2 = H$, febrifugine 2



 $R_1 = Br, R_2 = Cl$, isohalofuginone 3 $R_1 = R_2 = H$, isofebrifugine 4

Figure 1. Selected examples of halofuginone family compounds.

Our retrosynthetic analysis is shown in **Scheme 1**. Halofuginone (\pm)-1 may be obtained through the isomerization of isohalofuginone 3 inspired by a similar transformation from isofebrifugine 4 to febrifugine 2 reported by Takeuchi *et al.*^{5f} and applied to the synthesis of halofuginone by Li *et*

al.^{5h} Compound **3** may be assembled from bromohemiacetal **14** and commercially available quinazolinone **15** under basic conditions. Bromination of compound **13**, which may be derived from the β -keto-ester **9** through a series of transformations, may lead to compound **14** under aqueous conditions, while compound **9** may be obtained from compound **8**, an intermediate that may be assembled with commercially available materials (including diethyl acetamidomalonate, 2,3-dichloropropene and ethyl 4-bromobutyrate) through a series of conventional reactions, by using the Dieckmann condensation. **Scheme 1**. Retrosynthetic Analysis of Halofuginone.



Based on the above analysis, the total synthesis of halofuginone is shown in **Scheme 2**. Compound **5** was prepared from diethyl acetamidomalonate via alkylation with 2,3dichloropropene. Decarboxylation of one ester group of compound **5** in refluxing aqueous HCl solution afforded α -amino acid hydrochloride **6**. After esterification, compound **7** was obtained, the purity of which was determined to be 87% using ¹H NMR. With compound **7** in hand, we next carried out its N-substitution reaction with ethyl 4-bromobutyrate. While the reaction did not work well when an organic base (Et₃N, DIPEA or DBU) was used in an organic solvent (acetonitrile or toluene), a high conversion rate (92%) was observed when anhydrous Na₂CO₃ was used in the

presence of a catalytic amount of TBAI in toluene. After the completion of the N-substitution reaction, Cbz-Cl was added dropwise directly to the reaction mixture to afford compound **8**.⁶





Reagents and conditions: (a) 2,3-Dichloropropene, K₂CO₃, cat. KI, cat. TBAB, acetonitrile, 85-90 °C; (b) 6M HCl(aq), refluxing; (c) conc. H₂SO₄, DEC/EtOH, refluxing; DEC = diethyl carbonate; (d) Ethyl 4-bromobutyrate, Na₂CO₃, cat. TBAI, toluene, 75-85 °C, then Cbz-Cl, 25 °C; (e) t-BuONa, THF, 0±5 °C; (f) LiCl, DMF, H₂O, 120 °C; (g) NaBH₄, EtOH, 5-10 °C; (h) 6M HCl(aq)/EtOH, refluxing; (i) Na₂CO₃, dioxane, H₂O, Fmoc-Cl, 20 °C; (j) NBS, acetonitrile, H₂O, 0-5 °C; (k) quinazolinone **15**, LiOH, DMF, 0-5 °C, then diethylamine, 0-5 °C; (l) EtOH, refluxing.

It was surprising that the seemingly straightforward Dieckmann condensation turned out to be problematic since no condensation product was observed when compound **8** was treated with NaOMe in MeOH or with NaOEt in EtOH. However, treating compound **8** (5g scale) with either NaOMe or NaOEt in either THF or toluene afforded the desired product **9** in less than 50% yield.

Inspired by this result, we thought that a polar aprotic solvent may be a better choice for the Dieckmann condensation of compound **8**. After a brief screening, we found that *t*-BuONa in THF gave 89% yield (15g scale compound **8**). Thus, *t*-BuONa was used as the base in the following scale-up experiments, in which 5.44 kg of compound **9** was obtained in a single batch in 75% HPLC purity.⁷ The removal of the ethoxycarbonyl functionality of compound **9** was achieved by Krapcho decarboxylation to afford 4.19 kg of compound **10** in 73% HPLC purity. Reduction of the carbonyl group with NaBH₄ with the hydride attacking the carbonyl group from the sterically less congested side led to the *cis* isomer, and 3.50 kg of compound **11** was obtained in 81% HPLC purity.^{5c, 5d} The Cbz group was removed by using aqueous HCl and compound **12** was obtained in over 98% ¹H NMR purity after one crystallization from acetonitrile.





It was necessary for us to switch the protecting group from Cbz to Fmoc, the rationale for which is illustrated in Scheme **3**. Compound **17** was initially prepared via a two-step transformation from

11, but the removal of the Cbz group of 17 was problematic. Hydrogenolysis of the Cbz of 17 was accompanied by debromination leading to 18, which was isomerized to product 19 in refluxing EtOH. Treating 17 with a Bronsted acid afforded compound 20 and 21 as the two major products, while treating 17 with Lewis acids afforded compound 22 as the major product. The results obtained from the reaction of Cbz-protected isohalofuginone 17 with acid were in agreement with those previously reported.^{5k, 8} Based on the above observations, it was realized that the protecting group of the piperidine nitrogen played an extremely important role in the last few steps of the entire synthetic route. An ideal protecting group here should not only tolerate the reaction conditions of the next two steps, i.e., NBS bromination and base-promoted substitution with quinazolinone 15, but also be easily removable without affecting the piperidine ring. Fortunately, this purpose was served when Fmoc was used to protect the piperidine nitrogen. Protection of the N atom of **12** with Fmoc-Cl afforded compound **13**, which was treated with NBS in the presence of water to provide 14.^{5h} Slow addition of the DMF solution of 14 to the preformed quinazolinone 15 lithium salt in DMF produced the coupling product, which upon deprotection of the Fmoc group with diethylamine afforded isohalofuginone **3** containing a small amount of halofuginone 1 (< 5%). It was found that **3** was isomerized to **1** during the solvent evaporation process because the ratio of these two compounds gradually changed (e.g., isohalofuginone 3/halofuginone 1 = 2/1). The total weight of the resulting mixture was 1.84 kg (77% over 3 steps), of which 98% is isohalofuginone 3 and halofuginone 1. Isohalofuginone 3 was further isomerized to halofuginone 1 in refluxing EtOH in 94% conversion. After crystallization, 1.65 kg of halofuginone 1 (> 98.5%)HPLC purity) was obtained in 90% yield. Starting from 5 kg diethyl acetamidomalonate, 1.65 kg halofuginone was obtained in high purity (>98.5% HPLC purity), which corresponds to an overall yield of 17% over 12 steps.

This synthetic route may also enable the large scale production of enantiomerically pure halofuginone **1**. As shown in **Scheme 4**, resolution of racemic-**12** using dibenzoyl-L-tartaric acid provided (+)-**12** in 32% yield, and its ee value was determined to be 96.2% in the form of its Fmoc derivative (**13**).

Scheme 4. Synthesis of (+)-halofuginone 1.



In conclusion, we have accomplished a kilogram-scale total synthesis of halofuginone in high purity over 12 steps. No column chromatographic purification was involved in the entire process. This synthetic route may also be used for the preparation of enantiomerically pure halofuginone. The reaction conditions are mild, safe and environmentally benign. The production operations are quite simple and convenient. All the materials in this route are commercially available and inexpensive. The success of this approach is dependent on a series of key chemical transformations, including (1) Dieckmann condensation to efficiently set up the piperidine ring; (2) Fmoc was chosen as a key protecting group for the piperidine nitrogen-atom at the late stage; (3) high conversion rate of isomerization of isohalofuginone in ethanol to halofuginone; and (4) resolution of racemic-**12** by using dibenzoyl-L-tartaric acid to provide (+)-**12** in 96.2% ee value.

General Procedures:

¹H NMR spectra were recorded on 400 MHz or/and 500 MHz (126 MHz for ¹³C NMR) Bruker FT-NMR spectrometers and calibrated using residual undeuterated solvent as an internal reference (CHCl₃ @ δ 7.26 ppm ¹H NMR, δ 77.16 ppm ¹³C NMR; DMSO @ δ 2.50 ppm ¹H NMR, δ 39.52 ppm ¹³C NMR). The following abbreviations (or combinations thereof) were used to explain ¹H NMR multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, p = quintet, m = multiplet, br = broad. High Resolution Mass (MS) analysis was obtained using on an Agilent 6210 LC/MSD TOF spectrometer system with Electrospray Ionization (ESI).

Experimental procedure:

Note: compound **5**, **6**, **7**, **8**, **9**, **10**, **11**, **13**, **14** were prepared as crude materials in the entire process. However, it was counted as 100% purity when calculating the stoichiometery.



Synthesis of **5**. A 50 L glass reactor was charged with diethyl acetamidomalonate (5.00 kg, 23.02 mol), anhydrous K_2CO_3 (6.35 kg, 46.04 mol), KI (0.76 kg, 4.58 mol), TBAB (0.37 kg, 1.15 mol), and acetonitrile (25 L). After being stirred for 20 min, 2,3-dichloropropene (3.07 kg, 27.62 mol) was added. The reaction mixture was stirred at 85–90 °C for about 10 h, at which time the starting material was consumed over 95% (HPLC). After being cooled to 25 °C, dilute $HCl_{(aq)}$ (1 M) was

slowly added to neutralize the reaction mixture to pH 7–7.5. The reaction mixture was allowed to stand at r.t. for a few minutes . The layers were separated. The organic layer was concentrated under reduced pressure at 50 °C to provide a crude slurry that was dissolved in EtOH–H₂O (1:10, 20 L). The mixture was stirred at 25 °C for 1 h and compound **5** crystallized out of the solution. After filtration through a Buchner funnel, the crystals were washed with water (2 × 5 L). The final product weighted 10.50 kg (wet weight), which was used directly in the next step. Data for compound **5**: ¹H NMR (500 MHz, Chloroform-*d*) δ 5.28 (d, *J* = 1.2 Hz, 1H), 5.17 (d, *J* = 1.1 Hz, 1H), 4.26 (qd, *J* = 7.1, 2.4 Hz, 4H), 3.47 (s, 2H), 2.03 (s, 3H), 1.55 – 1.51 (m, 1H), 1.27 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 169.3, 167.3, 136.5, 117.8, 65.2, 63.0, 41.6, 23.0, 14.0. HRMS (m/z): calc. for C₁₂H₁₉ClNO₅ [M+H]⁺ = 292.0952; found, 292.0954.

Synthesis of **6**. Compound **5** [10.50 kg (wet weight)] was charged to a 100 L glass reactor. $HCl_{(aq)}$ (6 M, 56.5 L) was added. The mixture was refluxed for about 8–12 h at 100 °C. After completion of the reaction, activated charcoal (650 g) was added. After being cooled to below 50 °C, the reaction mixture was filtered through a Buchner funnel. The filter cake was washed with water, and the combined filtrates were concentrated under reduced pressure at 80–85 °C to provide crude **6** (4.05 kg) as a yellowish solid, which was used directly in the next step.

Synthesis of 7. A 50 L glass reactor was charged with crude 6 (4.05 kg), diethyl carbonate (12 L), and anhydrous ethanol (4 L). To the stirred mixture, was added H₂SO₄ (1.13 kg, 98%, 11.51 mol) in 20–30 min. The reaction mixture was heated for about 20–25 h at 85–95 °C. After completion of the reaction, the mixture was concentrated under reduced pressure at 50–60 °C. The residual liquid was diluted with water. After being cooled to below 10 °C, NaOH_(aq) (30% wt, 4.88kg) was slowly added to neutralize the mixture to pH 8–9. After being warmed to 25 °C, the mixture was extracted with DCM (3 × 10 L).The combined organic layers were dried over

anhydrous Na₂SO₄, and filtered through a Buchner funnel. The filtrate was concentrated under reduced pressure at 40 °C to provide crude **7** as a brownish oil (3.07 kg, in 87% ¹H-NMR purity), which was used directly in the next step. Data for compound **7**: ¹H NMR (500 MHz, Chloroform-*d*) δ 5.30 (d, *J* = 1.3 Hz, 1H), 5.26 (q, *J* = 1.1 Hz, 1H), 4.20 (q, *J* = 7.1 Hz, 2H), 3.78 (dd, *J* = 8.6, 4.8 Hz, 1H), 2.81 (ddd, *J* = 14.2, 4.8, 1.1 Hz, 1H), 2.56 (dd, *J* = 14.2, 8.6 Hz, 1H), 1.28 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 174.5, 138.5, 115.9, 61.4, 52.3, 44.6, 14.3. HRMS (m/z): calc. for C₇H₁₃ClNO₂ [M+H]⁺ = 178.0635; found, 178.0628.



Synthesis of 8. A 50 L glass reactor was charged with the crude compound 7 (3.07 kg, 87% by ¹H-NMR), anhydrous Na₂CO₃ (5.49 kg, 51.79 mol), TBAI (0.64 kg, 1.73 mol), and toluene (9 L). After being stirred for 20 min, ethyl 4-bromobutyrate (3.37 kg, 17.26 mol) dissolved in toluene (6 L) was added. The reaction mixture was stirred at 75–80 °C for about 72 h, at which time compound 7 was consumed over 90% (monitored by HPLC). After being cooled to 20–25 °C, water (9 L) was added. After being stirred for 10–15 min, benzyl chloroformate (2.94 kg, 17.26 mol) was added dropwise in 2–3 h. The reaction mixture was stirred at 25 °C for about 2 h before addition of water (10 L) and toluene (10 L). The layers were separated. The organic layer was washed with NaOH_(aq) (5% wt, 15 L), water (20 L), HCl_(aq) (5% wt, 15 L), and water (20 L) respectively. After the organic layer was stirred at 25 °C for 1 h with activated charcoal (250 g), it was filtered through a Buchner funnel. The filtrate was concentrated under reduced pressure at 60 °C to give crude 8 (8.10 kg) as a brownish oil, which was used directly in the next step. Data for compound 8: ¹H NMR (500 MHz, Chloroform-*d*) δ 7.39 – 7.27 (m, 5H), 5.19 (d, *J* = 1.3 Hz,

1H), 5.17 - 5.07 (m, 2.6H), 5.02 (s, 0.4H), 4.27 - 3.89 (m, 5H), 3.68 - 3.56 (m, 1H), 3.23 - 3.10 (m, 1.6H), 3.02 - 2.90 (m, 1.4H), 2.47 - 2.26 (m, 2H), 1.99 - 1.84 (m, 2H), 1.27 - 1.17 (m, 4.8H), 1.13 (t, J = 7.2 Hz, 1.2H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 173.3, 173.2, 170.3, 170.2, 155.6, 138.8, 138.4, 136.7, 136.2, 128.7, 128.6, 128.4, 128.2, 127.9, 116.3, 116.0, 67.6, 67.4, 61.7, 60.5, 59.6, 58.7, 49.2, 48.9, 40.5, 39.4, 31.6, 31.4, 24.2, 23.7, 14.4, 14.14, 14.07. HRMS (m/z): calc. for $C_{21}H_{29}CINO_6$ [M+H]⁺ = 426.1683; found, 426.1678.



Synthesis of **9**. A 100 L glass reactor was charged with *t*-BuONa (3.31 kg, 34.53 mol) and anhydrous THF (38 L). After being cooled to -5 °C, crude compound **8** (8.10 kg) dissolved in THF (15 L) was added dropwise in 4–5 h while keeping the temperature below 0°C. The reaction mixture was stirred at 0–5 °C for about 3 h. After completion of the reaction, HCl_(aq) (1 M) was slowly added to neutralize the reaction mixture to pH 5–6. After addition of EA (10 L), the reaction mixture was stirred at 25 °C for a few minutes. The layers were separated and the organic layer was washed with saturated NaCl_(aq) (2 × 20 L), while the aqueous layer was extracted with EA (10 L). After addition of activated charcoal (500 g), the combined organic layers were stirred at 25 °C for 1 h, and then filtered through a Buchner funnel. The filtrate was concentrated under reduced pressure to give crude **9** (5.44 kg) as a light brown oil, which was used directly in the next step. Data for compound **9**: ¹H NMR (500 MHz, Chloroform-*d*) δ 12.23 (s, 1H), 7.41 – 7.28 (m, 5H), 5.27 – 5.05 (m, 4H), 5.01 (s, 0.4H), 4.94 – 4.82 (m, 0.6H), 4.30 (dd, *J* = 13.6, 5.7 Hz, 0.6H), 4.23 (q, *J* = 7.1 Hz, 2H), 4.19 – 4.12 (m, 0.4H), 3.09 – 2.66 (m, 3H), 2.46 – 2.23 (m, 2H), 1.30 (t, *J* =

7.1 Hz, 3H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 172.0, 171.9, 168.8, 168.2, 155.2, 155.1, 138.4, 138.3, 136.7, 136.3, 128.5, 128.2, 128.0, 115.8, 115.6, 97.7, 97.4, 67.7, 67.5, 61.0, 52.7, 52.5, 41.5, 40.7, 38.2, 37.3, 22.8, 22.3, 14.3. HRMS (m/z): calc. for C₁₉H₂₃ClNO₅ [M+H]⁺ = 380.1265; found, 380.1266.



Synthesis of **10**. A 50 L glass reactor was charged with crude **9** (5.44 kg), LiCl (0.61 kg, 14.33 mol), H₂O (2.7 L), and DMF (16.3 L). The reaction mixture was stirred at 120 °C for about 14 h. After completion of the reaction, the mixture was cooled to 25 °C. After addition of H₂O (25 L) and MTBE (30 L), the layers were separated. The organic layer was washed with water (2 × 25 L). The combined aqueous layers were extracted with MTBE (25 L). Then the combined organic layers were concentrated under reduced pressure at 45 °C to give crude **10** (4.19 kg) as a brownish oil, which was used directly in the next step. Data for compound **10**: ¹H NMR (500 MHz, Chloroform-*d*) δ 7.40 – 7.29 (m, 5H), 5.32 – 5.03 (m, 4H), 4.86 (s, 1H), 4.34 – 3.98 (m, 1H), 3.23 (s, 1H), 2.94 – 2.59 (m, 2H), 2.59 – 2.39 (m, 2H), 2.05 (s, 1H), 2.01 – 1.92 (m, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 206.7, 155.5, 128.6, 128.3, 116.1, 67.8, 61.8, 37.1, 22.5. HRMS (m/z): calc. for C₁₆H₁₉ClNO₃ [M+H]⁺ = 308.1053 ; found, 308.1057.



Synthesis of 11. A 50 L glass reactor was charged with EtOH (18 L). After being cooled to 5– 10 °C, NaBH₄ (0.51 kg, 13.61 mol) was added and crude 10 (4.19 kg) dissolved in EtOH (9 L) was added dropwise while keeping the temperature below 10 °C. The reaction mixture was stirred at 5-10 °C for about 1 h. After completion of the reaction, H₂O (20 L) was slowly added followed by MTBE (25 L). After standing at r.t. for a few minutes, the layers were separated. The organic layer was washed with NaOH_(aq) (10% wt, 10L), HCl_(aq) (5% wt, 2×10 L), and water (10 L), respectively. The combined aqueous layers were extracted with MTBE (25 L). After addition of activated charcoal (500 g), the combined organic layers were stirred at 25 °C for 1 h before filtration through a Buchner funnel. The filtrate was concentrated under reduced pressure at 45 °C to give crude 11 (3.50 kg) as a brown oil, which was used directly in the next step. Data for compound **11**: ¹H NMR (500 MHz, Chloroform-*d*) δ 7.42 – 7.27 (m, 5H), 5.21 – 5.06 (m, 4H), 4.76 (s, 1H), 4.05 (d, J = 14.0 Hz, 1H), 3.92 - 3.79 (m, 1H), 2.84 - 2.60 (m, 3H), 1.87 - 1.76 (m, 1H), 1.76 – 1.65 (m, 1H), 1.58 – 1.42 (m, 2H). ¹³C NMR (126 MHz, Chloroform-d) δ 155.8, 139.8, 136.7, 128.5, 128.2, 128.1, 114.5, 68.7, 67.5, 54.2, 37.9, 33.5, 27.9, 24.3. HRMS (m/z): calc. for $C_{16}H_{21}CINO_3 [M+H]^+ = 310.1210$; found, 310.1208. , OH (h)



Synthesis of **12**. A 50 L glass reactor was charged with crude **11** (3.50 kg), $HCl_{(aq)}$ (6 M, 18.83 L) and EtOH (20 L). The reaction mixture was refluxed for about 30 h. After completion of the reaction, the mixture was concentrated under reduced pressure at 50–55 °C to remove EtOH. To the residual liquid was added with MTBE (2 × 20 L), and resulting mixture was stirred for a few

minutes. The layers were separated. The aqueous layer was neutralized with NaOH_(aq) (40% wt) to pH >11, and then extracted with EA (2 × 20 L). The combined organic layer was dried over anhydrous MgSO₄ before filtration through a Buchner funnel. The filtrate was concentrated under reduced pressure to give crude **12** that was dissolved in acetonitrile (5 L) at 70 °C. After standing at 25 °C, compound **12** crystallized out of the solution. After filtration through a Buchner funnel, the collected crystals were dried in vacuum. The final product weighted 1.01 kg (> 98% ¹H NMR purity, 25% yield for 8 steps from diethyl acetamidomalonate). Data for compound **12**: ¹H NMR (500 MHz, Chloroform-*d*) δ 5.25 (d, *J* = 1.1 Hz, 1H), 5.23 (t, *J* = 1.0 Hz, 1H), 3.65 (s, 1H), 3.03 (ddt, *J* = 11.5, 4.3, 2.0 Hz, 1H), 2.88 (ddd, *J* = 7.5, 6.3, 1.4 Hz, 1H), 2.65 (td, *J* = 11.9, 2.9 Hz, 1H), 2.49 (d, *J* = 6.8 Hz, 2H)., 1.91 (dtt, *J* = 13.4, 4.0, 2.0 Hz, 1H), 1.73 (qt, *J* = 13.1, 4.3 Hz, 1H), 1.54 (tdd, *J* = 13.3, 4.7, 2.5 Hz, 1H), 1.45 (ddq, *J* = 12.9, 4.9, 2.6 Hz, 1H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 139.8, 115.2, 67.1, 57.7, 47.2, 43.0, 32.2, 20.4. HRMS (m/z): calc. for C₈H₁₅CINO [M+H]⁺ = 176.0842; found, 176.0837.



Synthesis of 13. A 50 L glass reactor was charged with compound 12 (1.00 kg, 5.69 mol), anhydrous Na₂CO₃ (0.91 kg, 8.54 mol), H₂O (5 L), and dioxane (5 L). After being cooled to 5–10 °C, Fmoc-Cl (1.47 kg, 5.69 mol) dissolved in dioxane (2 L) was added dropwise while keeping the temperature below 20 °C. The reaction mixture was stirred at 25 °C for about 1 h. After completion of the reaction, the reaction mixture was added with EA (20 L) and water (20 L). The layers were separated. The organic layer was washed with saturated NaCl_(aq) (2 × 5 L), while the aqueous layer was extracted with EA (10 L). To the combined organic layers were added activated

 charcoal (500 g) and then stirred at 25 °C for 1 h. After filtration through a Buchner funnel, the organic layer was concentrated under reduced pressure to give crude **13** (2.81 kg) as a yellow slurry, which was used directly in the next step. Data for compound **13**: ¹H NMR (500 MHz, Chloroform-*d*) δ 7.81 – 7.71 (m, 2H), 7.65 – 7.53 (m, 2H), 7.40 (td, *J* = 7.5, 2.4 Hz, 2H), 7.31 (t, *J* = 7.4 Hz, 2H), 5.17 (s, 1H), 5.14 (s, 1H), 4.93 – 4.51 (m, 1H), 4.49 (dd, *J* = 10.7, 6.7 Hz, 1H), 4.41 (dd, *J* = 10.7, 6.5 Hz, 1H), 4.25 (t, *J* = 6.5 Hz, 1H), 3.92 (s, 1H), 3.76 (s, 1H), 2.85 – 2.54 (m, 3H), 1.87 – 1.74 (m, 1H), 1.73 – 1.58 (m, 1H), 1.57 – 1.33 (m, 2H). ¹³C NMR (126 MHz, Chloroform-*d*) δ 155.8, 144.1, 141.5, 141.4, 139.8, 127.73, 127.70, 127.15, 127.10, 125.09, 125.06, 120.02, 119.99, 114.4, 68.4, 67.5, 54.3, 47.5, 37.9, 33.3, 27.7, 24.2. HRMS (m/z): calc. for C₂₃H₂₅ClNO₃ [M+H]⁺ = 398.1523; found, 398.1530



Synthesis of 14. A 50 L glass reactor was charged with compound 13 (2.81 kg), H₂O (5 L), and acetonitrile (10 L). After being cooled to 0–5 °C, NBS (1.01 kg, 5.69 mol) was added in portions while keeping the temperature below 5 °C. The reaction mixture was stirred at 0–5 °C for about 0.5 h. After completion of the reaction, Na₂SO_{3(aq)} (10%wt, 10 L) was added. The mixture was stirred for 0.5 h before extraction with EA (2 × 10 L). The combined organic layers were washed with saturated NaHCO_{3(aq)} (5 L) and saturated NaCl_(aq) (2 × 5 L), respectively. After addition of activated charcoal (320 g), the organic layer was stirred at 25 °C for 1 h. After addition of MgSO₄ (1.00 kg) the mixture was stirred for another 0.5 h. The mixture was filtered through a Buchner

funnel, and the filtrate was concentrated under reduced pressure to give crude **14** (2.85 kg) as a slurry, which was used directly in the next step.

Synthesis of 3. A 100 L glass reactor was charged with compound 15 (1.40 kg, 5.41 mol), LiOH (0.15 kg, 6.26 mol), and DMF (28 L). The reaction mixture was stirred at 25 °C for 1 h before being cooled to 0–5 °C. To the stirred mixture, crude compound 14 (2.85 kg) dissolved in DMF (2.8 L) was added dropwise in 4–5 h, while keeping the temperature below 5°C. The reaction mixture was stirred at 0–5 °C for 24 h before diethylamine (1 L) was added. The reaction mixture was stirred at 0–5 °C for another 12 h. After addition of H₂O (25 L) and EA (30 L), the layers were separated. The aqueous layer was extracted with EA (3×20 L). The combined organic layers were concentrated under reduced pressure. To the residual product was added Lactic acid_(aq) (85% wt, 1.5 kg). The mixture was stirred at 25 °C for 1 h before being extracted with MTBE (2×10 L). The aqueous layer (containing compound 3) was neutralized to pH 8-9 with K₂CO₃ before extraction with EA (3 \times 15 L). The combined organic layers (containing compound 3) were concentrated under reduced pressure to give crude 3, to which EA (8 L) was added. The mixture was stirred at 25 °C for 0.5 h before being filtered through a Buchner funnel. The filter cake was dried in vacuum. The final product weighted 1.84 kg as a white solid. Data for compound 3: 1 H NMR (400 MHz, Chloroform-*d*) δ 8.32 (s, 1H), 8.26 (s, 1H), 7.98 (s, 1H), 4.34 (d, J = 13.9 Hz, 1H), 4.16 (d, J = 13.9 Hz, 1H), 3.88 (t, J = 3.1 Hz, 1H), 3.29 (t, J = 3.4 Hz, 1H), 2.97 (d, J = 10.9 Hz, 1H), 2.52 (t, J = 11.8 Hz, 1H), 2.10 (d, J = 15.1 Hz, 1H), 2.03 (dd, J = 13.1, 3.7 Hz, 2H), 1.83 (d, J = 13.2 Hz, 1H), 1.81 - 1.72 (m, 1H), 1.54 (ddt, J = 15.0, 12.0, 3.4 Hz, 2H). ¹³C NMR (126) MHz, Chloroform-d) δ 160.1, 149.5, 147.2, 133.4, 132.7, 129.4, 127.9, 122.1, 105.3, 78.0, 55.8, 50.3, 44.67, 43.7, 26.9, 20.2. HRMS (m/z): calc. for $C_{16}H_{18}BrClN_3O_3$ [M+H]⁺ = 414.0220/416.0200; found, 414.0216/416.0197.



Synthesis of **1**. A 50 L glass reactor was charged with above obtained white solid **3** (1.84 kg) and EtOH (20 L). The mixture was refluxed for about 24 h, at which time the conversion to halofuginone was over 94% (HPLC). After being cooled to 55–60 °C, the reaction mixture was filtered through a Buchner funnel. The filter cake was washed with EtOH and then dried in vacuum. The final product weighted 1.65 kg as a white solid in 98.5% HPLC purity (69% yield for 4 steps from compound **12**, and 17% overall yield for 12 steps). Data for compound **1**: ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.23 (s, 1H), 8.22 (s, 1H), 8.15 (s, 1H), 4.99 (d, *J* = 2.8 Hz, 2H), 4.79 (d, *J* = 5.8 Hz, 1H), 2.98 (dt, *J* = 15.3, 4.7 Hz, 2H), 2.78 (d, *J* = 12.3 Hz, 1H), 2.64 (td, *J* = 8.9, 3.8 Hz, 1H), 2.44 (dd, *J* = 15.5, 8.7 Hz, 1H), 2.36 (td, *J* = 12.1, 2.7 Hz, 1H), 1.95 – 1.83 (m, 1H), 1.56 (dt, *J* = 13.3, 3.2 Hz, 1H), 1.34 (qt, *J* = 12.4, 3.7 Hz, 1H), 1.28 – 1.13 (m, 1H). ¹³C NMR (126 MHz, DMSO-*d*₆) δ 200.7, 158.6, 149.5, 147.2, 132.4, 131.8, 128.4, 126.8, 121.7, 66.7, 56.2, 54.4, 43.0, 30.5, 20.1. HRMS (m/z): calc. for C₁₆H₁₈BrClN₃O₃ [M+H]⁺ = 414.0220/416.0200; found, 414.021/4/16.0195.



Synthesis of (+)-12. A 500 mL glass reactor was charged with compound (\pm)-12 (10.0 g, 0.057 mol) and acetonitrile (100 mL). After being warmed to 60 °C, dibenzoyl-L-tartaric acid (21.4, 0.06 mol) dissolved in acetonitrile (60 mL) was added dropwise in 10 min. After being stirred at 60 °C for 20–30 min, the reaction mixture was cooled to 25 °C and stirred for 2 h. The white solid

crystallized out of the solution. After filtration through a Buchner funnel, the collected crude solid was washed with acetonitrile (20 ml) and then dried in vacuum. This salt was then mixed with H₂O–acetonitrile (1:4, 150 mL). The stirred mixture was warmed to 80 °C until most of the solid was dissolved. After filtration through a Buchner funnel, the filtrate was cooled to 25 °C and stirred for 2 h. The white solid crystallized out of the solution. After filtration through a Buchner funnel, the white solid was washed with acetonitrile (50 ml), and then dried in vacuum. The collected product weighted 9.7 g as a white solid. A 500 mL glass reactor was charged with the above obtained solid (9.7 g), H₂O (50 mL), and EA (50 mL). NaOH_(aq) (1 M) was slowly added to neutralize the stirred mixture to pH 12–14. After standing at r.t. for a few minutes, the layers were separated. The aqueous layer was extracted with EA (2 × 50 mL). The combined organic layers were dried on anhydrous Mg₂SO₄. After filtration through a Buchner funnel, the filtrate was concentrated under reduced pressure at 50 °C to provide compound (+)-**12** (3.2 g, 32%) as a white solid.

Following the procedures for the synthesis of racemic halofuginone, the preparations of compound (+)-13, (+)-14, (+)-3, and (+)-1 starting from compound (+)-12 were completed.

Optical rotation values of compound (+)-12, (+)-13, (+)-14, (+)-3, and (+)-1 (*Optical rotation were obtained at 20 °C, measured at 589 nm*) *are as follows:*

compound (+)-12: $[\alpha]_{D} = +8.4^{\circ}$ (*c* = 0.46, MeOH);

compound (+)-13: $[\alpha]_D = +25.4^{\circ}$ (*c* = 0.35, CHCl₃);

compound (+)-14: $[\alpha]_D = +36.7^{\circ}$ (*c* = 0.30, CHCl₃);

compound (+)-3: $[\alpha]_D = +81.2^\circ$ (*c* = 0.53, CHCl₃);

compound (+)-1: $[\alpha]_D = +18.5^{\circ}$ (*c* = 0.53, DMSO).

ASSOCIATED CONTENT

Supporting Information

¹H NMR, ¹³C NMR, HPLC analysis

AUTHOR INFORMATION

Corresponding Author

*jinyehua@launch-pharma.com

*qiufayang@launch-pharma.com

Author Contributions

[‡]H.X. and W.Y. contributed equally to this work.

Funding Sources

This work was supported by Launch-Pharma Technologies, Ltd.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENT

The authors thank Prof. Zhi Li (ShanghaiTech University, China) for helpful discussions and assistance in the preparation of the manuscript.

REFERENCES

(1) (a) Koepfli, J. B.; Mead, J. F.; Brockman, J. A. An Alkaloid with High Antimalarial Activity from Dichroa Febrifuga. *J. Am. Chem. Soc.* 1947, *69*, 1837. (b) Kuehl, F. A.; Spencer, C. F.; Folkers, K. Alkaloids of Dichroa Febrifuga Lour. *J. Am. Chem. Soc.* 1948, *70*, 2091–2093. (c) Koepfli, J. B.; Mead, J. F.; Brockman, J. A. Alkaloids of Dichroa Febrifuga. I. Isolation and Degradative Studies. *J. Am. Chem. Soc.* 1949, *71*, 1048–1054. (d) Koepfli, J. B.; Brockman, J. A.; Moffat, J. The Structure of Febrifugine and Isofebrifugin. *J. Am. Chem. Soc.* 1950, *72*, 3323. (e) Ablondi, F.; Gordon, S.; Morton, J.; Williams, J. H. An Antimalarial Alkaloid from Hydrangea. II. Isolation. *J. Org. Chem.* 1952, *17*, 14–18.

(2) (a) Edgar, S. A.; Flanagan, C. Efficacy of Stenorol® (Halofuginone): I. Against Recent Field Isolates of Six Species of Chicken Coccidia. *Poult. Sci.* 1979, *58*, 1469–1475. (b) Angel, S.; Weinberg, Z. G.; Polishuk, O.; Heit, M.; Plavnik, I.; Bartov, I. A Connection Between a Dietary Coccidiostat and Skin Tears of Female Broiler Chickens. *Poult. Sci.* 1985, *64*, 294–296. (c) Folz, S. D.; Lee, B. L.; Nowakowski, L. H.; Conder, G. A. Anticoccidial Evaluation of Halofuginone, Lasalocid, Maduramicin, Monensin and Salinomycin. *Vet. Parasitol.* 1988, *28*, 1–9. (d) Granol, I.; Bartov, I.; Plavnik, I.; Wax, E.; Hurwitz, S.; Pines, M. Increased Skin Tearing in Broilers and Reduced Collagen Synthesis in Skin *In Vivo* and *In Vitro* in Response to the Coccidiostat Halofuginone. *Poult. Sci.* 1991, *70*, 1559–1563. (e) Ernst, R. A.; Vohra, P.; Kratzer, F. H.; Kuhl, H. J. Effect of Halofuginone (Stenorol) on Chukar Partridge (Alectoris chukar). *Poult. Sci.* 1996, *75*, 1493–1495.

(3) (a) Jarvie, B. D.; Trotz-Williams, L.A.; McKnight, D. R.; Leslie, K. E.; Wallace, M. M.; Todd, C. G.; Sharpe, P. H.; Peregrine, A. S. Effect of Halofuginone Lactate on the Occurrence of *Cryptosporidium parvum* and Growth of Neonatal Dairy Calves. *J. Dairy Sci.* 2005, 88,

1801–1806. (b) De Waele, V.; Speybroeck, N.; Berkvens, D.; Mulcahy, G.; Murphy, T.M. Control of Cryptosporidiosis in Neonatal Calves: Use of Halofuginone Lactate in Two Different Calf Rearing Systems. *Prev. Vet. Med.* **2010**, *96*, 143–151.

(4) (a) McGaha, T. L.; Phelps, R. G.; Spiera, H.; Bona, C. Halofuginone, an Inhibitor of Type-I Collagen Synthesis and Skin Sclerosis, Blocks Transforming-Growth-Factor-β-Mediated Smad3 Activation in Fibroblasts. J. Invest. Dermatol. 2002, 118, 461–470. (b) McGaha, T. L.; Kodera, T.; Spiera, H.; Stan, A. C.; Pines, M.; Bona, C. A. Halofuginone Inhibition of COL1A2 Promoter Activity Via a c-Jun–Dependent Mechanism. Arthritis Rheumatol. 2002, 46, 2748–2761. (c) Sheffer, Y.; Leon, O.; Pinthus, J. H.; Nagler, A.; Mor, Y.; Genin, O.; Iluz, M.; Kawada, N.; Yoshizato, K.; Pines, M. Inhibition of Fibroblast to Myofibroblast Transition by Halofuginone Contributes to the Chemotherapy-Mediated Antitumoral Effect. Mol. Cancer Ther. 2007, 6, 570-577. (d) Sundrud, M. S.; Koralov, S. B.; Feuerer, M.; Calado, D. P.; Kozhaya, A. E.; Rhule-Smith, A.; Lefebvre, R. E.; Unutmaz, D.; Mazitschek, R.; Waldner, H.; Whitman, M.; Keller, T.; Rao, A. Halofuginone Inhibits Th17 Cell Differentiation by Activating the Amino Acid Starvation Response. Science. 2009, 324, 1334–1338. (e) Sato, S.; Kawamura, H.; Takemoto, M.; Maezawa, Y.; Fujimoto, M.; Shimoyama, T.; Koshizaka, M.; Tsurutani, Y.; Watanabe, A.; Ueda, S.; Halevi, K.; Saito, Y.; Yokote, K. Halofuginone Prevents Extracellular Matrix Deposition in Diabetic Nephropathy. Biochem. Biophys. Res. Commun. 2009, 379, 411–416. (f) Nelson, E.F.; Huang, C. W.; Ewel, J. M.; Chang, A. A.; Yuan, C. Halofuginone Down-Regulates Smad3 Expression and Inhibits the TGFbeta-Induced Expression of Fibrotic Markers in Human Corneal Fibroblasts. Mol. Vis. 2012, 18, 479–487. (g) Keller1, T. L.; Zocco1, D.; Sundrud, M. S.; Hendrick, M.; Edenius, M.; Yum, J.; Kim, Y.-J.; Lee, H.-K.; Cortese, J. F.; Wirth, D. F.; Dignam, J. D.; Rao, A.; Yeo, C.-Y.; Mazitschek, R.; Whitman, M. Halofuginone and Other Febrifugine Derivatives Inhibit ProlyltRNA Synthetase. *Nat. Chem. Biol.* **2012**, *8*, 311–317. (h) Carlson, T. J.; Pellerin, A.; Djuretic, I. M.; Trivigno, C.; Koralov, S. B.; Rao, A.; Sundrud, M. S. Halofuginone-Induced Amino Acid Starvation Regulates Stat3-Dependent Th17 Effector Function and Reduces Established Autoimmune Inflammation. *J. Immunol.* **2014**, *192*, 2167–2176. (i) Park, M. K.; Park, J. S.; Park, E. M.; Lim, M. A.; Kim, S. M.; Lee, D. G.; Baek, S. Y.; Yang, E. J.; Woo, J. W.; Lee, J.; Kwok, S. K.; Kim, H. Y.; Cho, M. L.; Park, S. H. Halofuginone Ameliorates Autoimmune Arthritis in Mice by Regulating the Balance Between Th17 and Treg Cells and Inhibiting Osteoclastogenesis. *Arthritis Rheumatol.* **2014**, *66*, 1195–1207. (j) Cui, Z.; Crane, J.; Xie, H.; Jin, X.; Zhen, G.; Li, C.; Xie, L.; Wang, L.; Bian, Q.; Qiu, T.; Wan, M.; Xie, M.; Ding, S.; Yu, B.; Cao, X. Halofuginone Attenuates Osteoarthritis by Inhibition of TGF-β Activity and H-Type Vessel Formation in Subchondral Bone. *Ann. Rheum. Dis.* **2016**, *75*(9), 1714–1721.

(5) (a) McLaughlin, N. P.; Evans, P.; Pines, M. The Chemistry and Biology of Febrifugine and Halofuginone. *Bioorg. Med. Chem.* 2014, 22, 1993–2004. (b) Smullen, S.; McLaughlin, N. P.; Evans, P. Chemical Synthesis of Febrifugine and Analogues. *Bioorg. Med. Chem.* 2018, 26, 2199–2220. (c) Takeuchi, Y.; Abe, H.; Harayama, T. Total Synthesis of *dl*-Febrifugine and *dl*-Isofebrifugine. *Chem. Pharm. Bull.* 1999, 47, 905–906; (d) Takeuchi, Y.; Hattori, M.; Abe, H.; Harayama, T. Synthesis of D/L-Febrifugine and D/L-Isofebrifugine. *Synthesis* 1999, 10, 1814–1818;
(e) Kobayashi, S.; Ueno, M.; Suzuki, R.; Ishitani, H.; Kim, H.-S.; Wataya, Y. Catalytic Asymmetric Synthesis of Antimalarial Alkaloids Febrifugine and Isofebrifugine and Their Biological Activity. *J. Org. Chem.* 1999, 64, 6833–6841. (f) Takeuchi, Y.; Azuma, K.; Takakura, K.; Abe, H.; Kim, H.-S.; Wataya, Y.; Harayama, T. Asymmetric Synthesis of (+)-Febrifugine and (+)-Isofebrifugine Using Yeast Reduction. *Tetrahedron.* 2001, *57*, 1213–1218. (g) McLaughlin, N. P.; Evans, P. Dihydroxylation of Vinyl Sulfones: Stereoselective Synthesis of (+)- and (-)-

Febrifugine and Halofuginone. J. Org. Chem. 2010, 75, 518–521. (h) Li, W.; Qin, T.; Chen, L.;
Chen, G. Method for Synthesis of Halofuginone Hydrobromide and its Analogs. CN101987843,
A, Mar 23, 2011. (i) Smullen, S.; Evans, P. An Asymmetric Synthesis of Febrifugine,
Halofuginone and Their Hemiketal Isomers. *Tetrahedron* 2017, 73, 5493–5499. (j) Wang, C.; Liu,
Y.-W.; Zhou, Z.; Si, C.-M.; Sun, X.; Wei, B.-G. Diastereoselective Approach to *trans*-5-Hydroxy6-Substitutedethanone-2-Piperidinones: Scalable Syntheses of (+)-febrifugine and (+)halofuginone. *Tetrahedron*. 2018, 74, 2158–2165. (k) Maiden, T. M. M.; Mbelesi, N.; Procopiou,
P. A.; Swanson, S.; Harrity, J. P. A. A Convergent Strategy towards Febrifugine and Related
Compounds. Org. Biomol. Chem. 2018, 16, 4159–4169.

(6) (6a) Bosch, J.; Bonjoch, J. Synthetic Route to 6-Functionalized 2-Azabicyclo[3.3.1]nonanes. *J. Org. Chem.* 1981, 46, 1538–1543. (6b) Sunagawa, M.; Itoh, M.; Kubota, K.; Sasaki, A.; Ueda,
Y.; Angehrn, P.; Bourson, A.; Goetschi, E.; Hebeisen, P.; Then, R. L. New Anti-MRSA and AntiVRE Carbapenems; Synthesis and Structure-activity Relationships of 1β-Metyl-2-(thiazol-2ylthio) carbapenems. *J. Antibiotics* 2002, *55*, 722–757.

(7) Meredith, E. L.; Mainolfi, N.; Poor, S.; Qiu, Y.; Miranda, K.; Powers, J.; Liu, D.; Ma, F.;
Solovay, C.; Rao, C.; Johnson, L.; Ji, N.; Artman, G.; Hardegger, L.; Hanks, S.; Shen, S.;
Woolfenden, A.; Fassbender, E.; Sivak, J. M.; Zhang, Y.; Long, D.; Cepeda, R.; Liu, F.;
Hosagrahara, V. P.; Lee, W.; Tarsa, P.; Anderson, K.; Elliott, J.; Jaffee, B. Discovery of Oral
VEGFR-2 Inhibitors with Prolonged Ocular Retention That Are Efficacious in Models of Wet
Age-Related Macular Degeneration. *J. Med. Chem.* 2015, *58*, 9273–9286.

(8) Takeuchi, Y.; Azuma, K.; Abe, H.; Harayama, T. Reaction of N-Acylated Isofebrifugine with Acid. *Heterocycles*. **2000**, *53*, 2247–2252.