

Available online at www.sciencedirect.com



Journal of Fluorine Chemistry 126 (2005) 297-300



www.elsevier.com/locate/fluor

# Synthesis and fungicidal activity of fluorine-containing phenylimino-thiazolidines derivatives

Xiaoyong Xu<sup>a</sup>, Xuhong Qian<sup>b,\*</sup>, Zhong Li<sup>a</sup>, Gonghua Song<sup>a</sup>, Weidong Chen<sup>a</sup>

<sup>a</sup>Shanghai Key Laboratory of Chemical Biology, Institute of Pesticides and Pharmaceuticals, East China University of Science and Technology, P.O. Box 544, 130 Meilong Road, Shanghai 200237, PR China <sup>b</sup>State Key Laboratory of Fine Chemicals, Dalian University of Technology, Dalian 116012, PR China

> Received 20 January 2004; accepted 26 October 2004 Available online 28 November 2004

#### Abstract

Nine new fluorine-containing phenylimino-thiazolidines derivatives were prepared. The structures of all compounds were confirmed by <sup>1</sup>H NMR, mass and high resolution mass spectroscopy. The antifungicidal activities of the title compounds on *Phytophthoza capsici* L., *Pyriculazia ozyzae* C., *Fusazium* spp. at 100 ppm were screened.

© 2004 Elsevier B.V. All rights reserved.

Keywords: Phenylimino-thiazolidines; Synthesis; Antifungicidal activity

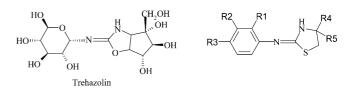
#### 1. Introduction

Recently, a variety of the reports regarding synthetic studies of the trehazolin derivatives have been presented due to the chemical and biological interests to the trehazolins [1–5]. Trehazolin, just like Validamycin A which has been commercialized and is used to control blight sheath of rice caused by the plant pathogenic fungus Rhizoctonia solani, is a slow, tight-binding inhibitor of trehalase [2,6]. It also shows potential fungicidal activity to control R. solani at 100 ppm [7]. But, the total synthesis of trehazolin is not an easily case, Masao shiozaki reported trehezolin was obtained from an 15-step synthesis starting from D-glucose, which was a convergent strategy [8-12]. It is difficult to make Trehezolin be commercialized, thus, to modify and simplify trehezolin's structure and find commercialized compound is our interest. It is well known that many fluorine-containing compounds exhibit significant agricultural bioactivities owing to fluorine atom's unique

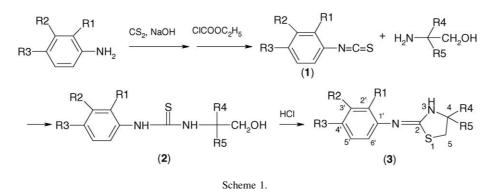
\* Corresponding author. E-mail address: xhqian@ecust.edu.cn (X. Qian).

0022-1139/\$ – see front matter  $\odot$  2004 Elsevier B.V. All rights reserved. doi:10.1016/j.jfluchem.2004.10.018

properties, such as high thermal stability and lipophilicity [13]. Previously, we ever reported fluorine-containing N,N'-diphenylcarbaminidothioates which exhibited fungicidal activity toward *R. solani* and *Pyricoraria orizae* [14]. In this report, based on the structure features of trehazolin, we designed and synthesized fluorine-containing phenylimino-thiazolidines derivatives and investigated their bioactivities



By a facile and convenient method, novel fluorine-containing phenylimino-thiazolidines (**3a**–**i**) were synthesized. Very interestingly, the preliminary bioassay tests showed that some compounds (**3f**, **3g**) exhibited good fungiticidal activity on *Phytophthoza capsici* L., *Pyriculazia ozyzae* C., *Fusazium* spp. at 100 ppm.



#### 2. Results and discussion

The designed compounds were prepared by Scheme 1. The aryl isothiocyanates were commonly prepared from arylamines by treatment with carbon disulfide, aqueous sodium hydroxide and chloroformate according to our reported procedure [15].

Reaction of arylamines and carbon disulfide in NaOH solution was carried out to give intermediate dithiocarbamates. After adding ethyl chloroformate, aryl isothiocyanates (1) was obtained. Compounds (2) was synthesized by reaction of aryl isothiocyanates (1) with substituted aminoethanol in ethyl acetate under reflux or at room temperature, the reaction rate depended on the nature of corresponding amine. When the substituents were -H,  $-CH_3$ , the corresponding compounds (3) could be synthesized easily at room temperature for a few minutes or half an hour, respectively. When the substituents were  $-CH_2OH$  or  $-CH_2CH_3$ , the products could be obtained under reflux for 5-6 h. The target compounds (3) were obtained by the heating of compounds (2) in hydrochloric acid at 90 °C. It is shown in Scheme 1.

Compound no.	R1	R2	R3	R4	R5
3a	Н	F	F	CH <sub>3</sub>	CH <sub>3</sub>
3b	Н	F	F	CH <sub>2</sub> OH	CH <sub>2</sub> OH
3c	Н	F	F	CH <sub>2</sub> OH	CH <sub>2</sub> CH <sub>3</sub>
3d	Н	Н	F	CH <sub>3</sub>	$CH_3$
3e	Н	Н	F	CH <sub>2</sub> OH	CH <sub>2</sub> CH <sub>3</sub>
3f	Н	Н	F	Н	Н
3g	F	F	F	Н	Н
3h	F	F	F	CH <sub>2</sub> OH	CH <sub>2</sub> CH <sub>3</sub>
3i	F	F	F	CH <sub>3</sub>	CH <sub>3</sub>

The fungicide activities of compound 3a-i were screened at the concentration of 100 ppm according to the same method as literature [16]. The fungicidal data are listed in Table 1.

The data indicated that the fungicidal activities of compounds **3a–i** against *R. solani* and *P. orizae* were lower than the series of N,N'-diphenylcarboamimidothioates.

But, we surprised to find that our compounds(**3f** and **3g**), especially the compound **3f**, were highly toxic to the test fungi *P. capsici* L., *Fusazium* spp. Their fungicidal activities depended upon the position of the fluorine on the aryl rings and the substituent on five-member heterocycle. Introduction of fluorine into the *para* position of the aryl rings increased greatly the their fungi toxicities. Meanwhile, when increasing the hydrophilicity of the group on five-member heterocycles being hydro, showed the significant biological activity, e.g. compound **3f** and **3g**. But, their antifungicidal activities decreased very quickly with the decrease of concentration.

#### 3. Experimental

Melting points were obtained with an electrothermal digital apparatus made in Beijing and are uncorrected. The infrared (IR) spectra were recorded on a Nicolet 470 infrared Fourier transform spectrometer using a potassium bromide pellets. The proton nuclear magnetic resonance(<sup>1</sup>H NMR, 500 MHz) spectra were recorded on a Bruker WP-500SY spectrometers with CD<sub>3</sub>Cl as the solvent and TMS as internal standard. High resolution mass spectra were obtained on MicroMass GCT CA 055 spectrometers. Analytical thin-layer chromatography (TLC) was carried out on precoated plate (silica gel 60 F<sub>254</sub>), and spots were visualized with ultraviolet light. All chemicals or reagents were purchased from standard commercial suppliers.

#### 3.1. Synthesis of phenylisothiocyanate (1)

To a stirred solution of sodium hydroxide (2.40 g, 0.06 mol) in 30 ml H<sub>2</sub>O, carbon disulfide (4.57 g, 0.06 mol) was added at 2–5 °C, then, substituted aniline (0.06 mol) was added over a period of 30 min. After the mixture was refluxed for 24 h, ethyl chloroformate (6.51 g, 0.06 mol) was added dropwise in 35–40 °C and the resulting mixture were stirred for about 40 min at the same temperature. The organic phase was separated and washed with water, dried over anhydrous magnesium and concentrated under reduced pressure and the residue was distilled to obtain colorless liquid.

Table 1 Fungicidal percent mortality (%) of  $I_{a-i}$  at 100 ppm

Compound	P. capsici L.	P. ozyzae C.	Fusazium spp.	R. Solani K.
3a	5.1	1.8	2.3	9.8
3b	3.0	6.7	0.7	2.0
3c	50.0	57.0	1.0	0.7
3d	4.4	4.3	2.1	5.3
3e	8.2	1.3	3.7	4.6
3f	100, 42.9 (50 ppm)	87.5, 25.0 (50 ppm)	100, 33.3 (50 ppm)	60.0
3g	79.2, 21.4 (50 ppm)	85.7, 12.5 (50 ppm)	57.9, 22.2 (50 ppm)	80.0
3h	6.0	3.3	2.5	1.6
3i	2.2	1.3	8.9	11.3

#### 3.2. Substituted thiourea preparation procedure (2)

Appropriate amine was dissolved in ethyl acetate, and appropriate isothiocyanate was added dropwise with stirring and the mixture was kept at room temperature for 30–60 min or refluxed for 2–8 h. Then, the solvent was evaporated under reduced pressure to give the crude product, it could be used to the next reaction directly.

# 3.3. General synthetic procedure of substituted phenylimino-thiazolidine (3)

Appropriate thiourea (10 mmol) was dissolved in concentrated HCl (10 ml) and heated at 90 °C for 45 min. The cooled mixture was basified with 10N NaOH in an ice bath. The solid was filtered and recrystallized or the precipitated gummy residue was extracted with  $Et_2O$ . The extract was washed with brine, dried and evaporated to dryness.

# *3.3.1.* Syntheses of 2-(3,4-difluorophenylimino)-4, 4-dimethyl-thiazolidine (**3a**)

Yield 68%, mp 183–184 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 1.40 (s, 6H, CH<sub>3</sub>), 3.16 (s, 2H, H-5), 7.00 (br S, 1.08H, H-2' and H-6'), 7.18 (q, 1H,  $J_{3'5'} = 9.0$  Hz,  $J_{5'6'} = 9.0$  Hz,  $J_{4'5'} = 9.1$  Hz, H-5'). IR (KBr) (cm<sup>-1</sup>): 3125 (NH), 2920, 2875 (C–H), 1630 (C=N), 1600, 1500 (Ph), 1280 (C–S), 1140, 870, 820, 780, 650. MS (EI, 70 eV): m/z (%): 242 (44.38) [*M*], 227 (100.00) [*M* – CH<sub>3</sub>], 154 (22.18) [*M* – C<sub>4</sub>H<sub>8</sub>S], 129 (8.89), 88 (15.40), 55 (15.00). HRMS Calcd. for C<sub>11</sub>H<sub>12</sub>F<sub>2</sub>N<sub>2</sub>S 242.0689. Found: 242.0701.

#### 3.3.2. Syntheses of 2-(3,4-difluorophenylimino)-4,4dihydroxymethyl-thiazolidine (**3b**)

Yield 73%, mp 207–208 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 3.32 (s, 2H, H-5), 3.66 (d, 2H, *J* = 11.0 Hz, CH<sub>2a(b)</sub>OH), 3.69 (d, 2H, *J* = 11.0 Hz, CH<sub>2b(a)</sub>OH), 7.18 (q, 1H, *J*<sub>3'5'</sub> = 9.1 Hz, *J*<sub>4'5'</sub> = 10.5 Hz, *J*<sub>5'6'</sub> = 9.1 Hz, H-5'), H-2' and H-6' (no signal). IR (KBr) (cm<sup>-1</sup>): 3300 (OH), 3125 (NH), 2875, 1630 (C=N), 1600, 1510, 1200, 1050, 780. MS (EI, 70 eV): *m/z* (%): 274 (11.02) [*M*], 243 (100.00) [*M* – CH<sub>2</sub>OH], 213 (56.94) [*M* – C<sub>2</sub>H<sub>5</sub>O<sub>2</sub>], 155 (9.50), 128 (6.64), 113 (5.72), 88 (5.60). HRMS Calcd. for C<sub>11</sub>H<sub>12</sub> F<sub>2</sub>N<sub>2</sub>O<sub>2</sub>S 274.0588. Found: 274.0585.

# 3.3.3. Syntheses of 2-(3,4-difluorophenylimino)-4-ethyl-4-hydroxymethyl-thiazolidine (**3c**)

Yield 51%, mp 143–144 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 1.00 (t, 3H, J = 7.5 Hz, CH<sub>3</sub>), 1.76 (m, 2H, CH<sub>2</sub>), 3.10 (d, 1H, J = 10.9 Hz, H<sub>a(b)</sub>-5), 3.40 (d, 1H, J = 10.9 Hz, H<sub>b(a)</sub>-5), 3.57 (d, 1H, J = 10.9 Hz, H<sub>a(b)</sub>-CH<sub>2</sub>OH), 3.61 (d, 1H, J = 10.9 Hz, H<sub>b(a)</sub>-CH<sub>2</sub>OH), 7.10 (br S, 1H, H-2'), 7.19 (q, 1H',  $J_{4'5'} = J_{6'5'} = 9.0$  Hz,  $J_{5'3'} = 9.0$  Hz, H-5'), 7.60 (br S, 1H, H-6'). IR (KBr) (cm<sup>-1</sup>): 3300, 3100, 2875, 1640 (C–N), 1600, 1500 (Ph), 1280 (C–S), 1140, 870, 820, 780, 650. MS (EI, 70 eV): *m/z* (%): 272 (614) [*M*], 243 (6.12) [*M* – C<sub>2</sub>H<sub>5</sub>], 242 (11.41) [*M* – HCHO], 241 (100.00) [*M* – CH<sub>2</sub>OH], 102 (30.04), 102 (9.56), 87 (9.01). HRMS Calcd. for C<sub>12</sub>H<sub>14</sub>F<sub>2</sub>N<sub>2</sub>OS 272.0795. Found: 272.0791.

## 3.3.4. Syntheses of 2-(4-fluorophenylimino)-4,4-dimethylthiazolidine (**3d**)

Yield 61%, mp 209–210 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 1.39 (s, 6H, CH<sub>3</sub>) 3.18 (s, 2H, H-5), 7.00 (t, 2H,  $J_{2'3'} = J_{5'6'} = 8.3$  Hz,  $J_{4'3'} = J_{4'5'} = 8.8$  Hz, H-3' and H-5'), 7.36 (br S, 1.2H, H-2' and H-6'). IR (KBr) (cm<sup>-1</sup>): 3100 (NH), 2950, 2825 (C–H), 1630 (C–N), 1590, 1500 (Ph), 1320, 1210 (C–S), 1190, 1170, 850, 760, 650. MS (EI, 70 eV): m/z (%): 225 (23.84) [M + 1], 224 (45.53) [M], 209 (100.00) [M – CH<sub>3</sub>], 137 (11.80), 136 (27.87), 88 (12.72). HRMS Calcd. for C<sub>11</sub>H<sub>13</sub>FN<sub>2</sub>S 224.0783. Found: 224.0788.

## 3.3.5. Syntheses of 2-(4-fluorophenylimino)-4-ethyl-4hydroxymethyl-thiazolidine (**3e**)

Yield 42%, mp 154–155 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 0.99 (t, 3H, J = 7.5 Hz, CH<sub>3</sub>), 1.69 (m, 2H, CH<sub>2</sub>), 3.06 (d, 1H, J = 10.9 Hz, H<sub>a(b)</sub>-5), 3.36 (d, 1H, J = 10.9 Hz, H<sub>b(a)</sub>-5), 3.52 (d, 1H, J = 10.8 Hz, H<sub>a(b)</sub>-CH<sub>2</sub>OH), 3.59 (d, 1H, J = 10.8 Hz, H<sub>b(a)</sub>-CH<sub>2</sub>OH), 7.00 (t, 2H,  $J_{2'3'} = J_{6'5'} = 8.9$  Hz,  $J_{4'5'} = J_{4'3'} = 8.8$  Hz, H-3'and H-5'), 7.49 (br S, 1.7H, H-2' and H-6'). IR (KBr) (cm<sup>-1</sup>): 3300 (OH), 3100 (NH), 2875, 1640 (C–N), 1500 (Ph), 1200, 1050, 850. MS (EI, 70 eV): m/z (%): 254 (5.61) [*M*], 223 (100.00) [*M* - CH<sub>2</sub>OH], 195 (4.31) [*M* - HCHO-C<sub>2</sub>H<sub>5</sub>], 102 (23.28). HRMS Calcd. for C<sub>12</sub>H<sub>15</sub>FN<sub>2</sub>OS 254.0889. Found: 254.0880.

## 3.3.6. Syntheses of 2-(4-fluorophenylimino)-tetrahydrothiazolidine (3f)

Yield 84%, mp 153–154 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 3.32 (t, 2H, *J* = 7.2 Hz, H-5), 3.92 (s, 2H, H-4), 7.01 (m, 2H, H-3' and H-5'), 7.40 (br S, 2H, H-2' and H-6'). IR (KBr) (cm<sup>-1</sup>): 3125 (NH), 2800, 1630 (C=N), 1600, 1500 (Ph), 1300, 1200, 1180, 780, 620. MS (EI, 70 eV): *m/z* (%): 196 (93.04) [*M*], 168 (11.72) [*M* – CH<sub>2</sub>=CH<sub>2</sub>], 149 (12.52) [*M* – SCH<sub>3</sub>], 136 (100.00) [*M* – C<sub>2</sub>H<sub>4</sub>S], 122 (32.36) [*M* – C<sub>2</sub>H<sub>4</sub>NS], 109 (22.65) [*M* – C<sub>3</sub>H<sub>5</sub>NS], 95 (17.34). HRMS Calcd. for C<sub>9</sub>H<sub>9</sub> FN<sub>2</sub>S 196.0470. Found: 196.0479.

## 3.3.7. Syntheses of 2-(2,3,4-trifluorophenylimino)tetrahydro-thiazolidine (**3g**)

Yield 80%, mp 151–152 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 3.39 (t, 2H, J = 7.0 Hz, H-5), 3.74 (t, 2H, J = 7.0 Hz, H-4), 6.90 (br S, 1H, H-6'), 7.06 (q, 1H,  $J_{4'5}$  = 10.4 Hz,  $J_{6'5'}$  = 10.4 Hz,  $J_{3'5'}$  = 9.2 Hz, H-5'). IR (KBr) (cm<sup>-1</sup>): 3150 (NH), 2850, 1650 (C=N), 1630, 1600, 1500, 1450 (Ph), 1320, 1230, 1040, 980. MS (EI, 70 eV): m/z (%): 232 (100.00) [M], 213 (77.11) [M – F], 185 (13.55) [M – SCH<sub>3</sub>], 172 (88.86) [M – SC<sub>2</sub>H<sub>4</sub>], 158 (25.37): [M – C<sub>2</sub>H<sub>4</sub>NS], 145 (12.94), 61 (9.61). HRMS Calcd. for C<sub>9</sub>H<sub>7</sub>F<sub>3</sub>N<sub>2</sub>S 232.0282. Found: 232.0296.

#### *3.3.8.* Syntheses of 2-(2,3,4-trifluorophenylimino)-4-ethyl-4-hydroxymethyl-thiazolidine (**3h**)

Yield 52%, mp 165–166 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz)  $\delta$ : 1.00 (t, 3H, J = 7.5 Hz, CH<sub>3</sub>), 1.78 (q, 2H, J = 7.5 Hz, CH<sub>2</sub>), 3.31 (d, 1H, J = 11.1 Hz, H<sub>a(b)</sub>-5), 3.53 (d, 1H, J = 11.1 Hz, H<sub>b(a)</sub>-5), 3.56 (d, 1H, J = 11.1 Hz, CH<sub>2a(2b)</sub>OH), 3.71 (d, 1H, J = 11.1 Hz, CH<sub>2b(2a)</sub>OH), 6.87 (br S, 0.6H, H-6'), 7.05 (q, 1H,  $J_{2'5'} = 9.1$  Hz,  $J_{4'5'} = 10.1$  Hz,  $J_{3'5'} = 8.4$  Hz, H-5'). IR (KBr) (cm<sup>-1</sup>): 3300 (OH), 3100, 2875, 1630 (C–N), 1500 (Ph), 1250 (C–S), 1220, 1050, 980, 820. MS (EI, 70 eV) m/z (%): 290 (7.33) [M], 259 (100.00) [M - CH<sub>2</sub>OH], 231 (36.64): [M - C<sub>3</sub>H<sub>7</sub>O], 172 (6.08), 102 (24.29), 87 (9.46). HRMS Calcd. for C<sub>12</sub>H<sub>13</sub>F<sub>3</sub>N<sub>2</sub>OS 290.0701. Found: 290.0702.

# *3.3.9.* Syntheses of 2-(2,3,4-trifluorophenylimino)-4,4dimethyl-thiazolidine (**3i**)

Yield 73%, mp 195–196 °C. <sup>1</sup>H NMR (CD<sub>3</sub>COCD<sub>3</sub>, 500 MHz) δ: 1.43 (s, 6H, CH<sub>3</sub>), 3.22 (s, 2H, H-5), 6.81 (br S,

0.36H, H-2' and H-6'), 7.10 (q, 1H,  $J_{4'5} = 8.9$  Hz,  $J_{6'5'} = 8.9$  Hz,  $J_{3'5'} = 8.3$  Hz, H-5'). IR (KBr) (cm<sup>-1</sup>): 3150 (NH), 2950, 2875, 1620 (C=N), 1600, 1500, 1300, 1260, 1230 (C–S), 1000, 820. MS (EI, 70 eV): m/z (%): 260 (79.16) [*M*], 245 (100.00) [*M* – CH<sub>3</sub>], 213 (6.70) [*M* – SCH<sub>3</sub>], 172 (28.62), 147 (12.96), 88 (29.87), 55 (29.98). HRMS Calcd. for C<sub>11</sub>H<sub>11</sub>F<sub>3</sub>N<sub>2</sub>S 260.0595. Found: 260.0612.

#### Acknowledgement

This work was partly supported by Shanghai Education Committee, National Natural Science Foundation of China, and Minister of Science and Technology (2001AA235011; 2003CB114405).

#### References

- O. Ando, H. Satake, K. Itoil, A. Sato, M. Nakajima, H. Haruyama, Y. Ohkuma, T. Kinoshita, R. Enokita, J. Antibiot. 44 (1991) 1165–1168.
- [2] O. Ando, M. Nakajima, M. Kifune, H. Fang, K. Tanzawa, Biochim. Biophys. Acta 1244 (1995) 295–302.
- [3] Y. Kobayashi, Carbohydr. Res. 315 (1999) 3-15.
- [4] Y. Kobayashi, H. Miyazaki, M. Shiozaki, J. Antibiot. 47 (1994) 932– 938.
- [5] T. Nakayama, T. Amachi, S. Murao, T. Sakai, T. Shin, P.T. Kenny, T. Iwashita, M. Zagorski, H. Komura, K. Nomoto, J. Chem. Soc., Chem. Commun. (1991) 919–920.
- [6] T. Iwase, E. Higashide, H. Yamamoto, M. Shitata, J. Antibiot. 24 (1971) 107–113.
- [7] O. Ando, M. Klfune, M. Nakajima, Biosci. Biotechnol. Biochem. 59 (1995) 711–712.
- [8] B.M. Trost, D.L. Van Vrankon, J. Am. Chem. Soc. 115 (1993) 444– 458.
- [9] J. Li, F. Lang, B. Ganem, J. Org. Chem. 63 (1998) 3403-3410.
- [10] S. Ogawa, C. Uchida, Chem. Lett. (1993) 173-176.
- [11] C. Uchida, T. Yamagishi, S. Ogawa, J. Chem. Soc., Prekin Trans. I (1994) 589–602.
- [12] K. Yoshiyuki, M. Hideki, S. Masao, J. Org. Chem. 59 (1994) 813–822.
- [13] M. Hudlicky, Chemistry of Organic Fluorine Compounds II, American Chemical Society, Washington, DC, 1995, p. 979, 1145.
- [14] Z. Li, X. Qian, G. Song, Z. Li, J. Fluorine Chem. 108 (2001) 143-146.
- [15] Z. Li, X. Qian, Org. Prep. Proc. Int. 32 (2000) 571–573.
- [16] K. Hostettmann, J.B. Harborne, P.M. Dey, Methods in Plant Biochemistry, vol. 6, Academic Press, London, UK, 1991, pp. 33–46.