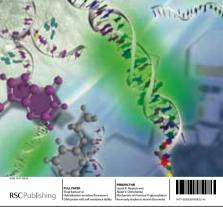
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Chemistry

Tandem One-Pot Synthesis of Flavans by Recyclable Silica-HClO₄ catalyzed Knoevenagel Condensation and [4+2]-Diels-Alder Cycloaddition^{#¤}

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An efficient one-pot multi-component synthesis of flavans using perchloric acid supported on silica as a recyclable heterogeneous catalyst has been described. This is the first report of

10 direct one-step construction of flavan skeleton from phenolic precursor. The method involves Knoevenagel-type condensation leading to in-situ formation of transient O-quinone methide which further undergoes [4+2]-Diels-Alder cycloaddition with styrene to yield flavan skeleton. The method provides easy access to wide range of bio-active natural products viz. flavonoids, anthocyanins and catechins.

Introduction

15

Flavans are a set of naturally occurring flavonoids possessing a 2phenyl-3,4-dihydro-2H-chromene nucleus. They are widely distributed in the plant kingdom¹ with >17000 natural flavans ²⁰ isolated so far² and many exhibit interesting and useful biological activities.^{1, 3} Amongst different flavans, flavan-3-ols are most abundant and occur in monomeric, oligomeric and polymeric forms, which are also known as condensed tannins or proanthocyanidins. They exhibit high degree of structural

- ²⁵ diversity varying according to the type of constitutive units, type of interflavan linkages, and degree of polymerization. Most widely known flavans are catechins (1-2, 2R,3S and 2S,3R) and epicatechins (3-4, 2R,3R and 2S,3S), which occur in plants such as catechu, tea cocoa etc. and are known to possess various
- 30 biological properties such as anticarcinogenic, anti-inflammatory, antioxidant, immunomodulatory, inhibition of bone resorption etc.⁴ Fully substituted monomeric flavans sideroxylonal B (5) and grandinal (6) have been reported from Eucalyptus sideroxylon and E. grandis which exhibited antibacterial activities against
- 35 gram-positive bacteria and inhibition of aldose reductase.⁵ Recently benzodipyran and benzotripyran type of compounds have been reported to posses potent angiogenic activities.⁶ Structures of flavans 1-6 are shown in Figure 1.

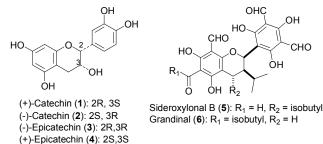


Figure 1 Examples of naturally occurring biologically active flavans

Flavans have received considerable amount of attention in last few decades because of their numerous bioactivities. However, 45 due to their structural complexity, very little synthetic work has been reported. Currently available protocols for synthesis of flavan skeleton include Heck-arylation of chromene intermediate with arenediazonium salt,⁷ reaction of tris-O-boc formyl phloroglucinol with styrene in presence of magnesium bromide ⁵⁰ dietherate and lithium aluminum hydride,⁸ reduction of dihydroflavones or dihydroflavonols,9 and cinnamylation of phenolic compounds with cinnamyl alcohol in presence of H₃PO₄.¹⁰ Synthesis of polyfunctional flavans sideroxylonals **5**¹¹ and grandinal 6^{12} have been reported *via* cycloaddition reaction 55 between in situ generated O-quinone methide from ortho-alkyl substituted phloroglucinols (using DDQ) and substituted styrene type compound. Most of the reported protocols require multiple steps to construct flavan skeleton; many of them require

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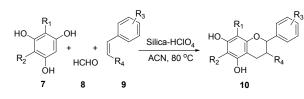
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[&]quot;Electronic supplementary information (ESI) available: NMR spectra of all new compounds. See DOI: 10.1039/xxxx

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expensive, hazardous chemicals and require anhydrous conditions. Several protocols comprise protection/ deprotection steps and uses homogeneous catalytic systems. Direct one-step construction of a flavan skeleton is not known in the literature.

5 In recent years, great number of short and efficient strategies have been discovered by chemists to facilitate synthesis of complex natural products.¹³ In this regard, several domino 'one-pot' strategies which consist of merging compatible single bond forming processes so as to allow multiple bond forming events 10 between several substrates, a concept generally termed as multicomponent reactions (MCRs) are coming up.¹⁴ In the present communication, we report synthesis of flavans directly from phloroglucinol or mono- or di-substituted phloroglucinols 7 using one-pot MCR between phloroglucinol 7, formaldehyde 8 and 15 styrene 9 in presence of recyclable heterogeneous solid acid catalyst in excellent yields. This one-pot protocol involves Knoevenagel type condensation of formaldehyde 8 with phloroglucinol 7 leading to in situ formation of transient Oquinone methide which further undergoes [4+2]-Diels-Alder ²⁰ cycloaddition with styrene 9 to yield flavan 10 (Scheme 1).



Scheme 1 One-pot MCR approach to flavan synthesis

Results and discussion

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- Initially, an experimental exploration on reaction parameters, including catalyst, solvent, temperature and reaction time, was conducted using the model MCR between 2,4-diacetyl ³⁰ phloroglucinol (**7c**), formaldehyde (**8**) and styrene (**9a**) (Table 1). Catalyst-free MCR was first attempted by varying solvent, reaction temperature and time; however no product was formed. Catalytic effect of various cation-exchange resins and silica based acid catalysts¹⁵⁻¹⁷ was investigated. Amberlyst-15 and amberlite ³⁵ catalysts led to formation of flavan **10c** in 50% yield. A brief examination of solvents showed ACN as a suitable solvent (entries 1 *vs.* 4 and 5), which was selected for further optimization studies. After numerous trials, we eventually found that all three silica based catalysts, silica-I₂, silica-FeCl₃ and
- ⁴⁰ silica-HClO₄ are competent, affording the flavan **10c** in >55% yield (entries 7-9). Further optimization revealed that the reaction could proceed smoothly by decreasing the catalyst loading as low as 10 %w/w (entry 14); however, 50% w/w silica-HClO₄ appeared to be superior in terms of efficiency and reaction time.
- ⁴⁵ Recyclability of the catalyst was checked to prove the heterogeneous nature and its repeated use. The MCR between 7c, 8 and 9a in presence of silica-HClO₄ (50 %w/w) led to formation of flavan 10c in 84, 65, 58 and 48% in 3 h reaction time over four cycles respectively. However, when reaction time was increased
- ⁵⁰ to 8, 12, 16 h for cycles 2-4, product yields were improved to 78, 70 and 65% respectively. In order to understand the reason for decreased catalytic activity of Silica-HClO₄ catalyst after its use,

View Online we determined total acidity of fresh and used catalyst (after first use) by ammonia-TPD experiment. The total acidity of the fresh

⁵⁵ catalyst was found to be 10.95 mmol NH₃/g, however it was decreased to 7.6 mmol NH₃/g after first use. This decreased acidity is a clear indication of the catalyst leaching after its use, which justifies the reason for decrease in the product yield from 84% (fresh catalyst) to 65% (first recycle). However, the catalyst ⁶⁰ could be recycled upto three times producing >50% yield of desired flavan.

We also performed control experiment with neat HClO₄ instead of Silica-HClO₄ catalyst. When 100 mol% of HClO₄ in ACN as a solvent was used, 20% product was formed after 3 h reflux (entry ⁶⁵ 16), however after reflux for 10 h, 60% of the desired flavan **10c** was formed (entry 17). The reaction when performed in neat HClO₄ (excess) without ACN, reaction did not proceed (entry 18). These results indicated that apart from ease of product isolation and its recyclable nature; Silica-HClO₄ catalyst is much ⁷⁰ more efficient in comparison to neat HClO₄.

Table 1 Solvent and catalyst optimization studies^a

Table I Solvent and catalyst optimization studies"					
	+ + Н	Catalyst Solvent	HO HO O	С ОН 10с	
Entry	Catalyst (% w/w)	Solvent	Temp.	Time	Yield ^b
		~~~~	(°C)	(h)	(%)
1	Amberlyst-15 (50)	ACN	80	3	50
2	Amberlite-IR-50 (50)	ACN	80	3	50
3	Amberlite-IR-140 (50)	ACN	80	3	50
4	Amberlyst-15 (50)	$H_2O$	100	3	20
5	Amberlyst-15 (50)	Dioxane	100	6	30
6	Silica gel (50)	ACN	80	6	0
7	Silica-I ₂ (50)	ACN	80	3	55
8	Silica-FeCl ₃ (50)	ACN	80	3	60
9	Silica-HClO ₄ (50)	ACN	80	3	84
10	Silica-HClO ₄ (50)	$H_2O$	100	3	50
11	Silica-HClO ₄ (50)	Dioxane	100	3	50
12	Silica-HClO ₄ (20)	ACN	80	3	70
13	Silica-HClO ₄ (10)	ACN	80	3	50
14	Silica-HClO ₄ (10)	ACN	80	12	80
15	Silica-HClO ₄ $(5)$	ACN	80	3	35
16	$HClO_{4}(100)$	ACN	70	3	20
17	HClO ₄ (100)	ACN	70	10	60
18	HClO ₄	HClO ₄	70	3	0

 a  Reagents and conditions: 7c (1 mmol), 8 (3 mmol), 9a (1.5 mmol) and 75 catalyst;  b  Isolated yields.

Next, we studied scope of the MCR. Phloroglucinol (7a) and its mono- 7b and di-substituted analogs 7c-7e,^{18, 19} styrenes (9a), 4-methyl styrene (9b), 4-tert-butyl styrene (9c), 2-vinyl napthalene ⁸⁰ (9d) and isosaffrol (9e) were investigated (Table 2). Under optimized reaction conditions, MCR of phloroglucinol (7a) with 8 and 9a led to formation of hexahydrobenzotripyran 10a in 35% yield (entry 1). Similarly, mono-substituted phloroglucinol 7b (entry 2) produced tetrahydrodipyran 10b in 52% yield. Like 2,4-⁸⁵ diacetyl phloroglucinol 7c (entry 3), other disubstituted

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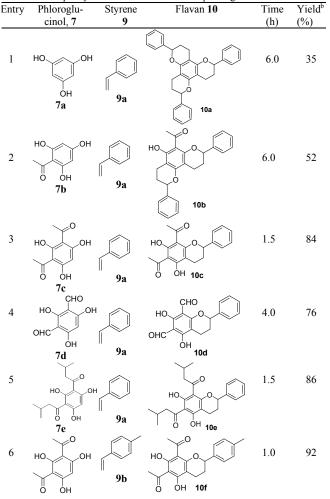
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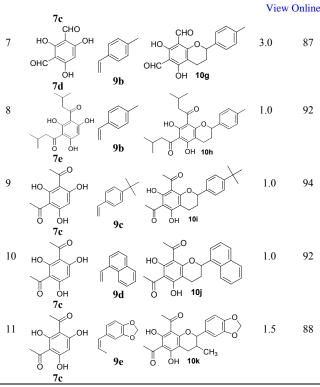
phloroglucinols **7d** and **7e** also produced desired flavans **10d** and **10e** in excellent yields (entries 4 and 5). 4-Methyl styrene **9b**, 4tert-butyl styrene **9c** as well as 2-vinyl naphthalene **9d** participated well in this reaction producing desired flavans in s good yields (entries 6-10). The MCR protocol also worked well with  $\beta$ -substituted styrene isosaffrol (**9e**). Reaction of **7c** with isosaffrol (**9e**) produced 3-methyl substituted flavan **10k** in 88% yield (entry 11). It is evident from the Table 2 that disubstituted phloroglucinols showed better reactivity in MCR compared with

¹⁰ mono-substituted followed by non-substituted phloroglucinol. Amongst different styrenes used, 4-methyl **9b** and t-butyl **9c** styrenes produced higher yields compared with unsubstituted styrene **9a**.

All synthesized flavans have been fully characterized using ¹⁵ melting point, ¹H NMR, ¹³C NMR, MS, IR and HRMS data. The CH proton at C₂ position shows typical 'dd (J = 2-6 and 8-15 Hz)' in ¹H NMR spectrum, which is in consistence with literature values.²⁰ The H₂ proton of flavan **10k** showed  $J_{2-3} = 9.9$  Hz indicating 2,3-trans relative stereochemistry. 2,3-Trans relative ²⁰ stereochemistry was further confirmed by NOESY experiment. The applicability of MCR protocol for substituted flavan **10k**, indicates its promise in total synthesis of variety of biologically important 3-substituted flavans (e.g. grandinal, catechins).

25 Table 2 One-pot synthesis of flavans 10 from phloroglucinols 7





^a Reagents and conditions: **7** (1 mmol), **8** (3 mmol), **9** (1.5 mmol) and catalyst silica-HClO₄ (50 %w/w) in ACN at 80 °C; ^b Isolated yields.

- Further, the utility of this protocol was explored for flavonoid 30 synthesis. Flavan 10c on treatment with DDQ in dioxane (1% water) led to formation of corresponding flavone 11 in 75% yield. Next, we attempted one-pot conversion of disubstituted phloroglucinol 7c to flavone 11. The mixture of 7c, 8, 9a and silica-HClO₄ in acetonitrile was refluxed for 1.5 h, followed by 35 addition of a DDQ (4 equiv). Reaction was continued to reflux for 6 h, which led to formation of desired flavone 11, but only in 10% yield. After further optimization studies, we found that flavan 10 synthesis works well in ACN solvent; however oxidation/dehydrogenation reaction requires dioxane solvent. ⁴⁰ Thus an optimized one-pot protocol for flavone synthesis directly from phloroglucinols 7 include, evaporation of ACN after flavan 10c formation followed by addition of DDQ and dioxane (1% water) in the same pot, which leads to formation of flavone 11 in 65% yield (Figure 2).
- ⁴⁵ The formation of flavone 11 via one-pot protocol from phoroglucinol precursor 7c involves cascade of 4 reactions namely, Knoevenagel-type condensation, [4+2] Diels-Alder cycloaddition, DDQ-mediated oxidation and DDQ-mediated dehydrogenation. The plausible mechanism for this one-pot MCR
  ⁵⁰ is depicted in Figure 3. Under acidic conditions, formaldehyde gets protonated (structure I) which enhances its electrophilicity. 2,4-Disubstituted phloroglucinol being an active hydrogen species, it undergoes Knoevenagel-type condensation with protonated formaldehyde, producing transient *o*-quinone methide
  ⁵⁵ III. This *o*-quinone methide III quickly undergoes [4+2]-Diels-Alder cycloaddition reaction with dienophile 9a to produce flavan 10c. Further DDQ-mediated oxidation of the benzylic CH₂,

produces 2,3-dihydro flavone **VIII**. Here, activation of benzylic CH₂ of flavan **10c** occurs via hydride ion abstraction by DDQ.^{21, 22} 2,3-Dihydroflavone **VIII** further undergoes DDQ-mediated dehydrogenation leading to formation of flavone **11** (Figure 3).

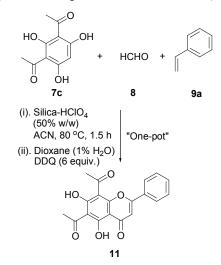


Figure 2 One-pot synthesis of flavone 11 from phloroglucinol 7c

#### Conclusion

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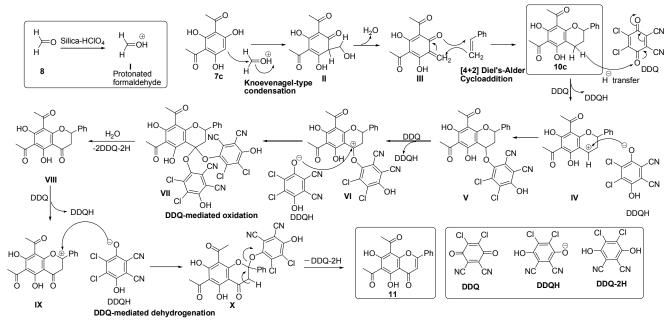
In summary, we have developed a very simple and economical tandem one-pot protocol for synthesis of flavans **10** and flavones **11** directly from substituted phloroglucinol precursors. Key View Online features of our developed methodology are (a). no protection/deprotection steps required; (b). inexpensive, easy to prepare, non-hazardous, easy to separate from reaction mixture, 15 reusable catalyst and (c). diversity-oriented synthesis. The developed protocol provides shorter route to access variety of flavan natural products such as catechins, flavonoids, anthocyanins, grandinal etc. using appropriate dienophile, aldehyde and phloroglucinol precursor.



#### **Experimental Section**

#### General

All chemicals were obtained from Sigma-Aldrich Company and used as received. ¹H, ¹³C and DEPT NMR spectra were recorded ²⁵ on Brucker-Avance DPX FT-NMR 500 and 400 MHz instruments. Chemical data for protons are reported in parts per million (ppm) downfield from tetramethylsilane and are referenced to the residual proton in the NMR solvent (CDCl₃, 7.26 ppm). Carbon nuclear magnetic resonance spectra (¹³C ³⁰ NMR) were recorded at 125 MHz or 100 MHz: chemical data for carbons are reported in parts per million (ppm, δ scale) downfield from tetramethylsilane and are referenced to the carbon resonance of the solvent (CDCl₃, 77 ppm). ESI-MS and HRMS spectra were recorded on Agilent 1100 LC-Q-TOF and HRMS-6540-UHD ³⁵ machines. IR spectra were recorded on Perkin-Elmer IR spectrophotometer. Melting points were recorded on digital melting point apparatus.



40 Figure 3 Plausible mechanism of one-pot multi-component synthesis of flavan 10c and flavone 11

**Procedures for preparation of silica catalysts.** (a). Silica-I₂:¹⁶ Commercial iodine (1 g) and silica gel (#230–400) (1 g) were taken in a culture tube and mixed thoroughly for 30 min. The ⁴⁵ prepared catalyst was kept in the closed tube. (b). Silica-FeCl₃:¹⁷ To a solution of FeCl₃ 6.H₂O (1.2 g) in acetone (20 mL) was

added silica gel (10 g, #230–400) at room temperature. The solvent was evaporated under the reduced pressure and the resulting yellow powder was kept in a closed container. (c). ⁵⁰ Silica-HClO₄.¹⁵ Perchloric acid (1.25 g, as a 70% aqueous solution) was added to the suspension of silica gel (23.75 g,

#230-400) in Et₂O. The mixture was concentrated and the residue heated at 100 °C for 72 h under vacuum to afford HClO₄-SiO₂ as a free flowing powder.

- Procedure for determination of acidity of Silica-HClO₄  $_{5}$  catalyst. Acidity of the catalyst before and after use was determined by CHEMBET-3000 TPD/TPR/TPO instrument, containing a quartz reactor and TCD detector. Prior to TPD studies, samples were pretreated at 250 °C for 2 h with continuous flow of pure nitrogen (99.9%), then cooled to room
- ¹⁰ temperature. After pre-treatment, samples were saturated with NH₃ gas until saturated adsorption. Temperature increased to 80 °C and kept there for 2 h, while a helium flow of 20 cm³/min to remove the physisorbed ammonia. Finally the system was heated from 80 °C to 1000 °C at the rate of 10 °C/min and desorbed gas
- ¹⁵ monitored with TCD detector. All the flow rates were maintained at normal temperature and pressure.

Procedure for preparation of 1-acetylphloroglucinol (7b). A solution of phloroglucinol (7a, 10 g, 79.36 mmol) and anhydrous aluminum chloride (21.7 g, 237.9 mmol) in carbon disulphide ²⁰ (100 mL) was stirred at room temperature for 20 min. Nitrobenzene (150 mL) was added and temperature of the reaction mixture was allowed to increase up to 50 °C. Acyl chloride (18.6 mL, 237.9 mmol) was added and reaction mixture was stirred further for 30 min. On cooling, reaction mixture was ²⁵ diluted with ethyl acetate. Water was added to the resultant mixture leading to formation of white precipitate in aqueous layer. Organic layer was decanted out and the remaining solid residue was washed 5-6 times with ethyl acetate. The combined ethyl acetate layer was evaporated under reduced pressure and the ³⁰ remaining viscous oil was purified by silica gel column

- chromatography using hexane-EtOAc as eluent to yield monoacetyl phloroglucinol **7b** (9.5 g). Yield: 72%; cream colored solid; mp. 134-136 °C; ¹H NMR (CD₃OD, 400 MHz):  $\delta$  5.80 (s, 2H), 2.60 (s, 3H); ESI-MS: *m/z* 169 [M+1]^{+.19}
- ³⁵ General procedure for preparation of diacylphloroglucinols
  7c and 7e. A solution of phloroglucinol (7a, 10 g, 79.36 mmol) and acetic acid or isovaleric acid (3 equiv.) in BF₃-etherate (100 mL) were refluxed at 100 °C for 2.5 h. Reaction mixture was cooled to room temperature, poured into crushed ice and
  ⁴⁰ extracted with ethyl acetate (100 mL x 3). Combined organic layers were evaporated on rotary evaporator. Crude product was purified by silica gel (#100-200) column chromatography to get
- diacyl phloroglucinols 7c (11.6 g) and 7e (17.2 g). 1,3-Diacetyl-2,4,6-trihydroxy benzene (7c): Yield: 70%; cream colored solid;
   45 mp. 172-174 °C; ¹H NMR (CD₃OD, 400 MHz): δ 5.84 (s, 1H),
- 2.65 (s, 6H); ESI-MS: m/z 211 [M+1]⁺. 1,3-Di-(3-methylbutanoyl)-2,4,6-trihydroxy benzene (7e): Yield: 75%; yellow solid; mp. 114-116 °C; ¹H NMR (CDCl₃, 200 MHz):  $\delta$  5.85 (s, 1H), 2.99 (d, J = 6.7 Hz, 4H), 2.26 (m, 2H), 0.99 (d, J = 6.7 Hz, 50 12H); ESI-MS: m/z 295 [M+1]^{+.23}
- **Procedure for preparation of 2,4-diformyl phloroglucinol** (7d). Phosphoryl chloride (1.6 mL, 16.7 mmol) was added drop-wise to DMF (1.3 mL, 16.7 mmol) with strong stirring, at room temperature under a nitrogen atmosphere. Stirring was continued
- ss for 30 minutes. This Vilsmeier reagent was then slowly added to a stirred solution of anhydrous phloroglucinol (7a, 1g, 7.9 mmol)

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in dioxane (5 mL) at room temperature, under a nitrogen atmosphere. This solution was then stirred at room temperature for 12 hours, whereupon it turned into a yellow amorphous solid.

⁶⁰ This solid mixture was cooled to 0 °C before being added to icewater slurry (~40 mL). The solution was allowed to slowly warm to room temperature and stirring was continued for a further 4 hours, during which time a cream precipitate formed. This precipitate was then filtered off and washed with more water, to ⁶⁵ get 2,4-diformyl phloroglucinol **7d** (1.22 g). Yield: 85%; cream colored solid; mp. 218-220 °C; ¹H NMR (CDCl₃, 400 MHz): δ 10.09 (s, 2H), 5.83 (s, 1H); ESI-MS: *m/z* 183 [M+1]⁺.²⁰

# General procedure for one-pot multi-component synthesis of flavans 10a-k. To a solution of substituted phloroglucinol (7, 100 ⁷⁰ mg) in acetonitrile were added formaldehyde (8, 3 equiv.),

- ⁷⁰ mg) in accontinue were added formatdenyde (**8**, 5 equiv.), substituted styrene (**9**, 1.5 mmol) and Silica-HClO₄ (50 %w/w). The mixture was then refluxed at 80 °C for 1-6 h. Completion of the reaction was monitored by TLC. After completion of reaction, reaction mixture was cooled to room temperature and was filtered ⁷⁵ through Whatman filter paper. Filtrate was concentrated on vacuo rotavapor to get crude product. Crude products were purified by silica gel (#100-200) column chromatography to get flavans **10ak** in 35-92% yield.
- **Tris-(2-phenyl-2,3-dihydro benzopyran) (10a**, Table 2, entry ⁸⁰ 1): 131 mg; Yield: 35%; white sticky solid; ¹H NMR (CDCl₃, 400 MHz): δ 7.46-7.31 (m, 15H), 5.07-5.00 (m, 3H), 2.85 (m, 3H), 2.74 (m, 3H), 2.23 (m, 3H), 2.05 (m, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 151.28, 151.18, 142.39, 142.32, 142.29, 128.38, 127.51, 125.79, 125.76, 102.17, 102.15, 77.42, 29.81, 29.79, 29.67, ⁸⁵ 19.45, 19.44, 19.36; IR (CHCl₃):  $v_{max}$  3451, 3030, 3064, 2925, 2852, 1732, 1614, 1496, 1443, 1310, 1218, 1126, 1085 cm⁻¹; ESI-MS: m/z 475 [M+1]⁺; HRMS: m/z 475.2266 calcd for C₃₃H₃₀O₃+H⁺ (475.2268).

8-Acetyl-bis-(2-phenyl-2,3-dihydro benzopyran) (10b, Table 2, 90 entry 2): 123 mg; Yield: 52%; white sticky mass; ¹H NMR (CDCl₃, 500 MHz):  $\delta$  14.23 (s, 1H), 7.39 (m, 10H), 5.09 (m, 2H), 2.80-2.69 (m, 4H), 2.57 (s, 3H), 2.32 (m, 2H), 2.02 (m, 2H); ¹³C NMR (CDCl₃, 125 MHz):  $\delta$  203.44, 162.37, 159.08, 155.99, 141.22, 141.12, 128.60, 128.55, 126.02, 125.71, 105.25, 102.20, 95 101.14, 78.78, 78.72, 33.47, 29.31, 29.12, 19.27, 18.59; IR (CHCl₃):  $v_{max}$  3450, 2927, 1615, 1424, 1369, 1276, 1131, 1105 cm⁻¹; ESI-MS: *m/z* 401 [M+1]⁺; HRMS: *m/z* 401.1732 calcd for C₂₆H₂₄O₄+H⁺ (401.1753).

**6,8-Diacetyl-5,7-dihydroxyflavan (10c**, Table 2, entry 3): 130 mg; Yield: 84%; white crystalline solid; m.p. 101-103 °C; ¹H NMR (CDCl₃, 500 MHz):  $\delta$  16.15 (s, 1H), 15.18 (s, 1H), 7.40 (m, 5H), 5.15 (dd, J = 2.3, 10.4 Hz, 1H), 2.76 (m, 1H), 2.72 (s, 3H), 2.66 (m, 1H), 2.53 (s, 3H), 2.24 (m, 1H), 2.02 (m, 1H); ¹³C NMR (CDCl₃, 125 MHz):  $\delta$  204.5, 203.4, 171.6, 170.9, 162.6, 140.0, ¹⁰⁵ 128.8, 128.5, 127.8, 125.8, 104.6, 103.9, 101.1, 79.99, 33.34, 33.10, 28.54, 18.49; IR (CHCl₃): v_{max} 3400, 2924, 1615, 1423, 1364, 1169, 1105, 1024 cm⁻¹; ESI-MS: *m/z* 327 [M+1]⁺; HRMS: *m/z* 349.1045 calcd for C₁₉H₁₈O₅+Na⁺ (349.1015).

**6,8-Diformyl-5,7-dihydroxyflavan (10d**, Table 2, entry 4): 124 ¹¹⁰ mg; Yield: 76%; light yellow solid; m.p. 236-238 °C; ¹H NMR (CDCl₃, 500 MHz):  $\delta$  13.51 (s, 1H), 13.30 (s, 1H), 10.19 (s, 1H), 10.07 (s, 1H), 7.39 (m, 5H), 5.22 (dd, J = 6.0, 15 Hz, 1H), 2.76

(m, 1H), 2.66 (m, 1H), 2.31 (m, 1H), 2.04 (m, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 192.0, 191.9, 169.0, 168.0, 164.0, 139.6, 128.8, 128.5, 125.8, 104.1, 103.9, 101.2, 79.7, 28.3, 17.6; IR (CHCl₃):  $v_{max}$  2924, 1640, 1442, 1305, 1154 cm⁻¹; ESI-MS: m/z $5299 [M+1]^+$ ; HRMS: m/z 299.0888 calcd for  $C_{17}H_{14}O_5+H^+$ (299.0918).

6,8-Di-(3-methylbutyryl)-5,7-dihydroxyflavan (10e, Table 2, entry 5): 119 mg; Yield: 86%; light yellow solid; m.p. 117-119 °C; ¹H NMR (CDCl₃, 400 MHz):  $\delta$  16.39 (s, 1H), 15.37 (s, 1H), ¹⁰ 7.42 (m, 5H), 5.09 (dd, J = 2.3, 10.7 Hz, 1H), 3.03 (d, J = 6.8 Hz, 2H), 2.82 (d, J = 6.7 Hz, 2H), 2.62 (m, 2H), 2.28 (m, 2H), 2.04 (m, 2H), 1.00 (d, J = 6.7 Hz, 6H), 0.72 (m, 6H);  13 C NMR (CDCl₃, 100 MHz): δ 207.0, 206,0, 170.1, 169.7, 162.3, 139.8, 128.7, 128.6, 126.4, 104.7, 103.8, 101.1, 80.2, 53.1, 53.0, 28.2, 15 25.6, 25.1, 22.8, 22.7, 22.4, 22.3, 18.8; IR (CHCl₃): v_{max} 3400, 2956, 2926, 1614, 1417, 1294, 1192, 1161, 1023 cm⁻¹; ESI-MS: m/z 411 [M+1]⁺; HRMS: m/z 411.2139 calcd for C₂₅H₃₀O₅+H⁺ (411.2166).

6,8-Diacetyl-5,7-dihydroxy-4'-methyl-flavan (10f, Table 2, 20 entry 6): 148 mg; Yield: 92%; white crystalline solid; m.p. 157-159 °C; ¹H NMR (CDCl₃, 400 MHz):  $\delta$  16.15 (s, 1H), 15.16 (s, 1H), 7.29-7.20 (m, 4H), 5.13 (dd, J = 2.0, 10.4 Hz, 1H), 2.81 (m, 1H), 2.78 (m, 1H), 2.64 (m, 1H), 2.52 (s, 3H), 2.40 (s, 3H), 2.26 (m, 1H), 2.07 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz):  $\delta$  204.5, 25 203.5, 170.0, 169.5, 162.7, 138.3, 137.0, 129.4, 126.1, 104.5, 103.9, 101.1, 79.9, 33.3, 33.1, 29.7, 28.5, 21.2, 18.6; IR (CHCl₃):  $v_{max}$  3400, 2921, 2851, 1620, 1591, 1422, 1365, 1169, 1109 cm⁻¹; ESI-MS: *m/z* 341 [M+1]⁺, 363 [M+Na]⁺; HRMS: *m/z* 341.1385 calcd for  $C_{20}H_{20}O_5+H^+$  (341.1389).

30 6,8-Diformyl-5,7-dihydroxy-4'-methyl-flavan (10g, Table 2, entry 7): 149 mg; Yield: 87%; white crystalline solid; m.p. 115-117 °C; ¹H NMR (CDCl₃, 400 MHz):  $\delta$  13.49 (s, 1H), 13.30 (s, 1H), 10.19 (s, 1H), 10.06 (s, 1H), 7.28-7.22 (m, 4H), 5.18 (d, J = 10 Hz, 1H), 2.76 (m, 1H), 2.64 (m, 1H), 2.40 (s, 3H), 2.30 (m, ³⁵ 1H), 2.04 (m, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 192.0, 191.8, 168.9, 168.0, 164.1, 138.4, 136.6, 129.4, 129.1, 125.8, 125.7, 104.0, 103.9, 101.2, 79.7, 28.2, 21.2, 17.6; IR (CHCl₃): v_{max} 2924, 1643, 1443, 1306, 1272, 1182, 1154 cm⁻¹; ESI-MS: m/z 313  $[M+1]^+$ , 335  $[M+Na]^+$ ; HRMS: m/z 313.1072 calcd for  $_{40} C_{18}H_{16}O_5 + H^+ (313.1076).$ 

#### 6.8-Di-(3-methyl-butyryl)-5,7-dihydroxy-4'-methyl-flavan

(10h, Table 2, entry 8): 132 mg; Yield: 92%; light greyish crystalline solid; m.p. 100-102 °C; ¹H NMR (CDCl₃, 400 MHz):  $\delta$  16.38 (s, 1H), 15.36 (s, 1H), 7.30 (d, J = 7.7 Hz, 2H), 7.23 (d, J⁴⁵ = 7.7 Hz, 2H), 5.07 (d, *J* = 10.5 Hz, 1H), 3.03 (d, *J* = 6.7 Hz, 2H), 2.84 (m, 2H), 2.68 (m, 2H), 2.38 (s, 3H), 2.16 (m, 2H), 2.06 (m, 2H), 0.99 (d, J = 6.2 Hz, 6H), 0.74 (d, J = 6.3 Hz, 3H), 0.70 (d, J = 6.6 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz):  $\delta$  206.9, 206.0, 170.1, 169.6, 162.3, 138.4, 136.8, 129.3, 126.3, 104.6, 103.8, 50 101.1, 80.0, 53.1, 53.0, 28.2, 25.6, 25.0, 22.4, 22.3, 21.2, 18.8; IR (CHCl₃): v_{max} 2957, 2929, 2870, 1614, 1435, 1367, 1294, 1193, 1161, 1126 cm⁻¹; ESI-MS: m/z 425 [M+1]⁺, 447 [M+Na]⁺; HRMS: m/z 425.2323 calcd for C₂₆H₃₂O₅+H⁺ (425.2323).

6,8-Diacetyl-5,7-dihydroxy-4'-tert-butyl-flavan (10i, Table 2, 55 entry 9): 170 mg; Yield: 92%; light yellow crystalline solid; m.p. 119-121 °C; ¹H NMR (CDCl₃, 400 MHz): δ 16.15 (s, 1H), 15.16 View Online

(s, 1H), 7.44 (d, J = 6.8 Hz, 2H), 7.32 (d, J = 6.8 Hz, 2H), 5.16 (dd, J = 2.0, 10.0 Hz, 1H), 2.80 (m, 1H), 2.76 (s, 3H), 2.64 (m, 1H))1H), 2.52 (s, 3H), 2.27 (m, 1H), 2.08 (m, 1H), 1.35 (s, 9H); ¹³C 60 NMR (CDCl₃, 125 MHz): δ 204.4, 203.5, 170.0, 169.5, 162.7, 151.4, 136.9, 125.7, 125.6, 104.5, 103.9, 101.1, 79.8, 34.6, 33.4, 33.0, 31.3, 28.5, 18.5; IR (CHCl₃): v_{max} 3401, 2961, 1615, 1424, 1364, 1292, 1169, 1104 cm⁻¹; ESI-MS: *m/z* 383 [M+1]⁺, 405  $[M+Na]^+$ ; HRMS: m/z 383.1882 calcd for  $C_{23}H_{26}O_5+H^+$ 65 (383.1853).

#### 6,8-Diacetyl-3,4-dihydro-2-(naphthalen-1-yl)-2H-chromene-

5,7-diol (10j, Table 2, entry 10): 164 mg; Yield: 92%; white crystalline solid; m.p. 161-163 °C; ¹H NMR (CDCl₃, 500 MHz):  $\delta$  16.20 (s, 1H), 15.22 (s, 1H), 8.21 (d, J = 8.0 Hz, 1H), 7.94 (d, J  $_{70} = 6.6$  Hz, 1H), 7.90 (d, J = 8.2 Hz, 1H), 7.65 (d, J = 7.0 Hz, 1H), 7.58 (m, 3H), 5.91 (d, J = 10.4 Hz, 1H), 2.44 (m, 1H), 2.38 (m, 1H), 2.36 (s, 3H), 2.22 (s, 3H), 2.14 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz): δ 204.6, 203.5, 170.1, 169.6, 162.8, 135.2, 133.9,

130.3, 129.2, 129.1, 126.6, 125.9, 125.4, 123.6, 122.7, 104.7, 75 104.0, 101.3, 77.2, 33.2, 33.1, 27.5, 18.9; IR (CHCl₃): v_{max} 3400, 2923, 2852, 1615, 1423, 1364, 1293, 1172, 1105 cm⁻¹; ESI-MS: m/z 377  $[M+1]^+$ , 399  $[M+Na]^+$ ; HRMS: m/z 377.1384 calcd for  $C_{23}H_{20}O_5 + H^+$  (377.1384).

#### 6,8-Diacetyl-3,4-dihydro-2-(3,4-methylene-dioxy-phenyl)-3-

- 80 methyl-2H-chromene-5,7-diol (10k, Table 2, entry 11): 160 mg; Yield: 88%; white sticky solid; ¹H NMR (CDCl₃, 500 MHz):  $\delta$ 16.12 (s, 1H), 15.16 (s, 1H), 6.85 (m, 3H), 6.01 (s, 2H), 4.59 (d, J = 9.9 Hz, 1H), 2.93 (dd, J = 5.0, 14.5 Hz, 1H), 2.73 (s, 3H), 2.45 (s, 3H), 2.25 (dd, J = 11.2, 16.5 Hz, 1H), 2.08 (m, 1H), 0.89 (d, J
- $_{85} = 6.6$  Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz):  $\delta$  204.5 (-CO), 203.5 (-CO), 170.0 (C7), 169.2 (C5), 162.4 (C8a), 148.1 (C4'), 147.9 (C4'), 147.8 (C5'), 133.5 (C5'), 133.5 (C1'), 133.3 (C1'), 121.2 (C6'), 121.1 (C6'), 108.2 (C2'), 108.0 (C2'), 107.6 (C3'), 107.0 (C4a), 104.5 (C6), 103.7 (C8), 101.3 (O-CH₂-O), 101.2
- 90 (O-CH₂-O), 86.0 (C2), 85.8 (C2), 33.2 (CO<u>C</u>H₃), 33.1 (CO<u>C</u>H₃), 32.0 (C3), 27.2 (C4), 17.5 (CH-CH₃); IR (CHCl₃): v_{max} 2927, 1618, 1505, 1445, 1425, 1381, 1364, 1247, 1175, 1039 cm⁻¹; ESI-MS: m/z 385  $[M+1]^+$ ; HRMS: m/z 385.1255 calcd for C₂₁H₂₀O₇+H⁺ (385.1282).
- 95 Synthesis of flavone 11 from flavan 10c. To the solution of 5,7dihydroxy-6,8-diacetyl-flavan (10c, 100 mg, 0.306 mmol) in dioxane (1% H₂O) was added DDQ (0.278 mg, 1.2 mmol). Resulting mixture was refluxed at 100 °C for 16 h. Reaction mixture was allowed to cool to room temperature and then
- 100 filtered through Whatman filter paper. Filtrate was concentrated on vacuo rotavapor to get crude product, which on silica gel (#100-200) column chromatography gave flavone 11 (78 mg) in 75% yield. 6,8-Diacetyl-5,7-dihydroxyflavone (11, Figure 2): White sticky solid; ¹H NMR (CDCl₃, 400 MHz):  $\delta$  16.04 (s, 1H),
- ¹⁰⁵ 15.72 (s, 1H), 7.97 (d, J = 8.1 Hz, 2H), 7.67 (m, 2H), 7.64 (t, J = 7.8 Hz, 2H), 2.94 (s, 3H), 2.82 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 205.9, 201.3, 182.6, 171.4, 167.9, 159.5, 151.4, 137.0, 133.0, 129.1, 128.7, 115.5, 110.9, 107.0, 100.6, 32.8, 30.9; IR (CHCl₃): v_{max} 3400, 2923, 2852, 1614, 1611, 1455, 1372, 1304,
- 110 1172 cm⁻¹; ESI-MS: m/z 339 [M+1]⁺, 361 [M+Na]⁺; HRMS: m/z339.0878 calcd for  $C_{19}H_{14}O_6+H^+$  (339.0863).

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**Procedure for one-pot synthesis of 6,8-diacetyl-5,7dihydroxyflavone (11) from phloroglucinol precursor 7.** To the solution of 2,4-diacetylphloroglucinol (7c, 100 mg, 0.476 mmol) in acetonitrile were added formaldehyde (8, 3 equiv.), s styrene (9a, 0.074 g, 0.714 mmol) and Silica-HClO₄ (0.05 g, 50 %w/w). The mixture was then refluxed at 80 °C for 1.5 h.

- % w/w). The mixture was then refluxed at 80 °C for 1.5 n. Completion of the reaction was monitored by TLC. After completion of reaction, acetonitrile was evaporated on vacuo rotavapor to dryness. Dioxane (5 mL), water (10  $\mu$ L, 5% of
- ¹⁰ dioxane) and DDQ (0.649 g, 2.856 mmol) was then added to the dry reaction mixture and was refluxed at 100 °C for 16 h. Reaction mixture was allowed to cool to room temperature and then filtered through whatman filter paper. Filtrate was concentrated on vacuo rotavapor to get crude product, which on 15 silica gel (#100-200) column chromatography gave flavone 11
- (104 mg) in 65% yield.

**Recyclability studies of silica-HCIO₄ catalyst**. To a solution of 2,4-diacetylphloroglucinol (7c, 500 mg, 2.38 mmol) in acetonitrile (10 mL) were added formaldehyde (**8**, 3 equiv.), ²⁰ styrene (**9**, 3.57 mmol) and Silica-HCIO₄ (250 mg, 50 %w/w). The mixture was then refluxed at 80 °C for 1.5 h. After completion of reaction, catalyst was recovered by filtration followed by washing with acetonitrile. Recovered catalyst was recycled in oven and reused in next cycle. The catalyst was recycled ²⁵ 3 times and the amount of catalyst recovered and percentage yield

25 3 times and the amount of catalyst recovered and percentage yield of the flavan **10c** were determined.

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