

Synthesis, crystal structure, and anticancer properties of cyclic monocarbonyl analogs of curcumin

Natesan Sundarmurthy Karthikeyan ·
Kulathu Iyer Sathiyarayanan · Paduthapillai Gopal Aravindan ·
P. Giridharan

Received: 12 May 2009 / Accepted: 4 December 2009 / Published online: 25 December 2009
© Springer Science+Business Media, LLC 2010

Abstract A series of novel indan-2-one and dibenzylindene-1,3-dione compounds were synthesized and screened for anticancer activities. The compounds are symmetrical and they have conjugated double bonds. They closely resemble the curcumin analogs which are found to possess anticancer properties. The structure of the compounds was confirmed by single crystal study and wherever the compound is a powder, the structures were confirmed by spectral data (IR, NMR, and Mass).

Keywords 2-Indanone · 1-Benzyl-4-piperidone · Aromatic aldehyde · Curcumin derivatives · Molecular docking

Introduction

Curcumin, a compound isolated from turmeric—the “Golden spice,” possesses many biological properties. It is a traditional medicine for liver diseases, indigestion, rheumatic arthritis, and insect bite in India. Owing to its anticancer and antiangiogenic properties, low molecular weight and lack of toxicity, curcumin could be an ideal candidate for a chemotherapeutic agent (Adams *et al.*,

2004; Aggarwal *et al.*, 2003; Zeng *et al.*, 2009). In the last few years, a number of natural and synthetic compounds have proven to inhibit HIV-1 IN in *in vitro* assays, but have failed to provide antiviral efficacy in HIV-1 infected cells. Curcumin derivatives are integrase inhibitors *in vitro* endowed with antiviral activity in cell-based assays (Santo *et al.*, 2005). Curcumin is capable of inducing apoptosis in various cancer cell lines (Karunagaran *et al.*, 2005; Lakshmi *et al.*, 2008) and can suppress cancer formation (Li *et al.*, 2002). Both phase I and II clinical trials show that curcumin is well tolerated and is effective for cancer patients, (Dhillon *et al.*, 2006) suggesting curcumin as a potential drug in cancer prevention and therapy (Dube *et al.*, 2008; Chandru *et al.*, 2008). Further, it also enhances wound healing (Panchatcharam *et al.*, 2006). Curcumin is nontoxic even at high dosage. It has been classified as “generally recognized as safe” (GRAS) by the National Cancer Institute (Osawa *et al.*, 1995).

The Curcumin analogs (1,5-diaryl-3-oxo-1,4-pentadienyl group) have been mounted on a variety of cyclic scaffolds leading to the discovery of a number of potent cytotoxins (Pati *et al.*, 2009; Fadda *et al.*, 2009). This group is considered to react at a primary binding site (Fig. 1a). However, the magnitude of the bioactivity observed will be influenced by the presence of other structural units in the molecule, which align at an auxiliary site (Fig. 1b).

Several studies have revealed that compounds in which R is an acyl group possess increased cytotoxic potencies compared to the analogs when R is a hydrogen atom (Das *et al.*, 2007). Thus, by expanding the size of the molecules, there is a possibility of additional binding of the ligand at a receptor which results in the lowering of the IC₅₀ values (Pati *et al.*, 2009). In curcumin analogs, the substitution pattern of the aromatic moiety is of importance. The most active compounds have a fluorine atom or a methoxyl

N. S. Karthikeyan · K. I. Sathiyarayanan (✉)
Chemistry Division, School of Advanced Sciences, VIT
University, Vellore 632 014, India
e-mail: sathiya_kuna@hotmail.com

P. G. Aravindan
Physics Division, School of Advanced Sciences, VIT University,
Vellore 632 014, India

P. Giridharan
Piramal Life Sciences Ltd., Mumbai 400 063, India

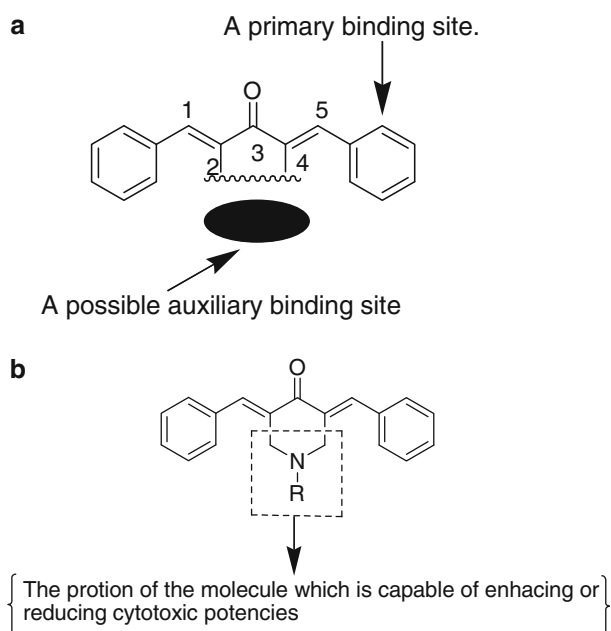


Fig. 1 a Primary binding sites. b A possible auxiliary binding site

group in the *ortho* position of the OH-group of the *p*-hydroxyphenyl moiety (Gafner *et al.*, 2004). *Ortho*-substituents consistently exhibit better activity than their *meta*- and *para*-counterparts (Adams *et al.*, 2004).

Experimental

Chemistry

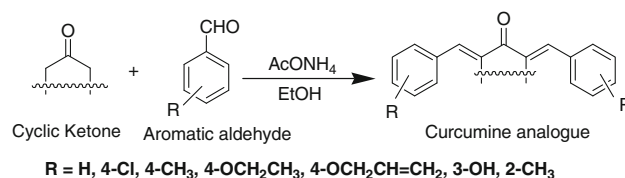
The synthesis and biological evaluation of curcumin analogs have been reported previously. In this study, the authors wish to report a new synthetic method (Scheme 1), using ammonium acetate, for monocarbonyl curcumin analogs (symmetrical aromatic moieties) with more stable chemical structures, as well as their anticancer screening in various cell lines.

This procedure is more convenient than the literature method (Adams *et al.*, 2004; Pati *et al.*, 2008, 2009; Liang *et al.*, 2008a), easy to synthesis, isolate, and less expensive.

A mixture of cyclic ketone (0.01 mol) and the appropriate aldehyde (0.02 mol) in alcoholic ammonium acetate (15 ml, 0.01 mol) was gently heated and left overnight at room temperature. The separated solid was purified by column chromatography.

Biology

The two series (IND—Fig. 2 and BP—Fig. 3) of eight monocarbonyl curcumin analogs were synthesized and screened for anticancer activity. Recently, antibacterial activities of monocarbonyl analogs of curcumin have also been discussed (Liang *et al.*, 2008b). To the best of our knowledge, this is the first time an investigation has been made for the anticancer activity of IND series (Fig. 2). The anticancer screening was performed using cell lines, HCT-116 (Colon cancer), Panc-1 (Pancreatic cancer), H460 (Non small lung cancer), Calu-1 (Lung cancer), ACHN (Renal cancer). The results compare well with W138 (Normal lung fibroblast cells).



Scheme 1 General synthesis of curcumin analogs (1–8)

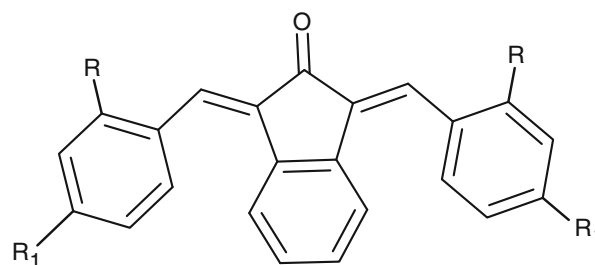


Fig. 2 Compound 1–3

Fig. 3 Compound 4–8

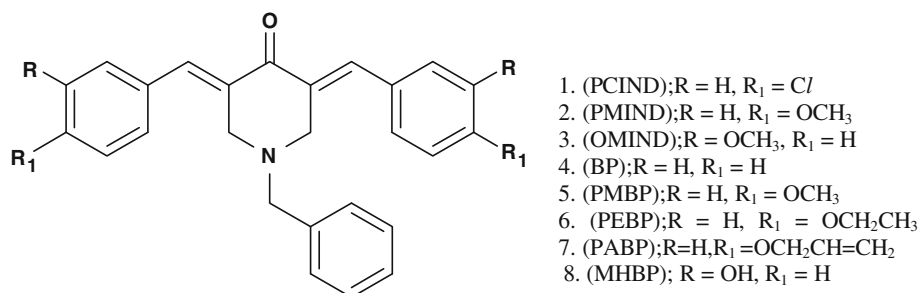


Chart 1 Cell mortality for OMIND, BP, and PMBP in various cell lines at different concentration (μM)

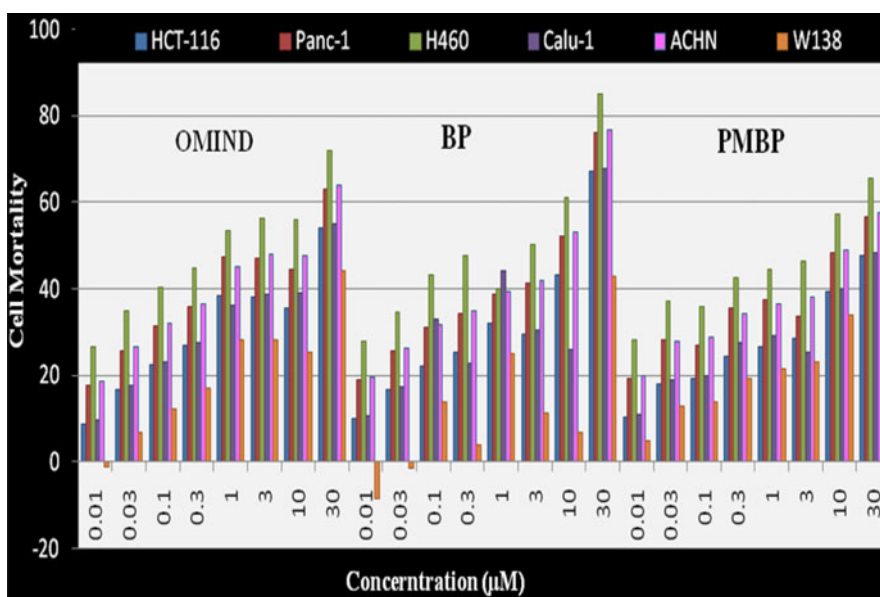
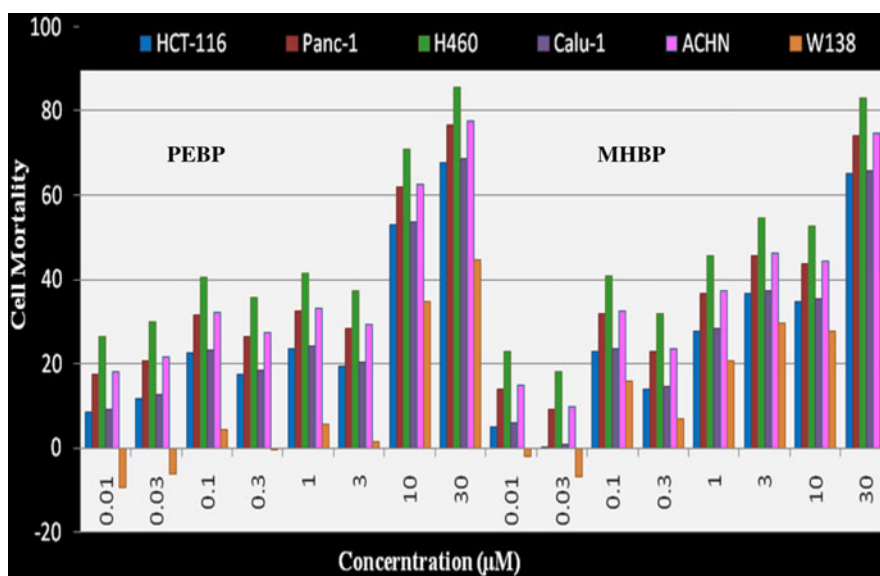


Chart 2 Cell mortality for PEBP and MHBP in various cell lines at different concentration (μM)



Results and discussion

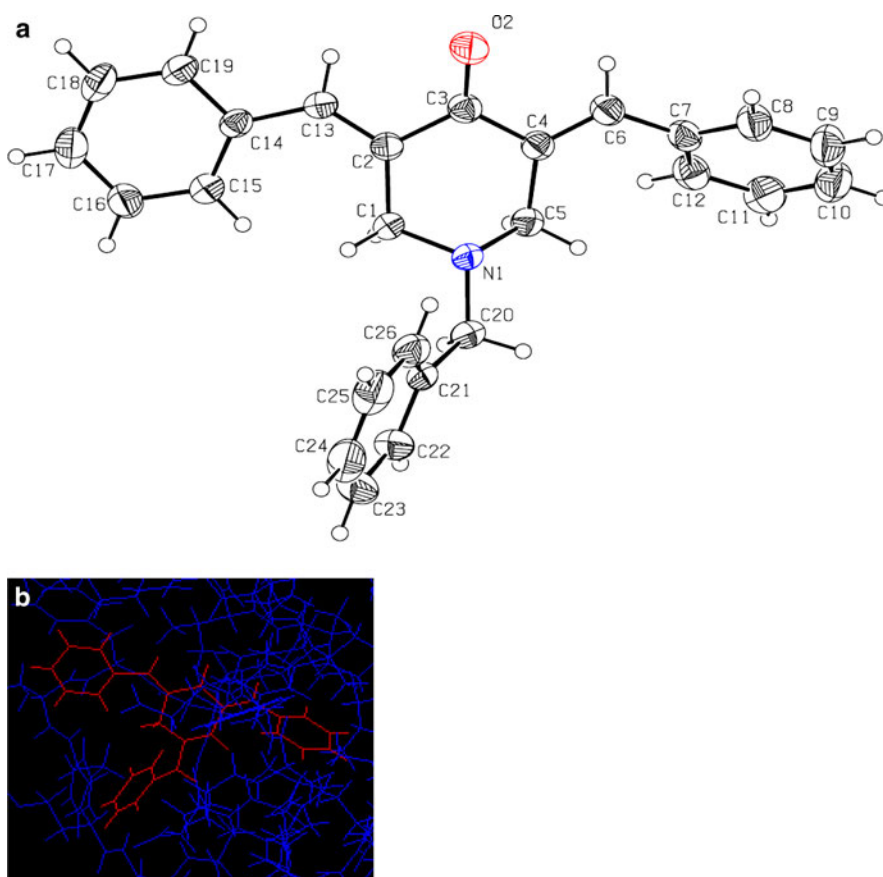
Chemistry

The structures of curcumin analogs (1–8) were in accordance with their crystal data and spectroscopic data. The FT-IR spectrum of the compounds in general exhibited the absorption band at around $1705\text{--}1569\text{ cm}^{-1}$ indicating the carbonyl group and olefinic bonds of α,α' -(EE)-bis(substituted benzylidene)cycloalkanones are in conjugation. The mass spectrum of the compounds showed its respective peaks. In the $^1\text{H-NMR}$ spectrum of the compounds, the vinylic protons either occur as singlet at around δ 7.61–7.49 ppm or as merged ones with the multiplet of aromatic protons ranging from δ 8.25 to 6.96 ppm.

Biology

The CCK-8 nonradioactive colorimetric assay was carried out (Jing-yan *et al.*, 2006; Cell Counting Kit-8 technical manual) to characterize the in vitro growth of human tumor cell lines as well as to test the cytotoxic activity of these compounds. The compound PEBP showed the highest anticancer activity among the compounds tested, and it showed IC_{50} of $8.3\text{ }\mu\text{M}$ in Colon cancer, $7.2\text{ }\mu\text{M}$ in Pancreatic cancer, $5.5\text{ }\mu\text{M}$ for Non small lung cancer, $8.1\text{ }\mu\text{M}$ for Lung cancer, and $6.9\text{ }\mu\text{M}$ for Renal cancer cells (Chart 2). The activity was tested in the concentration ranges of $0.01\text{--}30\text{ }\mu\text{M}$. The activity was found to increase with increase in concentration. Fortunately, none of the compounds is found to be toxic in normal primary lung

Fig. 4 **a** ORTEP diagram of BP. **b** Docking of receptor model—1m3g with BP



fibroblast cells. If we see the order of activity, the order is roughly as follows OMIND < PMBP < BP < MHBP < PEBP (Charts 1, 2). Out of the two series, BP derivatives (Fig. 3) are more active than IND series (Fig. 2). This may be due to the reason that the auxiliary binding site in BP is capable of enhancing the activity. However, in the case of IND series, the auxiliary binding site is not that capable of increasing the activity. This is clear from the Fig. 1a and b. This gets support from the earlier observation in which some BP compounds were reported for selective toxicity for malignant cells (Pati *et al.*, 2008). However, the activity of BP series is moderately affected by the substitution present in it. Within the BP series, the highest activity is shown by PEBP followed by MHBP, BP, and PMBP (Charts 1, 2). This order may be due to the reason that, out of ethoxy and methoxy, ethoxy is more active than the methoxy due to the large chain of ethyl group. The moderately good activity of MHBP is due to the reason that OH derivatives normally increase the anticancer activity.

In the case of Colon cancer, the order follows OMIND < MHBP < BP < PEBP < PMBP. Here, again the ethyl group in the substitution plays a major role. With Pancreatic cancer, the order follows OMIND < MHBP < PMBP < BP < PEBP. Here also the ethyl substitution is responsible for the highest activity. However, in the case of

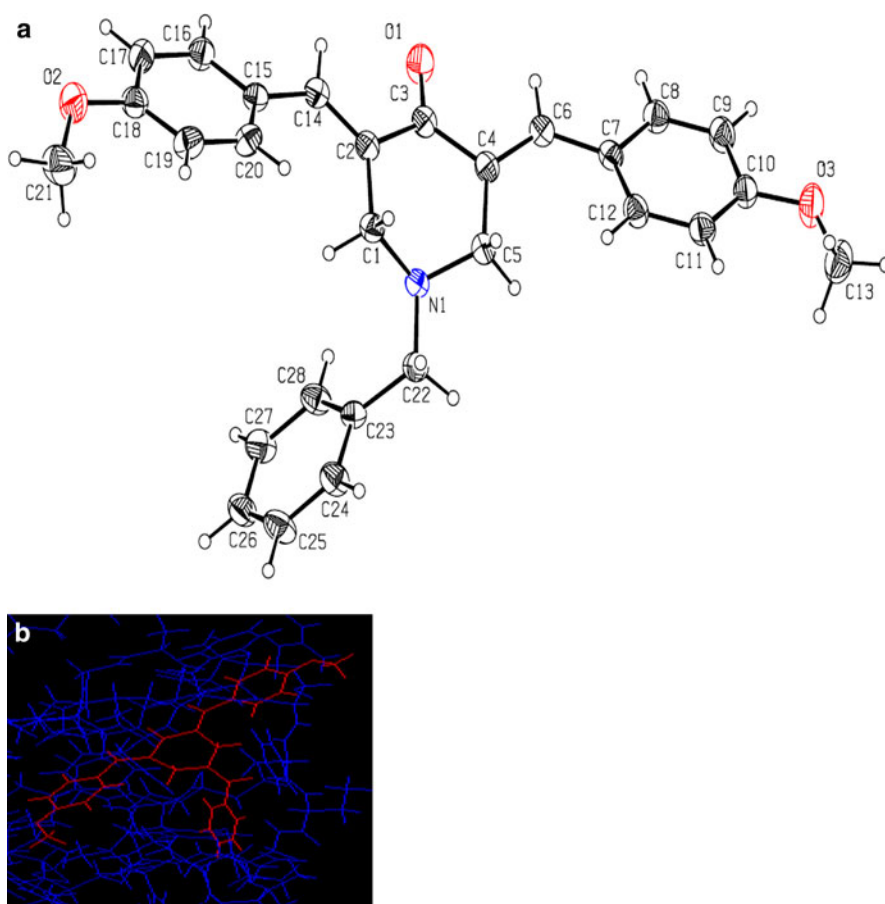
Non small lung cancer, the order of the activity is changed completely, and here, the most active one is OMIND and the least active is PEBP. The order follows PEBP < PMBP < BP < MHBP < OMIND. Here, the activity of all the compounds increases tremendously, when compared to other cancers. In the case of Lung cancer, again the order of activity resembles the order followed in Colon cancer PMBP < OMIND < MHBP $\approx$ BP < PEBP. The activity here is not as pronounced as in the case of Non small lung cancer. In the case of Renal cancer, the activity follows the order PMBP < OMIND < MHBP < BP < PEBP.

From the above observations, the PEBP (Chart 2) is found to be more active than all other compounds.

Molecular modeling

Before the compounds were tested in in vitro, molecular modeling studies were performed. Docking studies were undertaken by means of the patch dock program (Patch-Dock Server 2008), with the aim of better understanding the binding mode of the curcumin analogs to the biological target. We got crystalline structure for only two compounds, namely, BP (Fig. 4a) and PMBP (Fig. 5a), and these two were subjected to docking studies.

Fig. 5 **a** ORTEP diagram of PM. **b** Docking of receptor model—1m3g with PMBP



Tyrosine phosphorylation of proteins is required for signal transduction in cells and for growth regulation. A mitogen-induced gene (PAC-1) has been cloned from human T cells and encodes a 32-kDa protein that contains a sequence that defines the enzymatic site of known protein phosphotyrosine phosphatases (PTPases). We subjected these two compounds for molecular modeling studies against catalytic domain of MAPK protein phosphatase (PAC-1) (residue 170–314) (Wu *et al.*, 2007; RCSB Protein Data Bank—Receptor 1m3g).

Initial data reveal that the compound binds to PAC-1 and modulates its expression (Figs. 4b, 5b). The PAC-1 is a direct target of E2F-1, which plays a main role in cell cycle regulation. The enhanced activity of this compound in cancer cells may be due to PAC-1 modulation in cancer cells.

Conclusion

The above discussion reveals that compounds in the BP (Fig. 2) and IND series (Fig. 3) have anticancer properties. These heterocycles could be considered as useful templates for future development and further derivatization to obtain more potent and selective anticancer agents. Substantial

structural modifications of curcumin have an important potential for drug development.

Spectral data

1,3-Bis(4-chlorobenzylidene)-indan-2-one (PCIND)

Yellow amorphous powder; mp 182°C; 83% Yield; IR (KBr) cm^{-1} 2918, 1713, 1623, 776; $^1\text{H-NMR}$ (DMSO-d_6 , 400 MHz): δ 7.27–7.74 (14H, olefinic protons and ArH); GC–MS: m/z = 376 ($\text{M} - 1$) $^+$.

1,3-Bis(4-methoxybenzylidene)-indan-2-one (PMIND)

Yellow crystal; mp 156–160°C; 72% Yield; IR (KBr) cm^{-1} : 2958, 2838, 1705, 1593; $^1\text{H-NMR}$ (CDCl_3 , 300 MHz): δ = 3.89 (6H, s, OCH_3), 6.96–8.29 (14H, olefinic protons and ArH); $^{13}\text{C NMR}$ (CDCl_3 , 500 MHz): δ = 55.49, 113.71, 114.07, 114.12, 119.52, 123.10, 123.16, 127.65, 127.74, 127.87, 128.60, 129.07, 130.05, 131.04, 131.09, 133.04, 133.31, 133.71, 134.03, 134.84, 140.82, 160.44, 160.60, 161.38, 192.76. LC–MS: m/z = 369 ($\text{M} + 1$) $^+$.

1,3-Bis(2-methoxybenzylidene)-indan-2-one (OMIND)

Yellow amorphous powder; mp 108–112°C; 78% Yield; IR (KBr) cm^{-1} 3070, 2933, 2836, 1708, 1596; ^1H -NMR (DMSO- d_6 , 400 MHz): δ 3.84 (6H, s, OCH_3), 6.98–7.80 (14H, olefinic protons and ArH); ^{13}C NMR (CDCl_3 , 500 MHz): δ = 55.58, 55.69, 110.24, 110.96, 111.01, 120.23, 123.48, 124.37, 128.56, 129.46, 129.58, 130.99, 158.13, 158.16, 194.41. GC–MS: m/z = 368 (M^+).

1-Benzyl-3,5-dibenzylidenepiperidin-4-one (BP)

Yellow block Crystal; mp 152–154°C; 97% Yield; IR (KBr) cm^{-1} 3061, 3019, 2798, 2755, 1671, 1569; ^1H -NMR (DMSO- d_6 , 300 MHz): δ 3.69 (2H, s), 3.81 (4H, s), 7.18–7.42 (15H, ArH), 7.61 (2H, s, olefinic protons); GC–MS: m/z = 365 (M^+).

3,5-Bis(4-methoxybenzylidene)-1-benzylpiperidin-4-one (PMBP)

Yellow block Crystal; mp 150–154°C; 83% Yield; IR (KBr) cm^{-1} 3063, 2935, 2836, 2743, 1668, 1577; ^1H -NMR (DMSO- d_6 , 300 MHz): δ 3.71 (2H, s), 3.78 (6H, s, OCH_3), 3.85 (4H, s), 6.97–7.89 (15H, olefinic protons and ArH); LC–MS: m/z = 426 ($\text{M} + 1$) $^+$.

3,5-Bis(4-ethoxybenzylidene)-1-benzylpiperidin-4-one (PEBP)

Yellow needle Crystal; mp 110–114°C; 81% Yield; IR (KBr) cm^{-1} 3062, 3031, 2764, 1670, 1577; ^1H -NMR (DMSO- d_6 , 300 MHz): δ 1.29–1.33 (6H, t, CH_2CH_3) 3.70 (2H, s), 3.77 (4H, s), 4.01–4.08 (4H, q, OCH_2) 6.94–7.39 (13H, ArH) 7.54 (2H, s, olefinic protons); LC–MS: m/z = 454 ($\text{M} + 1$) $^+$.

3,5-Bis(4-allyloxybenzylidene)-1-benzylpiperidin-4-one (PABP)

Yellow Crystal; mp 114–116°C; 91% Yield; IR (KBr) cm^{-1} 3063, 3028, 2887, 2743, 1670, 1600, 1577; ^1H -NMR (DMSO- d_6 , 300 MHz): δ 3.70 (2H, s), 3.78 (4H, s), 4.59–4.61 (4H, d, OCH_2) 5.24–5.41 (4H, CH_2), 5.98–6.04 (2H, CH), 6.94–7.40 (13H, ArH) 7.54 (2H, s, olefinic protons); MS: m/z = 478 ($\text{M} + 1$) $^+$.

3,5-Bis(3-hydroxybenzylidene)-1-benzylpiperidin-4-one (MHBP)

Pale Yellow Crystal; mp 180–184°C; 83% Yield; IR (KBr) cm^{-1} 3299, 2772, 1666; ^1H -NMR (DMSO- d_6 , 300 MHz):

δ 3.69 (2H, s), 3.78 (4H, s), 6.76–7.26 (13H, ArH) 7.49 (2H, s, olefinic protons), 9.55 (2H, OH); LC–MS: m/z = 398 ($\text{M} + 1$) $^+$.

Acknowledgments The authors wish to thank Dr. Arun Balakrishnan, Vice President, External Liaison & Screening, Piramal Life Sciences Limited, Mumbai, for the anticancer screening. We also thank Dr. R. Sethumadhavan, Professor, School of biosciences and technology, VIT University, Vellore, for his valuable suggestions.

References

- Adams BK, Ferstl EM, Davis MC, Herold M, Kurtkaya S, Camalier RF, Hollingshead MG, Kaur G, Sausville EA, Rickles FR, Snyder JP, Liotta DC, Shoji M (2004) Synthesis and biological evaluation of novel curcumin analogs as anti-cancer and anti-angiogenesis agents. *Bioorg Med Chem* 12:3871–3883. doi: [10.1016/j.bmc.2004.05.006](https://doi.org/10.1016/j.bmc.2004.05.006)
- Aggarwal BB, Kumar A, Bharti AC (2003) Anticancer potential of curcumin: preclinical and clinical studies. *Anticancer Res* 23:363–398. doi: [0250-7005/2003](https://doi.org/10.2500/7005/2003)
- Cell viability assessment was performed by using Cell Counting Kit (CCK-8), which was purchased from Dojindo Laboratory, Japan. Experiment design was guided by the CCK-8 technical manual available at <http://www.dojindo.com/newimages/CCK-8TechnicalInformation.pdf>
- Chandru H, Sharada AC, Kumar CSA, Rangappa KS (2008) Antiangiogenic and growth inhibitory effects of synthetic novel 1,5-diphenyl-1,4-pentadiene-3-one-3-yl-ethanone pyridine curcumin analogues on Ehrlich ascites tumor in vivo. *Med Chem Res* 17:515–529. doi: [10.1007/s00044-008-9095-2](https://doi.org/10.1007/s00044-008-9095-2)
- Das U, Alcorn J, Shrivastav A, Sharma RK, De Clercq E, Balzarini J, Dimmock JR (2007) Design, synthesis and cytotoxic properties of novel 1-[4-(2-alkylaminoethoxy)phenylcarbonyl]-3,5-bis(arylidene)-4-piperidones and related compounds. *Eur J Med Chem* 42:71–80. doi: [10.1016/j.ejmech.2006.08.002](https://doi.org/10.1016/j.ejmech.2006.08.002)
- Dhillon N, Wolf RA, Abbruzzese JL, Hong DS, Camacho LH, Li L, Braithel FS, Kurzrock R (2006) Phase II clinical trial of curcumin in patients with advanced pancreatic cancer. *J Clin Oncol* 24:14151
- Dube D, Kukshal V, Srivastava SK, Tripathi RP, Ramachandran R (2008) NAD^+ -dependent DNA ligase (Rv3014c) from *M. tuberculosis*: strategies for inhibitor design. *Med Chem Res* 17:189–198. doi: [10.1007/s00044-007-9052-5](https://doi.org/10.1007/s00044-007-9052-5)
- Fadda AA, Badria FA, El-Attar KM (2009) Synthesis and evaluation of curcumin analogues as cytotoxic agents. *Med Chem Res*. doi: [10.1007/s00044-009-9199-3](https://doi.org/10.1007/s00044-009-9199-3)
- Gafner SK, Lee SK, Cuendet M, Barthelemy S, Vergnes L, Labidalle S, Mehta RG, Boone CW, Pezzuto JM (2004) Biologic evaluation of curcumin and structural derivatives in cancer chemoprevention model systems. *Phytochemistry* 65:2849–2859. doi: [10.1016/j.phytochem.2004.08.008](https://doi.org/10.1016/j.phytochem.2004.08.008)
- Jing-yan T, Guo L, Yan-yun G, Hong-li Z, Wen-zhong Z, Xiao W, Hong-da Z, Tian-hong L, Min L (2006) Role and mechanism of rosiglitazone on the impairment of insulin secretion induced by free fatty acids on isolated rat islets. *Chin Med* 119(7):574–580
- Karunakaran D, Rashmi R, Kumar TR (2005) Induction of apoptosis by curcumin and its implications for cancer therapy. *Curr Cancer Drug Target* 5:117–129. doi: [10.2174/1568009053202081](https://doi.org/10.2174/1568009053202081)
- Lakshmi S, Dhanya GS, Joy B, Padmaja G, Remani P (2008) Inhibitory effect of an extract of *Curcuma zedoariae* on human cervical carcinoma cells. *Med Chem Res* 17:335–344. doi: [10.1007/s00044-007-9069-9](https://doi.org/10.1007/s00044-007-9069-9)

- Li N, Cehn X, Liao J, Yang G, Wang S, Jossephson Y, Han C, Chen J, Huang MT, Yang CS (2002) Inhibition of 7,12-dimethylbenz[a]anthracene (DMBA)-induced oral carcinogenesis in hamsters by tea and curcumin. *Carcinogenesis* 23:1307–1313. doi:[10.1093/carcin/23.8.1307](https://doi.org/10.1093/carcin/23.8.1307)
- Liang G, Li X, Chen L, Yang S, Wu X, Studer E, Gurley E, Hylemon PB, Ye F, Li Y, Zhou H (2008a) Synthesis and anti-inflammatory activities of mono-carbonyl analogues of curcumin. *Bioorg Med Chem Lett* 18:1525–1529. doi:[10.1016/j.bmcl.2007.12.068](https://doi.org/10.1016/j.bmcl.2007.12.068)
- Liang G, Yang S, Jiang L, Zhao Y, Shao L, Xiao J, Ye F, Li Y, Li X (2008b) Synthesis and anti-bacterial properties of mono-carbonyl analogues of curcumin. *Chem Pharm Bull* 56(2):162–167. doi:[10.1248/cpb.56.162](https://doi.org/10.1248/cpb.56.162)
- Osawa T, Sugiyama Y, Inayoshi M, Kawakishi S (1995) Antioxidative activity of tetrahydrocurcuminoids. *Biosci Biotechnol Biochem* 59:1609–1612 PMID: 8520105
- Panchatcharam M, Miriyala S, Gayathri VS, Suguna L (2006) Curcumin improves wound healing by modulating collagen and decreasing reactive oxygen species. *Mol Cell Biochem* 290:87–96. doi:[10.1007/s11010-006-9170-2](https://doi.org/10.1007/s11010-006-9170-2)
- PatchDock Server (2008) An automatic server for molecular docking. <http://bioinfo3d.cs.tau.ac.il/PatchDock/>
- Pati HN, Das U, Quail JW, Kawase M, Sakagami H, Dimmock JR (2008) Cytotoxic 3,5-bis(benzylidene)piperidin-4-ones and *N*-acyl analogs displaying selective toxicity for malignant cells. *Eur J Med Chem* 43:1–7. doi:[10.1016/j.ejmech.2007.03.010](https://doi.org/10.1016/j.ejmech.2007.03.010)
- Pati HN, Das U, Das S, Bandy B, Clercq ED, Balzarini J, Kawase M, Sakagami H, Quail JW, Stables PJ, Dimmock JR (2009) The cytotoxic properties and preferential toxicity to tumour cells displayed by some 2,4-bis(benzylidene)-8-methyl-8-azabicyclo [3.2.1] octan-3-ones and 3,5-bis(benzylidene)-1-methyl-4-piperidones. *Eur J Med Chem* 44:54–62. doi:[10.1016/j.ejmech.2008.03.015](https://doi.org/10.1016/j.ejmech.2008.03.015)
- RCSB Protein Data Bank. Solution structure of the catalytic domain of mapk phosphatase pac-1: insights into substrate-induced enzymatic activation. Receptor 1m3g available at <http://www.rcsb.org/pdb/explore/explore.do?structureId=1M3G>
- Santo RD, Costi R, Artico M, Ragno R, Greco G, Novellino E, Marchand C, Pommier Y (2005) Design, synthesis and biological evaluation of heteroaryl diketohexenoic and diketobutanoic acids as HIV-1 integrase inhibitors endowed with antiretroviral activity. *IL Farmaco* 60:409–417. doi:[10.1016/j.farmac.2005.03.008](https://doi.org/10.1016/j.farmac.2005.03.008)
- Wu J, Jin YJ, Calaf GM, Huang WL, Yin Y (2007) PAC1 is a direct transcription target of E2F-1 in apoptotic signaling. *Oncogene* 26(45):6526–6535. doi:[10.1038/sj.onc.1210484](https://doi.org/10.1038/sj.onc.1210484)
- Zeng JH, Xu GB, Chen X (2009) Application of the chromatographic fingerprint for quality control of essential oil from GuangXi *Curcuma kwangsiensis*. *Med Chem Res* 18:158–165. doi:[10.1007/s00044-008-9115-2](https://doi.org/10.1007/s00044-008-9115-2)