DOI 10.1002/aoc.3694

FULL PAPER

Novel macrocyclic Schiff base and its complexes having N₂O₂ group of donor atoms: Synthesis, characterization and anticancer screening

Ehab M. Zayed¹ | Mohamed A. Zayed² | Asmaa M. Fahim¹ | Fatma A. El-Samahy¹

¹Green Chemistry Department, Research Centre,
 33 EL Bohouthst (former EL Tahrirst), Dokki,
 12622 Giza, Egypt

²Chemistry Department, Faculty of Science, Cairo University, 12613 Giza, Egypt

Correspondence

M. A. Zayed, Chemistry Department, Faculty of Science, Cairo University, 12613 Giza, Egypt. Email: mazayed429@yahoo.com

Funding information Chemistry Department at Cairo University and Green Chemistry Department Novel Schiff base [N',N'''-(((ethane-1,2-diylbis(oxy))bis(2,1-phenylene))bis(methanylylidene))di(benzohydrazide)] was formed by the condensation reaction of benzohydrazide with 2,2'-(ethane-1,2-diylbis(oxy))dibenzaldehyde. Its reaction with various metal ions was studied and the structures of the new products were characterized using common analytical and spectroscopic methods. All the metal complexes have pronounced anticancer activities. The antimicrobial activities against Gram-negative and Gram-positive bacteria were investigated.

KEYWORDS

anticancer screening, antimicrobial activities, complexes, kinetic, metal ions, novel Schiff's base, spectroscopic analysis, thermal analysis

1 | INTRODUCTION

The physicochemical and pharmacological properties of heterocyclic compounds such as benzimidazoles are improved upon reaction with transition metal chlorides to give complexes.^[1–4] Positively charged metal centre combined with heteroaromatic periphery forms well-defined geometries. which facilitate the interaction with biomolecules and transacross membranes in biological systems.^[5,6] port Thiosemicarbazone heterocyclic compounds and their metal complexes have attracted considerable attention due to their coordination chemistry and broad range of pharmacological properties.^[7,8] Hydrazones are characterized by the presence of azomethine group (-CH=N-); they are good polydentate chelating agents that can form a variety of complexes with various transition metals and inner transition metals.^[9–17] Metal complexes containing improved organic ligands are widely used in cancer chemotherapy. The great success in the clinical treatment of human malignancies has stimulated research in the area of inorganic antitumor agents, the application of which can be hampered by severe toxicity and

development of resistance during therapy.^[6,18,19] To avoid these disadvantages, current strategies for the development of novel metallo drugs have focused on the use of transition metal complexes.^[11,20] Various coordination compounds have been synthesized and the effects of metal, ligand and substituent on biological and anticancer activities have been investigated.^[21–23] However, side effects, toxicity, cancer specificity and especially acquired resistance are still significant problems.

The main goal of the research reported here was to prepare and characterize a biologically active heterocyclic ligand. This was reacted with metal chlorides to yield complexes having anticancer and other biological activities.

2 | EXPERIMENTAL

2.1 | Materials and reagents

All chemicals used in this study were of analytical reagent grade and of the highest purity available. They included Cu(II) chloride (Sigma), Co(II) chloride hexahydrate and

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2017 The Authors Applied Organometallic Chemistry Published by John Wiley & Sons Ltd.



SCHEME 1 Synthesis of metal complexes 5

Ni(II) chloride hexahydrate (BDH), ferric chloride hexahydrate (Prolabo), zinc chloride (Ubichem) and Cd(II) chloride (Aldrich). The other materials included 2,2'-(ethane-1,2-diylbis(oxy))dibenzaldehyde, salicylaldehyde, 1,2dibromoethane (Sigma) and benzohydrazide (Aldrich). Organic solvents used included absolute ethyl alcohol and dimethylformamide (DMF). These solvents were spectroscopically pure from BDH. Double-distilled water collected using all-glass equipment was used in all preparations.

2.2 | Instrumentation

Elemental microanalyses of the separated solid chelates for C, H, N and Cl were performed at the Microanalytical Centre, Cairo University, using a CHNS-932 (LECO) Vario elemental analyser. The analyses were repeated twice to check the accuracy of the data. The molar conductance of solid chelates in DMF was measured using a Jenway 4010 conductivity meter. Fourier transform infrared (FT-IR) spectra were recorded with a PerkinElmer FT-IR type 1650 spectrophotometer in the wavenumber region 400–4000 cm^{-1} . The spectra were recorded as KBr pellets. Solid reflectance spectra were measured with a Shimadzu 3101pc spectrophotometer. The molar magnetic susceptibility was measured with powdered samples using the Faraday method. Diamagnetic corrections were made using Pascal's constant and Hg[Co(SCN)₄] was used as a calibrant. Mass spectra were recorded with the EI technique at 70 eV using an MS-5988 GS-MS Hewlett-Packard instrument at the Microanalytical Centre, Cairo University. ¹H NMR spectra were recorded using a 300 MHz Varian-Oxford Mercury. The solvent used was deuterated dimethylsulfoxide (DMSO- d_6) and the spectra extended from 0 to 15 ppm. Thermal analyses (thermogravimetry (TG) and differential thermogravimetry (DTG)) were carried out in a dynamic nitrogen atmosphere (20 ml min⁻¹) with a heating rate of 10 °C min⁻¹ using a DTG-60 H Shimadzu simultaneous DTA/TG apparatus.

2.3 | Synthesis of metal complexes 5

The metal complexes **5** were prepared by the addition of a hot solution (60 °C) of the appropriate metal chloride **4** in absolute ethanol (15 ml) to a hot solution (60 °C) of the organic ligand **3** (0.3 g) in ethanol and DMF (15 ml). The resulting mixture was heated with stirring to evaporate all the solvents to afford a precipitate. The precipitate was dried and weighed to calculate the yield. All the above steps were repeated for all the selected transition metal complexes.

2.4 | Biological activity

Testing was done using the diffusion agar technique. Spore suspension (0.5 ml, 10^6 – 10^7 spores ml⁻¹) of each of the investigated organisms was added to a sterile agar medium just before solidification, then poured into sterile Petri dishes (9 cm in diameter) and left to solidify. Using a sterile cork borer (6 mm in diameter), three holes (wells) were made into each dish, and then 0.1 ml of the test compound dissolved in $DMF (100 \text{ mg ml}^{-1})$ was poured into these holes. The dishes were incubated at 37 °C for 48 h where a clear or inhibition zone was detected around each hole. DMF (0.1 ml) was used as a control under the same conditions. By subtracting the diameter of the inhibition zone resulting from DMF from that obtained from each metal complex or the free Schiff base, antibacterial activities were calculated as a mean of three replicates. MIC₅₀ was determined, defined as the lowest compound concentration that inhibits growth by 50%.

2.5 | Pharmacology materials and methods

MCF-7 breast cancer cell line was obtained from the National Cancer Institute (Cairo, Egypt). MCF-7 cells were grown in RPMI-1640. Media were supplemented with 10% heatinactivated foetal bovine serum, 50 U ml⁻¹ penicillin and 50 g ml⁻¹ streptomycin and maintained at 37 °C in a humidified atmosphere containing 5% CO2. The cells were maintained as 'monolayer culture' by serial sub-culturing. Cytotoxicity was determined using the sulforhodamine B (SRB) method as previously described.^[21] Exponentially growing cells were collected using 0.25% trypsin-EDTA and seeded in 96-well plates at 1000-2000 cells per well in RPMI-1640 supplemented medium. After 24 h, cells were incubated for 72 h with various concentrations of the test compounds as well as doxorubicin as reference drug. Following 72 h of treatment, the cells were fixed with 10% trichloroacetic acid for 1 h at 4 °C. Wells were stained for 10 min at room temperature with 0.4% SRB stain dissolved in 1% acetic acid. The plates were air-dried for 24 h and the dye was solubilized with Tris-HCl for 5 min on a shaker at 1600 rpm. The optical density of each well was measured spectrophotometrically at 564 nm with an ELISA microplate

ZAYED ET AL

reader (ChroMate-4300, FL, USA). The IC_{50} values were calculated according to the equation for Boltzmann sigmoidal concentration–response curve using nonlinear regression fitting models (GraphPad Prism, Version 5).

3 | RESULTS AND DISCUSSION

3.1 | Characterization of metal complexes

Ligand 3 was formed by the condensation reaction of dibenzaldehyde derivative 1 with benzohydrazide $(2)^{[24]}$ (Scheme 1). The reactions of ligand 3 with metal ions 4 in equal molar ratio afforded metal complexes 5 (Scheme 1); their elemental analyses, yields and melting points are presented in Table 1. The ¹H NMR spectra of metal complexes 5 show the absence of a signal of NH group in the free ligand at 8.71 ppm due to the chelating process with metal. The characteristic signals of the ¹³C NMR spectrum for complex **5b** as an example show that metal ions influence the electronic charge distribution around particular carbons which appear at 72.2 ppm (CH₂) and 150.4 ppm (CH=), and slightly change that of other carbons at 113.2 (CH), 117 (CH), 121.3 (CH), 125 (CH), 127 (CH), 134.7 (CH), 135.1 (CH), 157 (C-O) and 162 ppm (C=O). This means that the electron densities around the referred carbons are affected by the ligand interaction with metal cations to form their metal complexes.^[25–28]

3.2 | Molar conductance measurements

Conductivity measurement of metal chelates in non-aqueous solutions has been used in structural studies within the limits of their solubility. This method gives the degree of ionization of compounds **5**, the molar conductivity increasing with increasing amount of ions that a complex liberates in solution. The molar conductivities of 10^{-3} molar solutions of the metal chelates at $25 \pm 2 \,^{\circ}$ C are given in Table 1. From the molar conductance values ($108-150 \,\Omega^{-1} \,\text{mol}^{-1} \,\text{cm}^2$) of Co(II), Ni(II), Cu(II), Mn(II), Cd(II) and Zn(II) complexes, it is concluded that these complexes are 1:2 electrolytes. On

TABLE 1 Analytical and physical data of metal complexes 5

the other hand, the Fe(III) complex has a molar conductance value of $250 \ \Omega^{-1} \ \text{mol}^{-1} \ \text{cm}^2$, indicating its ionic nature and it is considered as a 1:3 electrolyte.^[29–31]

3.3 | FT-IR spectra and mode of bonding

The FT-IR spectra of all complexes **5** show absorption bands ν (C=N) at 1586–1656 cm⁻¹ which are shifted by 50–52 cm⁻¹ to lower energy regions compared to compound **3**.^[32–35] This is due to the coordination of azomethine nitrogen to metal ion.^[36,37] Also, a broad ν (H₂O) band of **5** is found at 3260 cm⁻¹.^[38,39] The stretching band of ν (C–O–C) is observed at 1226 cm⁻¹ in the spectrum of **3** which is shifted to higher wavenumbers (1253–1261 cm⁻¹) in the spectra of **5** due to the participation of the oxygen atom in chelation. New absorption bands appear in the spectra of complexes corresponding to stretching vibrations ν (M–O) and ν (M–N) in the regions 505–609 and 435–484 cm⁻¹, respectively.^[40]

3.4 | Electronic spectral and magnetic susceptibility studies (Table 2)

Electronic spectra of complexes **5b** and **5f** show bands at 26 000 and 24 500 cm⁻¹, respectively, which may be due to ligand–metal charge transfer. Molar conductivities of **5b** and **5f** are over 100 Ω^{-1} cm² mol⁻¹ as expected for 1:2 electrolytes of Zn(II) and Cd(II) complexes.^[41–44]

3.5 | Thermal analyses (TG and DTG) and thermodynamic calculations

TG analysis of the ligand shows two successive steps of decomposition. The first mass loss of 16% (15.41%) in the temperature range 50–250 °C may be due to the decomposition of benzene molecule. At 250–400 °C the mass loss of 34% (34.13%) may be for the decomposition of $C_8H_7N_3O_2$ molecule. In the final stage from 400 to 600 °C, the estimated mass is loss of 50% (50.30%) is due to $C_{16}H_{13}NO_2$ molecule with complete decomposition.

			% Found (calcd)					II.ce	$\Lambda_{\rm m} (\Omega^{-1} {\rm mol}^{-1} {\rm cm}^2)$
Complex	Colour (% yield)	M.p. (°C)	С	Н	Ν	0	М	(BM)	n _m (n mor em)
$C_{30}H_{30}Cl_2CuN_4O_6$ [Cu(L).2H ₂ O].Cl ₂ (5a)	Brown (77)	220	53.22 (53.11)	4.47 4.25)	8.28 (8.13)	14.18 (13.78)	9.39	1.5	108
$\begin{array}{l} C_{30}H_{30}Cl_2ZnN_4O_6 \\ [Zn(L).2H_2O].Cl_2 \ (\textbf{5b}) \end{array}$	Yellowish white (80)	215	53.08 (52.89)	4.45 (4.55)	8.25 (8.12)	14.14 (13.98)	9.63	Dia.	120
$\begin{array}{l} C_{30}H_{30} \; FeCl_2N_4O_6 \\ [Fe(L).2H_2O].Cl_3 \; (\textbf{5c}) \end{array}$	Brown (75)	284	47.37 (47.21)	3.98 (3.54)	7.37 (7.26)	12.62 (12.52)	14.68	3.8	250
$\begin{array}{l} C_{30}H_{30}Cl_2NiN_4O_6 \\ [Ni(L).2H_2O].Cl_2 \; \textbf{(5d)} \end{array}$	Greenish yellow (88)	255	53.60 (53.42)	4.50 (4.23)	8.34 (8.22)	14.28 (14.16)	8.73	2.4	118
$\begin{array}{l} C_{30}H_{30}Cl_{2}CoN_{4}O_{6}\\ [Co(L).2H_{2}O].Cl_{2} \ (\textbf{5e}) \end{array}$	Dark red (78)	280	53.59 (53.58)	4.50 (4.26)	8.33 (7.89)	14.28 (13.87)	8.76	3.06	130
$\begin{array}{l} C_{30}H_{30}Cl_2CdN_4O_6 \\ [Cd(L).2H_2O].Cl_2 \ (\mathbf{5f}) \end{array}$	Pale yellow (80)	265	49.64 (49.32)	4.17 (4.56)	7.72 (7.48)	13.22 (13.54)	15.49	Dia.	125
$C_{30}H_{30}Cl_2MnN_4O_6$ [Mn(L).2H ₂ O].Cl ₂ (5 g)	Pale yellow (86)	250	53.91 (53.28)	4.52 (4.31)	8.38 (8.26)	14.36 (14.12)	8.22	3.69	150

TABLE 2 Electronic spectral data and magnetic susceptibility of complexes 5a, 5c-e and 5 g

Compound	Bands (cm ⁻¹)	Transitions	Magnetic moment (BM)	Geometry
5a	15 331 16 588 21 188	$\begin{array}{l} 2B1g \rightarrow 2B2g \\ 2B1g \rightarrow 2Eg \\ 2B1g \rightarrow 2A1g \end{array}$	1.5	Octahedral ^[45,46]
5c	22 754 16 550–18 321 13 822–17 436	$\begin{array}{l} 6A1g \rightarrow T2 \ g(G) \\ 6A1g \rightarrow 5T1g \end{array}$	3.8	Octahedral ^[42]
5d	15 785 17 406 19 588	$\begin{array}{l} 3A2g \rightarrow 3T2g \\ 3A2g \rightarrow 3T1g \ (F) \\ 3A2g \rightarrow 3T1g \ (P) \end{array}$	2.4	Octahedral ^[43,44]
5e	17 452 19 569 24 459	$\begin{array}{l} 4\text{T1}g(\text{F}) \rightarrow 4 \text{ T2 } g(\text{F}) \\ 4\text{T1}g(\text{F}) \rightarrow 4\text{A2}g(\text{F}) \\ 4\text{T1}g(\text{F}) \rightarrow 4 \text{ T2 } g(\text{P}) \end{array}$	3.06	Octahedral ^[43,44]
5 g	15 810 17 513	$\begin{array}{c} 4\text{T1g} \rightarrow 6\text{A1g} \\ 4 \text{ T2 } g(\text{G}) \rightarrow 6\text{A1g} \\ 4\text{T1g}(\text{D}) \rightarrow 6\text{A1g} \end{array}$	3.69	Octahedral ^[41]

The TG analysis of complex **5a** as an example shows decomposition at 44–800 °C (Table 3). At 44–193 °C the complex loses 2HCl, O_2 and NO molecules with a mass loss of 20% (20.08%) as a first step. The second step corresponds to 2H₂O, N₂ and 4CH₄ molecules with a mass loss of 18% (18.9%) within the range 196–364 °C. The third step of decomposition occurs at 366–457 °C for loss of C₂H₅ molecule with a mass loss of 4% (4.28%). The final stage within the range 457–715 °C shows loss of C₂H₃N with a mass loss of 45% (45.79%) leaving CuO as a residue.

Three decomposition steps appear in the thermal analysis of complex **5c** as an example of trivalant complex (Table 3). The first one corresponds to $2H_2O$ and $1/2O_2$ molecules at 24–194 °C with mass loss of 7% (6.84%). The second (198–407 °C) corresponds to the loss of some of organic ligand, 3HCl and CH₄ molecules with mass loss of 16% (16.31%). The final stage at 407–699 °C corresponds to the loss of $C_{29}H_{19}N_4$ molecule with mass loss of 56% (56.18%) and Fe₂O₃ as a residue.

TABLE 3 Thermal analysis (TG and DTG) results for complexes 5

Compound	TG range (°C)	DTG _{max} (°C)	n*	Mass loss, calcd (estim.) (%)	Total mass loss, calcd (estim.) (%)	Assignment	Residue
[Cu(L).2H ₂ O]Cl ₂ (5a)	44–193 196–364 366–457 457–715	84 301 406 561	1 1 1	20.08 (20) 18.9 (18) 4.28 (4) 45.79 (45)	88.37 (88)	Loss of 2HCl,O ₂ and NO Loss of 2H ₂ O, N ₂ and 4CH ₄ Loss of C_2H_5 Loss of C_24H_3N	CuO 11.7 (11)
[Zn(L).2H ₂ O]Cl ₂ (5b)	37–108 112–229 231–429 431–997	75 170 307 615	1 1 1 1	5.30 (5) 12.97 (13) 13.56 (14) 55.75 (55)	87.58 (87)	Loss of 2(H ₂ O) Loss of 2(HCl) and $1/2O_2$ Loss of 2(C ₂ H ₅) and NO Loss of C ₂₆ H ₁₄ N ₃ O	ZnO 12.42 (12)
[Fe(L).2H ₂ O]Cl ₃ (5c)	24–194 198–407 407–699	138 308 499	1 1 1	6.84(7) 16.31(16) 56.18 (56)	79.33(79)	Loss of $2(H_2O)$ and $1/2O_2$ Loss of $3(HCl)$ and CH_4 Loss of $C_{29}H_{19}N_4$	Fe ₂ O ₃ 20.67 (21)
$[Ni(L).2H_2O]Cl_2$ (5d)	23–97 98–354 355–452 453–654	56 286 401 518	1 1 1 1	5.35 (6) 20.23 (20) 13.69 (13) 50.44 (50)	89.71 (89)	Loss of 2(H ₂ O) Loss of 2(HCl) and 2(NO) Loss of C ₂ H ₅ , N ₂ O and CH ₄	NiO 11.11 (11)
[Co(L).2H ₂ O]Cl ₂ (5e)	26–115 117–204 206–393 395–710	74 155 294 517	1 1 1 1	4.76 (5) 5.35 (5) 14.77 (14) 63.98 (64)	88. 86 (88)	Loss of 2(H ₂ O) Loss of HCl Loss of HCl and NO Loss of C ₃₀ H ₂₄ N ₂ O	CoO 11.14 (12)
$[Cd(L).2H_2O]Cl_2$ (5f)	23–238 240–461 461–792	152 341 605	1 1 1	4. 95 (5) 23. 17 (23) 54.82 (54)	82.94 (82)	Loss of 2(H ₂ O) Loss of 2(HCl), 2(NO) and 2(CH ₄) Loss of $C_{28}H_{16}N_2O$	CdO 17.06 (18)
$[Mn(L).2H_{2}O]Cl_{2}\ ({\bf 5}\ {\bf g})$	24–204 204–386	79 311	1	5.38 (5) 17.81 (18)	89. 8 (89)	Loss of $C_2H_4N_4S_2$ Loss of 2(HCl) and 1/2O ₂ Loss of $C_4H_{10}O_2N_2$	MnO C ₁₂ H ₈
	387-658	514	1	66.61 (66)			10.2 (11)

3.6 | Calculation of activation thermodynamic parameters

The activation energies of decomposition of the new compounds are found to be in the range $80.01-964.90 \text{ kJ mol}^{-1}$, these high values of the activation energies reflecting the thermal stability of the complexes (Table 4). On the other hand, the entropy of activation has a negative value for all the complexes, which indicates that the decomposition reactions proceed with a lower rate than the normal ones.

3.7 | Biological activity

The Schiff base **3** and its metal complexes **5** were tested in terms of antibacterial activity using the diffusion agar method.^[45–52] The reference compound for antibacterial activities was streptomycin and more than one test organism was used. The antibacterial activity data for compounds **3** and **5** have a wide degree of variation (Table 5).

The bis-Schiff base ligand **3** has more sensitivity towards Gram-positive than Gram-negative bacteria and has high MIC₅₀ (>100 mg ml⁻¹) for both types of bacteria. It is found that Cu(II) compound **5a** and Ni(II) compound **5d** have the highest inhibition zones and high MIC₅₀ (>100 mg ml⁻¹) against Gram-negative bacteria (*E. coli* and *P. vulgaris*). On the other hand, complex **5e** has moderate inhibitory activity (20 and 22 mm) for *E. coli* and *P. vulgaris*, respectively, but lowest MIC₅₀ (>50 mg ml⁻¹) for both microorganisms. Mn (II) compound **5** g poorly inhibits *E. coli* and *P. vulgaris* against two Gram-negative bacteria; whereas forms **3**, **5b**

TABLE 4	Thermodynamic	data for	complexes	5

-WILEY-Organometallic 5 of 7 Chemistry

 TABLE 5
 Antibacterial activity data for compounds 3 and 5

	Inhibition zone (mm mg^{-1} sample)/MIC ₅₀ (mg ml^{-1})						
	Gram nega	tive	Gram positive				
Sample	Escherichia coli	Proteus vulgaris	Bacillus subtilis	Streptococcus pyogenes			
3	25	26	25	25			
5a	30/>100	31/>100	31/>100	34/>100			
5b	25/>100	28/>100	29/>100	27/>100			
5c	18/>100	19/>100	20/>100	22/>100			
5d	32/>100	30/>100	31/>100	34/>100			
5e	20/>50	22/>50	23/>25	25/>50			
5f	28/>100	29/>100	27/>100	25/>100			
5 g	24/>100	22/>100	23/>100	21/>100			

and **5f** are found to possess slightly inhibited significant activity. Fe (III) compound **5c** shows comparatively weak inhibition against Gram-positive bacterium *S. pyogenes* at MIC50 (>100 mg ml⁻¹).

3.8 | *In vitro* cytotoxic activity and anticancer screening studies

The results are reported in Table 6 for three separate experiments. Statistical differences were analysed according to one-way ANOVA tests wherein the differences were considered to be significant at p < 0.05.

Anti-proliferative activities of the new metal complexes of Co, Cu, Zn, Cd, Fe, Ni and Mn were examined in MCF-

Complex	Decomposition temp. (°C)	$A (s^{-1})$	E (kJ mol ⁻¹)	$\begin{array}{c}\Delta S\\ (J \ K^{-1} \ mol^{-1})\end{array}$	$\frac{\Delta H}{(\text{kJ mol}^{-1})}$	$\frac{\Delta G}{(\text{kJ mol}^{-1})}$
[Cu(L).2H ₂ O]Cl ₂ (5a)	44–193	4.10×10^{5}	52.86	-107.26	46.46	128.97
	196–364	9.02×10^{6}	28.72	-244.18	280.82	92.96
	366–457	7.46×10^{8}	98.35	-82.95	97.71	104.94
	457–715	7.56×10^{7}	79.51	-101.98	155.12	80.01
$[Zn(L).2H_2O]Cl_2~(\textbf{5b})$	37–108	4.68×10^{6}	229.16	-39.18	222.76	786. 83
	112–229	1.93×10^{7}	278.90	-34.61	272.08	569.73
	231–429	9.46×10^{8}	385.32	-34.05	378.92	117.16
	431–997	1.57×10^{5}	711.82	15.33	647.82	182.74
[Fe(L).2H ₂ O]Cl ₃ (5c)	24–197	2.87×10^{6}	307.53	-44. 52	301.13	413. 78
	198–407	2.96×10^{7}	469.44	-48. 37	46.30	908.69
	407–699	6.57×10^{6}	134.81	-64.85	128.41	178.31
$[Ni(L).2H_2O]Cl_2~(\textbf{5d})$	23–97	1.17×10^{5}	308.29	-64.83	301.89	196.91
	98–354	6.36×10^{6}	918.06	-136.37	858.68	183. 56
	355–452	2.81×10^{7}	803.99	-90.47	797.59	101.60
	453–654	4.84×10^{8}	689.79	-105.67	497.36	817.96
$[C_0(L).2H_2O]Cl_2$ (5e)	26–115 117–204 206–393 395–710	$\begin{array}{c} 6.\ 81\times10^{6}\\ 1.54\times10^{7}\\ 1.35\times10^{8}\\ 1.91\times10^{11} \end{array}$	608.29 485.92 390.11 127.61	-131. 40 -76. 55 -32.40 -36.90	601.76 479.52 383.71 636.66	409. 12 109.40 134.39 347.47
$[Cd(L).2H_2O]Cl_2$ (5f)	23–238	1.91×10^{6}	53.80	39.68	537.97	537.67
	240–461	9.63×10^{7}	96.55	72.35	965.45	964.90
	461–792	6.36×10^{8}	144.60	77.71	144.60	144.60
$[Mn(L).2H_2O]Cl_2 (5 g)$	24–204	1.68×10^{7}	110.48	195.53	104.08	890.39
	204–386	4.09×10^{8}	246.79	21.84	240.39	723.64
	387–658	7.22×10^{9}	523.47	35.71	517.07	242.307

^aE, activation energy; H, enthalpy; S, entropy; G, Gibbs free energy.

 TABLE 6
 In vitro
 anti-proliferative activities of the newly prepared derivatives against various cell lines

	$IC_{50} \ (\mu g \ \mu l^{-1})^b$					
Compound ^a	MCF-7	HepG2	НСТ			
5e	14.8 ± 0.02	10.6 ± 0.01	8.2 ± 0.012			
5a	10.3 ± 0.01	4.7 ± 0.007	3.7 ± 0.005			
5b	4.8 ± 0.007	3.1 ± 0.004	11.2 ± 0.016			
5f	4.4 ± 0.006	3.3 ± 0.004	5.9 ± 0.008			
5c	11.9 ± 0.017	9.4 ± 0.014	8.6 ± 0.013			
5d	12.2 ± 0.018	5.3 ± 0.007	3.9 ± 0.005			
5 g	4.4 ± 0.006	10.5 ± 0.015	5.7 ± 0.008			
DOX	4.6 ± 0.008	ND	ND			
DOX	ND	1.2 ± 0.002	ND			
DOX	ND	ND	4.69 ± 0.008			

^aDOX, doxorubicin (standard drug)

 ${}^{b}IC_{50}$ values are mean \pm SD of three separate experiments. ND, not detected.

7 (breast cancer) cell line (Figure 1), HepG2 (perpetual) cell line and HCT (colon cancer) cell line using doxorubicin colorimetric assay as described previously.^[53]

Doxorubicin was used as a reference cytotoxic compound for the MCF-7, HepG2 and HCT cell lines. The growth inhibitory concentration (IC₅₀) values, which refer to the concentration of compound required to produce a 50% inhibition of cell growth after 72 h of incubation compared to untreated controls, are summarized in Table 6. The complexes for which cell growth was inhibited by more than 50% are assigned as active. Almost all heterocyclic transition metal(II) complexes did not show cytotoxic activity and did not enter the secondary screening. Transition metal complexes with Zn, Cd and Mn show high specificity and are more potent for MCF-7 cell line compared with doxorubicin analogue with IC₅₀ = 4.6 μ M. This is due to chelation of Schiff bases of our heterocyclic compounds containing an azomethine group (--CH=N--) bond.

Also, the heterocyclic transition metal complexes were screened against HepG2 cells (Figure 2). The results show moderate activity for the Zn metal complex at $IC_{50} = 3.1 \mu M$ compared with doxorubicin analogue with $IC_{50} = 1.2 \mu M$. This means that none of the complexes



FIGURE 1 The growth inhibitory IC_{50} values [μ M] of metal (II) complexes **5** at the concentration of 20 μ M for MCF-7 (breast cancer cells)



FIGURE 2 The growth inhibitory IC₅₀ values $[\mu M]$ of metal (II) complexes **5** at the concentration of 20 μ M for HEPG-2 (liver cancer cells)



FIGURE 3 The growth inhibition IC_{50} values [μ M] of metal (II) complexes **5** at the concentration of 20 μ M for HCT (colony cancer cells)

possesses the ability to inhibit the growth of cancer cell lines at 20 $\mu M.$

Finally the transition metal(II) complexes were tested against HCT cell line (Figure 3). The Cu(II) complex shows higher activity at $IC_{50} = 3.7 \mu M$ compared with doxorubicin analogue at $IC_{50} = 4.69 \mu M$, which indicates that the Cu(II) complex exhibits cytotoxic activity against HCT cell line.

4 | CONCLUSIONS

In this study a novel Schiff base and novel synthesized heterocyclic transition metal complexes were developed via a delivery system emulated by self-assembly of doxorubicin. Their biological, *in vitro* cytotoxic and anticancer activities were investigated. The Cu(II) complex increased the accumulation of doxorubicin in tumour cells (HCT). The Cd, Zn and Mn complexes increased the efficacy in breast cancer cells (MCF-7). The metal complexes have greater antimicrobial effect than the free ligand.

ACKNOWLEDGMENTS

The authors acknowledge the support of this research given by the Chemistry Department at Cairo University and Green Chemistry Department and National Research Centre Egypt. Thanks are also due to the staff of the Microanalytical Centre of Cairo University at which all analyses were made.

REFERENCES

- R. P. Bakale, G. N. Naik, C. V. Mangannavar, I. S. Muchchandi, I. N. Shcherbakov, C. Frampton, K. B. Gudasi, *Eur. J. Med. Chem.* 2014, 73, 38.
- [2] A. Inam, S. M. Siddiqui, T. S. Macedo, D. R. Magalhaes, A. C. Lima Leite, M. B. Soares, A. Azam, *Eur. J. Med. Chem.* **2014**, *75*, 67.
- [3] W. B. Júnior, M. S. Alexandre-Moreira, M. A. Alves, A. Perez-Rebolledo, G. L. Parrilha, E. E. Castellano, O. E. Piro, E. J. Barreiro, L. M. Lima, H. Beraldo, *Molecules* **2011**, *16*, 6902.
- [4] F. A. Muregi, A. Ishih, Drug Dev. Res. 2010, 71, 20.
- [5] A. Almeida, B. L. Oliveira, J. D. G. Correia, G. Soveral, A. Casini, *Coord. Chem. Rev.* 2013, 275, 2689.
- [6] B. F. Ruan, Y. Z. Zhu, W. D. Liu, B. A. Song, Y. P. Tian, *Eur. J. Med. Chem.* 2014, 72, 46.
- [7] M. X. Li, C. L. Chen, D. Zhang, J. Y. Niu, B. S. Ji, Eur. J. Med. Chem. 2010, 45, 3169.
- [8] T. S. Raji, M. Zec, T. Todorovi, K. Celkovi, S. Radulovi, Eur. J. Med. Chem. 2011, 46, 3734.
- [9] P. G. Avaji, C. H. V. Kumar, S. A. Patil, K. N. Shivananda, C. Nagaraju, *Eur. J. Med. Chem.* **2009**, *44*, 3552.
- [10] B. Murukan, K. Mohanan, J. Enzyme Inhib, Med. Chem. 2007, 22, 65.
- [11] T. Suksrichavalit, S. Prachayasittikul, C. Nantasenamat, C. I. Ayudhya, V. Prachayasittikul, *Eur. J. Med. Chem.* 2009, 44, 3259.
- [12] M. Valko, D. Leibfritz, J. Moncol, M. T. Cronin, M. Mazur, J. Telser, Int. J. Biochem. Cell Biol. 2007, 39, 44.
- [13] K. B. Gudas, M. S. Patil, R. S. Vadavi, Eur. J. Med. Chem. 2008, 43, 2436.
- [14] A. Juneja, T. S. Macedo, D. R. M. Moreira, M. B. P. Soares, A. C. L. Leite, J. K. Andrade, L. Neves, V. R. A. Pereira, F. Avecilla, A. Azam, *Eur. J. Med. Chem.* **2014**, *75*, 203.
- [15] D. Esteban-Fernandez, E. Moreno-Gordaliza, B. Canas, M. A. Palaciosa, M. M. Gomez-Gomez, *Metallomics* 2010, 2, 19.
- [16] V. Milacic, Q. P. Dou, Coord. Chem. Rev. 2009, 253, 1649.
- [17] J. Tan, B. Wang, L. C. Zhu, Bioorg. Med. Chem. 2009, 17, 614.
- [18] A. Tarushi, C. P. Raptopoulou, V. Psycharis, A. Terzis, G. Psomas, D. P. Kessissoglou, *Bioorg. Med. Chem.* 2010, 18, 2678.
- [19] E. M. Zayed, M. A. Zayed, M. El-Desawy, Spectrochim. Acta A 2015, 134, 155.
- [20] M. I. Hossain, M. Switalska, W. Peng, M. Takashima, N. Wang, M. Kaise, J. Wietrzyk, S. Dan, T. Yamor, T. Inokuchi, *Eur. J. Med. Chem.* 2013, 69, 294.
- [21] P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. McMahon, D. Vistica, J. T. Warren, H. Bokesch, S. Kenney, M. R. Boyd, *J. Natl Cancer Inst.* 1990, 82, 1107.
- [22] Y. Cao, S. Lindström, F. Schumacher, V. L. Stevens, D. Albanes, S. I. Berndt, H. Boeing, H. Bas Bueno-de-Mesquita, F. Canzian, S. Chamosa, S. J. Chanock, W. R. Diver, S. M. Gapstur, J. M. Gaziano, E. L. Giovannucci, C. A. Haiman, B. Henderson, M. Johansson, L. L. Marchand, D. Palli, B. Rosner, A. Siddiq, M. Stampfer, D. O. Stram, R. Tamimi, R. C. Travis, D. Trichopoulos, W. C. Willett, M. Yeager, P. Kraft, A. W. Hsing, M. Pollak, X. Lin, J. Ma, J. Natl Cancer Inst. 2014, 106, dju218.
- [23] B. K. Killelea, J. B. Long, A. B. Chagpar, X. Ma, R. Wang, J. S. Ross, C. P. Gross, J. Natl Cancer Inst. 2014, 106, dju159.
- [24] E. M. Zayed, M. A. Zayed, Spectrochim. Acta A 2015, 143, 81.
- [25] R. Anbazhagan, K. R. Sankaran, J. Mol. Struct. 2013, 1050, 73.
- [26] O. A. El-Gammal, G. M. Abu El-Reash, S. E. Ghazy, A. H. Radwan, J. Mol. Struct. 2012, 1020, 6.

- [27] E. M. Zayed, M. A. Zayed, A. M. M. Hindy, J. Therm. Anal. Calorim. 2014, 116, 391.
- [28] J. A. Dean, Lange's Handbook of Chemistry, Vol. 14, McGraw-Hill, New York 1992.
- [29] H. Alyar, S. Alyar, A. Unal, N. Ozbek, E. Sahin, N. Karacan, J. Mol. Struct. 2012, 1028, 116.
- [30] N. Raman, S. Sobha, A. Thamaraichelvan, Spectrochim. Acta A 2011, 78, 888.
- [31] W. M. I. Hassan, E. M. Zayed, A. K. Elkholy, H. Moustafa, G. G. Mohamed, Spectrochim. Acta A 2013, 103, 378.
- [32] R. A. A. Ammar, A. M. A. Alaghaz, Int. J. Electrochem. Sci. 2013, 8, 8686.
- [33] M. B. Halli, R. B. Sumathi, M. Kinni, Spectrochim. Acta A 2012, 99, 46.
- [34] E. M. Zayed, A. M. M. Hindy, G. G. Mohamed, J. Therm. Anal. Calorim. 2015, 120, 893.
- [35] S. Ilhan, H. Temel, I. Yilmaz, M. Sekerci, J. Organometal. Chem. 2007, 692, 3855.
- [36] E. M. Zayed, E. H. Ismail, G. G. Mohamed, M. M. H. Khalil, A. B. Kamel, *Monatsh. Chem.* 2014, 145, 755.
- [37] a) M. M. H. Khalil, G. G. Mohamed, E. H. Ismail, E. M. Zayed, A. B. Kamel, *Egyptian J. Pure Appl. Sci.* 2011, 29–37; b) M. M. H. Khalil, G. G. Mohamed, E. H. Ismail, E. M. Zayed, A. B. Kamel, *Open J. Inorg. Chem.* 2012, 2, 13.
- [38] E. M. Zayed, H. H. Sokker, H. M. Albishri, A. M. Farag, *Ecol. Eng.* 2013, 61, 390.
- [39] M. M. H. Khalil, G. G. Mohamed, E. H. Ismail, E. M. Zayed, A. B. Kamel, *Chin. J. Inorg. Chem.* **2012**, 28, 1495.
- [40] E. M. Zayed, G. G. Mohamed, A. M. M. Hindy, Spectrochim. Acta A 2015, 145, 76.
- [41] F. A. Cotton, G. Wilkinson, C. A. Murillo, M. Bochmann, Advanced Inorganic Chemistry, 6th ed., Wiley, New York 1999.
- [42] G. G. Mohamed, M. H. Solimanm, Spectrochim. Acta A 2010, 76, 341.
- [43] G. G. Mohamed, N. E. A. El-Gamel, F. Teixidor, Polyhedron 2001, 20, 2689.
- [44] M. A. Zayed, M. F. Hawash, M. A. Fahmey, A. M. A. El-Gizouli, J. Therm. Anal. Calorim. 2012, 108, 315.
- [45] M. S. Karthikeyan, D. J. Parsad, B. Poojary, K. S. Bhat, B. S. Holla, N. S. Kumari, *Bioorg. Med. Chem.* 2006, 14, 7482.
- [46] N. Shahabadi, Z. Ghasemian, S. Hadidi, *Bioinorg. Chem. Appl.* 2012, 2012, 126451.
- [47] S. Sen, N. A. Farooqui, S. Dutta, T. S. Easwari, V. Gangwar, K. Upadhya, S. Verma, A. Kumar, *Pharm. Chem.* 2013, 5, 128.
- [48] K. Singh, M. S. Barwa, P. Tyagi, Eur. J. Med. Chem. 2006, 41, 147.
- [49] T. Mosmann, J. Immunol. Methods 1983, 65, 55.
- [50] P. Vijayan, C. Raghu, G. Ashok, S. A. Dhanaraj, B. Suresh, *Indian J. Med. Res.* 2004, 120, 24.
- [51] S. W. C. Leuthauser, L. W. Oberley, T. D. Oberley, J. R. J. Sorenson, K. Ramakrishna, J. Natl Cancer Inst. 1981, 66, 1077.
- [52] L. R. de Alvare, K. Goda, T. Kimura, Biochem. Biophys. Res. Commun. 1976, 69, 687.
- [53] H. Tamura, H. Imai, J. Am. Chem. Soc. 1987, 109, 6870.

How to cite this article: Zayed EM, Zayed MA, Fahim AM, El-Samahy FA. Novel macrocyclic Schiff base and its complexes having N_2O_2 group of donor atoms: Synthesis, characterization and anticancer screening. *Appl Organometal Chem.* 2017;e3694. doi: 10.1002/aoc.3694