

# Cyanogen Bromide as Dehydrosulfurizing Agent for the Synthesis of *N*<sup>β</sup>-Fmoc-Amino Alkyl Isonitriles from *N*<sup>β</sup>-Fmoc-Amino Alkyl Thioformamides

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**Abstract:** Synthetically useful *N*<sup>β</sup>-Fmoc amino alkyl isonitriles are prepared conveniently from *N*<sup>β</sup>-Fmoc amino alkyl thioformamides via a cyanogen bromide mediated dehydrosulfurization. The reaction is fast, clean, and yields are good.

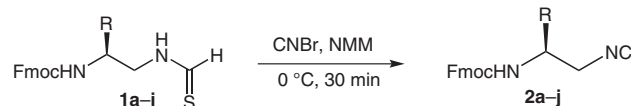
**Key words:** *N*<sup>β</sup>-Fmoc-amino alkyl thioformamides, isonitriles, dehydrosulfurization, cyanogen bromide

In recent years, the synthetic utility of isonitriles<sup>1</sup> in organic reactions has been exploited. Compounds containing an isocyano group find use as antibiotics,<sup>2</sup> antineoplastics,<sup>3</sup> and antianaesthetics.<sup>4</sup> The synthetic utility of isonitriles arises from their divergent character<sup>5</sup> and this functionality has wide utility, such as in the preparation of isothiocyanates,<sup>6</sup> *N,N*-dialkylcarbodiimides,<sup>7</sup> 1-substituted tetrazoles,<sup>8</sup> pyrazoles,<sup>9</sup> oxazoles,<sup>10</sup> and thiazoles.<sup>11</sup> The use of isonitriles in multicomponent reactions such as the Passerini<sup>12</sup> and Ugi<sup>13</sup> reactions for generating a wide variety of biologically important derivatives of peptides,<sup>14</sup> glycopeptides,<sup>15</sup> and other peptidic molecules<sup>16–20</sup> has been well demonstrated.

In peptide chemistry, the application of isonitrile derivatives of amino acids has led to a variety of peptidomimetics<sup>21</sup> and other useful products such as β-lactams,<sup>22</sup> glycopeptides,<sup>23</sup> depsipeptides,<sup>24</sup> peptide nucleic acids,<sup>25</sup> and *N*-hydroxy peptides.<sup>26</sup> Amino acid ester derived isonitriles are the known entities, which are prepared from corresponding formyl amino acid esters.<sup>27</sup> A recent entry into this new class of amino acid derived isonitriles has been described by our group wherein *N*<sup>β</sup>-Fmoc amino alkyl isonitriles were obtained through carboxyl group modification.<sup>28</sup>

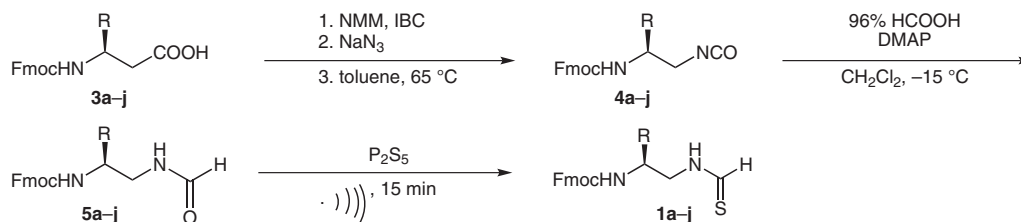
Of the many methods available for the preparation of isonitriles, dehydration of formamides is the widely followed route in amino acid chemistry, showing general applicability and reproducibility.<sup>29</sup> Reaction of alkyl halides with heavy metal cyanide salts,<sup>30</sup> addition of dichlorocarbene to amines,<sup>31</sup> reduction of isocyanates and isothiocyanates<sup>32</sup> are other available routes. Dehydration of formamides has been carried out by using tosyl chloride, phosphorus oxychloride (POCl<sub>3</sub>) in the presence of a base, cyanuric chloride, triphenyl phosphine–CCl<sub>4</sub>, phos-

gene, diphosgene, and triphosgene.<sup>33</sup> The method employing POCl<sub>3</sub> being quite harsh is found to cause racemization and therefore is not applicable in peptide chemistry, and isolation of enantiomerically pure isonitriles is always challenging and demanding. Danishefsky and co-workers have isolated enantiomerically pure α-isocyanoesters<sup>34</sup> derived from formyl amino acid esters, employing triphosgene/*N*-methylmorpholine (NMM) at –78 °C. Nenajdenko's group has synthesized the amino acid derived nonracemic isonitriles employing the less commonly used 4-methyl-2,6,7-trioxabicyclo[2.2.2]octyl-protected formyl amino ester.<sup>35</sup> Sureshbabu et al. reported the synthesis and isolation of enantiomerically pure *N*-Fmoc-amino alkyl isonitriles by the dehydration of the corresponding *N*<sup>β</sup>-Fmoc amino alkyl formamides under neutral conditions employing Burgess' reagent.<sup>28</sup> In a continuation of our current studies on isonitriles, we sought a simple and economically viable route involving dehydrosulfurization of *N*<sup>β</sup>-Fmoc amino alkyl thioformamides using cyanogen bromide (CNBr, Scheme 1).



**Scheme 1**

There has been a surge in the synthesis of thio analogues of peptides and related intermediates, the replacement of O with S yielding novel unnatural amino acids and peptidomimetics such as thiopeptides,<sup>36</sup> thioureas, and isothiocyanates<sup>37</sup> possessing therapeutic importance. Sulfur-containing compounds such as thioamides and thioesters have been found to be better reactants in the preparation of several heterocycle-tethered peptidomimetics and other related compounds compared to their oxygenated analogues.<sup>38,39</sup> Furthermore, some thioformamides show antifungal<sup>40</sup> and antibacterial activity,<sup>41</sup> and they have been employed in the synthesis of thiazoles via the Hantzsch protocol.<sup>42</sup> We envisaged the use of *N*-urethane-protected amino alkyl thioformamides for the synthesis of the corresponding isonitriles through a dehydrosulfurization reaction. In a patent, Ugi had demonstrated the conversion of thioformamides into their respective isonitriles using CNBr.<sup>43</sup> However, this protocol has yet to be exploited to its full advantage although the method shows promise.<sup>44,45</sup> Thus, in the present communication,



Scheme 2

we describe the synthesis of *N*<sup>β</sup>-Fmoc amino alkyl thioformamides and their conversion to isonitriles employing cyanogen bromide as the dehydrosulfurization reagent.

In the first part of this study, we undertook the preparation of *N*<sup>β</sup>-Fmoc amino alkyl thioformamides, a hitherto unreported class of molecules, by the thionation of the corresponding formamides. Starting with a *N*<sup>β</sup>-Fmoc amino acid, the carboxyl group was converted to its formamide under Gold–Schmidt–Wick formolysis of the corresponding isocyanate using formic acid/DMAP.<sup>46</sup> On treating a solution of *N*-Fmoc-Ala-Ψ[CH<sub>2</sub>NHCHO] **5b** in THF with an equimolar quantity of P<sub>2</sub>S<sub>5</sub> at room temperature, the respective thioformamide was obtained within an hour in satisfactory yield and purity after a flash chromatography. By way of optimization, a suspension of *N*-Fmoc-Ala-Ψ[CH<sub>2</sub>NHCHO] **5b** (1 mmol) and P<sub>2</sub>S<sub>5</sub> (0.7 mmol) in THF was subjected to ultrasonication at 25 °C. To our satisfaction, the reaction was complete in 15 minutes as judged by TLC, and the product was isolated after flash chromatography in excellent yield (Scheme 2) and enantiomeric purity.<sup>47</sup> The protocol was then successfully extended to other amino acids as well to prepare a series of *N*<sup>β</sup>-Fmoc-amino alkyl thioformamides **1a–j**,<sup>49,50</sup> and the

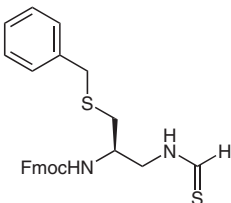
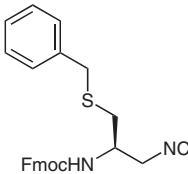
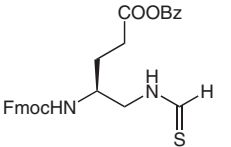
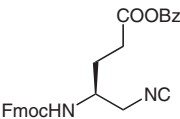
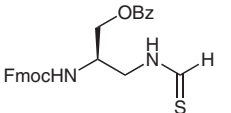
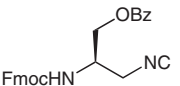
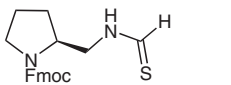
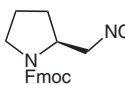
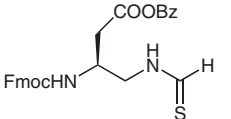
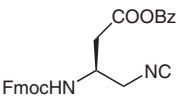
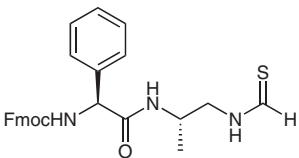
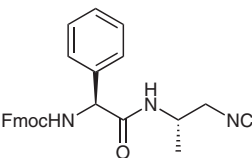
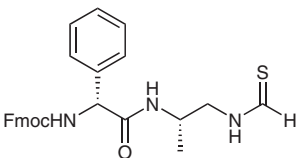
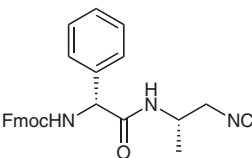
products were isolated as stable solids after purification in 92–96% yield and adequately characterized (Table 1).

In the next step, conversion of the thioformamides into isonitriles was undertaken.<sup>48</sup> Accordingly, a reaction of thioformamide **1** with CNBr (1.5 mmol) in the presence of NMM in dry CH<sub>2</sub>Cl<sub>2</sub> at 0 °C afforded the corresponding isonitrile **2**<sup>51,52</sup> within 30 minutes (Scheme 1). The reaction was rapid and complete, and the product was isolated after chromatography in excellent yield as a stable solid. When the reaction was carried out at room temperature, a considerable drop in the yield of isonitrile **1** (up to 20%) was incurred. Reproducible results were obtained when the protocol was employed for dehydrosulfurization of several other *N*<sup>β</sup>-Fmoc amino alkyl thioformamides. All the compounds were isolated as stable solids (Table 1) whose physical properties matched those previously reported by us<sup>28</sup> and were also further characterized spectroscopically. Thus dehydrosulfurization of thioformamides is more efficient when compared to the dehydration of formamides for the synthesis of isonitriles.

Table 1 List of Thioformamides **1** and Isonitriles **2**

Compd	Thioformamide <b>1</b>	Mp (°C)	HRMS [M + Na] <sup>+</sup> (found/calcd)	Isonitrile <b>2</b>	Mp (°C)	HRMS [M + H] <sup>+</sup> (found/ calcd)
<b>a</b>		141	349.0978/ 349.0987		165	293.1031/ 293.1200
<b>b</b>		105	363.1132/ 363.1143		112	307.1346/ 307.1368
<b>c</b>		118	439.1443/ 439.1456		128	383.1672/ 383.1681
<b>d</b>		126	391.1438/ 391.1456		101	335.1672/ 335.1681
<b>e</b>		117	405.1603/ 405.1613		109	349.1816/ 349.1838

**Table 1** List of Thioformamides **1** and Isonitriles **2** (continued)

Compd	Thioformamide <b>1</b>	Mp (°C)	HRMS [M + Na] <sup>+</sup> (found/calcd)	Isonitrile <b>2</b>	Mp (°C)	HRMS [M + H] <sup>+</sup> (found/ calcd)
<b>f</b>		129	485.1319/ 485.1333		115	429.1536/ 429.1558
<b>g</b>		97	511.1658/ 511.1667		56	455.1865/ 455.1893
<b>h</b>		103	469.1549/ 469.1562		76	413.1767/ 413.1787
<b>i</b>		124	389.1289/ 389.1300		104	333.1518/ 333.1525
<b>j</b>		107	497.1500/ 497.1511		80	441.1718/ 441.1736
<b>k</b>		167	459.1609/ 459.1617		160	440.1887/ 440.1896
<b>l</b>		165	459.1607/ 459.1617		160	440.1865/ 440.1896

The conditions employed in the synthesis of isonitriles were scrutinized to check for the possibility of racemization during the reaction. Two dipeptide thioformamides **1k,l**, prepared by the reaction of alanyl alkyl isonitrile with Fmoc-D/L-Phg-OH, were converted into their respective dipeptidyl isonitriles **2k,l** following the protocol described above. The <sup>1</sup>H NMR spectra possessed methyl doublets of the Ala residue of **2k** and **2l** at  $\delta = 1.16, 1.18$  ppm and at  $\delta = 1.23, 1.24$  ppm, respectively, indicating the presence of a single epimer in each sample, a conclusion which was also supported by HPLC data. Furthermore, the *N*-Fmoc-peptidyl isonitrile obtained by the reaction of an equimolar mixture of Fmoc-L/D-Phg-OH with alanyl alkyl thioformamide followed by dehydrosulfurization showed two separate doublets for the alanyl methyl group in the <sup>1</sup>H NMR spectrum, once again demonstrating the absence of epimerization.

In summary, we have described an efficient synthesis of *N*<sup>β</sup>-Fmoc-amino alkyl isonitriles by the dehydrosulfurization of corresponding thioformamides employing CNBr. This protocol provides an alternative efficient method for the preparation of the title compounds in high yields.

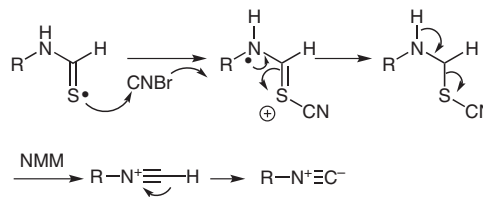
### Acknowledgment

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- (47) The two dipeptide thioformamides were synthesized in to verify the optical purity of *N*<sup>β</sup>-Fmoc-amino alkyl thioformamides **1**. For this, **1b** was treated with 20% Et<sub>3</sub>NH in CH<sub>2</sub>Cl<sub>2</sub> to deprotect the Fmoc group. The resulting amino-free *N*<sup>β</sup>-(1-amino alanine)-thioformamide was coupled both D and L isomers of Fmoc-Phg-OH using EDC/separately to obtain **1k** and **1l**. The <sup>1</sup>H NMR spectra of both Fmoc-L-Phg-Ala-Ψ[CH<sub>2</sub>NHCHS] and Fmoc-D-Phg-Ala-Ψ[CH<sub>2</sub>NHCHS] showed distinct methyl group doublets at δ = 1.13, 1.14 ppm and δ = 1.21, 1.23, respectively. However, the mixture of epimers prepared by coupling amino-free *N*<sup>β</sup>-(1-amino alanine)-thioformamide with epimeric mixture of Fmoc (L/D)-Phg-OH had two separate doublets corresponding to each diastereomer. This clearly confirmed the optical purity of thioformamides.
- (48) The mechanism of the dehydrosulfurization of thioformamides using CNBr is given in Scheme 3.



Scheme 3

(49) Typical Experimental Procedure for **1a–j**

To a stirred solution of *N*<sup>β</sup>-Fmoc amino alkyl formamide **5** (1 mmol) in THF (5 mL), P<sub>2</sub>S<sub>5</sub> (0.7 mmol) was added. The reaction mixture was subjected to ultrasonication for 15 min. After the completion of reaction (TLC), the solvent was evaporated in vacuo, and the crude was purified by a flash chromatography to obtain the thioformamides as solids.

## (50) Selected Spectroscopic Data.

Compound **1b**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.30 (d, 3 H, *J* = 6.53 Hz), 2.65 (d, 2 H), 3.90 (m, 1 H), 4.20 (t, 1 H, *J* = 13.04 Hz), 4.41 (d, 2 H, *J* = 6.62 Hz), 5.01 (br, 1 H), 7.31 (t, 2 H, *J* = 14.79 Hz), 7.39 (t, 2 H, *J* = 14.69 Hz), 7.57 (d, 2 H, *J* = 7.21 Hz), 7.76 (d, 2 H, *J* = 7.48 Hz), 8.20 (s, 1 H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ = 17.44, 46.83, 47.18, 56.18, 66.83, 122.24, 124.93, 127.08, 128.17, 141.34, 143.66, 155.77, 139.31.

Compound **1j**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 2.39–2.65 (m, 2 H), 3.12–3.51 (m, 2 H), 3.1 (m, 1 H), 4.10 (t, 1 H, *J* = 6.90 Hz), 4.12 (d, 2 H, *J* = 7.2 Hz), 4.74 (s, 2 H), 5.56 (br, 1 H), 7.15–7.80 (m, 13 H), 8.25 (s, 1 H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>; CCl<sub>4</sub>): δ = 36.91, 43.06, 46.82, 51.89, 65.83, 69.82, 119.15, 124.90, 126.25, 127.53, 128.12, 128.24, 141.67, 143.53, 143.81, 156.38, 139.22, 171.58.

Compound **1k**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ = 1.14 (d, 3

H,  $J = 5.0$  Hz), 2.65–3.16 (m, 2 H), 4.05–4.20 (m, 3 H), 4.22 (d, 2 H,  $J = 8.0$  Hz), 6.65 (br, 1 H), 6.88 (br, 1 H), 7.28–7.48 (m, 8 H), 7.56 (d, 2 H,  $J = 4.0$  Hz), 7.74 (d, 2 H,  $J = 8.0$  Hz), 8.16 (s, 1 H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 17.12, 43.09, 45.86, 53.69, 58.29, 67.18, 125.34, 126.22, 127.09, 127.77, 128.01, 128.40, 128.63, 135.54, 141.30, 143.75, 159.14, 156.29, 138.9$ .

(51) **Typical Experimental Procedure for 2a–j**

A solution of  $N^\beta$ -Fmoc amino alkyl thioformamide **1** (1 mmol) in dry  $\text{CH}_2\text{Cl}_2$  (10 mL) was cooled to 0 °C, NMM (2 mmol) and CNBr (1.5 mmol) were added, and the reaction mixture was stirred at this temperature for 30 min. After completion of reaction, it was diluted with  $\text{CH}_2\text{Cl}_2$  (10 mL) and was washed with  $\text{H}_2\text{O}$ , brine, and dried over anhyd  $\text{Na}_2\text{SO}_4$ . The solvent was evaporated under reduced pressure followed by chromatographic purification (silica gel, 100–200 mesh, 20% EtOAc in hexane) to afford the desired isonitriles **2** in excellent yield and purity as stable solids.

(52) **Selected Spectroscopic Data**

Compound **2c**: IR (KBr):  $\nu_{\text{max}} = 1715, 2149 \text{ cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 2.81$  (br, 2 H), 3.12–3.65 (m, 2 H), 3.95 (br, 1 H), 4.12 (br, 1 H), 4.29 (d, 2 H,  $J = 4.0$  Hz), 5.15

(br, 1 H), 7.17 (m, 9 H), 7.44 (d, 2 H,  $J = 8.0$  Hz), 7.69 (d, 2 H,  $J = 8.0$  Hz).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 37.30, 44.39, 47.19, 50.79, 66.99, 120.10, 125.04, 127.15, 127.35, 127.84, 129.03, 129.10, 135.98, 141.38, 143.65, 155.55, 158.45$ .

Compound **2f**: IR (KBr)  $\nu_{\text{max}} = 1710, 2151 \text{ cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 2.20$ – $2.71$  (m, 2 H), 3.00–3.59 (m, 4 H), 4.00 (br, 1 H), 4.12 (d, 1 H,  $J = 7.8$  Hz), 4.41 (br, 2 H), 6.02 (s, 1 H), 7.00–7.74 (m, 13 H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 32.10, 36.21, 44.00, 46.85, 49.12, 53.34, 66.88, 119.90, 124.82, 126.95, 127.08, 127.83, 128.15, 136.77, 141.00, 143.13, 155.11, 158.10$ .

Compound **2l**: IR (film/pellet)  $\nu_{\text{max}} = 1715, 2151 \text{ cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta = 1.24$  (d, 3 H,  $J = 4.0$  Hz), 3.12–3.79 (m, 2 H), 3.87–4.10 (m, 3 H), 4.11 (d, 2 H,  $J = 8.0$  Hz), 5.09 (br, 1 H), 5.89 (br, 1 H), 7.28–7.48 (m, 8 H), 7.56 (d, 2 H,  $J = 4.0$  Hz), 7.74 (d, 2 H,  $J = 8.0$  Hz), 8.18 (s, 1 H).  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta = 17.00, 43.69, 46.42, 54.10, 57.95, 67.10, 119.77, 125.45, 127.22, 127.75, 128.53, 128.40, 128.63, 135.09, 141.30, 143.75, 154.94, 156.49, 170.97$ .

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