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Synthesis of 2',3'-dideoxy-2'-monofluoromethyl azanucleosides

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Abstract—(2S,4S)-Methyl-*N-tert*-butoxycarbonyl-4-monofluoromethylpyroglutamate **6** was synthesized via a key dehydrofluorination followed by hydrogenation. Compound **6** was converted to (5S,3S)-*N*-benzyloxycarbonyl-5-*tert*-butyldimethylsilyloxymethyl-3monofluoromethyl-2-pyrrolidone **12** over four steps in 62% yield, which was used as a precursor for the synthesis of 2',3'-dideoxy-2'monofluoromethyl azanucleosides **17–18**.

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1. Introduction

In the last two decades, nucleosides and nucleotides, known to be DNA and RNA subunits, have become an important subject of research in the field of pharmaceutical science. Modification of the ribose moiety and base moiety of nucleosides exhibits two important ways to new high bioactive nucleoside analogues.¹ Recently, many modified sugar nucleosides,² including thionucleo-sides,³ carbocyclic nucleosides,⁴ and azanucleosides⁵ were synthesized and studied. In connection with special properties of fluorine, fluorinated nucleosides have received a great interest for their biological and medical applications.⁶ In fluorinated sugar nucleosides, fluorine stabilizes the glycosidic bond toward hydrolysis by alteration of the conformation of the sugar moiety.⁷ Some highly bioactive fluorinated nucleosides have been synthesized and used in cancer and herpes therapies, for example, 2'-deoxy-2',2'-difluorocytidine (Gemcitabine) has been approved as a drug for solid tumor treatment.⁸ Interestingly, sugar moleties of several high bioactive fluorinated nucleosides FMAU,⁹ FLT,¹⁰ FIAC,^{9b} F-ddC,¹¹ SFDC,¹² and DFDT¹³ contain a fluorine atom, which in some degree, reveals the special influences of monofluoro groups on the biological activities of nucleosides (Fig. 1). Azanucleosides, in which the oxygen atom of pyranose or furanose ring is replaced by nitrogen atom, represent an important class of modified nucleosides. The synthesis and biological activities

of azanucleosides were summarized by Yokoyama and Momotake.⁵ However, to the best of our knowledge, few fluorinated azanucleosides have been synthesized¹⁴ and up to date there is no corresponding report on the monofluorinated azanucleosides. Considering all these aspects and in connection with our studies on fluorinated sugar nucleosides; we decided to synthesize monofluorinated azanucleosides: 2',3'-dideoxy-2'-monofluoromethyl azanucleosides and investigate their biological activities.

2. Results and discussion

During the synthesis of 2',3'-dideoxy-2'-difluoromethyl azanucleosides,^{14b} we accidentally found that the dehydrofluorination reaction occurred when the amino group of 5-*tert*-butyldimethylsilyloxymethyl-3-difluoromethyl-pyrrolidin-2-one **1** was protected with *tert*-but-oxycarbonyl group (Boc) (Scheme 1). The dehydrofluorination was induced by the enhancement of acidity of 3-H as a result of the protection of the amino groups with electron-withdrawing group (Boc). In our opinion, compound **2** could be used as a precursor for the preparation of target molecules: 2',3'-dideoxy-2'-monofluoromethyl azanucleosides.

So, according to our previous reports, 14b,15 the methyl (2*S*)-*N*-tert-butoxycarbonyl-4-difluoromethylpyroglutamate **4** were synthesized from the *trans*-4-hydroxy-Lproline **3** in 22% yield over six steps (Scheme 2). Dehydrofluorination of the compound **4** under the

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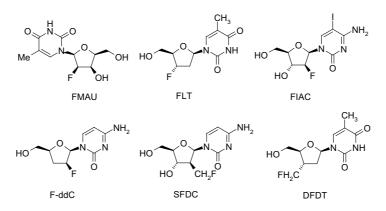
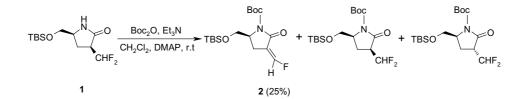
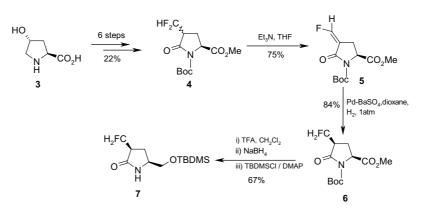


Figure 1. Several high bioactive monofluorinated nucleosides.



Scheme 1.

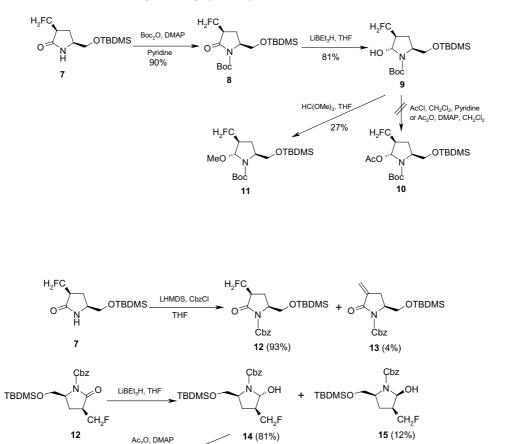


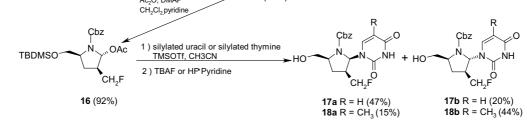
Scheme 2.

optimized condition (Et₃N, THF) smoothly provided the monofluoro olefin **5** in 75% yield. Hydrogenation of the compound **5** with Pd–BaSO₄ in dioxane gave the desired product **6** in 84% yield. Then deprotection of *N*-Boc group of **6** with trifluoroacetic acid in CH₂Cl₂ followed by reduction of the ester with NaBH₄, and further protection of the resulting hydroxyl group with TBDMSCl afforded the compound **7** in 67% yield over three steps.

Treatment of the lactam 7 with Boc₂O/pyridine/CH₂Cl₂ under the catalyst of DMAP provided (5*S*,3*S*)-*N*-tertbutoxycarbonyl-5-tert-butyldimethylsilyloxymethyl-3monofluoromethyl-2-pyrrolidione **8** in 90% yield and a few defluorinated byproduct could be detected by TLC (Scheme 3). Then, reduction of the compound **8** with Li-BEt₃H in THF at -78 °C yielded the desired compound **9** in 81% yield. However, acetylation of compound **9** under two reaction conditions (Ac₂O/DMAP/pyridine/CH₂Cl₂ and AcCl/pyridine/CH₂Cl₂) failed to give the desired compound **10**. Protection of hydroxyl group of **9** with methyl group only gave the desired product **11** in 27% yield.¹⁶ In our opinion, the big block of the *tert*-butoxycarbonyl group along with the existence of monofluoromethyl group may be responsible for the failure of the acetylation reaction and the low yield in preparation of the compound **11**.

In view of above failure, the protection of the amino group of compound 7 by benzyloxycarbonyl (Cbz) group instead of Boc group was investigated. We were pleased to find that treatment of 7 with CbzCl and 1.0 equiv LHMDS in THF at -78 °C for 10 min smoothly gave the desired lactam 12 in 93% yield along





Scheme 4.

Scheme 3.

with the dehydrofluorination product 13 in 4% yield (Scheme 4).¹⁷ Similarly, reduction of lactam 12 with LiBEt₃H yielded two diastereoisomers 14 (81% yield) and 15 (12% yield) (14/15 = 6.8:1.0). The two diastereoisomers could be separated on silica gel chromatography. As we expected, O-acetylation of the major diastereoisomer 14 with Ac₂O/DMAP/pyridine smoothly afforded the desired compound 16 in 92% yield. Finally, coupling of 16 with silvlated uracil and thymine under Vorbrüggen's condition (glycosylation reaction)¹⁸ gave the silyl-protected azanucluosides. Deprotection of the silvl protection groups with TBAF or HF pyridine afforded target molecules 17a, 17b, 18a, and 18b. The ratio of 17a/17b and that of 18a/18b were different and opposite. The absolute configuration of compound 17b was confirmed by X-ray (Fig. 2).¹⁹

We investigated the biological activities of the target molecules 17a, 17b, and 18b to the tumor cell U2OS. The experiments were carried out in the effects of 17a, 17b, and 18b on the cell life cycle of U2OS. Mimosine

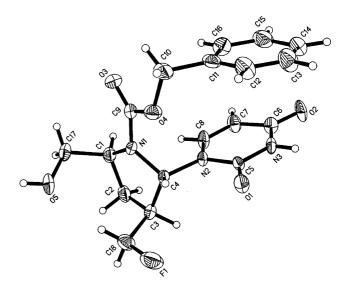


Figure 2. ORTEP drawing of the X-ray crystallographic structure of 17b.

 Table 1. Effects of azanucleosides 17a, 17b, and 18b to the cell cycles of U2OS

Cell cycles	Control	Mimosine	17a	17b	18b
Dip G1 (%)	46.20	76.78	47.18	49.16	49.16
Dip S (%)	43.23	20.93	45.33	41.73	41.73
Dip G2/M (%)	10.58	2.29	7.49	9.11	9.11

 $(500 \,\mu\text{M})$ was used as S phase block drug in control experiment and the sample concentrations (17a, 17b, and 18b) were 10 mmol/L. The biological results were summarized in Table 1. As shown in Table 1, the cell cycles had no evident change after azanucleosides 17a, 17b, and 18b were injected. So, none of azanucleosides 17a, 17b, and 18b exhibited a significant biological effect on the tumor cell U2OS.

In summary, we synthesized the novel fluorinated azanucleosides: 2',3'-dideoxy-2'-monofluoromethyl azanucleosides 17–18 from *trans*-4-hydroxy-L-proline via a novel dehydrofluorination found accidentally. Active tests shown that none of the synthesized azanucleosides 17a, 17b, and 18b had significant activity against U2OS. The related modified fluorinated analogues are in intensive synthesis and to be reported soon.

3. Experimental

3.1. (2*S*)-Methyl-*N*-*tert*-butoxycarbonyl-4-[(*Z*)-fluoromethylidene]-pyroglutamate (5)

To a solution of 4 (3.162g, 10.79 mmol) in CH_3CN (100 mL), Et₃N (7.0 mL) was added dropwise at room temperature. The mixture was stirred overnight at room temperature. Removal of the solvent in vacuo gave a residue and H_2O (50mL) and CH_2Cl_2 (50mL) were added. The organic phase was separated and the aqueous phase was extracted with CH_2Cl_2 (2×100 mL). The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent in vacuo, the resulting residue was purified by silica gel chromatography (hexane/ethyl acetate, 6:1) to give 5 as a white solid (2.082g, 75%): mp 118–119°C; $[\alpha]_{D}^{20}$ –8.6 (c 1.03, CHCl₃); ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3)^{\circ} \delta$ 7.46 (dt, J = 78.3, 2.7 Hz, 1 H), 4.66 (dd, J = 3.3, 3.6 Hz, 1H), 3.80 (s, 3H), 3.11–3.00 (m, 1H), 2.76 (dq, J = 18.0, 3.0 Hz, 1H), 1.51 (s, 9H); ¹⁹F NMR (282 MHz, CDCl₃) δ -119.76 to -120.07 $(dt, J = 78.5, 3.7 \text{ Hz}, 1\text{F}); {}^{13}\text{C}$ NMR $(75.5 \text{ MHz}, \text{CDCl}_3)$ δ 170.9, 165.0 (d, J = 19.3 Hz), 154.9 (d, J = 276.1 Hz), 149.0, 114.4 (d, J = 12.7 Hz), 83.7, 55.8, 52.4, 29.4, 27.6, 22.7 (d, J = 1.5 Hz). IR (thin film) 3109, 1790, 1747, 1706, 1687, 1288, 1152 cm⁻¹; MS (ESI) *m*/*z* 296.0 (M^++Na) ; Anal. Calcd for $C_{12}H_{16}FNO_5$: C, 52.75; H, 5.86; N, 5.13. Found: C, 52.82; H, 5.89; N, 5.10.

3.2. (2*S*,4*S*)-Methyl-*N*-*tert*-butoxycarbonyl-4-monofluoromethylpyroglutamate (6)

To a solution of compound 5 (1.028 g, 3.77 mmol) in dioxane (60 mL), Pd–BaSO₄ (250 mg) was added. Then,

the mixture was hydrogenated at room temperature for 24h. Filtration and removal of the solvent gave a residue, which was purified by silica gel chromatography (hexane/ethyl acetate, 8:1 then 6:1) to give **6** as a white solid (871mg, 84%): mp 83–84°C; $[\alpha]_D^{20}$ –29.3 (*c* 0.96, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.72 (ddd, J = 26.1, 6.0, 5.7Hz, 1H), 4.65–4.52 (m, 2H), 3.79 (s, 3H), 3.03–2.85 (m, 1H), 2.64–2.53 (m, 1H), 2.12–2.03 (m, 1H), 1.50 (s, 9H); ¹⁹F NMR (282 MHz, CDCl₃) δ –228.01 to –228.44 (m, 1F); ¹³C NMR (75.5 MHz, CDCl₃) δ 171.4, 171.2 (d, J = 8.8 Hz), 148.9, 83.9, 81.7 (d, J = 170.1 Hz), 57.2, 52.4, 43.7 (d, J = 21.1 Hz), 29.5, 27.7, 23.7 (d, J = 3.2 Hz); IR (thin film) 1776, 1745, 1702, 1339, 1296 cm⁻¹; MS (EI) *m/z* 276 (M⁺+1, <1), 260 (M⁺-15, <1), 57 (100); MS (ESI) *m/z* 298 (M⁺+Na); EI-HRMS *m/z* 260.09538 (M⁺-CH₃, C₁₁H₁₅FNO₅ required 260.09343).

3.3. (5*S*,3*S*)-5-*tert*-Butyldimethylsilyloxymethyl-3-mono-fluoromethylpyrolidin-2-one (7)

TFA (1.0 mL) was added dropwise to a solution of 6 (813 mg, 2.96 mmol) in CH_2Cl_2 (30 mL) at $-78^{\circ}C$. The mixture was then warmed up to room temperature and stirred overnight. The reaction mixture was cooled to 0°C and quenched with saturated aqueous NaHCO₃ (10 mL). The organic phase was separated and the aqueous phase was extracted with CH_2Cl_2 (3×40 mL). The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After removal of the solvent in vacuo, the residue was dissolved in MeOH (30 mL) and the solution was cooled to -78 °C. NaBH₄ (145 mg, 3.82 mmol) was added. The mixture was then warmed up to 0°C and stirred for 1.5h. The reaction was quenched with concentrated HCl. The mixture was then filtered and concentrated to give a yellowish solid. To a cooled solution of DMAP (26mg, 0.21 mmol) and imidazole (900 mg, 13.24 mmol) in CH_2Cl_2 (50mL), a solution of above yellowish solid in DMF (1mL) was added. Then, a solution of TBDMSCl (2.0 g, 13.27 mmol) in CH₂Cl₂ (10 mL) was added dropwise. After the mixture was stirred overnight at room temperature, it was quenched with saturated aqueous NH₄Cl (10mL) in cooled ice-bath. The organic phase was separated and the aqueous layer was extracted with CH_2Cl_2 (3 × 40 mL). The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After removal of the solvent in vacuo, the resulting residue was purified by flash chromatography (hexane/ ethyl acetate, 3:2) to give 7 as a clear oil (516mg, 67%). [α]_D²⁰ +51.4 (*c* 1.29, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 6.75 (br, 1H), 4.60 (ddd, J = 92.5, 4.2, 4.2 Hz, 1H), 4.62-4.60 (m, 1H), 3.76-3.68 (m, 1H), 3.62 (dd, J = 3.9, 3.9 Hz, 1H), 3.42 (dd, J = 7.8, 7.8 Hz,1H), 2.82-2.63 (m, 1H), 2.32 (ddd, J = 15.3, 7.2, 6.0 Hz, 1H), 1.70 (ddd, J = 14.7, 7.5, 3.6 Hz, 1H), 0.85 (s, 9H), 0.02 (2s, 6H); ¹⁹F NMR (282 MHz, CDCl₃) δ -228.52 to -228.96 (m, 1F); ¹³C NMR (75.5 MHz, CDCl₃) δ 175.4 (d, J = 7.5 Hz), 82.3 (d, J = 169.7 Hz), 66.7, 53.9, 42.6 (d, J = 21.7 Hz), 25.7, 25.3 (d, J = 3.2 Hz, 18.1 (d, J = 0.5 Hz), -5.6 (d, J = 0.8 Hz); IR (thin film) 3223, 1708, 1464, 1256, 1104 cm⁻¹; MS (EI) m/z 262 (M⁺+1, 3), 260 (M⁺-1, <1), 246 (M⁺-

15, 2), 110 (100); EI-HRMS m/z 246.13041 (M⁺-CH₃, C₁₁H₂₁FNO₂Si required 246.13256).

3.4. (5*S*,3*S*)-*N*-*tert*-Butoxycarbonyl-5-*tert*-butyldimethyl-silyloxymethyl-3-monofluoromethyl-2-pyrrolidone (8)

To a cooled solution of 7 (178 mg, 0.68 mmol), DMAP (5 mg, 0.04 mmol), and pyridine (0.31 mL) in CH_2Cl_2 (10mL), Boc₂O (500mg, 2.29mmol) in CH₂Cl₂ (5mL) was added dropwise. The mixture was warmed up to room temperature and stirred for 3h. Then the reaction was quenched with H₂O and the organic phase was separated. The aqueous layer was extracted with CH₂Cl₂ $(2 \times 30 \text{ mL})$ and the combined organic phases were washed with diluted aqueous HCl, brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent in vacuo, the resulting residue was purified by flash chromatography (hexane/ethyl acetate, 7:1) to give **8** as a white solid (222mg, 90%): mp 65–66 °C; $[\alpha]_D^{20}$ -53.2 (c 0.87, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 4.76 (ddd, J = 23.3, 5.1, 5.7 Hz, 1H), 4.59 (ddd, J = 22.5, 5.7, 5.4 Hz, 1H), 4.18–4.11 (m, 1H), 3.92 (dd, $J = 4.8, 4.8 \,\text{Hz}, 1 \text{H}$), 3.72 (dd, $J = 2.4, 2.7 \,\text{Hz}, 1 \text{H}$), 3.01-2.83 (m, 1H), 2.32 (ddd, J = 16.5, 8.9, 2.3 Hz, 1H), 2.09 (ddd, J = 13.7, 5.1, 5.3Hz, 1H), 1.54 (s, 9H), 0.88 (s, 9H), 0.05 (2s, 6H); ¹⁹F NMR (282 MHz, CDCl₃) δ -226.23 to -226.65 (m, 1F); ¹³C NMR (75.5 MHz, CDCl₃) δ 172.3 (d, J = 10.1 Hz), 150.0, 83.1, 83.0 (d, J = 170.2 Hz), 63.4 (d, J = 0.6 Hz), 57.1, 43.9 (d, J = 21.7 Hz), 28.0, 25.8, 22.7 (d, J = 2.5 Hz), 18.3, -5.5 (d, J = 0.8 Hz); IR (thin film) 1765, 1689, 1473, 1369, 836, $779 \,\mathrm{cm}^{-1}$; MS (EI) *m/z* 288 (M⁺-C₄H₉O, 7), 248 (51), 204 (100); MS (ESI) m/z 384.2 (M⁺+Na); ESI-HRMS *m*/*z* 384.19792 (M⁺+Na, C₁₇H₃₂FNO₄SiNa required 384.19768).

3.5. *tert*-Butyl (2*R*,3*S*,5*S*)-5-*tert*-butyldimethylsilyloxymethyl-3-monofluoromethyl-2-hydroxypyrrolidine-1-carboxylate (9)

To a solution of 8 (222 mg, 0.62 mmol) in THF (10 mL) at -78°C was added LiBEt₃H (1.70mL, 1M in THF, 1.70 mmol) dropwise. The mixture was stirred at -78 °C for 30 min. The reaction was then quenched with $H_2O(3mL)$ and warmed up to room temperature. H_2O (10mL) was added and the mixture was extracted with CH_2Cl_2 (3 × 40 mL). The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent, the resulting residue was purified by flash chromatography (hexane/ ethyl acetate, 20:1, then 15:1) to give 9 as a clear oil (181 mg, 81%). $[\alpha]_{\rm D}^{20}$ -51.2 (c 1.05, acetone); ¹H NMR (300 MHz, acetone- d_6) δ 5.44 (d, J = 12.6 Hz, 1H), 4.61 (dt, J = 46.5, 8.1 Hz, 1H), 4.40 (ddd, J = 47.1, 5.7, 6.0 Hz, 1 H), 3.95 - 3.54 (m, 3H), 2.47 - 2.30(m, 1H), 2.23–2.10 (m, 1H), 1.79–1.65 (m, 1H), 1.44 (s, 9H), 0.89 (s, 9H), 0.08, 0.07 (2s, 6H); ¹⁹F NMR (282 MHz, acetone- d_6) δ -221.72 to -222.34 (m, 1F); ¹³C NMR (75.5 MHz, acetone- d_6) δ 153.3 (d., $J = 37.0 \,\mathrm{Hz}$, 82.6 (d, $J = 81.6 \,\mathrm{Hz}$), 80.7 (d, $J = 5.7 \,\mathrm{Hz}$), 79.4, 65.4, 63.6, 57.9, 43.5, 27.7, 27.1, 25.4, 17.9. IR (thin film) 3441, 1700, 1474, 1390, 839 cm⁻¹; MS (ESI) *m/z* 386.2 (M^+ +Na); Anal. Calcd for $C_{17}H_{34}FNO_4Si$: C,

56.20; H, 9.37; N, 3.86. Found: C, 56.35; H, 9.71; N, 3.70.

3.6. *tert*-Butyl (2*R*,3*S*,5*S*)-2-methoxy-5-*tert*-butyldimethylsilyloxy-3-monofluoromethylpyrrolidine-1-carboxylate (11)

To a cooled mixture of 9 (86 mg, 0.24 mmol) and 4Å MS (131 mg, powder) in Et₂O (5mL), CH(OMe)₃ (0.06mL) was added dropwise followed by BF₃·Et₂O (10 µL, 1 M in Et_2O , 0.01 mmol). The mixture was then stirred at room temperature for 20min. The reaction was quenched with H₂O and extracted with Et₂O. The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. Filtration and removal of the solvent gave a residue, which was purified by flash chromatography to give 11 as an oil (24 mg, 27%). $\left[\alpha\right]_{D}^{20}$ -59.3 (c 0.87, acetone); ¹H NMR (300 MHz, acetone- d_6) δ 5.14 (br, 1H), 4.67–4.29 (m, 2H), 3.97–3.82 (m, 2H), 3.47 (br, 1H), 3.27 (2s, 3H), 2.51–2.34 (m, 1H), 2.15 (br, 1H), 1.74-1.63 (m, 1H), 1.46 (s, 9H), 0.89 (s, 9H), 0.07, 0.05 (2s, 6H); ¹⁹F NMR (282 MHz, acetone- d_6) δ -221.04 to -221.63 (m, 1F); ¹³C NMR (75.5 MHz, acetone- d_6) δ 154.2 (d, J = 32.8 Hz), 88.4 (d, J = 5.0 Hz), 82.3 (d, J = 162.0 Hz), 79.5, 65.6, 64.8, 58.5, 54.6, 43.4, 27.6, 25.3, 17.8, -6.1; IR (thin film) 1705, 1473, 1381, 839 cm⁻¹; MS (ESI) *m*/*z* 400.2 (M⁺+Na); ESI-HRMS m/z 400.22817 (M⁺+Na, C₁₈H₃₆FNO₄SiNa required 400.22898).

3.7. (5*S*,3*S*)-*N*-Benzyloxycarbonyl-5-*tert*-butyldimethylsilyloxymethyl-3-monofluoromethyl-2-pyrrolidone (12) and (5*S*)-*N*-benzyloxycarbonyl-5-*tert*-butyldimethylsilyloxymethyl-3-methylene-2-pyrrolidone (13)

To a solution of 7 (516mg, 2.00mmol) in THF (20mL) at -78°C was added LHMDS (2.0mL, 1M in THF, 2.0 mmol) dropwise. After the mixture was stirred for 10min, CbzCl (0.45mL, 3.16mmol) was added dropwise. The mixture was stirred for further 10min at -78 °C and quenched with saturated aqueous NH₄Cl (5mL). The mixture was then warmed up to room temperature and extracted with EtOAc $(3 \times 40 \text{ mL})$. The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent, the resulting residue was purified by flash chromatography (hexane/ethyl acetate, 10:1) to give 13 as a white solid (less polar, 28 mg, 4%) and 12 as a white solid (more polar, 727 mg, 93%). Compound 12: mp 80–82 °C; $[\alpha]_D^{20}$ –57.2 (*c* 0.84, CHCl₃); ¹H NMR (300 MHz, CDCl₃) δ 7.50–7.32 (m, 5H), 5.34–5.23 (m, 2H), 4.76 (ddd, J = 28.1, 5.3, 5.0 Hz, 1H), 4.61 (ddd, J = 27.2, 5.3, 5.4 Hz, 1H), 4.24–4.17 (m, 1H), 3.90 (dd, J = 4.8, 4.8 Hz, 1H), 3.70 (dd, J = 2.7, 2.4 Hz, 1H), 3.02-2.84 (m, 1H), 2.40-2.32 (m, 1H), 2.17–2.04 (m, 1H), 0.84 (s, 9H), -0.02, -0.03 (2s, 6H); ¹⁹F NMR (282 MHz, CDCl₃) δ –226.31 to -226.72 (m, 1F); IR (thin film) 2955, 1714, 1289, 834 cm⁻¹; MS (ESI) *m*/*z* 418.2 (M⁺+Na); Anal. Calcd for C₂₀H₃₀FNO₄Si: C, 60.76; H, 7.59; N, 3.54. Found: C, 60.70; H, 7.87; N, 3.40. Compound 13: mp 41-43 °C; $[\alpha]_{D}^{20}$ -33.8 (*c* 0.66, CHCl₃); ¹H NMR $(300 \text{ MHz}, \text{ CDCl}_3) \delta$ 7.47–7.32 (m, 5H), 6.16 (t,

J = 2.4 Hz, 1H), 5.46 (t, J = 2.1 Hz, 1H), 5.37–5.23 (m, 2H), 4.28–4.22 (m, 1H), 3.77 (dd, J = 4.8, 4.5 Hz, 1H), 3.69–3.66 (dd, J = 2.4, 2.7 Hz, 1H), 2.81–2.77 (m, 2H), 0.81 (s, 9H), -0.04, -0.07 (2s, 6H); ¹³C NMR (75.5 MHz, CDCl₃) δ 166.2, 152.1, 139.0, 135.4, 128.6, 128.3, 128.1, 119.3, 68.1, 63.7, 55.7, 27.8, 25.7, 18.1, -5.7; IR (thin film) 2956, 2929, 1739, 1700, 1658, 1301, 1014, 838 cm⁻¹; MS (ESI) *m*/*z* 376.2 (M⁺+1); ESI-HRMS *m*/*z* 398.17538 (M⁺+Na, C₂₀H₂₉NO₇SiNa required 398.17581).

3.8. Benzyl (2*R*,3*S*,5*S*)-5-*tert*-butyldimethylsilyloxymethyl-3-monofluoromethyl-2-hydroxypyrrolidine-1-carboxylate (14) and benzyl (2*S*,3*S*,5*S*)-5-*tert*-butyldimethylsilyloxymethyl-3-monofluoromethyl-2-hydroxypyrrolidine-1carboxylate (15)

To a solution of 12 (727 mg, 1.84 mmol) in THF (25 mL) at -78°C was added LiBEt₃H (6.0mL, 1M in THF, 6.0 mmol) dropwise. The mixture was stirred at -78 °C for 1 h. The reaction was quenched with H_2O (10mL) and warmed up to room temperature. The mixture was extracted with $Et_2O(3 \times 40 \text{ mL})$ and the combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent, the resulting residue was purified by flash chromatography (hexane/ethyl acetate, 5:1) to give 14 (less polar) as a clear oil (589mg, 81%) and 15 (more polar) as a clear oil (90 mg, 12%). Compound 14: $[\alpha]_D^{20}$ -48.1 (c 0.76, acetone); ¹H NMR (300 MHz, acetone d_6) δ 7.42–7.31 (m, 5H), 5.56 (t, J = 5.4 Hz, 1H), 5.21– 5.05 (m, 2H), 4.75–4.34 (m, 3H), 4.02–3.90 (m, 2H), 3.78-3.56 (br, 1H), 2.51-2.42 (m, 1H), 2.27-2.15 (m, 1H), 1.82–1.70 (m, 1H), 0.90 (s, 9H), 0.08, 0.01 (2s, 6H); ¹⁹F NMR (282 MHz, acetone- d_6) δ –221.99 to -222.37 (m, 1F); IR (thin film) 3429, 2956, 1708, 1412, 1101, 838 cm^{-1} ; MS (ESI) *m*/*z* 380.2 (M⁺-OH); MS (MALDI) m/z 420.2 (M⁺+Na); Anal. Calcd for C₂₀H₃₂FNO₄Si: C, 60.45; H, 8.06; N, 3.53. Found: C, 60.87; H, 8.20; N, 3.85. Compound **15**: $[\alpha]_D^{20}$ -47.6 (*c* 0.47, acetone); ¹H NMR (300 MHz, acetone-*d*₆) δ 7.44-7.34 (m, 5H), 5.45-5.07 (m, 4H), 4.61-4.34 (m, 2H), 3.99-3.60 (m, 3H), 2.46-2.36 (m, 2H), 1.79 (br, 1H), 0.88 (s, 9H), 0.04 (s, 6H); ¹⁹F NMR (282 MHz, acetone- d_6) δ -218.77 to -220.47 (m, 1F); IR (thin film) 3426, 2956, 1710, 1409, 1101, 838 cm^{-1} ; MS (ESI) m/z420.1 (M^+ +Na), 398.3 (M^+ +1), 380.2 (M^+ -OH); Anal. Calcd for C₂₀H₃₂FNO₄Si: C, 60.45; H, 8.06; N, 3.53. Found: C, 60.60; H, 8.37; N, 3.42.

3.9. Benzyl (2*R*,3*S*,5*S*)-2-acetyloxy-5-*tert*-butyldimethylsilyloxymethyl-3-monofluoromethylpyrrolidine-1-carboxylate (16)

To a mixture of 14 (589 mg, 1.48 mmol), DMAP (16 mg, 0.13 mmol), pyridine (2.0 mL, 24.56 mmol) in CH₂Cl₂ (25 mL) was added Ac₂O (1.30 mL, 13.89 mmol). After the mixture was stirred overnight at room temperature, the reaction was quenched with H₂O (10 mL) and extracted with CH₂Cl₂ (3×30 mL). The combined organic phases were washed with dilute HCl, brine and dried over anhydrous Na₂SO₄. After filtration and removal of the solvent, the resulting residue was purified by flash

chromatography (hexane/ethyl acetate, 5:1) to afford **16** as a white solid (597 mg, 92%): mp 74–76 °C; $[\alpha]_D^{20}$ –53.2 (*c* 0.98, acetone); ¹H NMR (300 MHz, acetone-*d*₆) δ 7.39–7.34 (m, 5H), 6.78 (d, *J* = 5.1 Hz, 1H), 5.14 (br, 2H), 4.55 (dd, *J* = 4.2, 3.0 Hz, 1H), 4.39 (dd, *J* = 3.6, 3.6 Hz, 1H), 3.96–3.54 (m, 3H), 2.76–2.65 (m, 1H), 2.27–2.15 (m, 1H), 2.04–1.87 (m, 4H), 0.90 (s, 9H), 0.03 (s, 6H); ¹⁹F NMR (282 MHz, acetone-*d*₆) δ –223.55 to –223.87 (m, 1F); IR (thin film) 2937, 1736, 1716, 1401, 1213, 1086, 837 cm⁻¹; MS (ESI) *m*/*z* 478.2 (M⁺+K), 462.2 (M⁺+Na); Anal. Calcd for C₂₂H₃₄FNO₅Si: C, 60.14; H, 7.74; N, 3.19. Found: C, 60.27; H, 7.73; N, 2.98.

3.10. Benzyl (2S,3S,5S)-5-hydroxymethyl-2-[2,4-dioxo-3,4-dihydro-pyrimidin-1(2*H*)-yl]-3-(monofluoromethyl)pyrrolidine-1-carboxylate (17a) and benzyl (2R,3S,5S)-5hydroxymethyl-2-[2,4-dioxo-3,4-dihydro-pyrimidin-1(2*H*)-yl]-3-(monofluoromethyl)-pyrrolidine-1-carboxylate (17b)

Typical procedure: To a stirred solution of 16 (128 mg, 0.29 mmol) and uracil (96 mg, 0.86 mmol) in anhydrous acetonitrile (30 mL) was added N,O-bis(trimethylsilyl)acetamide (0.44 mL, 1.33 mmol). The reaction mixture was stirred under reflux for 30 min. After cooled to 0°C, TMSOTf (0.15mL, 0.72mmol) was added dropwise and the solution was stirred at room temperature for further 40 min. The reaction was quenched with cold saturated aqueous NaHCO₃ (6mL) and the resulting mixture was extracted with CH₂Cl₂ $(3 \times 40 \text{ mL})$. The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After removal of the solvent, the resulting residue was purified by flash chromatography (hexane/ethyl acetate, 3:2) to give two compounds, the less polar compound (75 mg, white foam) and more polar compound (34mg, white foam). A stirred solution of above less polar compound (42mg) in THF (4.5mL) was treated with 1.0M solution of TBAF (0.11 mL, 0.11 mmol) at 0 °C. After stirred at room temperature for 8.5h, the reaction was quenched with H₂O and the mixture was extracted with CH_2Cl_2 (3 × 20 mL). The combined organic phases were washed with brine and dried over anhydrous Na₂SO₄. After removal of the solvent, the resulting residue was purified by flash chromatography (hexane/ethyl acetate, 1:3) to give **17a** as a white foam (29 mg, 47% from **16**). The more polar compound (34mg) was also treated with TBAF under the same conditions to give 17b as a white foam (22mg, 20% from 16). Deprotection of the silyl protection groups was also conveniently achieved using HF pyridine in CH₂Cl₂. Compound **17a**: $[\alpha]_{D}^{20}$ -5.3 (*c* 0.66, CHCl₃); ¹H NMR (300 MHz, MeOH- d_4) δ 8.46 (br, 1H), 7.29 (br, 5H), 6.44 (d, J = 7.2 Hz, 1 H), 5.56 (d, J = 7.8 Hz, 1 H), 5.21–5.17 (m, 1H), 5.10-4.97 (br, 1H), 4.56 (ddd, J = 25.0, 4.2, 4.2 Hz, 1H), 4.47–4.34 (m, 2H), 3.92 (t, J = 9.0 Hz, 1H), 3.66 (d, J = 11.4 Hz, 1H), 2.93–2.74 (m, 1H), 2.33–2.18 (m, 1H), 2.10–2.00 (m, 1H); ¹⁹F NMR $(282 \text{ MHz}, \text{ MeOH-}d_4) \delta - 228.87 \text{ to } -229.45 \text{ (m, 1F)};$ ¹³C NMR (75.5 MHz, MeOH- d_4) δ 166.3, 156.5, 152.9, 144.0, 137.4, 129.6, 129.3, 128.9, 101.9, 82.1 (d, $J = 168.1 \,\mathrm{Hz}$, 72.2, 68.7, 61.8, 61.0, 43.5 (d,

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J = 19.5 Hz, 27.8; IR (thin film) 3448, 3199, 1685, 1466, 1400, 1275 cm^{-1} ; MS (EI) *m*/*z* 266 (M⁺-uracil, 14), 222 (16), 91 (100); MALDI-HRMS m/z 400.1319 $(M^++Na, C_{18}H_{20}N_3O_5FNa required 400.1279)$. Compound **17b**: $[\alpha]_D^{20} - 25.2$ (*c* 0.65, CHCl₃); ¹H NMR (300 MHz, MeOH- d_4) δ 7.54–7.23 (m, 6H), 5.86 (br, 1H), 5.42-5.30 (m, 1H), 5.11 (br, 1H), 4.84-4.80 (m, 1H), 4.62–4.59 (m, 1H), 4.47–4.44 (m, 1H), 4.22 (br, 1H), 3.99-3.79 (m, 1H), 3.67-3.63 (m, 1H), 2.73 (br, 1H), 2.48–2.37 (m, 1H), 1.88–1.75 (m, 1H); ¹⁹F NMR (282 MHz, MeOH- d_4) δ -222.53 to -222.65 (m, 1F); ¹³C NMR (75.5 MHz, MeOH- d_4) δ 166.0, 155.2, 151.9, 143.5, 137.2, 129.7, 129.6, 129.5, 102.9, 84.6 (d, J = 170.3 Hz), 75.8, 68.7, 62.8, 61.8, 46.1 (d, J = 18.5 Hz), 28.5; IR (thin film) 3510, 3152, 3033, 1677, 1627, 1461, 1402, 1262, $699 \,\mathrm{cm}^{-1}$; MS (EI) m/z346 (M^+ -CH₂OH, <1), 266 (M^+ -uracil, 17), 222 (16), 91 (100); MALDI-HRMS m/z 400.1301 $(M^++Na, C_{18}H_{20}N_3O_5FNa required 400.1279).$

3.11. Benzyl (2*S*,3*S*,5*S*)-5-hydroxymethyl-2-[5-methyl-2,4-dioxo-3,4-dihydro-pyrimidin-1(2*H*)-yl]-3-(monofluoro-methyl)-pyrrolidine-1-carboxylate (18a) and benzyl (2*R*,3*S*,5*S*)-5-hydroxymethyl-2-[5-methyl-2,4-dioxo-3,4-dihydro-pyrimidin-1(2*H*)-yl]-3-(monofluoromethyl)-pyrrolidine-1-carboxylate (18b)

Compounds 18a (27mg, 15%) and 18b (78mg, 44%) were prepared as white foams from compound 16 (119mg, 0.45mmol) and thymine (178mg, 1.41mmol) using the same conditions as described for compounds **17a** and **17b**. Compound **18a**: $[\alpha]_{D}^{20}$ +9.4 (*c* 0.72, CHCl₃); ¹H NMR (300 MHz, MeOH- d_4) δ 8.41 (br, 1H), 7.28 (br, 5H), 6.43 (d, J = 5.1 Hz, 1H), 5.23–4.99 (m, 2H), 4.60–4.33 (m, 3H), 3.90 (t, J = 9.0 Hz, 1H), 3.66 (d, J = 11.7 Hz, 1H), 2.93–2.74 (m, 1H), 2.34– 2.22 (m, 1H), 2.10–2.01 (m, 1H), 1.79 (s, 3H); ¹⁹F NMR (282 MHz, MeOH- d_4) δ -229.95 to -230.72 (m, 1F); ¹³C NMR (75.5 MHz, MeOH- d_4) δ 166.5, 156.3, 153.0, 139.9, 137.5, 129.6, 129.3, 128.8, 110.7, 82.1 (d, J = 168.1 Hz), 71.8, 68.6, 61.8, 60.8, 43.5 (d, J = 19.6 Hz), 27.7, 12.4; IR (thin film) 3448, 3192, 3064, 1685, 1472, 1404, 1273 cm^{-1} ; MS (EI) m/z391 (M^+ , <1), 266 (M^+ -thymine, 16), 222 (18), 91 (100); MALDI-HRMS m/z 414.1468 (M⁺+Na, $C_{19}H_{22}N_3O_5FNa$ required 414.1436). Compound **18b**: $[\alpha]_{D}^{22}$ -24.7 (*c* 0.62, CHCl₃); ¹H NMR (300 MHz, MeOH- d_4) δ 7.27 (m, 5H), 7.07 (br, 1H), 5.83 (br, 1H), 5.35-5.31 (d, J = 11.4 Hz, 1H), 4.78-4.75 (m, 1H), 4.60-4.55 (m, 1H), 4.44-4.43 (m, 1H), 4.24 (br, 1H), 3.99-3.95 (m, 1H), 3.79-3.63 (m, 1H), 2.72 (br, 1H), 2.47–2.37 (m, 1H), 1.86–1.64 (m, 4H); ¹⁹F NMR (282MHz, MeOH- d_4) δ -220.83 to -223.35 (m, 1F); ¹³C NMR (75.5MHz, MeOH- d_4) δ 166.2, 155.2, 152.0, 139.2, 137.1, 129.7, 129.6, 129.5, 111.8, 84.5 (d, J = 169.7 Hz), 76.7, 75.5, 68.6, 62.7, 46.0 (d, J = 18.6 Hz), 28.6, 12.4; IR (thin film) 3440, 3188, 3038, 1689, 1470, 1404 cm⁻¹; MS (EI) m/z 391 (M⁺, <1), 266 (M⁺-thymine, 19), 222 (21), 91 (100); MAL-DI-HRMS m/z 414.1474 (M⁺+Na, C₁₉H₂₂N₃O₅FNa required 414.1436).

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