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Synthesis of Thelepamide via Catalyst-Controlled 1,4-Addition of Cysteine Derivatives and Structure Revision of Thelepamide

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Supporting Information

ABSTRACT: The first enantioselective total synthesis and structural reassignment of (-)-thelepamide, a cytotoxic tetraketide—amino acid from the marine worm *Thelepus crispus*, is reported. A convergent approach provides access to all thelepamide diastereomers in six steps from four simple building blocks. Key features of the synthesis include the application of Melchiorre's organocatalytic thia-Michael reaction and a sonication-assisted assembly of an unprecedented *N*,*O*-acetal—hemiacetal moiety. The corrected structure was confirmed by NMR–DFT analysis.





Figure 1. Structures of (-)-thelepamide (1: proposed) and spongiacysteine (2).

The relative configurations of thelepamide's four stereogenic centers were established on the basis of ROESY, J-based configurational analysis, and by comparison of calculated and experimental NMR shifts using Goodman's DP4 method.⁵ Motivated by the unique 1,3-oxazolidin-2-one, the selective yet moderate biological activity against the leukemia cell line CCRF-CEM (IC₅₀ = 13.2 μ M), we decided to pursue its chemical synthesis.

In order to validate the structural assignment and probe structure-activity by synthetic analogues, a flexible synthetic access to thelepamide was sought. We identified the carbonsulfur bond at C3 in thelepamide as the key disconnection



between the tetraketide part of the molecule and a suitable cysteine derivative. In synthetic direction, two approaches to form this bond in a stereoselective fashion were considered. First, an $S_N 2$ substitution of a tosylate leaving group under inversion of configuration was investigated. As a second option, we envisaged an asymmetric catalyst-controlled thia-Michael addition⁶ of cysteine derivatives to (*E*)-oct-5-en-4-one and a subsequent diastereoselective reduction of the keto group.

The first synthetic route commenced with a Ru-catalyzed reduction⁷ of β -ketoester 3 followed by Weinreb amide formation⁸ to afford β -hydroxyamide 4 in 70% yield over two steps and 99% ee (Scheme 1). This material was subjected to ethylmagnesium bromide in Et₂O at 0 °C to give hydroxy ketone 5 in 64% yield.⁹ The C3 stereogenic center was generated using an Evans-Tishchenko reduction.¹⁰ The expected anti-configuration in 6 was confirmed using Rychnovsky's acetonide method.¹¹ After the free hydroxyl group had been converted to the corresponding tosylate 7, ¹² we carried out S_N^2 reactions with D- and L-N-Boc-Cys-OMe. Due to partial epimerization at C2' during the S_N2-reaction and problems occurring in the subsequent steps toward thelepamide, we decided to abandon this route in favor of the diastereoselective thia-Michael approach. Nevertheless, thioethers 9 and 10 turned out to be useful as reference standards for the following transformations.

In our second-generation approach, the synthetic sequence commenced with a stereoselective 1,4-addition of L-8 to known enone 11^{13} (Table 1). Enantioselective thia-Michael additions have been studied by Wynberg since the late 1970s.¹⁴ For our substrate, 20 mol % of unmodified cinchona alkaloids 12a-d gave only moderate selectivities using dichloromethane as the solvent (entries 1–4). With Takemoto's thiourea catalyst $12e^{15}$

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Scheme 1. First-Generation Route to Thelepamide

under the same conditions, the selectivity increased to 6.6:1 in favor of 13 (entry 5).¹⁶ Surprisingly, using the enantiomeric Takemoto catalyst 12f, no selectivity toward 14 could be induced (entry 6). In 2008, Melchiorre¹⁷ reported that 1:2 adducts of cinchona alkaloid amines¹⁸ and \hat{N} -Boc-phenylglycine (Phg) derivatives¹⁹ provide superior selectivities in thia-Michael reactions. Using their conditions at 0 °C provided 13 with 12.9:1 selectivity (entry 7). Lowering the temperature to -28 °C increased the selectivity to 24.5:1 but also raised the reaction time to 168 h (entry 8). Changing the amino acid to its enantiomer lowered the selectivity to 16.7:1, demonstrating its minor yet relevant influence (entry 9). Additionally, and in order to gain access to 14, we screened the catalysts based on the pseudoenantiomeric cinchona alkaloids. Catalyst 12h provided 14 in 9.3:1 ratio at 0 °C which increased to 12.9:1 at -28 °C (entries 10 and 11). The cinchonine-based catalyst 12i gave 14 in 12:1 selectivity at 0 $^{\circ}$ C and 17.4:1 at $-28 ^{\circ}$ C (entries 12 and 13). Again, the mismatching alkaloid-Phg combination lowered the selectivity to a 5:1 ratio (entry 14). On a preparative scale, 13 and 14 were obtained in 98% and 99% yield, respectively. It should be noted that this approach might also be useful in the stereoselective preparation of cysteine-S conjugates, which play an important role as precursor of plant aromas as well as in the formation of natural scents of cats and humans.²⁰

Having developed a catalyst-controlled access to both diastereomers 13 and 14, we then turned our attention to the diastereoselective reduction of the keto group and the completion of the synthesis of the proposed stereoisomer of thelepamide. As shown in Scheme 2, diastereoselective reduction of β -substituted ketone 13 with zinc borohydride²¹ yielded a separable 3.8:1 mixture (89%). The relative and absolute configuration of the minor diastereomer 15a was confirmed by X-ray-crystallography. The major diastereomer 15b was acylated with propionic acid using the Steglich protocol²² to give *ent*-10,



^{*a*}Reaction conditions (entries 1–6): 1.10 equiv of 11, 1.00 equiv of L-8, cat. (20 mol %), CH₂Cl₂ (0.10 M), Reaction conditions (entries 7– 14): 2.00 equiv of 11, 1.00 equiv L-8, cat. (20 mol %), PhMe (0.24 M). ^{*b*}Reaction was run until starting material L-8 was fully consumed. ^{*c*}The diastereomeric ratio was determined via GC analysis. ^{*d*}Reaction was carried out with L-N-Boc-phenylglycine. ^{*e*}Reaction was carried out with D-N-Boc-phenylglycine.

the enantiomer of **10** obtained in the initial route (Scheme 1). The Boc group of **15b** was removed with TFA in CH_2Cl_2 (**15b** \rightarrow **16b**), followed by acylation with 2-ketoisovaleric acid to give α -ketoamide **17b**. Subsequently, methyl ester cleavage with lithium hydroxide afforded carboxylic acid **18b** in 75% yield. The reaction was accompanied by epimerization at C2'. Formation of the 5-hydroxyoxazolidin-4-one was achieved via *N*,*O*-acetal formation with formaldehyde by sonicating the reaction under



Scheme 2. Second-Generation Route to Thelepamide

basic conditions.²³ At this step it was necessary to maintain the reaction between 24 and 26 °C in order to suppress the formation of side products. Thus, the proposed structure of thelepamide (1) was obtained in 19% yield over six steps as 1:1 diastereomeric mixture at C2″.

In an analogous fashion to the route described above, 3,5-epithelepamide (3,5-epi-1) obtained from 14 and 5-epi-thelepamide (5-epi-1) obtained from 15a were synthesized in six steps with 23% and 9% overall yield, respectively (Figure 2). Surprisingly, a comparison of the original thelepamide spectra with those of all obtained diastereomers showed a clear mismatch in both the ¹H and ¹³C NMR spectra. The differences manifested in the magnitude of the chemical shifts as well as the diastereomeric



Figure 2. Structures of thelepamide diastereomers 3,5-epi-1 and 5-epi-1.

ratios. From a previous project,²⁴ we were alerted that NMR spectra can be strongly affected by deprotonation of a carboxylic acid moiety. Gratifyingly, the ¹H NMR spectrum of 3,5-*epi*-1 sodium carboxylate was superimposable with the original thelepamide spectrum.

In the ${}^{13}C$ NMR spectrum, the 44–71 ppm region was most indicative for comparing the thelepamide diastereomers (Figure 3). The sodium carboxylate of 3,5-*epi*-1 and the reported



Figure 3. Comparison of the 44–71 ppm ¹³C NMR spectral region of three synthetic thelepamide diastereomers with that of isolated material.

spectrum were nearly identical with an overall median ¹³C NMR shift deviation of <0.1 ppm. These results were supported by HPLC comparison with an authentic thelepamide sample.²⁵

While the previous DFT-NMR analysis² was carried out assuming that **1** was present as free carboxylic acid, we repeated the computational analysis with the corresponding carboxylates. The sodium carboxylates of four possible thelepamide diastereomers (**1a**–**d**, see Figure 4) were initially submitted to a conformational search with the Macromodel program using the protocol of Hoye et al.²⁶ Thus, 85 conformers for **1a**, 67 for **1b**, 90 for **1c**, and 51 for **1d** were found within a 3.0 kcal/mol window. All conformers were classified by energy and frequencies using the B3LYP²⁷/6-31+G(d,p) functional. Finally,



Figure 4. Proposed thelepamide (1) and four diastereometic carboxylates (1a-d) used in the DFT calculations.

the combination MPW1PW91²⁸/6-311+G(2d,p)-polarizable continuum model was used for proton and carbon chemical shift calculations.²⁹ The sets of ¹H and ¹³C chemical shifts were compared by mean absolute error (MAE), R^2 of $\delta_{calcd}/\delta_{expt}$ by the linear regression of calculated (δ_{scaled}) and by the statistical DP4+ parameter developed by Sarotti and co-workers.³⁰ A 100% probability DP4+ value for both carbon and proton chemical shifts in favor to 1d was in agreement with the synthetic 3,5-epi-1 configuration.

Comparison of the optical rotation of (-)-thelepamide and 3,5-*epi*-1 showed them to be equal in sign and magnitude further confirming the configuration to be $3S_{5}S_{R,2}'R$. From the NMR data, we could not make a direct assignment of the C2"-configuration of the major thelepamide diastereomer. However, DFT calculations suggest the configuration to be 2"R (1d).

In order to probe the reported biological activity for the lepamide, the three synthetic diastereomers were tested against the CCRF-CEM cell line. Unfortunately, no cytotoxicity for concentrations up to 100 μ M could be detected.

In conclusion, we have completed a short and efficient synthesis of (-)-thelepamide. Key steps include a catalyst controlled thia-Michael addition and a sonication-assisted oxazolidinone synthesis. Comparison of the spectra of three synthetic diastereomers with those of isolated material led to a revision of the structure of thelepamide from 1 to 1d, the sodium carboxylate of 3,5-*epi*-1. In addition, DFT calculations support the revised structure by including the correct protonation state of pH-sensitive functional groups (e.g., carboxylic acid–carboxylate) in the computational analysis.³¹

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.7b03706.

Procedures and spectroscopic data (PDF)

Accession Codes

CCDC 1587470 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) (a) Blunt, J. W.; Copp, B. R.; Keyzers, R. A.; Munro, M. H. G.; Prinsep, M. R. *Nat. Prod. Rep.* **2017**, *34*, 235. (b) Newman, D. J.; Cragg, G. M. *Planta Med.* **2016**, *82*, 775.

(2) Rodríguez, J.; Nieto, R. M.; Blanco, M.; Valeriote, F. A.; Jiménez, C.; Crews, P. Org. Lett. **2014**, *16*, 464.

(3) Lubell, W. D., Ed. *Peptidomimetics I*; Topics in Heterocyclic Chemistry 48; Springer International Publishing: Cham, 2017.

(4) Kobayashi, K.; Šhimogawa, H.; Sakakura, A.; Teruya, T.; Suenaga, K.; Kigoshi, H. *Chem. Lett.* **2004**, *33*, 1262.

- (5) Smith, S. G.; Goodman, J. M. J. Am. Chem. Soc. 2010, 132, 12946.
- (6) Chauhan, P.; Mahajan, S.; Enders, D. Chem. Rev. 2014, 114, 8807.
- (7) Noyori, R.; Ohkuma, T. Angew. Chem., Int. Ed. 2001, 40, 40.

(8) (a) Nahm, S.; Weinreb, S. M. Tetrahedron Lett. 1981, 22, 3815.
(b) Roche, C.; Labeeuw, O.; Haddad, M.; Ayad, T.; Genet, J.-P.; Ratovelomanana-Vidal, V.; Phansavath, P. Eur. J. Org. Chem. 2009, 2009, 3977.

(9) Wang, L.; Xu, Z.; Ye, T. Org. Lett. 2011, 13, 2506.

- (10) Evans, D. A.; Hoveyda, A. H. J. Am. Chem. Soc. 1990, 112, 6447.
- (11) Rychnovsky, S. D.; Skalitzky, D. J. Tetrahedron Lett. 1990, 31, 945.

(12) Yoshida, Y.; Sakakura, Y.; Aso, N.; Okada, S.; Tanabe, Y. Tetrahedron 1999, 55, 2183.

(13) (a) Ishii, Y.; Sakata, Y. J. Org. Chem. **1990**, 55, 5545. (b) Zörb, A.; Brückner, R. Eur. J. Org. Chem. **2010**, 2010, 4785.

(14) Helder, R.; Arends, R.; Bolt, W.; Hiemstra, H.; Wynberg, H. Tetrahedron Lett. 1977, 18, 2181.

- (15) Okino, T.; Hoashi, Y.; Takemoto, Y. J. Am. Chem. Soc. 2003, 125, 12672.
- (16) Li, B.-J.; Jiang, L.; Liu, M.; Chen, Y.-C.; Ding, L.-S.; Wu, Y. Synlett **2005**, 603.
- (17) Ricci, P.; Carlone, A.; Bartoli, G.; Bosco, M.; Sambri, L.; Melchiorre, P. Adv. Synth. Catal. 2008, 350, 49.
- (18) Xie, J.-W.; Chen, W.; Li, R.; Zeng, M.; Du, W.; Yue, L.; Chen, Y.-C.; Wu, Y.; Zhu, J.; Deng, J.-G. Angew. Chem., Int. Ed. **2007**, 46, 389.
- (19) Bartoli, G.; Bosco, M.; Carlone, A.; Pesciaioli, F.; Sambri, L.; Melchiorre, P. Org. Lett. **200**7, *9*, 1403.
- (20) Starkenmann, C.; Troccaz, M.; Howell, K. Flavour Fragrance J. 2008, 23, 369.
- (21) González-Rodríguez, C.; Parsons, S. R.; Thompson, A. L.; Willis, M. C. *Chem. Eur. J.* **2010**, *16*, 10950.
- (22) Neises, B.; Steglich, W. Angew. Chem., Int. Ed. Engl. 1978, 17, 522.
- (23) Jouglet, B.; Oumoch, S.; Rousseau, G. Synth. Commun. 1995, 25, 3869.
- (24) Seitz, T.; Fu, P.; Haut, F.-L.; Adam, L.; Habicht, M.; Lentz, D.; MacMillan, J. B.; Christmann, M. Org. Lett. **2016**, *18*, 3070.
- (25) See the Supporting Information for details.
- (26) Willoughby, P. H.; Jansma, M. J.; Hoye, T. R. Nat. Protoc. 2014, 9, 643.
- (27) Becke, A. D. J. Chem. Phys. 1993, 98, 5648.
- (28) Adamo, C.; Barone, V. J. Chem. Phys. 1998, 108, 664.
- (29) Lodewyk, M. W.; Siebert, M. R.; Tantillo, D. J. Chem. Rev. 2012, 112, 1839.
- (30) (a) Grimblat, N.; Sarotti, A. M. Chem. Eur. J. 2016, 22, 12246.
- (b) Grimblat, N.; Zanardi, M. M.; Sarotti, A. M. J. Org. Chem. 2015, 80, 12526.
- (31) Saunders, C. M.; Tantillo, D. J. Mar. Drugs 2017, 15, 171.