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Rational Design, Synthesis and Structure–Activity Relationships of a Cyclic Succinate Series of TNF-α Converting Enzyme Inhibitors. Part 2: Lead Optimization

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Abstract—Modifications of the lead TACE inhibitor 1 (*N*-hydroxy-*trans*-2-{[4-(4-quinolinyloxymethyl)anilinyl]carbonyl}-1-cyclohexanecarboxamide) at the cyclohexyl ring and the quinoline moiety led to the identification of a series of piperidine containing TACE inhibitors with potent activity in the inhibition of TNF- α release in the whole blood assay (WBA). The most potent analogue IM491 [*N*-hydroxy-(5*S*,6*S*)-1-methyl-6-{[4-(2-methyl-4-quinolinylmethoxy)anilinyl]carbonyl}-5-piperidinecarboxamide] exhibited an IC₅₀ value of 20 nM in WBA with excellent selectivity over MMP-1, -2 and -9 and is orally bioavailable with an *F* value of 43% in beagle dogs.

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Tumor necrosis factor- α (TNF- α)¹ is a key proinflammatory cytokine that plays a pivotal role in a number of autoimmune diseases such as rheumatoid arthritis (RA).² Over-expression of TNF- α has been shown in the joint of RA patients.³ The clinic success of anti-TNF- α biologics, which neutralize TNF- α activity by forming biologically inactive complex with the cytokine,⁴ has validated TNF- α as a target for the treatment of RA and stimulated considerable interest in small molecule TNF- α converting enzyme (TACE) inhibitors recently.⁵ TACE⁶ is responsible for the processing of inactive membrane-anchored proform of TNF- α to its active soluble form. It is believed that inhibition of TNF- α through inhibition of TACE may represent a new potential therapeutic target for the treatment of RA.⁷

In the preceding communication,⁸ we described the rational design of TACE inhibitors based on the Ciba-Geigy's sulfonamide CGS 27023A,⁹ which led to the identification of a novel series of TACE inhibitors as

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exemplified by compound 1 (Fig. 1). Compound 1 is a six-membered cyclic succinate derivative with 4-[(4-quinolinyl)oxymethyl]aniline as a P1' residue and as a racemic mixture, showed potent porcine TACE (pTACE) activity with an IC₅₀ of 8 nM and excellent selectivity over MMP-1, -2, -9 and aggrecanase. Unfortunately, compound 1 is ineffective in the inhibition of TNF- α release in the cellular assay. In this communication, we report that optimization at the six-membered carbocycle and the quinoline moiety led to the identification of a new series of selective TACE inhibitors with potent anti-TNF- α activity in the cellular assay as represented by **4b** (IM491, Fig. 1).

Synthesis

Compounds 2a-v were synthesized using the procedures described in Scheme 1. Optically pure (3R,4S)-4-(*tert*butoxycarbonyl)-3-piperidinecarboxylic acid (5), which was prepared as described in the literature,¹⁰ was reacted with allyl chloroformate in a mixed solvent of water and THF using *N*-methylmorpholine as a base to provide the *N*-allyloxycarbonyl intermediate **6** in 75% yield. Alkylation of **6** with allyl bromide in the presence

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Scheme 1. Reagents and conditions: (a) allyl chloroformate, NMM, THF, H₂O, 0°C, 75%; (b) allyl bromide, K₂CO₃, acetone, reflux, 85%; (c) TFA, CH₂Cl₂, 100%; (d) an aniline derivative, BOP, DIEA, DMF; (e) tetrakis(triphenylphosphine)palladium, pyrrolidine, THF; (f) aldehyde, Na(OAc)₃BH, DIEA, THF or chloroformate, NMM, THF, H₂O or acid chloride, NMM, THF, H₂O or sulfonyl chloride, NMM, THF, H₂O or isocyanate, DIEA, THF; (g) hydroxylamine hydrochloride, BOP, DIEA, DMF.

of K_2CO_3 provided the diester 7 which was treated with TFA to give the carboxylic acid 8. Coupling of the carboxylic acid 8 with an aniline derivative using BOP gave rise to the anilide product 9. After removal of the allyloxycarbonyl and allyl groups by treatment with pyrrolidine/tetrakis(triphenylphosphine)palladium, the resulting amine was subjected to a reductive amination, acylation, sulfonylation, reaction with a chloroformate or an isocyanate to give the intermediate 10. Conversion of the carboxylic acid to a hydroxamic acid by coupling with hydroxylamine hydrochloride using BOP afforded the final products 2a-v.



Scheme 2. Reagents and conditions: (a) 4-[(2-methyl-4-quinolinyl)methoxy]aniline, BOP, DIEA, DMF, 78%; (b) TFA, CH₂Cl₂, 100%; (c) an aldehyde, Na(OAc)₃BH, DIEA, THF or 2-furoyl chloride, NMM, THF, H₂O; (d) hydroxylamine hydrochloride, DIEA, BOP, DMF.

Compounds **3a–e** were synthesized using the protocols outlined in Scheme 2. Optically pure (3R,4S)-1,4-bis-(*tert*-butoxycarbonyl)-3-piperidinecarboxylic acid (**11**), which was synthesized as described in the literature,¹⁰ was condensed with 4-[(2-methyl-4-quinolinyl)methoxy] aniline¹¹ using BOP to provide the anilide **12** in 78% yield. Treatment of **12** with TFA removed the Boc and *tert*-butyl groups. The final products **3a–e** were obtained by reductive amination of the resulting amine with an aldehyde or by acylation with 2-furoyl chloride followed by coupling of the resulting carboxylic acids with hydroxylamine using BOP.

Compounds **4a**–e were synthesized using the sequence depicted in Scheme 3. Without protection of the secondary piperidne amine, the enantiopure (2S,3S)-3-(tert-butoxycarbonyl)-2-piperidinecarboxylic acid (14), which was prepared as described in the literature,¹⁰ was coupled with 4-[(2-methyl-4-quinolinyl)methoxy]aniline using BOP to give the anilide 15 in 68% yield. Reductive amination of 15 with an aldehyde using Na(OAc)₃BH provided the tertiary amine 16. Following removal of the *tert*-butyl group using TFA, the resulting carboxylic acid was converted to a hydroxamic acid as described above.

Results and Discussion

Since compound 1 had high affinity for pTACE¹² (IC₅₀ = 8 nM), we reasoned that the poor inhibition of TNF- α release in WBA (IC₅₀ > 50 μ M) might be caused by the lipophilicity of the cyclohexyl ring which could make the molecule highly protein-bound. Thus the cyclohexyl ring in 1 was replaced with a piperidine ring to provide a new series of 3,4-piperidinedicarboxamide derivatives (Table 1). With a cyclopropanecarbonyl group on the piperidine nitrogen, analogue **2a** showed 4-fold improvement in pTACE activity relative to 1 with maintenance of selectivity profile over MMP-1, -2 and -9. More importantly, this modification significantly boosted the cellular potency: **2a** displayed an IC₅₀ value of 0.71 μ M in WBA, which is > 70-fold improvement



R1=2-methyl-4-quinolinylmethoxy

Scheme 3. Reagents and conditions: (a) 4-[(2-methyl-4-quinolinyl)methoxy]aniline, BOP, DIEA, DMF, 68%; (b) aldehyde, Na(OAc)₃BH, DIEA, THF; (c) TFA, CH₂Cl₂; (d) hydroxylamine hydrochloride, BOP, DIEA, DMF.

over compound 1. To optimize the P1' residue, several quinoline-containing P1' groups were investigated using the piperidine template of 2a. While exchanging the positions of the oxygen and methylene between the quinoline and aniline in 2a basically maintained the potency and selectivity profile (2b), removal of the methylene residue at that position resulted in complete loss in pTACE and cellular activity as seen in 2c, suggesting the importance of the methylene for the orientation of the quinoline residue in the S1' pocket.¹³ Comparisons in pTACE and WBA potency between 2d and 2e and between 2f and 2g indicate that substitution at the 7-position with trifluoromethyl or at the 2-position with methyl on the quinoline is detrimental to pTACE and cellular activities. However, exchanging the positions of the oxygen and methylene between the quinoline and aniline in 2g afforded a potent analogue **2h**, which exhibited IC_{50} values of 66 picomolar and 170 nM in pTACE and whole blood assays, respectively. Although analogue 2h displayed a submicromolar potency for MMP-9, it is still > 7000-fold selective for pTACE versus MMP-9. Unfortunately, analogue **2h** had a low water solubility (0.01 mg/mL), low Caco-2 permeability ($P_{app} = 0.1 \times 10^{-6}$ cm/s) and low oral bioavailability (5%) when dosed to beagle dogs (Table 2).

To improve the water solubility, Caco-2 permeability and oral bioavailability of 2h, a variety of substituents on the piperidine nitrogen were examined while keeping the 4-[(2-methyl-4-quinolinyl)methoxy]anilinyl at P1' constant (Table 2). The N-acyl analogues 2i-k, urea analogue 21 and carbamate analogues 20-s all showed WBA potency comparable to 2h but no improvement in permeability, solubility and oral biavailability over 2h, whereas a slight loss in cellular potency was observed with the sulfonamide analogues 2m-n and the secondary and tertiary amino analogues 2t-v. Comparison in water solubility of analogues 2t - v with other analogues in Table 2 indicates that the basic piperidine nitrogen in the molecule in 2t-v significantly improved water solubility (>10 mg/mL), which prompted us to explore new analogues bearing a basic piperidine nitrogen (vide infra).

We next carried out SAR studies by moving the piperidine nitrogen one atom away from the hydroxamic acid in **2h** to provide a new series of 4,5-piperidinedicarboxamide derivatives (Table 3). In contrast to the 3,4piperidinedicarboxamide series in which the 2-furoyl analogue 2h is much more potent than analogues 2t-v with a basic piperidine nitrogen (Table 2), analogues 3b-e with a basic piperidine nitrogen in this series exhibited better WBA potency than the 2-furoyl analogue 3a (Table 3). Of particular note in this series is that the N-methyl analogue 3c is very potent in the inhibition of TNF- α release, with an IC₅₀ value of 100 nM in the cellular assay. Despite its good water solubility, 3c showed low Caco-2 permeability and low oral bioavailability (9%) when dosed to beagle dogs. Attempts to improve the Caco-2 permeability and oral bioavailability by increasing the size of the substituent on the piperidine nitrogen resulted in a slight loss in WBA potency as seen in the cyclopropylmethyl and 2-thiazolemethyl analogues **3d** and **3e**.

Table 1.	$(3R, 4S) - N^2$	³ -Hydroxy-3	,4-pip	eridined	icarboxam	ide de	rivatives	with	different	P1′	residues ^{a-c}
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		\sim R^2					
Compd	R ¹	R ²	PTACE IC ₅₀ , nM	WBA IC ₅₀ , µM	$\frac{MMP-1}{K_i, nM}$	MMP-2 <i>K</i> _i , nM	MMP-9 <i>K</i> _i , nM
1			8.0	> 50	> 4949	> 3333	> 2128
2a	4-[(4-Quinolinyl)oxymethyl]phenyl	Cyclopropanecarbonyl	2.0	0.71	> 4949	> 3333	>2128
2b	4-[(4-Quinolinyl)methoxy]phenyl	Cyclopropanecarbonyl	1.0	1.3	>4949	> 3333	>2128
2c	4-[(4-Quinolinyl)oxy]phenyl	Cyclopropanecarbonyl	>1000	> 50	>4949	> 3333	>2128
2d	4-[(4-Quinolinyl)oxymethyl]phenyl	Acetyl	1.0	0.56	>4949	> 3333	>2128
2e	4-[(7-Trifluoromethyl-4-Quinolinyl)oxymethyl]phenyl	Acetyl	3.2	2.6	>4949	> 3333	>2128
2f	4-[(4-Quinolinyl)oxymethyl]phenyl	2-Furoyl	0.80	0.70	>4949	> 3333	>2128
2g	4-[(2-Methyl-4-Quinolinyl)oxymethyl]phenyl	2-Furoyl	3.0	3.4	>4949	> 3333	>2128
2h	4-[(2-Methyl-4-Quinolinyl)methoxy]phenyl	2-Furoyl	0.066	0.17	>4949	1011	465

^aSee ref 12 for pTACE, WBA and MMP assay protocols.

^bpTACE IC₅₀ and MMP K_i values are from single determination.

^cInhibition of TNF-α release in whole blood assay (WBA) was determined with three donors.

Table 2. (3R,4S)-N³-Hydroxy-3,4-piperidinedicarboxamide derivatives with 4-[(2-methyl-4-quinolinyl)methoxy] anilide at P1^{'a-c}



Compd	R	PTACE IC ₅₀ , nM	WBA IC ₅₀ , µM	MMP-1 <i>K</i> i, nM	MMP-2 <i>K</i> _i , nM	MMP-9 <i>K</i> _i , nM	Caco-2 Papp $(\times 10^{-6} \text{ cm/s})$	Solubility mg/mL	Dog PK F%
2h	2-Furoyl	0.066	0.17	> 4949	1011	465	0.1	0.01	5
2i	Cyclopropanecarbonyl	0.45	0.39	> 4949	2454	1656	0.2	0.05	8
2j	Butyryl	0.26	0.33	> 4949	> 3333	>2128			
2k	2-Methylpropionyl	0.22	0.27	> 4949	> 3333	>2128	0.3	0.03	9
21	tert-Butylaminocarbonyl	0.41	0.56	> 4949	> 3333	>2128	0.7	< 0.01	5
2m	Methanesulfonyl	0.48	1.36	>4949	656	306	0.2	< 0.01	7
2n	1-Methyl-4-imidazolesulfonyl	0.31	1.5	>4949	1002	366			
20	Methoxycarbonyl	0.28	0.57	> 4949	1111	767			
2p	Isopropyloxycarbonyl	0.44	0.21	>4949	771	252			
2q	Cyclopropyloxycarbonyl	0.83	0.70	> 4949	642	209	0.2	< 0.01	8
2r	2-Thiazolemethoxycarbonyl	< 0.3	0.45	>4949	1537	768	0.1	< 0.01	3
2s	4-Thiazolemethoxycarbonyl	< 0.3	0.53	>4949	476	296			
2t	Н	1.5	4.3	>4949	> 3333	>2128		>10	
2u	Isopropyl	1.2	1.2	>4949	> 3333	>2128		>10	
2v	Cyclobutyl	1.6	1.1	> 4949	> 3333	>2128		>10	

^apTACE IC₅₀ and MMP K_i values are from single determination.

^bInhibition of TNF-α release in whole blood assay (WBA) was determined with three donors.

^cOral bioavailability (*F*%) from dog pharmacokinetics studies.

Table 3. (45,55)-N⁴-Hydroxy-4,5-piperidinedicarboxamide derivatives with 4-[(2-methyl-4-quinolinyl)methoxy]anilide at Pl^{/a-c}



Compd	R	PTACE IC ₅₀ , nM	WBA IC ₅₀ , µM	MMP-1 <i>K</i> _i , nM	MMP-2 <i>K</i> _i , nM	MMP-9 <i>K</i> _i , nM	Caco-2 Papp $(\times 10^{-6} \text{ cm/s})$	Solubility mg/mL	Dog PK F%
3a	2-Furoyl	< 0.3	1.3	> 4949	> 3333	> 2128	0.2	< 0.1	
3b	Н	3.0	0.53	> 4949	> 3333	> 2128		> 10	
3c	Me	5.0	0.10	> 4949	> 3333	> 2128	0.1	> 10	9
3d	Cyclopropylmethyl	1.0	0.30	> 4949	> 3333	> 2128		> 10	
3e	2-Thiazolemethyl	0.80	0.56	> 4949	> 3333	>2128			

^apTACE IC₅₀ and MMP K_i values are from single determination.

 b Inhibition of TNF- α release in whole blood assay (WBA) was determined with three donors.

^cOral bioavailability (F%) from dog pharmacokinetics studies.

To further improve the cellular potency and oral bioavailability, SAR studies were next conducted by moving the piperidine nitrogen next to the P1' residue to provide a new series of 5,6-piperidinedicarboxamide derivatives (Table 4). With no substitution and a methyl substituent on the piperidine nitrogen, analogues **4a** and **4b** (IM491) showed 4- and 5-fold improvement over the 4,5-piperidinedicarboxamide counterparts **3b** and **3c**, respectively, in WBA potency. IM491 is the most potent TACE inhibitor identified so far in the inhibition of TNF- α release from cells, with an IC₅₀ value of 20 nM in the whole blood assay. Accompanied by their excellent WBA potency were the good water solubility (>10 mg/ mL), excellent selectivity over MMP-1, -2, and -9 and the decent Caco-2 permeability of these two analogues, with P_{app} values of 2.2×10^{-6} and 2.1×10^{-6} cm/s for **4a** and IM491, respectively. The improved Caco-2 permeability of IM491 over **3c** probably can be explained by the less basicity and more steric hindrance of the piperidine nitrogen in IM491 compared to **3c** as the piperidine nitrogen is at a position α to the carboxamide in IM491 but β to the carboxamide in **3c**. In parallel to their good permeability, analogues **4a** and IM491 exhibited good oral bioavailability in beagle dogs, with *F* values of 24% and 43% for **4a** and IM491, respectively. In addition to its good oral absorption, the favorable oral bioavailability of IM491 may also be contributed from its low clearance (0.6 L/h/kg). The lower oral bioavailability

Table 4. $(5S,6S)-N^5$ -Hydroxy-5,6-piperidinedicarboxamide derivatives with 4-[(2-methyl-4-quinolinyl)methoxy]anilide at P1'a-c



Compd	R	PTACE IC ₅₀ , nM	WBA IC ₅₀ , µM	MMP-1 <i>K</i> _i , nM	MMP-2 <i>K</i> _i , nM	MMP-9 <i>K</i> _i , nM	Caco-2 Papp $(\times 10^{-6} \text{ cm/s})$	Solubility mg/mL	Dog PK F%
4a 4b (IM491) 4c 4d 4e	H Me Et Cyclopropylmethyl 2-Thiazolemethyl	1.0 6.2 8.0 1.0 1.0	0.12 0.020 0.10 0.40 1.4	> 4949 > 4949 > 4949 > 4949 > 4949 > 4949	> 3333 > 3333 > 3333 > 3333 > 3333 > 3333	>2128 >2128 >2128 >2128 >2128 >2128	2.2 2.1	>10 >10 >10	24 43

^apTACE IC₅₀ and MMP K_i values are from single determination.

^bInhibition of TNF- α release in whole blood assay (WBA) was determined with three donors.

^cOral bioavailability (F%) from dog pharmacokinetics studies.

of **4a** relative to IM491 is probably caused by its relatively high clearance (1.2 L/h/kg).

A more considerable effect on WBA potency was observed in the 5,6-piperidinedicarboxamide series (Table 4) compared with 4,5-piperidinedicarboxamide series (Table 3) by the increase in size of the substituent on the piperidine nitrogen. Replacement of the methyl group on the piperidine nitrogen in IM491 with ethyl (4c), cyclopropylmethyl (4d) and 2-thiazolemethyl (4e) resulted in 5-, 20- and 70-fold loss in cellular activity.

In summary, to improve the cellular activity of the lead TACE compound 1, modifications were carried out at the cyclohexyl ring and the quinoline P1' residue. Replacement of a methylene residue of the cyclohexyl ring with a nitrogen at three different positions provided a new series of piperidinedicarboxamide analogues which significantly improved the WBA potency. With 4-[(2-methyl-4-quinolinyl)methoxy]anilinyl at the P1' position, the 5,6-piperidinedicarboxamide analogue IM491 bearing a methyl group on the piperidine nitrogen was identified as the most potent TACE inhibitor in the inhibition of TNF- α release, with an IC₅₀ value of 20 nM in the cellular assay. IM491 also showed excellent selectivity over MMP-1, -2 and -9, decent Caco-2 permeability, excellent water solubility and good oral bioavailability in beagle dogs.

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References and Notes

1. Aggarwal, B.; Kohr, W.; Hass, P.; Moffat, B.; Spencer, S.; Henzel, W.; Bringman, T.; Nedwin, G.; Goeddel, D.; Harkins, R. J. Biol. Chem. **1985**, 260, 2345.

2. (a) Aggarwal, B.; Natarajan, K. Eur. Cytokine Network

1996, 7, 93. (b) Eigler, A.; Sinha, B.; Hartmann, G.; Endres, S. *Immunol. Today* **1997**, *18*, 487. (c) Newton, R.; Decicco, C. J. *Med. Chem.* **1999**, *42*, 2295.

3. Feldmann, M.; Brennan, F.; Elliott, M.; Katsikis, E.; Maini, R. *Circ. Shock* **1994**, *43*, 179.

4. (a) Elliott, M.; Maini, R.; Feldmann, M.; Kalden, J.; Antoni, C.; Smolen, J.; Leeb, B.; Breedveld, F.; Macfarlane, J.; Bijl, H.; Woody, J. *Lancet* **1994**, *344*, 1105. (b) Elliott, M.; Maini, R.; Feldmann, M.; Long-Fox, A.; Charles, P.; Bijl, H.; Woody, J. *Lancet* **1994**, *344*, 1125. (c) Van Dullemen, H.; Van Deventer, S.; Hommes, D.; Bijl, H.; Jansen, J.; Tytgat, G.; Woody, J. *Gastroenterology* **1995**, *109*, 129. (d) Moreland, L.; Baumgartner, S.; Schiff, M.; Tindall, E.; Fleischmann, R.; Weaver, A.; Ettlinger, R.; Cohen, S.; Koopman, W.; Mohler, K.; Widmer, M.; Blosch, C. *N. Engl. J. Med.* **1997**, *337*, 141. (e) Cameron, A. *InPharma* **1998**, *1123*, 9.

5. (a) Xue, C.-B.; He, X.; Corbett, R.; Roderick, J.; Wasserman, Z.; Liu, R.-Q.; Jaffee, B.; Covington, M.; Qian, M.; Trzaskos, J.; Newton, R.; Magolda, R.; Wexler, R.; Decicco, C. J. Med. Chem. 2001, 44, 3351. (b) Rabinowitz, M.; Andrews, R.; Becherer, J.; Bickett, D.; Bubacz, D.; Conway, J.; Cowan, D.; Gaul, M.; Glennon, K.; Lambert, M.; Leesnitzer, M.; McDougald, D.; Moss, M.; Musso, D.; Rizzolio, M. J. Med. Chem. 2001, 44, 4252. (c) Holms, J.; Mast, K.; Marcotte, P.; Elmore, I.; Li, J.; Pease, L.; Glaser, K.; Morgan, D.; Michaelides, M.; Davidsen, S. Bioorg. Med. Chem. Lett. 2001, 11, 2907. (d) Chen, J.; Jin, G.; Sung, A.; Levin, J. Bioorg. Med. Chem. Lett. 2002, 12, 1195. (e) Levin, J.; Chen, J.; Du, M.; Nelson, F.; Killar, L.; Skala, S.; Sung, A.; Jin, G.; Cowling, R.; Barone, D.; March, C.; Mohler, K.; Black, R.; Skotnicki, J. Bioorg. Med. Chem. Lett. 2002, 12, 1199. (f) Letavic, M.; Axt, M.; Barberia, J.; Carty, T.; Danley, D.; Geoghegan, K.; Halim, N.; Hoth, L.; Kamath, A.; Laird, E.; Lopresti-Morrow, L.; McClure, K.; Mitchell, P.; Natarajan, V.; Noe, M.; Pandit, J.; Reeves, L.; Schulte, G.; Snow, S.; Sweeney, F.; Tan, D.; Yu, C. Bioorg. Med. Chem. Lett. 2002, 12, 1387. (g) Duan, J.; Chen, L.; Wasserman, Z.; Lu, Z.; Liu, R.-Q.; Covington, M.; Qian, M.; Hardman, K.; Magolda, R.; Newton, R.; Christ, D.; Wexler, R.; Decicco, C. J. Med. Chem. 2002, 45, 4954. (h) Duan, J.; Lu, Z.; Xue, C.-B.; He, X.; Seng, J.; Roderick, J.; Wasserman, Z.; Liu, R.-Q.; Covington, M.; Magolda, R.; Newton, R.; Trzaskos, J.; Decicco, C. Bioorg. Med. Chem. Lett. 2003, 13, 2035.

6. (a) Black, R.; Rauch, C.; Kozlosky, C.; Peschon, J.; Slack, J.; Wolfson, M.; Castner, B.; Stocking, K.; Reddy, P.; Srinivasan, S.; Nelson, N.; Bolani, N.; Schooley, K.; Gerhart, M.; Devis, R.; Fitzner, J.; Johnson, R.; Paxton, R.; March, C.;

Cerretti, D. *Nature* **1997**, *385*, 729. (b) Moss, M.; Jin, S.-L.; Milla, M.; Burkhart, W.; Carter, H.; Chen, W.; Clay, W.; Didsbury, J.; Hassler, D.; Hoffman, C.; Kost, T.; Lambert, M.; Leesnitzer, M.; McCauley, P.; McGeehan, G.; Mitchell, J.; Moyer, M.; Pahel, G.; Rocque, W.; Overton, L.; Schoenen, F.; Seaton, T.; Su, J.; Warner, J.; Willard, D.; Becherer, J. *Nature* **1997**, *385*, 733.

7. Nelson, F.; Zask, A. Expert Opin. Invest. Drugs 1999, 8, 383.

8. Xue, C.-B.; He, X.; Roderick, J.; Corbett, R. L.; Duan, J. J.-W.; Liu, R.-Q.; Covington, M. B.; Newton, R. C.; Trzaskos, J. M.; Magolda, R. L; Wexler, R. R.; Decicco, C. P. *Bioorg. Med. Chem. Lett.* **2003**, *13*, preceding communication. doi:10.1016/j.bmcl.2003.09.056.

9. (a) MacPherson, L.; Bayburt, E.; Capparelli, M.; Carroll, B.; Goldstein, R.; Justice, M.; Zhu, L.; Hu, S.-I.; Melton, R.; Fryer, L.; Goldberg, R.; Doughty, J.; Spirito, S.; Blancuzzi, V.; Wilson, D.; O'Byrne, E.; Ganu, V.; Parker, D. J. Med. Chem. **1997**, 40, 2525. (b) Li, Y.-C.; Zhang, X.; Melton, R.;

Ganu, V.; Gonnella, N. *Biochemistry* **1998**, *37*, 14048. (c) Nar, H.; Werle, K.; Bauer, M.; Dollinger, H.; Jung, B. J. Mol. Biol. **2001**, *312*, 743.

10. Xue, C.-B.; He, X.; Roderick, J.; Corbett, R.; Decicco, C. J. Org. Chem. 2002, 67, 865.

11. 4-[(2-Methyl-4-quinolinyl)methoxy] aniline was prepared by alkylation of 4-*tert*-butoxycarbonylaminophenol with 2methyl-4-chloromethylquinoline in the presence of K_2CO_3 followed by Boc deprotection with acid using procedures analogous to those described in the previous communication.⁸

12. Xue, C.-B.; Voss, M.; Nelson, D.; Duan, J.; Cherney, R.; Jacobson, I.; He, X.; Roderick, J.; Chen, L.; Corbett, R.; Wang, L.; Meyer, D.; Kennedy, K.; DeGrado, W.; Hardman,

K.; Teleha, C.; Jaffee, B.; Liu, R.-Q.; Copeland, R.; Covington, M.; Christ, D.; Trzaskos, J.; Newton, R.; Magolda, R.; Wexler, R.; Decicco, C. J. Med. Chem. 2001, 44, 2636.

Wexiel, K., Decicco, C. J. Med. Chem. 2001, 44, 2030.

13. Wasserman, Z.; Duan, J.; Voss, M.; Xue, C.-B.; Cherney, R.; Nelson, D.; Hardman, K.; Decicco, C. *Chem. & Biol.* **2003**, *10*, 215.