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Tetrahedron xxx (2014) 1-5



Contents lists available at ScienceDirect

Tetrahedron



journal homepage: www.elsevier.com/locate/tet

Development of the 8-aza-3-bromo-7-hydroxycoumarin-4-ylmethyl group as a new entry of photolabile protecting groups

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ARTICLE INFO

Article history Received 5 March 2014 Received in revised form 22 April 2014 Accepted 22 April 2014 Available online xxx

Keywords: Azacoumarin Photolabile protecting groups Caged compounds Photolytic efficiency Hydrophilicity

ABSTRACT

A significant substitution effect of the position of the bromo group on the photosensitivity of the 8azacoumarin chromophore leads to the development of a highly photosensitive 8-aza-3-bromo-7hydroxycoumarin-4-ylmethyl (aza-3-Bhc) group that shows excellent photolytic efficiency and hydrophilicity with long-wavelength absorption maxima. The newly identified aza-3-Bhc group can be applied to caged glutamates for ester-type and carbamate-type protections of carboxyl and amino functionalities. © 2014 Published by Elsevier Ltd.

1. Introduction

Photosensitive biologically active compounds (referred to as caged compounds) have attracted considerable attention due to their practical potentials as phototriggers of biological functions.¹ Progress in the field has been driven by the development of new photolabile protecting group types,² such as nitrobenzyl,³ benzoin,⁴ phenacyl,⁵ and coumarin.⁶ Although a number of chromophores have been applied to photolabile protecting groups, 8-azacoumarin derivatives have not been applied to the caged compounds until our identification of several advantages as an attractive chromophore for caging chemistry, such as excellent water solubility, high molar absorptivity, and efficient photorelease at low pH as described in our previous report.⁷ This report describes the unexpected substitution effects on the photosensitivity of 8-azacoumarin chromophore, leading to the development of a novel 8-aza-3-bromo-7hydroxycoumarin-4-ylmethyl caging group (aza-3-Bhc 1) that shows excellent photolytic efficiency and hydrophilicity with longwavelength absorption maxima and high molar absorptivity,

whose features are superior to those of the 6-bromo derivatives 2 (Fig. 1).



Fig. 1. Structures of azacoumarin- and coumarin-based Bhc groups.

2. Results and discussion

Our study on the development of the aza-3-Bhc group emerged from the bromination of azacoumarin derivative **3** that provided the 6-brominated compound **4a** in a 67% yield as a major product, 7accompanied with a 16% yield of the 3-brominated compound 4b as a minor product (Scheme 1a, Supplementary data). Bromination of the coumarin chromophore provides several advantages for photolysis reactions including lowering the pK_a of the adjacent hydroxyl group accelerating the formation of the strongly absorbing anion and the promotion of intersystem crossing to the triplet. which is considered to be the photochemically reactive state.^{6d}

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^{0040-4020/\$ -} see front matter © 2014 Published by Elsevier Ltd. http://dx.doi.org/10.1016/j.tet.2014.04.063

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H. Takano et al. / Tetrahedron xxx (2014) 1-5



Scheme 1. Synthesis of brominated 8-azacoumarin esters 4 and aza-3-Bhc-CH₂OAc 7.

Preliminary studies revealed that the 8-aza-7-hydroxycoumarin chromophore as well as the 6-bromo-7-hydroxycoumarin has a pK_a value, which is lower than 7.4, and mostly assumes the deprotonated (ionic) form under physiological conditions without additional electron-withdrawing groups. Therefore, the absorption spectrum of **9** has a single peak as the spectrum of **8** (Fig. 2). These features imply the possibility that the regioisomeric 3-brominated 8-azacoumarin chromophore would also work as a hydrophilic caging group. Furthermore, several papers⁸ that identified strong substitution effects on quinolone chromophores prompted us to invoke that the substitution pattern could positively affect photophysical and photochemical properties of 8-azacoumarin chromophore.



Fig. 2. Absorption spectra of Bhc derivative 8 (dash line) and 8-aza-hc 9 (solid line).

The 3-brominated acetate aza-3-Bhc-CH₂OAc **7** was synthesized from **3** in four steps (Scheme 1b). Briefly, reduction of **3** with LiBH₄ gave the corresponding alcohol **5** in moderate yield. In contrast with the bromination of **3** that gave a mixture of regioisomers, the bromination of alcohol **5** proceeded regioselectively to provide the 3-brominated alcohol **6** in 80% yield, which was subjected to acetylation followed by TFA treatment to give the desired aza-3-Bhc-CH₂OAc **7**.

Initially, we investigated the photophysical and hydrophilic behaviors of aza-3-Bhc-CH₂OAc 7 (Table 1). Compared with the

Table 1

Photophysical and hydrophilic properties of aza-3-Bhc-CH₂OAc **7**, aza-Bhc-CH₂OAc **10**, and Bhc-CH₂OAc **11**



Compd	$\lambda_{\max}^{a}(nm)$	ε_{\max}^{b} (M ⁻¹ cm ⁻¹)	$C_{\rm s}^{\rm c}$ (μM)	pK _a ^d
Aza-3-Bhc-CH ₂ OAc (7)	378	27,086	3260	5.08
Aza-Bhc-CH ₂ OAc (10)	362	21,107	10,832	4.22
Bhc-CH ₂ OAc (11)	370	16,584	602	5.88

^a Long-wavelength absorption maxima in PBS (0.1% DMSO).

^b Molar absorptivity at the absorption maxima.

^c Concentration at saturation in PBS (0.1% DMSO).

^d Determined using citric/phosphate buffer in the pH range 2.6–7.0.

original Bhc-CH₂OAc 11, the absorption maximum of 7 shifted to longer wavelength from 370 nm for 11 to 378 nm for 7, whereas that of **10** shifted to shorter wavelength (λ_{max} =362 nm). The molar absorptivity at the maximum wavelength (ε_{max}) of **7** is 27,086 M^{-1} cm⁻¹, which is also higher than those of **10** $(\varepsilon_{\text{max}}=21,107 \text{ M}^{-1} \text{ cm}^{-1})$ and **11** $(\varepsilon_{\text{max}}=16,584 \text{ M}^{-1} \text{ cm}^{-1})$. These results indicated that the 3-brominated 8-azacoumarin would be a superior chromophore to the 6-brominated coumarins. As expected, aza-3-Bhc-CH₂OAc **7** has a pK_a value below the physiological pH and also shows high aqueous solubility, which is important for the photorelease of high concentrations of the caged compounds under physiological conditions. HPLC monitoring of the hydrolysis stability revealed that while aza-3-Bhc-CH₂OAc 7 was more sensitive to hydrolysis in PBS (pH 7.4) than aza-Bhc-CH₂OAc 10, the hydrolysis stability of 7 in KMOPS buffer (10 mM MOPS; 3morpholinepropane-1-sulfonic acid, and 100 mM KCl, pH 7.1) is comparable to that of 10 (Fig. 3).



Fig. 3. Hydrolysis stability of compounds 7 and 10 at room temperature in dark. Time course of compounds 7 and 10 in PBS or KMOPS was analyzed by reversed-phase HPLC.

Having recognized promising photophysical and hydrophilic behaviors of the 3-brominated 8-azacoumarin chromophore, we evaluated the photochemical properties of aza-3-Bhc-CH₂OAc **7** under the photolysis in 5 μ M KMOPS buffer solution at pH 7.2 at 350 nm. The time course of photolysis reaction of **7** was monitored by HPLC in terms of the consumption of the starting materials (Table 2), and indicates that the photolytic reaction at 350 nm of **7** follows a single-exponential decay as in the result of the original Bhc compound **11**. The time to reach 90% conversion (t_{90}) for photolysis of **7** is 19 s, which is shorter than those of the 6-

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ARTICLE IN PRESS

H. Takano et al. / Tetrahedron xxx (2014) 1-5

Table 2

Time course for photolysis reactions of aza-3-Bhc-CH₂OAc **7**, aza-Bhc-CH₂OAc **10**, and Bhc-CH₂OAc **11** and selected photochemical properties



^a Molar absorptivity at 350 nm.

^b Quantum yields for the disappearance of starting materials upon irradiation at 350 nm.

^c Product of the photolysis quantum yield and molar absorptivity.

brominated coumarins **10** and **11** (t_{90} =42 s for **10** and 28 s for **11**, respectively). The photolysis quantum yields of disappearance of starting materials were calculated from the single decay curves using the equation $\Phi = 1/(I \times 10^3 \epsilon t_{90})$ as reported by Tsien.⁹ Notably, the quantum yield of disappearance of **7** (ϕ_{chem} =0.17) is approximately three times higher than that of **10** (Φ_{chem} =0.059) and significantly higher than that of **11** (ϕ_{chem} =0.13). In addition, the photolytic efficiency,¹⁰ the product of the photolysis quantum yield (Φ_{chem}) and molar absorptivity (ε) of **7** $(\varepsilon_{350} \cdot \Phi_{\text{chem}} = 2667)$, is approximately 1.5–2.2-fold higher than that of **10** (ε_{350} · Φ_{chem} =1211) and **11** (ε_{350} · Φ_{chem} =1806). These results indicate that the bromo substitution on position 3 of the 8-azacoumarin chromophore leads not only to improve the photophysical behavior but also to increase the photosensitivity. Although the reason for the significant enhancement of photochemical reactivity of 7 is not fully understood at this stage, these observations suggest that the aza-3-Bhc group has a powerful set of photophysical, photochemical, and hydrophilic properties for caging chemistry.

Next, we examined the synthetic methods for the introduction of the aza-3-Bhc group to biologically relevant compounds (Scheme 2). The aza-3-Bhc group is not limited to the caging group for ester-types. In addition to the α -Glu ester **12**, the aza-3-Bhc group can be applied to the α -Glu carbamate **13** for the protection of an amino functionality, which can be removed efficiently with light of wavelength of 365 nm to produce the corresponding alcohol.¹¹ Compared to the corresponding Bhc-protected compounds 14 and 15,^{6d} both aza-3-Bhc-protected compounds 12 and 13 showed improved photosensitivity (Table 3). The time to reach 90% conversion (t_{90}) for photolysis of **12** is 32 s, which corresponds to be approximately 20% faster than that of the corresponding Bhcprotected α -Glu ester **14** (t_{90} of **14**=38 s). A similar superiority of the aza-3-Bhc group was observed with the carbamate substrates 13 and 15. In accordance with the greater quantum yields of disappearance and comparable molar absorptivities, the photolytic efficiency of both 12 and 13 is higher than those of 14 and 15 (relative



Scheme 2. Synthesis of aza-3-Bhc-caged glutamates.

value of $\varepsilon \cdot \Phi$ of **12**/**14**=1.19 and of **13**/**15**=1.27). These results indicate that the newly identified aza-3-Bhc chromophore can serve as a variant of Bhc chromophore in biologically relevant compounds. Further studies for the application of the 8-aza-3-Bhc group to the protection of other functionalities, such as alcohols, phosphoric acids, and thiols are in progress.

3. Conclusion

We have reported the development of the aza-3-Bhc group as a new entry of photolabile protecting groups for caging chemistry through the strong influence of the position of a bromo substituent on the photosensitivity of the 8-azacoumarin chromophore. The 3brominated 8-azacoumarin 7 is considerably more efficient than the 6-brominated regioisomer 10. Aza-3-Bhc-CH₂OAc 7 shows excellent photolytic efficiency with a bathochromic shift of the absorption maximum of 8-azacoumarin from 362 to 378 nm. Moreover, we have disclosed the potentials of the aza-3-Bhc group as protecting groups of carboxyl and amino functionalities for caged glutamates. A key to the development of the aza-3-Bhc group is the acidic azacoumarin chromophore, whose feature can provide a new opportunity to improve the photosensitivity by chemical modifications that are distinct from those in previous approaches. Efforts to elucidate the reason for the markedly enhanced photosensitivity by the 3-bromo substituent and studies on the twophoton sensitivity of the 8-azacoumarin chromophore are currently in progress.

4. Experimental section

4.1. General methods

All reactions utilizing air- or moisture-sensitive reagents were performed in dried glassware under an atmosphere of nitrogen, using commercially supplied solvents and reagents unless otherwise noted. CH₂Cl₂ was distilled from CaH₂ and stored over molecular sieves. Thin-layer chromatography (TLC) was performed on Merck 60F₂₅₄ precoated silica gel plates and were visualized by fluorescence quenching under UV light and by staining with phosphomolybdic acid, *p*-anisaldehyde, or ninhydrin, respectively. Flash column chromatography was carried out using silica gel 60 N (Kanto Chemical Co., Inc.).

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4

ARTICLE IN PRESS

H. Takano et al. / Tetrahedron xxx (2014) 1-5

Table 3

Selected photophysical and photochemical properties of aza-3-Bhc-caged glutamates 12 and 13, and Bhc-caged glutamates 14 and 15

Compd	$\lambda_{\max}^{a}(nm)$	$(M^{-1} cm^{-1})$	$\epsilon_{365}^{c} (\mathrm{M}^{-1} \mathrm{cm}^{-1})$	$t_{90}^{d}(s)$	$\Phi_{\rm chem}{}^{\sf e}$	$\epsilon_{365} \cdot \Phi_{\rm chem}{}^{\rm f}$	rel. $\varepsilon \cdot \Phi^{g}$
Aza-3-Bhc-Glu-ester (12)	378	13,648	12,153	32	0.17	2063	1.19
Aza-3-Bhc-Glu-carbamate (13)	376	17,068	15,623	10	0.43	6717	1.27
Bhc-Glu-ester (14)	370	16,912	16,466	38	0.11	1740	_
Bhc-Glu-carbamate (15)	370	17,822	17,636	13	0.30	5280	_

^a Long-wavelength absorption maxima in PBS (0.1% DMSO).

^b Molar absorptivity at the absorption maxima.

^c Molar absorptivity at 365 nm.

^d Time to reach 90% conversion.

^e Quantum yields for the disappearance of starting materials upon irradiation at 365 nm.

^f Photolytic efficiency: product of the photolysis quantum yield and molar absorptivity.

^g Relative value of photolytic efficiency [aza-3-Bhc/Bhc]. For full experimental protocol, see Supplementary data.

4.2. Characterization data

 ^{1}H NMR (400 or 500 MHz) and ^{13}C NMR (125 MHz) spectra were recorded using a Bruker Avance II spectrometer with a CryoProbe. Chemical shifts are reported in δ (ppm) relative to Me₄Si (in CDCl₃) as internal standard. Infrared (IR) spectra were recorded on a JASCO FT/IR 4100, and are reported as wavenumber (cm⁻¹). Low- and high-resolution mass spectra were recorded on a Bruker Daltonics micrOTOF (ESI-MS) spectrometers in the positive and negative detection modes.

4.3. HPLC condition

For analytical HPLC, a Cosmosil C18-ARII column (4.6×250 mm, Nacalai Tesque, Inc., Kyoto, Japan) was employed with a linear gradient of MeCN containing 0.1% (v/v) TFA at a flow rate of 1 cm³ min⁻¹ an Agilent HP 1100 system with DAD detection (Agilent Technologies JAPAN Ltd., Tokyo, Japan) and JASCO PU-2086 plus (JASCO corporation, Ltd., Tokyo, Japan), and eluting products were detected by UV at 340 nm.

4.4. Experimental procedures of 8-azacoumarin derivatives 5, 6, 7, 12, and 13

4.4.1. 7-(tert-Butoxy)-4-(hydroxymethyl)-2H-pyrano[2,3-b]pyridin-2-one (5). A suspension of LiBr (1.09 g, 12.6 mmol) and NaBH₄ (410.2 mg, 10.8 mmol) in THF (36.0 mL) was stirred under nitrogen at 50 °C for 2 h, producing the solution of LiBH₄ (ca. 0.3 M). To the solution was added compound 3 (1.03 g, 3.72 mmol) in THF (36.0 mL), and the mixture was stirred at -20 °C for 2 h. The reaction mixture was quenched by 1 M HCl aq, and the organic layer was removed under reduced pressure. The residue was extracted with EtOAc, washed with brine, and dried over Na2SO4. Concentration under reduced pressure followed by flash column chromatography over silica gel with n-hexane/EtOAc (1:1) gave the title compound 5 (411.8 mg, 45% yield) as white powder. Mp: 236–244 °C (dec); ¹H NMR (500 MHz, CDCl₃) δ 1.64 (s, 9H), 4.85 (m, 2H), 6.45 (m, 1H), 6.60 (m, 1H), 7.74 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 28.3, 60.9, 82.5, 104.9, 109.1, 110.5, 134.6, 153.9, 157.4, 161.5, 164.6; IR (ATR) v 3395 (OH), 1704 (CO); HRMS (ESI), m/z calcd for C₉H₈NO₄ [M-*tert*-Bu+2H]⁺ 194.0453, found 194.0450.

4.4.2. 3-Bromo-7-(tert-butoxy)-4-(hydroxymethyl)-2H-pyrano[2,3b]pyridin-2-one (**6**). To a solution of compound **5** (121.4 mg, 0.487 mmol) in CH₃CN (1.37 mL) was added NBS (428.6 mg, 125 mmol), and the mixture was stirred at room temperature for 4 h. After being concentrated under reduced pressure, the residue was dissolved in EtOAc, washed with H₂O, and dried over Na₂SO₄. Concentration under reduced pressure followed by flash column chromatography over silica gel with *n*-hexane/EtOAc (1:1) gave the title compound **6** (127.1 mg, 80% yield) as a dark green solid. Mp: 254–257 °C (dec); ¹H NMR (500 MHz, CDCl₃) δ 1.64 (s, 9H), 5.02 (s, 2H), 6.67 (m, 1H), 8.13 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 28.7, 61.9, 82.9, 106.1, 109.3, 111.3, 136.8, 150.3, 156.23, 157.7, 164.7; IR (ATR) *ν* 3465 (OH) 2977 (CH), 2929 (CH), 1707 (CO), HRMS (ESI), *m/z* calcd for C₁₃H₁₄BrNNaO₄ [M+Na]⁺ 350.0004, found 350.0000.

4.4.3. (3-Bromo-7-hydroxy-2-oxo-2H-pyrano[2,3-b]pyridin-4-yl) methylacetate (7). To a solution of compound 6 (127.1 mg, 0.389 mmol) and DMAP (6.20 mg, 0.0507 mmol) in CH₂Cl₂ (5.6 mL) were added sequentially pyridine (313.1 µL, 3.89 mmol) and acetic anhydride (183.7 µL, 1.94 mmol), and the mixture was stirred at room temperature for 1 h. The reaction mixture was diluted with CH₂Cl₂, washed with NaHCO₃ aq, and dried over Na₂SO₄. Concentration under reduced pressure gave the corresponding acetate (138.1 mg, 96% yield). To a solution of the corresponding acetate (138.1 mg, 0.374 mmol) in CH₂Cl₂ (1 mL) was added trifluoroacetic acid (1 mL), and the mixture was stirred at room temperature for 30 min. Concentration under reduced pressure followed by flash column chromatography over silica gel with CHCl₃/MeOH (5:1) to give the title compound 7 (80.1 mg, 68% yield) as a white solid. Mp: $273-276 \circ C (dec); {}^{1}H NMR (500 MHz, CD_{3}OD) \delta 1.99 (s, 3H), 5.35 (s, 3H)$ 2H), 6.53 (d, J=9.0 Hz, 1H), 7.97 (d, J=9.0 Hz, 1H); ¹³C NMR (125 MHz, CD₃OD) § 20.5, 61.1, 102.0, 106.1, 109.4, 136.2, 150.9, 158.4, 160.4, 166.8, 170.0; IR (ATR) v 2923 (OH) 1736 (CO), HRMS (ESI), m/z calcd for C₁₁H₈BrNNaO₅ [M+Na]⁺ 335.9484, found 335.9485.

4.4.4. (S)-4-Amino-5-((3-bromo-7-hydroxy-2-oxo-2H-pyrano[2,3*bpyridin-4-yl)methoxy)-5-oxopentanoic acid* (**12**). To a solution of Boc-Glu(OBn)-OH (158.0 mg, 0.468 mmol), EDCI · HCl (541.2 mg, 2.82 mmol), and DMAP (13.8 mg, 0.113 mmol) in CH₂Cl₂ (10.2 mL) was added compound 6 (100.7 mg, 0.308 mmol), and the mixture was stirred at room temperature for 24 h. The mixture was poured into water and extracted with EtOAc, and dried over Na₂SO₄. Concentration under reduced pressure gave the crude compound (321.9 mg), which was used in the next step without further purification. A solution of the crude compound (321.9 mg, 0.498 mmol), 1 M TMS-Br/TFA (3.32 mL), and 1 M thioanisole/TFA (3.32 mL) was stirred at room temperature for 3 h. Purification by preparative HPLC (Gradient: 0 min, 0% CH₃CN in H₂O; 90 min, 40% CH₃CN in H₂O) followed by lyophilization to give a title compound **12** (23.7 mg, 16% yield) as a pale purple solid. Mp: 149–154 °C (dec); ¹H NMR (500 MHz, CD₃OD) δ 2.06 (m, 2H), 2.39 (m, 2H), 4.10 (m, 1H), 5.56 (m, 2H), 6.60 (d, J=8.5 Hz, 1H), 8.06 (d, J=8.5 Hz, 1H); ¹³C NMR (125 MHz, CD₃OD) δ 26.6, 30.2, 53.2, 64.7, 106.3, 110.4, 112.0, 139.1, 147.5, 157.9, 158.3, 166.6, 169.9, 175.4; IR (ATR) v 3505 (NH), 2981 (OH), 2865 (CH), 1737 (CO), 1690 (CO) 1682 (CO), HRMS (ESI), m/z calcd for $C_{14}H_{14}BrN_2O_7$ $[M+H]^+$ 400.9984, found 400.9981.

4.4.5. (S)-2-((((3-Bromo-7-hydroxy-2-oxo-2H-pyrano[2,3-b]pyridin-4-yl)methoxy)carbonyl)amino)-pentanedioic acid (**13**). To a solution

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of CDI (25.4 mg, 0.157 mmol) of CH₂Cl₂ was added 6 (48.5 mg, 0.148 mmol), and the mixture stirred at 0 °C for 1 h. After H-Glu(O^tBu)-O^tBu (90.4 mg, 0.306 mmol) and Et₃N (52.3 µL, 0.375 mmol) were added, the mixture was stirred at room temperature for additional 12 h. The reaction mixture was concentrated with reduced pressure. The residue was diluted with Et₂O, and washed with water and brine, dried over Na₂SO₄ Concentration under reduced pressure gave the crude compound (168.1 mg). which was used in the next step without further purification. To a solution of the crude compound (168.1 mg, 0.275 mmol) in CH₂Cl₂ (0.90 mL) was added TFA (2.7 mL), and the mixture was stirred at room temperature for 1 h. Purification by preparative HPLC (Gradient: 0 min, 5% CH₃CN in H₂O; 90 min, 40% CH₃CN in H₂O) followed by lyophilization to give a title compound **13** (17.6 mg, 22%) yield) as a pale green solid. Mp: 196–198 °C (dec); ¹H NMR (500 MHz, CD₃OD) δ 1.88 (m, 1H), 2.16 (m, 1H), 2.38 (m, 2H), 4.19 (m, 1H), 5.46 (s, 2H), 6.68 (m, 1H), 8.19 (m, 1H); ¹³C NMR (125 MHz, CD₃OD) § 27.8, 31.1, 54.8, 64.0, 106.3, 110.4, 110.8, 139.6, 149.4, 157.6 (2C), 158.4, 166.3, 175.2, 176.3; IR (ATR) v 3306 (NH), 2981 (OH), 1742 (CO), 1698 (CO), HRMS (ESI), m/z calcd for C₁₅H₁₃BrN₂NaO₉ [M+Na]⁺ 466.9702, found 466.9701.

4.5. Determination of saturated concentrations

Saturated concentrations of compounds were calculated from the standard curves that related peak area (340 nm) against known concentration of compounds in PBS.

4.6. Determination of the pK_a values

The p K_a values of compounds were estimated from the titration curves of absorbance against pH using 10 μ M substrate solution in citric/phosphate buffer solution in the pH range 2.6–7.0 by adjusting the acidity with 10 μ L of 2 M NaOH sequentially. The pH values were analyzed by a sensitive pH meter (HORIBA, F51).

4.7. Photolysis and quantum efficiency measurement

Into a Pyrex test tube of 12 mm diameter was placed 2 mL of 10 μ M substrate solution in KMOP solution (pH 7.2) containing 0.1% DMSO. The solution was irradiated at 350 nm using four RPR 350 nm lamps (10 mJ s⁻¹). Aliquots of 10 μ M were removed periodically and analyzed by HPLC. The light output for the quantum efficiencies measurement was performed using ferrioxalate actinometry.

Acknowledgements

This research was supported by the Naito Foundation, KEN10000322 (Natural Science Scholarship) and in part by a Grantin-Aid for Young Scientist (B) from the Ministry of Education, Culture, Sports, Science, and Technology.

Supplementary data

This section presents the experimental details, and 1 H and 13 C NMR spectra. Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.tet.2014.04.063.

References and notes

- (a) *Caged Compounds*; Marriot, G., Ed.Methods in Enzymology; Academic: New York, NY, 1998; Vol. 291; (b) Mayer, G.; Heckel, A. *Angew. Chem., Int. Ed.* **2006**, *45*, 4900–4921; (c) Ellis-Davies, G. C. R. *Nat. Methods* **2007**, *4*, 619–628; (d) Lee, H. M.; Larson, D. R.; Lawrence, D. S. ACS Chem. Biol. **2009**, *4*, 409–427.
- (a) Brieke, C.; Rohrbach, F.; Gottschalk, A.; Mayer, G.; Heckel, A. *Angew. Chem., Int. Ed.* 2012, *51*, 8446–8476; (b) Klán, P.; Solomek, T.; Bochet, C. G.; Blanc, A.; Givens, R.; Rubina, M.; Popik, V.; Kostikov, A.; Wirz, J. *Chem. Rev.* 2013, *113*, 119–191.
 Nitrobenzyl-type: (a) Engels, J.; Schlaeger, E.-J. J. Med. Chem. 1977, *20*, 907–911;
- Nitrobenzyl-type: (a) Engels, J.; Schlaeger, E.-J. J. Med. Chem. 1977, 20, 907–911; (b) Kaplan, J. H.; Forbush, B., III; Hoffman, J. F. Biochemistry 1978, 17, 1929–1935.
- Benzoin-type: (a) Sheehan, J. C.; Wilson, R. M. J. Am. Chem. Soc. 1964, 86, 5277–5281; (b) Givens, R. S.; Athey, P. S.; Kueper, L. W., III; Matuszewski, B.; Xue, J. J. Am. Chem. Soc. 1992, 114, 8708–8710; (c) Hansen, K. C.; Rock, R. S.; Larsen, R. W.; Chan, S. I. J. Am. Chem. Soc. 2000, 122, 11567–11568.
- Phenacyl-type: (a) Sheehan, J. C.; Umezawa, K. J. Org. Chem. 1973, 38, 3771–3774; (b) Givens, R. S.; Weber, J. F. W.; Jung, A. H.; Park, C.-H. Methods Enzymol. 1998, 291, 1–29; (c) Conrad, P. G., II; Givens, R. S.; Weber, J. F. W.; Kandler, K. Org. Lett. 2000, 2, 1545–1547.
- 6. Coumarin-type: (a) Givens, R. S.; Matuszewski, B. J. Am. Chem. Soc. 1984, 106, 6860–6861; (b) Furuta, T.; Torigai, H.; Sugimoto, M.; Iwamura, M. J. Org. Chem. 1995, 60, 3953–3956; (c) Furuta, T.; Iwamura, M. Methods Enzymol. 1998, 291, 50–63; (d) Furuta, T.; Wang, S. S. H.; Dantzker, J. L.; Dore, T. M.; Bybee, W. J.; Callaway, E. M.; Denk, W.; Tsien, R. Y. Proc. Natl. Acad. Sci. U.S.A. 1999, 96, 1193–1200; (e) Hagen, V.; Bendig, J.; Fring, S.; Eckardt, T.; Helm, S.; Reuter, D.; Kaupp, U. B. Angew. Chem., Int. Ed. 2001, 40, 1045–1048; (f) Eckardt, T.; Hagen, V.; Schade, B.; Schmidt, R.; Schweitzer, C.; Bendig, J.; Org. Chem. 2002, 67, 703–710; (g) Geisler, D.; Kresse, W.; Wiesner, B.; Bendig, J.; Kettenmann, H.; Hagen, V. ChemBioChem 2003, 4, 162–170; (h) Nomura, W.; Narumi, T.; Ohashi, N.; Serizawa, Y.; Lewin, N. E.; Blumberg, P. M.; Furuta, T.; Tamamura, H. ChemBioChem 2011, 12, 535–539.
- 7. Narumi, T.; Takano, H.; Ohashi, N.; Suzuki, A.; Furuta, T.; Tamamura, H. Org. Lett. 2014, 16, 1184–1187.
- (a) Petit, M.; Tran, C.; Roger, T.; Gallavardin, T.; Dhimane, H.; Palma-Cerda, F.; Blanchard-Desce, M.; Acher, F. C.; Ogden, D.; Dalko, P. I. Org. Lett. 2012, 14, 6366–6369; (b) Zhu, Y.; Pavlos, C. M.; Toscano, J. P.; Dore, T. M. J. Am. Chem. Soc. 2005, 128, 4267–4276; (c) Davis, M. J.; Kragor, C. H.; Reddie, K. G.; Wilson, H. C.; Zhu, Y.; Dore, T. M. J. Org. Chem. 2009, 74, 1721–1729; (d) Laras, Y.; Hugues, V.; Chandrasekaran, Y.; Blanchard-Desce, M.; Acher, F. C.; Pietrancosta, N. J. Org. Chem. 2012, 77, 8294–8302.
- 9. Tsien, R. Y.; Zuker, R. S. Biophys. J. 1986, 50, 843-853.
- Brown, E. B.; Shear, J. B.; Adams, S. R.; Tsien, R. Y.; Webb, W. W. Biophys. J. 1999, 76, 489–499.
- 11. See Supplementary data for details.