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Expedient synthesis of 2-alkylthio-N⁶-aryladenosines from guanosine

Miao Tian, Ning Chen 🗅, Fangming Xu, Xiuxiu Li, Shunlai Li and Hongguang Du 🕒

Faculty of Science, Beijing University of Chemical Technology, Beijing, People's Republic of China

ABSTRACT

A general approach for the synthesis of 2-alkylthio- N^6 -aryladenosine was developed from the commercially available guanosine through the acetyl protection, chlorination, diazotization-alkylthionation, aromatic nucleophilic substitution and deacetylation. Two approaches were designed for the transformation of 2-amino-6-chloroguanosine to 2-alkylthio- N^6 -aryladenosines but only the one with diazotization-alkylthionation first could afford the target molecules. Both electron-rich and deficient anilines can afford the desired products in moderate to good yield. Finally, under the optimized condition, 20 2-alkylthio- N^6 -aryladenosines were synthesized, 5 of which exhibit poor antiplatelet aggregation activities.



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Adenosine; guanosine; purine riboside; diazotizationalkylthionation; antiplatelet aggregation

1. Introduction

Adenosine derivatives, with various substituents at C^2 , C^6 , C^8 , and N^9 positions, have attracted much attention due to their broad spectrum of biological activities [1–6]. In 1999, 2-alkylthio-6-aminopurine riboside derivatives were first reported as inhibitors for P2Y₁₂

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CONTACT Ning Chen Chenning@mail.buct.edu.cn; Hongguang Du Chenning@mail.buct.edu.cn Faculty of Science, Beijing University of Chemical Technology, Beijing 100029, People's Republic of China

receptor [7]. Since then, great endeavors have been devoted to developing various outstanding antiplatelet inhibitors through the modification of the substituents on the purine moiety [8–11]. In particular, adenosine derivatives with various substituents on the purine motif, including the alkylthio group at the C^2 position, the alkyl substituent at the N^6 position and the triphosphate side chain at the 5' position, possess unique biological effects, such as Cangrelor [7] (Figure 1, I) and Ticagrelor [12] (Figure 1, II). In view of the significant antiplatelet activities exhibited by purine derivatives, it is still highly desirable to search for more effective antiplatelet aggregation molecules from purine derivatives.

General methods to produce adenosine derivatives varied from guanosine or 6-halopurine riboside through alkylation, amination, and Buchwald–Hartwig amination [9,13–15]. In recent years, our research group synthesized a series of adenosine derivatives with alkoxy or alkylthio groups at position 2 and alkylamine at position 6 using guanosine as a raw material [16,17]. Meanwhile, we also studied the antiplatelet activities of these compounds and found that molecules BF061 and BF081 (Figure 1, III and IV) show excellent antiplatelet activities [16,18]. As the ongoing work, we planned to shorten the N^6 -arylalkyl substituent to an aryl group in adenosine derivatives to investigate their bioactivities. Herein, we report our recent progress for the synthesis and the antiplatelet aggregation evaluation of various substituted 2-alkylthio- N^6 -aryladenosines.

2. Results and discussion

Following our previous method [16], commercially available guanosine (1) was used as a starting material (Scheme 1). Initially, after acetylation with acetyl anhydride, 1 afforded the corresponding acetyl guanosine 2 with 85% yield and the following chlorination with POCl₃ provided key intermediates 6-chlordine purine nucleoside 3 in 78% yield (Scheme 1).



Figure 1. Structures of Cangrelor, Tricagrelor, BF061, and BF0801.



Scheme 1. The synthesis of 6-chloropurine nucleoside 3.



Scheme 2. Two approaches for the synthesis of 6-arylamino-2-alkylthiopurines.

In order to afford 2-alkylthio- N^6 -arylpurine riboside **5** (Scheme 2), two possible ways were designed: C6-amination first (route 1) or C2-arylthio-substitution first (route 2). Both approaches were examined, but only the latter one produced the target molecule. Take 2-ethylthio- N^6 -phenylamino-purine acetyl nucleoside **5'a** as an example; in route 1, amination of **3a** from aniline gave corresponding 6-arylaminopurine riboside **4a** in 44% yield, but only trace yields of the target **5'a** was detected from thin-layer chromatogra-phy (TLC) in the followed diazotization-alkylthionation process with *iso*-amylnitrite and diethyl disulfide [17]. The result demonstrates that electron-donating group (alkylamino) on purine motif makes the purine too electron-rich to stabilize the followed aryl radical [19], thus inhibiting the further radical alkylthionation approach of **3** was carried out with 65% yield of isolated **4'a**, which further demonstrated that electron-withdrawing group C6-position facilitated the radical coupling. To our gratification, the subsequent amination of **4'a** with aniline in the presence of triethylamine afforded the substitution product **5'a** in 81% yield.

Having obtained the model molecule **5b'**, we next examined the application of this method. During the diazotization-alkylthionation process, we have exploited that ultrasound irradiation highly facilitated the radical coupling, which not only promoted the reaction efficiency but also avoid the use of inert gas [17]. Herein, beside the diethyl disulfide, dimethyl- and dipropyl disulfide could also convert the muriatic purine nucleoside



Scheme 3. The reaction of 6-chloropurine **3** and dialkyl disulfide, and the radical trapping experiment with TEMPO.

3 to the corresponding 2-alkylthiopurine derivatives in 66% (**4'a**) and 60% (**4'c**) isolated yield under ultrasound irradiation at 65°C in acetonitrile (Scheme 3, a). When TEMPO, a common radical scavenger, was added to the model reaction, both TEMPO-trapping product **6** (35%) and target product **4'b** (28%) were isolated (Scheme 3, b), revealing that the reaction was a radical process.

In our previous work, alkyl amine could directly react with 6-chloropurines to afford the corresponding substitution molecules in excellent yield under the catalysis of Et_3N in EtOH (b.p. 78°C) [16]. However, in the current system, only 61% yield of **5'a** was obtained (Table 1, entry 1). As is generally known, the nucleophicity of arylamine (aniline) is much lower than that of alkylamine. We thus changed the solvent to *n*-propanol (b.p. 97°C), and to our gratification the yield of **5'a** was promoted to 81% under refluxing. Further screening of the organic bases (Table 1, entries 2–8) indicated that trimethylamine, pyridine, and diisopropylethylamine (DIPEA) prompted the reaction with high efficiency, and DIPEA was the best one (Table 1, entry 6), while DMAP and DBU showed no catalytic activity. Moreover, focused on the DIPEA, ethanol and *n*-butanol were also investigated as solvents but the reaction yields were diminished (Table 1, entries 7 and 8).

Under the optimized conditions, a series of substituted anilines, including electrondonating (methyl and methoxy) and electron-withdrawing (fluoro-, cholro-, and nitro-) substituents, were investigated. Moreover, **4'a** could be easily deacetylation by adding 3.0 equiv. of sodium methoxide [17]. To simplify the reaction operation, we tried the one-pot strategy. Followed by the substitution, sodium methoxide was added to the reaction solution directly, and to our gratification, the target deacetylation 2-ethylthio-6-phenylaminopurine nucleoside **5a** was obtained in 66% yield (Table 2, **5a**). On the further scope for *para*-substituted anilines, we found that electron-rich anilines shows better reactivity than that of electron-withdrawing ones probably due to their nucleophilicity nature (**5b**, **5c** vs. **5d**, **5e**). However, all *meta*-substituted anilines provided moderate to good yield (**5f**-**5i**). It is noteworthy that *para*-nitroaniline shows no reactivity in the current condition even though we made every endeavor to develop it, but *meta*-nitroaniline could produce the target **5j** in 45% yield because of the great difference in electronic influence between the *para*and *meta*-substituents. Furthermore, other purine substrates with 2-methylthio-(**4'b**) and



Table 1. Optimization of substitution of aniline and 6-purine 4'a.

^aReaction condition: **4' a** (0.2 mmol), PhNH₂ (0.4 mmol), base (0.4 mmol) solvent (4.0 mL), reflux 3 h; yields were determined by ¹H NMR obtained by using dimethyl maleate as an internal standard.

2-propylthio- (4'c) substituents also investigated and the results were in line with 4'a (5k-5t).

Finally, the antiplatelet aggregation of all synthetic compounds was assayed by adenosine-5'-diphosphate (ADP) and arachidonic acid (AA) with Ticagrelor and Cangrelor as positive controls (see Supplemental Materials, Table S1, http://dx.doi.org/10.1080/ 17415993.2017.1391812) [20]. Generally, most of them show poor antiplatelet aggregation compared with Ticagrelor and Cangrelor.

3. Conclusion

Herein, we have developed a general approach for the synthesis of 2-alkylthio- N^6 aryladenosine from the commercially available guanosine. After acetyl protection and chlorination, a key intermediate 2',3',5'-tri-O-acetyl-2-amino-6-chloropurine riboside **3** was obtained. Two approaches were designed for the transformation of **3** to 2-alkylthio- N^6 -aryladenosines **5** but only the one with diazotization-alkylthionation first could afford the target molecules. The result demonstrates that electron-rich purine prohibits the diazotization-alkylthionation reaction. In the aromatic substitution, DIPEA can highly prompt the aromatic nucleophilic substitution of 6-chloropurine with arylamines and both electron-rich and deficient anilines afford the desired products in moderate to good yield. Moreover, a wide array of functional groups, such as halogens, ethers, esters, hydroxyl, and nitro, are also tolerated in this strategy.

4. Experimental section

4.1. General

Commercially available reagents were used without further purification. Melting points were determined on a hot-stage melting point apparatus (uncorrected). Nuclear magnetic resonance (NMR) spectra were recorded in the CDCl₃ or DMSO- d_6 with TMS as an internal standard (¹H: 400 MHz, ¹³C: 100 MHz). The chemical shifts (δ) are reported





^a yields of the isolated products are given.

in parts per million (ppm) and coupling constants J are in Hz. The signal multiplicities were distinguished with the common abbreviations with s (singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), br s (broad singlet), and m (multiplet). IR spectra are recorded on an FT-IR instrument. HRMS measurements were carried out on an Liquid Chromatography/Mass Spectrometer Detector QTOF mass spectrometer. ADP, AA, and Aspirin were purchased from Sigma. Ticagrelor was purchased from Dalian Meilun Biotech Co., Ltd. 2',3',5'-Tri-O-acetyl-2-amino-6-chloropurine riboside **3** were synthesized according to the literature [21,22].

4.2. General procedure for the synthesis of 2-alkythio-6-chloro-9-(2',3',5'-tri-O-acetyl-b-D-ribofuranosyl)-9H-purine (4' a-4' c)

2-Amino-6-chloropurine riboside **3** (3.0 g, 7.01 mmol) and dialkyl disulfides (21.04 mmol) were suspended in 15 mL anhydrous acetonitrile in a 50-mL vial. Then, isopentylnitrite (2.6 mL, 21.04 mmol) was then added to the mixture. The reaction was stirred at 65°C under the condition of ultrasound for 30 min. Upon completion of the reaction, solvent was evaporated in vacuo and the resulting residue was purified by silica gel column chromatography (EtOAc/PE = 2:3, ν/ν) to afford corresponding product **4** as yellow oil.

4.2.1. 6-Chloro-2-ethylthio-9-(2',3',5'-tri-O-acetyl- β -D-ribofuranosyl)-9H-purine (4'a) [16]

2.09 g, yield = 65%; a yellow oil; ¹H NMR (400 MHz, CDCl₃): δ 8.11 (1H, s), 6.13 (1H, d, J = 4.8 Hz), 5.95 (1H, t, J = 4.8 Hz), 5.61 (1H, t, J = 5.2 Hz), 4.45–4.42 (2H, m), 4.35–4.31 (1H, m), 3.23 (2H, q, J = 7.2 Hz), 2.15 (3H, s), 2.11 (6H, app s), 1.43 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 170.3, 169.5, 169.3, 166.9, 152.0, 151.3, 142.1, 129.1, 86.9, 80.1, 73.0, 70.1, 62.7, 26.0, 20.7, 20.5, 20.4, 14.1.

4.2.2. 6-Chloro-2-methylthio-9-(2',3',5'-tri-O-acetyl-β-D-ribofuranosyl)-9H-purine (4'b) [16]

2.18 g, yield = 66%; a yellow oil; ¹H NMR (400 MHz, CDCl₃): δ 8.11 (1H, s), 6.12 (1H, d, J = 4.4 Hz), 6.00 (1H, t, J = 5.2 Hz), 5.65 (1H, t, J = 5.2 Hz), 4.46–4.40 (2H, m), 4.35–4.30 (1H, m), 2.65 (3H, s), 2.16 (3H, s), 2.11 (3H, s), 2.10 (3H, s); ¹³C NMR (100 MHz, CDCl₃): δ 170.3, 169.5, 169.3, 167.4, 151.9, 151.3, 142.2, 129.1, 87.1, 80.1, 73.0, 70.2, 62.7, 20.7, 20.5, 20.4, 14.9.

4.2.3. 6-Chloro-2-propyllthio-9-(2',3',5'-tri-O-acetyl-β-D-ribofuranosyl)-9H-purine (4'c) [16]

2.05 g, yield = 60%; a yellow oil; ¹H NMR (400 MHz, CDCl₃): δ 8.10 (1H, s), 6.14 (1H, d, J = 4.8 Hz), 5.92 (1H, t, J = 5.2 Hz), 5.59 (1H, t, J = 5.2 Hz), 4.47–4.41 (2H, m), 4.36–4.32 (1H, m), 3.20 (2H, t, J = 7.2 Hz), 2.15 (3H, s), 2.11 (3H, s), 2.10 (3H, s), 1.80 (2H, sext, J = 7.2 Hz), 1.07 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃): δ 170.3, 169.5, 169.3, 167.1, 152.0, 151.2, 141.9, 129.1, 86.8, 80.1, 73.1, 70.2, 62.8, 33.6, 22.3, 20.7, 20.5, 20.4, 13.4.

4.3. General procedure for the synthesis of 2-alkylthio-N⁶-aryladenosines (5a-5t)

To a suspend solution of **4** (1.06 mmol) and 15 mL *n*-propanol, DIPEA (2.11 mmol) and anilines (2.11 mmol) were added to the mixture. The reaction was stirred under the refluxing condition till the disappearance of intermediate **4** (about 3-11 h). When the starting material disappeared as monitored by TLC (EtOAc/PE = 2:3, ν/ν), the system was cooled

to room temperature, and sodium methoxide (3.0 equiv.) was added into the mixture. The whole system was then heated to reflux for 1.5 h. The solvent was evaporated in vacuo. Then, the mixture was dissolved with 12 mL EtOAc and H₂O (1:1) and was neutralized with dilute hydrochloric acid to pH = 7. The aqueous phase was extracted with EtOAc (3×20 mL). The combined organic phase was dried over by anhydrous MgSO₄. After filtrating and evaporating the solvent, the resulting residue was purified by silica gel column chromatography (EtOAc/MeOH = 15:1, *v*/*v*) to afford corresponding product **5** as white crystals.

4.3.1. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(2-(ethylthio)-6-(phenylamino)-9H-purin-9yl)tetrahydrofuran-3,4-diol (5a)

282 mg, yield = 66%; white solid; m.p. = $171-172^{\circ}$ C; $R_{\rm f}$ = 0.48 (EtOAc:MeOH = 15:1, v/v). ¹H NMR (400 MHz, DMSO- d_6): δ 9.99 (1H, s), 8.42 (1H, s), 7.91–7.89 (2H, m), 7.37–7.33 (2H, m), 7.08–7.05 (1H, m), 5.90 (1H, d, J = 5.6 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 5.2 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.66–4.62 (1H, m), 4.19–4.16 (1H, m), 3.98–3.95 (1H, m), 3.71–3.66 (1H, m), 3.53.60–3.55(1H, m), 3.16–3.11 (2H, m), 1.35 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 151.5, 150.4, 139.6, 139.2, 128.3, 122.9, 120.9, 117.8, 87.4, 85.6, 73.4, 70.5, 61.5, 24.7, 14.9; FT-IR (film, cm⁻¹): 3348, 3291, 2966, 2924, 1616, 1578, 1498, 1444, 1334, 1191, 1078, 748, 691. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₂N₅O₄S⁺ m/z 404.1387, found 404.1388.

4.3.2. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-(p-tolylamino)-9H-purin-9-yl)-5-(hydroxy methyl)tetrahydrofuran-3,4-diol (**5b**)

301 mg, yield = 68%; white solid; m.p. = $182-183^{\circ}$ C; $R_{\rm f}$ = 0.50 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.90 (1H, s), 8.40 (1H, s), 7.76 (2H, d, J = 8.4 Hz), 7.15 (2H, d, J = 8.0 Hz), 5.89 (1H, d, J = 6.0 Hz), 5.48 (1H, br s), 5.22 (1H, br s), 5.07 (1H, br s), 4.64 (1H, br s), 4.18 (1H, br s), 3.96 (1H, dd, J = 4.0, 7.6 Hz), 3.70-3.66 (1H, m), 3.58-3.56 (1H, m), 3.14-3.10 (2H, m), 2.29 (3H, s), 1.35 (3H, t, J = 7.6 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 151.5, 150.3, 139.4, 136.6, 131.9, 128.7, 121.0, 117.7, 87.4, 85.5, 73.4, 70.5, 61.5, 24.7, 20.4, 14.9. FT-IR (film, cm⁻¹): 3333, 3232, 3104, 2921, 2863, 1625, 1580, 1511, 1454, 1334, 1234, 1083, 811. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₄N₅O₄S⁺ m/z 418.1544, found 418.1550.

4.3.3. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((4-methoxyphenyl)amino)-9H-purin-9-yl)-5-(hydroxymethyl)tetrahydrofuran-3,4-diol (5c)

312 mg, yield = 68%; white solid; m.p. = $158-160^{\circ}$ C; $R_{\rm f}$ = 0.44 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.85 (1H, s), 8.37 (1H, s), 7.75 (2H, d, J = 9.2 Hz), 6.93 (2H, d, J = 8.8 Hz), 5.88 (1H, d, J = 6.0 Hz), 5.47 (1H, d, J = 6.0 Hz), 5.21 (1H, d, J = 4.8 Hz), 5.07 (1H, t, J = 5.6 Hz), 4.63 (1H, dd, J = 5.6, 11.2 Hz), 4.17 (1H, dd, J = 4.4, 8.4 Hz), 3.96–3.95 (1H, m), 3.76 (3H, s), 3.70–3.65 (1H, m), 3.60–3.55 (1H, m), 3.13–3.07 (2H, m), 1.33 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 155.2, 151.5, 150.1, 139.3, 132.2, 122.7, 117.6, 113.5, 87.4, 85.5, 73.4, 70.5, 61.5, 55.2, 24.7, 15.0. FT-IR (film, cm⁻¹): 3381, 3352, 3110, 2927, 1619, 1582, 1509, 1455, 1334, 1243, 1118, 1038, 828. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₄N₅O₅S⁺ m/z 434.1493, found 434.1495.

4.3.4. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((4-fluorophenyl)amino)-9H-purin-9-yl)-5- (hydroxymethyl)-tetrahydrofuran-3,4-diol (5d)

161 mg, yield = 36%; white solid; m.p. = $208-210^{\circ}$ C; $R_{\rm f} = 0.46$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.05 (1H, s), 8.41 (1H, s), 7.88 (2H, dd, J = 4.8, 8.8 Hz), 7.19 (2H, t, J = 8.8 Hz), 5.89 (1H, d, J = 6.0 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.65–4.61 (1H, m), 4.19–4.16 (1H, m), 3.97–3.94 (1H, m), 3.71–3.58 (1H, m), 3.57–3.54 (1H, m), 3.13–3.08 (2H, m), 1.33 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 158.0 (d, J = 238 Hz), 151.4, 150.4, 139.6, 135.6, 122.8, (d, J = 8 Hz), 117.7, 114.4 (d, J = 22 Hz), 87.4, 85.6, 73.4, 70.4, 61.5, 24.8, 14.9. ¹⁹F NMR (376 MHz, DMSO- d_6): $\delta = -119.90$. FT-IR (film, cm⁻¹): 3342, 3277, 3125, 2964, 2927, 1620, 1580, 1506, 1448, 1335, 1220, 1073, 838. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₁FN₅O₄S⁺ m/z 422.1293, found 422.1293.

4.3.5. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((4-chlorophenyl)amino)-9H-purin-9-yl)-5-(hydroxymethyl)-tetrahydrofuran-3,4-diol (5e)

200 mg, yield = 43%; white solid; m.p. = $188-190^{\circ}$ C; $R_f = 0.48$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.16 (1H, s), 8.43 (1H, s), 7.94 (2H, d, J = 9.2 Hz), 7.41 (2H, d, J = 8.8 Hz), 5.89 (1H, d, J = 6.0 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 4.8 Hz), 5.05 (1H, t, J = 5.6 Hz), 4.63 (1H, dd, J = 6.0, 11.2 Hz), 4.17 (1H, dd, J = 4.8, 8.4 Hz), 3.96 (1H, dd, J = 4.0, 8.0 Hz), 3.70–3.65 (1H, m), 3.59–3.54 (1H, m), 3.16–3.11 (2H, m), 1.35 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 151.2, 150.5, 139.8, 138.3, 128.2, 126.5, 122.3, 117.8, 87.4, 85.6, 73.4, 70.4, 61.5, 24.8, 14.8. FT-IR (film, cm⁻¹): 3342, 3288, 3122, 2961, 2928, 1614, 1573, 1491, 1443, 1334, 1192, 1069, 810. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₁ClN₅O₄S⁺ *m/z* 438.0997, found 438.1003.

4.3.6. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-(m-tolylamino)-9H-purin-9-yl)-5-(hydroxymethyl)tetrahydrofuran-3,4-diol (5f)

381 mg, yield = 86%; white solid; m.p. = $168-170^{\circ}$ C; $R_{\rm f}$ = 0.41 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.90 (1H, s), 8.40 (1H, s), 7.80 (1H, app s), 7.62 (1H, d, J = 8.4 Hz), 7.21 (1H, t, J = 8.0 Hz), 6.88 (1H, d, J = 7.2 Hz), 5.88 (1H, d, J = 6.0 Hz), 5.47 (1H, d, J = 6.4 Hz), 5.20 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.61 (1H, dd, J = 6.0, 11.2 Hz), 4.16 (1H, dd, J = 4.8, 8.4 Hz), 3.95 (1H, dd, J = 4.0, 7.6 Hz), 3.70–3.64 (1H, m), 3.59–3.53 (1H, m), 3.16–3.11 (2H, m), 2.31 (3H, s), 1.35 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.6, 151.5, 150.4, 139.5, 139.2, 137.4, 128.2, 123.6, 121.4, 118.1, 117.8, 87.4, 85.6, 73.4, 70.5, 61.5, 24.7, 21.2, 15.0. FT-IR (film, cm⁻¹): 3330, 3247, 3110, 2924, 2869, 1624, 1577, 1486, 1449, 1332, 1219, 1120, 1083, 779, 688. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₄N₅O₄S⁺ m/z 418.1544, found 418.1548.

4.3.7. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((3-methoxyphenyl)amino)-9H-purin-9-yl)-5- (hydroxymethyl)tetrahydrofuran-3,4-diol (5g)

299 mg, yield = 65%; white solid; m.p. = 95–96°C; $R_f = 0.47$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.95 (1H, s), 8.42 (1H, s), 7.65 (1H, t, J = 2.0 Hz), 7.52–7.50 (1H, m), 7.24 (1H, t, J = 8.0 Hz), 6.6–6.63 (1H, m), 5.90 (1H, d, J = 2.0 Hz), 5. 44–5.15 (3H, m), 4.64 (1H, t, J = 7.2 Hz), 4.19–4.17 (1H, m), 3.96–3.98 (1H, m), 3.77 (3H, s), 3.70–3.66 (1H, m), 3.59–3.55 (1H, m), 3.18–3.12 (2H, m), 1.36 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 159.4, 151.4, 150.3, 140.5, 139.6, 129.1, 117.9, 113.0,

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108.1, 106.7, 87.4, 85.5, 73.4, 70.4, 61.5, 55.0, 24.7, 14.9. FT-IR (film, cm⁻¹): 3350, 3235, 3113, 2927, 1624, 1577, 1491, 1454, 1330, 1225, 1157, 1046, 768, 694. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₄N₅O₅S⁺ *m/z* 434.1493, found 434.1497.

4.3.8. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((3-fluorophenyl)amino)-9H-purin-9-yl)-5- (hydroxymethyl)-tetrahydrofuran-3,4-diol (5h)

295 mg, yield = 66%; white solid; m.p. = 176–178°C; $R_{\rm f}$ = 0.48 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.24 (1H, s), 8.46 (1H, s), 7.98–7.94 (1H, m), 7.74–7.71 (1H, m), 7.40–7.34 (1H, m), 6.90–6.85 (1H, m), 5.91 (1H, d, J = 6.0 Hz), 5.50 (1H, d, J = 6.0 Hz), 5.23 (1H, d, J = 4.0 Hz), 5.06 (1H, br s), 4.64 (1H, dd, J = 5.6, 10.8 Hz), 4.18 (1H, m), 3.97 (1H, dd, J = 4.0, 7.6 Hz), 3.70–3.67 (1H, m), 3.59–3.56 (1H, m), 3.19–3.14 (2H, m), 1.38 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 162.0 (d, J = 239 Hz), 151.2, 150.6, 141.1, 139.9, 129.9, 117.9, 116.3, 109.0 (d, J = 21 Hz), 107.2 (d, J = 26 Hz), 87.4, 85.6, 73.4, 70.4, 61.5, 24.8, 14.9. ¹⁹F NMR (376 MHz, DMSO- d_6): δ = -112.59. FT-IR (film, cm⁻¹): 3357, 3303, 3140, 2969, 2928, 1623, 1578, 1490, 1445, 1333, 1253, 1225, 1124, 1078, 810, 681. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₁FN₅O₄S⁺ m/z 422.1293, found 422.1295.

4.3.9. (2R,3R,4S,5R)-2-(6-((3-chlorophenyl)amino)-2-(ethylthio)-9H-purin-9-yl)-5- (hydroxymethyl)-tetrahydrofuran-3,4-diol (5i)

348 mg, yield = 75%; white solid; m.p. = $168-170^{\circ}$ C; $R_{\rm f} = 0.47$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.23 (1H, s), 8.46 (1H, s), 8.24 (1H, t, J = 2.0 Hz), 7.82–7.80 (1H, m), 7.36 (1H, t, J = 8.0 Hz), 7.12–7.09 (1H, m), 5.91 (1H, d, J = 6.0 Hz), 5.50 (1H, d, J = 6.0 Hz), 5.23 (1H, d, J = 4.8 Hz), 5.07 (1H, t, J = 5.6 Hz), 4.63 (1H, dd, J = 5.6, 11.2 Hz), 4.18 (1H, dd, J = 4.4, 8.0 Hz), 3.97 (1H, dd, J = 4.0, 7.6 Hz), 3.71–3.66 (1H, m), 3.61–3.55 (1H, m), 3.19–3.14 (2H, m), 1.38 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 151.2, 150.6, 140.9, 139.9, 132.7, 130.0, 122.3, 120.0, 119.0, 117.9, 87.4, 85.6, 73.5, 70.4, 61.5, 24.8, 14.9. FT-IR (film, cm⁻¹): 3324, 3232, 3119, 2964, 2927, 1623, 1573, 1480, 1447, 1331, 1120, 1082, 777, 706. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₁ClN₅O₄S⁺ *m*/*z* 438.0997, found 438.0997.

4.3.10. (2R,3R,4S,5R)-2-(2-(ethylthio)-6-((3-nitrophenyl)amino)-9H-purin-9-yl)-5- (hydroxymethyl)-tetrahydrofuran-3,4-diol (5j)

214 mg, yield = 45%; white solid; m.p. = 115–116°C; $R_f = 0.34$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.56 (1H, s), 9.14 (1H, s), 8.49 (1H, app s), 8.25–8.23 (1H, m), 7.92–7.90 (1H, m), 7.66–7.62 (1H, m), 5.92 (1H, d, J = 6.0 Hz), 5.23 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.63 (1H, dd, J = 5.6, 11.2 Hz), 4.18 (1H, dd, J = 4.8, 8.4 Hz), 3.97 (1H, dd, J = 4.0, 7.6 Hz), 3.71–3.66 (1H, m), 3.60–3.55 (1H, m), 3.24–3.19 (2H, m), 1.36 (3H, t, J = 7.6 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 151.1, 150.7, 147.9, 140.7, 140.2, 129.7, 126.4, 118.0, 117.0, 114.4, 87.4, 85.6, 73.5, 70.4, 61.5, 24.9, 14.6. FT-IR (film, cm⁻¹): 3324, 3220, 3127, 2927, 1623, 1575, 1528, 1483, 1433, 1349, 1329, 1121, 1077, 789, 697. ESI-HRMS [M + H]⁺ calcd for C₁₈H₂₁N₆O₆S⁺ m/z 449.1238, found 449.1233.

4.3.11. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(2-(methylthio)-6-(phenylamino)-9H-purin-9-yl)tetrahydrofuran-3,4-diol (5k)

281 mg, yield = 68%; white solid, m.p. = 185–186°C; $R_{\rm f}$ = 0.44 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.99 (1H, s), 8.41 (1H, s), 7.91–7.89 (2H, m), 7.36–7.32 (2H, m), 7.07–7.04 (1H, m), 5.90 (1H, d, J = 6.0 Hz), 5.47 (1H, d, J = 6.4 Hz), 5.21 (1H, d, J = 4.8 Hz), 5.04 (1H, t, J = 6.0 Hz), 4.66–4.62 (1H, m), 4.19–4.16 (1H, m), 3.96–3.94 (1H, m), 3.70–3.33 (2H, m), 2.54 (3H, s); ¹³C NMR (100 MHz, DMSO- d_6): δ 164.3, 151.3, 150.4, 139.6, 139.2, 128.4, 122.8, 120.9, 117.7, 87.3, 85.6, 73.4, 70.5, 61.5, 14.0. FT-IR (film, cm⁻¹): 3327, 3286, 3095, 2907, 1618, 1578, 1498, 1444, 1339, 1298, 1076, 745, 691. ESI-HRMS [M + H]⁺ calcd for C₁₇H₂₀N₅O₄S⁺ *m/z* 390.1231, found 390.1239.

4.3.12. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(6-(phenylamino)-2-(propylthio)-9H-purin-9-yl)tetrahydrofuran-3,4-diol (5I)

279 mg, yield = 63%; white solid; m.p. = $159-160^{\circ}$ C; $R_{\rm f} = 0.42$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.00 (1H, s), 8.42 (1H, s), 7.89 (2H, d, J = 8.0 Hz), 7.35 (2H, t, J = 8.0 Hz), 7.07 (1H, t, J = 7.2 Hz), 5.89 (1H, d, J = 6.0 Hz), 5.49 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 4.8 Hz), 5.07 (1H, t, J = 5.2 Hz), 4.66–4.62 (1H, m), 4.18–4.17 (1H, m), 3.97–3.95 (1H, m), 3.71–3.67 (1H, m), 3.60–3.55 (1H, m), 3.13–3.08 (2H, m), 1.69 (2H, sext, J = 7.2 Hz), 0.98 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 151.4, 150.3, 139.6, 139.2, 128.3, 122.9, 120.9, 117.8, 87.4, 85.6, 73.4, 70.5, 61.5, 32.3, 22.8, 13.3. FT-IR (film, cm⁻¹): 3316, 3104, 2964, 2926, 1616, 1577, 1495, 1444, 1330, 1296, 1234, 1080, 750, 690. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₄N₅O₄S⁺ m/z 418.1544, found 418.1552.

4.3.13. (2R,3R,4S,5R)-2-(6-((4-fluorophenyl)amino)-2-(propylthio)-9H-purin-9-yl)-5- (hydroxymethyl)tetrahydrofuran-3,4-diol (5m)

291 mg, yield = 63%; white solid; m.p. = $171-173^{\circ}$ C; $R_{\rm f}$ = 0.41 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.05 (1H, s), 8.41 (1H, s), 7.89–7.86 (2H, m), 7.21–7.17 (2H, m), 5.88 (1H, d, J = 6.0 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.21 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.65–4.61 (1H, m), 4.17 (1H, dd, J = 4.8, 8.4 Hz), 3.96 (1H, dd, J = 4.0, 7.6 Hz), 3.71–3.65 (1H, m), 3.59–3.54 (1H, m), 3.10–3.06 (2H, m), 1.69 (2H, sext, J = 7.2 Hz), 0.98 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 158.0 (d, J = 238 Hz), 151.4, 150.3, 139.6, 135.5, 122.9 (d, J = 8 Hz), 117.7, 114.9 (d, J = 22 Hz), 87.4, 85.6, 73.4, 70.4, 61.5, 32.3, 22.7, 13.3. ¹⁹F NMR (376 MHz, DMSO- d_6): δ = -119.85. FT-IR (film, cm⁻¹): 3313, 3268, 3125, 2961, 2925, 1621, 1581, 1506, 1450, 1332, 1222, 1123, 1081, 835. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₃FN₅O₄S⁺ m/z 436.1449, found 436.1451.

4.3.14. (2R,3R,4S,5R)-2-(6-((4-chlorophenyl)amino)-2-(propylthio)-9H-purin-9-yl)-5-(hydroxymethyl)tetrahydrofuran-3,4-diol (5n)

273 mg, yield = 57%; white solid; m.p. = 175–177°C; $R_f = 0.42$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.15 (1H, s), 8.42 (1H, s), 7.92 (2H, d, J = 8.8 Hz), 7.39 (2H, d, J = 8.8 Hz), 5.88 (1H, d, J = 6.0 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.21 (1H, d, J = 5.2 Hz), 5.05 (1H, t, J = 5.6 Hz), 4.65–4.61 (1H, m), 4.16 (1H, dd, J = 4.8, 8.4 Hz), 3.95 (1H, dd, J = 4.0, 7.6 Hz), 3.69–3.64 (1H, m), 3.59–3.54 (1H, m), 3.12–3.06 (2H, m), 1.71 (2H, sext, J = 7.2 Hz), 0.97 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8,

151.2, 150.5, 139.8, 138.3, 128.2, 126.5, 122.4, 117.9, 87.4, 85.6, 73.4, 70.4, 61.5, 32.4, 22.6, 13.3. FT-IR (film, cm⁻¹): 3326, 3232, 3113, 2967, 2929, 2871, 1620, 1576, 1528, 1492, 1451, 1328, 1236, 1086, 825. ESI-HRMS $[M + H]^+$ calcd for $C_{19}H_{23}ClN_5O_4S^+$ m/z 452.1154, found 452.1155.

4.3.15. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(6-((4-methoxyphenyl)amino)-2-(propyl thio)-9H-purin-9-yl)tetrahydrofuran-3,4-diol (50)

280 mg, yield = 59%; white solid; m.p. = $119-121^{\circ}$ C; $R_{\rm f}$ = 0.38 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.85 (1H, s), 8.37 (1H, s), 7.73 (2H, d, J = 9.2 Hz), 6.93 (2H, d, J = 9.2 Hz), 5.87 (1H, d, J = 6.0 Hz), 5.47 (1H, d, J = 6.4 Hz), 5.20 (1H, d, J = 4.8 Hz), 5.07 (1H, t, J = 5.6 Hz), 4.65–4.61 (1H, m), 4.16 (1H, dd, J = 4.8, 8.0 Hz), 3.95 (1H, dd, J = 4.0, 7.6 Hz), 3.69–3.65 (1H, m), 3.59–3.54 (1H, m), 3.11–3.03 (2H, m), 1.69 (2H, sext, J = 7.2 Hz), 0.98 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 155.3, 151.5, 150.1, 139.3, 132.1, 122.8, 117.6, 113.5, 87.4, 85.5, 73.3, 70.5, 61.5, 55.2, 32.3, 22.8, 13.3. FT-IR (film, cm⁻¹): 3388, 3315, 3125, 2930, 2869, 1621, 1583, 1509, 1455, 1331, 1244, 1119, 1080, 828. ESI-HRMS [M + H]⁺ calcd for C₂₀H₂₆N₅O₅S⁺ *m/z* 448.1649, found 448.1656.

4.3.16. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(2-(propylthio)-6-(p-tolylamino)-9H-purin-9-yl)tetra-hydrofuran-3,4-diol (**5**p)

306 mg, yield = 67%; white solid; m.p. = 179–181°C; $R_f = 0.44$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.89 (1H, s), 8.38 (1H, s), 7.74 (2H, d, J = 8.8 Hz), 7.14 (2H, d, J = 8.4 Hz), 5.87 (1H, d, J = 5.6 Hz), 5.47 (1H, d, J = 6.0 Hz), 5.20 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.2 Hz), 4.64–4.60 (1H, m), 4.16–4.15 (1H, m), 3.95 (1H, dd, J = 4.0, 7.6 Hz), 3.69–3.66 (1H, m), 3.59–3.53 (1H, m), 3.10–3.06 (2H, m), 2.28 (3H, s), 1.70 (2H, sext, J = 7.2 Hz), 0.99 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.7, 151.5, 150.2, 139.4, 136.6, 131.9, 128.7, 121.1, 117.7, 87.4, 85.6, 73.4, 70.5, 61.5, 32.3, 22.8, 20.4, 13.3. FT-IR (film, cm⁻¹): 3325, 3235, 3116, 2922, 2869, 1624, 1580, 1512, 1454, 1335, 1236, 1124, 1083, 812. ESI-HRMS [M + H]⁺ calcd for C₂₀H₂₆N₅O₄S⁺ m/z 432.1700, found 432.1705.

4.3.17. (2R,3R,4S,5R)-2-(6-((3-fluorophenyl)amino)-2-(propylthio)-9H-purin-9-yl)-5-(hydroxymethyl) tetrahydrofuran-3,4-diol (5q)

328 mg, yield = 71%; white solid; m.p. = $155-157^{\circ}$ C; $R_{\rm f}$ = 0.44 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.23 (1H, s), 8.45 (1H, s), 7.96–7.91 (1H, m), 7.73–7.71 (1H, m), 7.40–7.34 (1H, m), 6.90–6.86 (1H, m), 5.90 (1H, d, J = 6.0 Hz), 5.49 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 3.2 Hz), 5.06 (1H, br s), 4.65–4.61 (1H, m), 4.17 (1H, br s), 3.97–3.94 (1H, m), 3.70–3.67 (1H, m), 3.59–3.56 (1H, m), 3.17–3.11 (2H, m), 1.73 (2H, sext, J = 7.2 Hz), 1.01 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 162.0 (d, J = 239 Hz), 151.2, 150.5, 141.2 (d, J = 11 Hz), 139.9, 129.9 (d, J = 10 Hz), 117.9, 116.4, 109.0 (d, J = 21 Hz), 107.3 (d, J = 26 Hz), 87.4, 85.6, 73.4, 70.4, 61.5, 32.4, 22.7, 13.2. ¹⁹F NMR (376 MHz, DMSO- d_6): δ = -112.45. FT-IR (film, cm⁻¹): 3319, 3226, 3134, 2931, 2869, 1621, 1578, 1489, 1444, 1331, 1226, 1124, 1079, 770, 680. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₃FN₅O₄S⁺ m/z 436.1449, found 436.1451.

4.3.18. (2R,3R,4S,5R)-2-(6-((3-chlorophenyl)amino)-2-(propylthio)-9H-purin-9-yl)-5- (hydroxymethyl) tetrahydrofuran-3,4-diol (**5**r)

307 mg, yield = 64%; white solid; m.p. = 137–139°C; $R_f = 0.44$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 10.22 (1H, s), 8.45 (1H, s), 8.19–8.18 (1H, m), 7.83–7.80 (1H, m), 7.38–7.34 (1H, m), 7.12–7.09 (1H, m), 5.89 (1H, d, J = 6.0 Hz), 5.48 (1H, d, J = 6.0 Hz), 5.22 (1H, d, J = 4.8 Hz), 5.06 (1H, t, J = 5.6 Hz), 4.65–4.61 (1H, m), 4.17 (1H, dd, J = 5.2, 8.8 Hz), 3.96 (1H, dd, J = 4.0, 7.6 Hz), 3.70–3.65 (1H, m), 3.60–3.57 (1H, m), 3.16–3.12 (2H, m), 1.73 (2H, sext, J = 7.2 Hz), 0.99 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 151.1, 150.6, 140.9, 139.9, 132.7, 130.0, 122.3, 120.1, 119.0, 117.9, 87.4, 85.6, 73.4, 70.4, 61.5, 32.4, 22.5, 13.2. FT-IR (film, cm⁻¹): 3321, 3225, 3155, 2962, 2929, 2872, 1624, 1573, 1479, 1447, 1331, 1237, 1120, 1079, 774,702. ESI-HRMS [M + H]⁺ calcd for C₁₉H₂₃ClN₅O₄S⁺ m/z 452.1154, found 452.1157.

4.3.19. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(6-((3-methoxyphenyl)amino)-2-(propylthio)-9H-purin-9-yl)tetrahydrofuran-3,4-diol (5s)

332 mg, yield = 70%; white solid; m.p. = 90–92°C; $R_f = 0.40$ (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.94 (1H, s), 8.41 (1H, s), 7.61–7.60 (1H, m), 7.52–7.50 (1H, m), 7.25–7.21 (1H, m), 6.66–6.63(1H, m), 5.88 (1H, d, J = 6.0 Hz), 5.12 (3H, br s), 4.64–4.61 (1H, m), 4.17–4.15 (1H, m), 3.96–3.93 (1H, m), 3.76 (3H, s), 3.69–3.65 (1H, m), 3.58–3.54 (1H, m), 3.12 (2H, dt, J = 4.8, 7.2 Hz), 1.71 (2H, sext, J = 7.2 Hz), 0.99 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 159.3,151.4, 150.3, 140.4, 139.6, 129.1, 117.9, 113.1, 108.1, 106.9, 87.4, 85.5, 73.4, 70.4, 61.5, 55.0, 32.3, 22.7, 13.2. FT-IR (film, cm⁻¹): 3345, 3238, 3107, 2958, 2928, 2869, 1624, 1577, 1492, 1454, 1329, 1287, 1226, 1157, 1084, 776, 685. ESI-HRMS [M + H]⁺ calcd for C₂₀H₂₆N₅O₅S⁺ *m/z* 448.1649, found 448.1648.

4.3.20. (2R,3S,4R,5R)-2-(hydroxymethyl)-5-(2-(propylthio)-6-(m-tolylamino)-9H-purin-9-yl)tetrahydrofuran-3,4-diol (5t)

366 mg, yield = 80%; white solid; m.p. = $182-184^{\circ}$ C; $R_{\rm f}$ = 0.40 (EtOAc:MeOH = 15:1, ν/ν). ¹H NMR (400 MHz, DMSO- d_6): δ 9.89 (1H, s), 8.39 (1H, s), 7.75 (1H, br s), 7.66–7.64 (1H, m), 7.23–7.19 (1H, m), 6.90–6.88 (1H, m), 5.88 (1H, d, J = 6.0 Hz), 5.47 (1H, d, J = 6.0 Hz), 5.20 (1H, d, J = 4.8 Hz), 5.07 (1H, t, J = 5.6 Hz), 4.64–4.61 (1H, m), 4.16 (1H, dd, J = 4.4, 8.0 Hz), 3.95 (1H, dd, J = 4.0,7.6 Hz), 3.69–3.65 (1H, m), 3.59–3.55 (1H, m), 3.14–3.09 (2H, m), 2.31 (3H, s), 1.70 (2H, sext, J = 7.2 Hz), 0.98 (3H, t, J = 7.2 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 163.8, 151.5, 150.3, 139.5, 139.1, 137.4, 128.2, 123.6, 121.6, 118.2, 117.8, 87.4, 85.6, 73.4, 70.5, 61.5, 32.4, 22.6, 21.3, 13.3. FT-IR (film, cm⁻¹): 3331, 3235, 3113, 2958, 2925, 2869, 1625, 1575, 1486, 1450, 1330, 1224, 1084, 779, 687. ESI-HRMS [M + H]⁺ calcd for C₂₀H₂₆N₅O₄S⁺ m/z 432.1700, found 432.1705.

4.4. Radical trapping experiment procedure for 6-chloropurine 3a, diethyl disulfide, and TEMPO

4.4.1. (2R,3R,4R,5R)-2-(acetoxymethyl)-5-(6-chloro-2-((2,2,6,6-tetramethylpiperidin-1-yl)oxy)-9H-purin-9-yl)tetrahydrofuran-3,4-diyl diacetate (6)

211 mg, yield = 35%; colorless oil; $R_{\rm f} = 0.51$ (EtOAc:PE = 1:1, ν/ν). ¹H NMR (400 MHz, CDCl₃): δ 8.01 (1H, s), 6.12 (1H, t, J = 5.2 Hz), 6.06 (1H, d, J = 5.2 Hz), 5.63 (1H, t,

 $J = 4.8 \text{ Hz}, 4.69-4.65 (1H, m), 4.52-4.43 (2H, m), 2.16 (3H, s), 2.11 (3H, s), 2.07 (3H, s), 1.66-1.51 (6H, m), 1.31 (3H, s), 1.30 (3H, s), 1.11 (3H, s), 1.04 (3H, s); ¹³C NMR (100 MHz, CDCl₃): <math>\delta$ 170.4, 169.4, 169.1, 165.2, 152.7, 152.3, 142.5, 128.2, 87.4, 80.4, 72.4, 70.8, 63.2, 61.0, 60.8, 39.4, 39.3, 32.1, 31.8, 20.8, 20.7, 20.5, 20.3, 17.1. FT-IR (film, cm⁻¹): 2975, 2936, 1751, 1608, 1565, 1506, 1436, 1381, 1225, 938. ESI-HRMS [M + H]⁺ calcd for C₂₅H₃₅ClN₅O₈⁺ *m/z* 568.2169, found 568.2299

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ORCID

Ning Chen http://orcid.org/0000-0001-6660-4892 *Hongguang Du* http://orcid.org/0000-0003-3237-4261

References

- Gao H, Mitra AK. Synthesis of acyclovir, ganciclovir and their prodrugs: a review. Synthesis. 2000;2000:329–351.
- [2] Ferrero M, Gotor V. Biocatalytic selective modifications of conventional nucleosides, carbocyclic nucleosides, and C-nucleosides. Chem Rev. 2000;100:4319–4348.
- [3] Priego E-M, Kuenzel JFD, Ijzerman AP, et al. Pyrido[2,1-f]purine-2,4-dione derivatives as a novel class of highly potent human A₃ adenosine receptor antagonists. J Med Chem. 2002;45:3337-3344.
- [4] Cragg GM, Grothaus PG, Newman DJ. Impact of natural products on developing new anticancer agents. Chem Rev. 2009;109:3012–3043.
- [5] Hwu JR, Lin S-Y, Tsay S-C, et al. Coumarin–purine ribofuranoside conjugates as new agents against hepatitis C virus. J Med Chem. 2011;54:2114–2126.
- [6] Barnett DS, Guy RK. Antimalarials in development in 2014. Chem Rev. 2014;114:11221-11241.
- [7] Ingall AH, Dixon J, Bailey A, et al. Antagonists of the platelet P₂T receptor: a novel approach to antithrombotic therapy. J Med Chem. 1999;42:213–220.
- [8] Cristalli G, Podda GM, Costanzi S, et al. Effects of 5'-phosphate derivatives of 2-hexynyl adenosine and 2-phenylethynyl adenosine on responses of human platelets mediated by P2Y receptors. J Med Chem. 2005;48:2763–2766.
- [9] Thomson PF, Lagisetty P, Balzarini J, et al. Palladium-catalyzed aryl amination reactions of 6-bromo- and 6-chloropurine nucleosides. Adv Synth Catal. 2010;352:1728–1735.
- [10] Xia R, Niu H-Y, Qu G-R, et al. Cui controlled C–C and C–N bond formation of heteroaromatics through C(sp³)-H activation. Org Lett. 2012;14:5546–5549.
- [11] Conroy S, Kindon N, Kellam B, et al. Drug-like antagonists of P2Y receptors—from lead identification to drug development. J Med Chem. 2016;59:9981–10005.
- [12] Springthorpe B, Bailey A, Barton P, et al. From ATP to AZD6140: The discovery of an orally active reversible P2Y₁₂ receptor antagonist for the prevention of thrombosis. Bioorg Med Chem Lett. 2007;17:6013–6018.
- [13] Urgaonkar S, Xu J-H, Verkade JG. Scope and limitations of Pd₂(dba)₃/P(*i*-BuNCH₂CH₂)₃Ncatalyzed Buchwald-Hartwig amination reactions of aryl chlorides. J Org Chem. 2004;69: 9135-9142.

- [14] Xia R, Xie M-S, Niu H-Y, et al. Radical route for the alkylation of purine nucleosides at C6 via Minisci reaction. Org Lett. 2014;16:444–447.
- [15] Wang D-C, Xia R, Xie M-S, et al. Synthesis of cycloalkyl substituted purine nucleosides via a metal-free radical route. Org Biomol Chem. 2016;14:4189–4193.
- [16] Liu G, Xu J, Chen N, et al. Synthesis of N⁶-alkyl(aryl)-2-alkyl(aryl)thioadenosines as antiplatelet agents. Eur J Med Chem. 2012;53:114–123.
- [17] Du H, Sun X, Yu M, et al. Synthesis of 8-alkoxy-6-alkylamino-2-alkylthiopurine nucleosides with a straightforward multiple-functionalization strategy. Tetrahedron Lett. 2016;57: 2949–2953.
- [18] Zhang S, Hu L, Du H, et al. BF0801, a novel adenine derivative, inhibits platelet activation via phosphodiesterase inhibition and P2Y12 antagonism. Thromb Haemost. 2010;104:845–857.
- [19] Nair V, Richardson SG. Utility of purinyl radicals in the synthesis of base-modified nucleosides and alkylpurines: 6-amino group replacement by hydrogen, chlorine, bromine, and iodine. J Org Chem. 1980;45:3969–3974.
- [20] Born GVR. Aggregation of blood platelets by adenosine diphosphate and its reversal. Nature. 1962;194:927–929.
- [21] Creech C, Kanaujia M, Causey CP. Synthesis and evaluation of 2-ethynyl-adenosine-5'-triphosphate as a chemical reporter for protein AMPylation. Org Biomol Chem. 2015;13:8550-8555.
- [22] Du H, He Q, Chen N, et al. Proton NMR investigations on 6-alkylamino-2-alkylthioadenosine derivatives. Magn Reson Chem. 2015;53:218–222.