Novel peptidomimetic structures: enantioselective synthesis of conformationally constrained lysine, ornithine and alanine analogues from pyroglutamic acid

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Conformationally constrained lysine and ornithine analogues, and an L-Ala-L-Ala dipeptide analogue, are available from pyroglutamic acid.

The synthesis of conformationally constrained amino acids is of considerable current interest;^{1–4} in addition to their intrinsic interest as ligands for a wide variety of biological receptors, incorporation of these structural elements into peptide chains can be used to generate novel structures of relevance to biological or materials application.⁵ Although structurally restricted analogues of a number of amino acids have been described,^{6–10} the ω -amino acids have generally only recently begun to attract attention. However, modified lysine chimeras, derived from pyroglutamic acid,¹¹ and from proline,⁹ and a peptidomimetic which includes a conformationally restricted lysine analogue¹² have all recently been reported, as have ornithine¹³ and arginine analogues.¹⁴ The synthesis of cyano¹⁵ or indole¹⁶ substituted glutamate analogues has also recently been described.

We have used the readily available lactam **1a** as a template for manipulation to a variety of functionalised pyrrolidinones^{17–20} and recently shown its application to the synthesis of conformationally restricted glutamates²¹ and aminopyrrolidones.²² We report here the extension of this versatile approach to the synthesis of several other conformationally restricted amino acids. The well-defined conformation of pyroglutamic acid has been investigated in detail²³ and its application as a template for peptidomimetics previously proposed;²⁴ the pyrrolidone ring simultaneously restricts τ_1 , τ_2 and τ_3 to very limited ranges (from a simple molecular modeling energy minimised structure,²⁵ these are -144, +23 and -17° respectively) and defines the *cis*-amide bond (Fig. 1).²⁶

The lysine chimera was obtained as follows: the enolate of lactam **1a** was treated with BrCH₂CN (Scheme 1), unusually to give exclusively the *endo* adduct **2** in 55% yield;²⁷ similar alkylations generally proceed under thermodynamic control to give the *exo* product.¹⁹ The *cis* stereochemistry of **2** and **4** was assigned on the basis of NOE data. Reduction of the nitrile function with NaBH₄–CoCl₂ gave the corresponding amine **3** in 76% yield, and this intermediate was easily converted to the product **4** in a four step (protection, deprotection, oxidation and *in situ* esterification) sequence in 16% overall yield.

The ornithine chimera was obtained from lactam **1b**. Selenenation and elimination to the known enone **5** followed by conjugate addition of the Reformatsky reagent derived from BrCH₂CN gave adduct **6a** in 69% yield as a single diastereomer, as shown by ¹³C NMR spectroscopic analysis. This strategy has proved to be very successful for manipulation of this position of a pyrrolidone ring.²¹ Hydrolysis and decarboxylation using $(Bu_3Sn)_2O^{28}$ readily afforded the product **6b** in 68% yield, and





Scheme 1 Reagents and conditions: i, LDA, THF, -78 °C then BrCH₂CN (55%); ii, NaBH₄, CoCl₂, EtOH; iii, ZCl, Et₃N; iv, TFA; v, RuO₂, NaIO₄ then CH₂N₂ (34%); vi, Zn, DMPU, BrCH₂CN (69%), room temp.; vii, (Bu₃Sn)₂O, toluene, Δ , 16 h (68%); viii, PDC, DMF, then CH₂N₂ (35%).

a similar sequence to that described above gave the product 7 in 14% yield over the four steps. In this case, however, application of RuO_4 in the final oxidation step did not give the desired product, and this step was successful only with PDC/DMF. The *trans* relative stereochemistry of **6a** and **7** was again shown by NOE data.

The presence of an internal amide bond suggested that amination at the C-7 position of **1a** could be used to generate an unusual dipeptide mimetic. Related aminopyrrolidones, aminopiperidones and larger ring heterocycles have recently attracted interest as enzyme inhibitors²⁹ and peptidomimetic structures.^{30–33} Amination of the enolate of lactam 1a with (PhO)₂P(O)N₃ followed by treatment with Boc₂O gave the amino lactam 8 in 50% yield (Scheme 2); surprisingly, the endo product, whose stereochemistry was subsequently established, was obtained exclusively. Acidic release of the protecting groups, and reprotection of the C-4 amino function as its benzyloxycarbonyl (Z) derivative, gave the product 9b in 51% yield over the two steps. Oxidation and esterification in the usual way then gave the product 10, which was a single stereoisomer at room temperature by NMR analysis, and whose cis stereochemistry was shown by NOE spectroscopy. Molecular modeling of the N-acetyl analogue $1\overline{1}$ of compound 10^{25} demonstrated that two well defined conformations existed, differing by 2.3 kcal mol⁻¹ in energy, with the two substituents either pseudodiequatorial or diaxial, and the latter being the more stable; since a 2.7 kcal mol⁻¹ energy difference corresponds to a 99:1 ratio of species at equilibrium,³⁴ the



Scheme 2 Reagents and conditions: i, LDA, THF, -78 °C, then (PhO)₂P(O)N₃, then Boc₂O, $-78 \rightarrow O$ °C (50%) ii, TFA, CH₂Cl₂, room temp., 1 h (quant.); iii, ZCl, DMF-THF, Et₃N, 0 °C, 3 h (51%); iv, PDC, DMF, 40 °C, 12 h, then CH₂N₂ (28%).



Fig. 2 Conformations of 12a and 12b.

minor diequatorial conformer would not be expected to be observable at room temperature by NMR analysis, and ¹H NMR VT analysis provided no evidence for the diequatorial conformer even at 223 K. The stability of the diaxial conformer could be attributed to the presence of A-strain³⁵ between the two substituents and the planar amide system; the importance of torsional strain in five- and six-membered ring heterocycles for the control of stereochemistry has been investigated in detail,³⁶ although its importance in lactams has only recently been appreciated.37 Thus, the diaxial conformer minimises the interations of the relatively bulky C(2) and C(4) substituents with the planar amide function (which would occur in the diequatorial conformer 12a) by placing C(2)-H and C(4)-H in an eclipsing conformation with the lactam system 12b (Fig. 2). Using the energy minimised structure for 11, some calculated dihedral angles are given in Table 1; the diaxial conformer most closely resembles a Type VIa (cis) β -turn.³⁸ Thus, this compound could be considered to be a conformationally restricted L-Ala-L-Ala dipeptide analogue, with the central amide bond constrained in the *cis* orientation, and the pyrrolidone ring capable of providing a reverse turn in an attached peptide chain; as such it represents a potential low molecular weight non-hydrophobic turn inducer. A related aminopyrrolidone has also been reported to induce Type II' β turn folding in a short peptide, and to exhibit hypoglycaemic activity.39

Table 1 Dihedral angles for energy minimised^a conformations of 11

AcHN Ψ_1 Ψ_1 Ψ_2 Ψ_2 Ψ_2 Θ_2 Ψ_2 Θ						
	Conformation	Ψ_1 (°)	${\it I}\!$			
	Diequatorial Diaxial	-139 +94	+137 -97			

^a Structures optimised with Chem3D Pro 3.5, available from Cambridge Scientific (MM2 parameters).

In view of the increasing interest in the use of variously modified proline derivatives for subtle conformational control in short peptide sequences,^{40–44} the ready synthetic accessibility of enantiopure functionalised pyrrolidones may enable their application as amino acid surrogates particularly where well-defined conformational restriction is required.

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Notes and references

- 1 M. D. Fletcher and M. M. Campbell, Chem. Rev., 1998, 98, 763.
- 2 A. Dutta, in *Specialist Periodical Reports 'Amino acids, Peptides and Proteins'*, ed. J. S. Davies, Royal Society of Chemistry, Cambridge, 1998, ch. 3.
- 3 S. Hanessian, G. McNaughton-Smith, H.-G. Lombart and W. D. Lubell, *Tetrahedron*, 1997, **53**, 12789.
- 4 J. Gante, Angew. Chem., Int. Ed. Engl., 1994, 1699.
- 5 S. H. Gellman, Acc. Chem. Res., 1998, 31, 173.

- 6 S. Hannessian, N. Bernstein, R.-Y. Yang and R. Maguire, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 1437.
- 7 S. Hannessian and R. Margarita, Tetrahedron Lett., 1998, 39, 5887.
- 8 S. Hannessian, R. Margarita, A. Hall and X. Luo, *Tetrahedron Lett.*, 1998, **39**, 5883.
- 9 Q. Wang, N.A. Sasaki and P. Potier, Tetrahedron, 1998, 54, 15759.
 - 10 R. Sharma and W. D. Lubell, J. Org. Chem., 1996, 61, 202.
 - 11 P. J. Murray, Tetrahedron Lett., 1998, 39, 6721.
 - 12 S. Cappelletti, M. Pegna, A. Zaliani and M. Pinori, *Lett. Pept. Sci.*, 1995, 2, 161.
 - 13 J. Eustache, A. Grob, C. Lam, O. Sellier and G. Schulz, *Bioorg. Med. Chem. Lett.*, 1998, 8, 2961.
 - 14 R. Zhang, A. Mamai and J. S. Madalengoitia, J. Org. Chem., 1999, 64, 547.
 - 15 C. Dugave, J. Cluzeau, A. Menez, M. Gaudry and A. Marquet, *Tetrahedron Lett.*, 1998, **39**, 5775.
 - 16 M. F. Brana, M. Garranzo and J. Perez-Castells, *Tetrahedron Lett.*, 1998, **39**, 6569.
 - 17 J. H. Bailey, D. T. Cherry, K. M. Crapnell, M. G. Moloney, S. B. Shim, M. Bamford and R. B. Lamont, *Tetrahedron*, 1997, 53, 11 731.
 - 18 M. Bamford, M. Beard, D. T. Cherry and M. G. Moloney, *Tetrahedron: Asymmetry*, 1995, 6, 337.
 - 19 M. J. Beard, J. H. Bailey, D. T. Cherry, M. G. Moloney, S. B. Shim, K. Statham, M. Bamford and R. B. Lamont, *Tetrahedron*, 1996, 52, 3719.
 - 20 J. Dyer, S. Keeling and M. G. Moloney, *Tetrahedron Lett.*, 1996, 37, 4573.
 - 21 J. Dyer, S. Keeling and M. G. Moloney, *Chem. Commun.*, 1998, 461.
 - 22 P. W. H. Chan, I. F. Cottrell and M. G. Moloney, *Tetrahedron Lett.*, 1997, **38**, 5891.
 - 23 P. K. C. Paul, D. J. Osguthorpe and M. M. Campbell, J. Chem. Soc., Perkin Trans. 1, 1990, 3363.
 - 24 P. K. C. Paul, P. A. Burney, M. M. Campbell and D. J. Osguthorpe, Bioorg. Med. Chem. Lett., 1992, 2, 141.
 - 25 Structures optimised with Chem3D Pro 3.5, available from Cambridge Scientific (MM2 Parameters).
 - 26 J. A. Monn, M. J. Valli, R. A. True, D. D. Schoepp, J. D. Leander and D. Lodge, *Bioorg. Med. Chem. Lett.*, 1993, 3, 95.
 - 27 All new compounds gave satisfactory spectroscopic and/or high resolution mass spectrometric or analytical data.
 - 28 C. J. Salomon, E. G. Mata and O. A. Mascarotti, J. Org. Chem., 1994, 59, 7259.
 - 29 R. Shankar and A. I. Scott, Heterocycles, 1996, 42, 145.
 - 30 T. Lehmann, D. Michel, M. Glanzel, R. Waibel and P. Gmeiner, *Heterocycles*, 1999, **51**, 1389.
 - 31 A. Nouvet, M. Binard, F. Lamaty, J. Martinez and R. Lazaro, *Tetrahedron*, 1999, 55, 4685.
 - 32 P. Benovsky, G. A. Stephenson and J. R. Stille, J. Am. Chem. Soc., 1998, 120, 2493.
 - 33 S. Derrer, N. Feeder, S. J. Teat, J. E. Davies and A. B. Holmes, *Tetrahedron Lett.*, 1998, **39**, 9309.
 - 34 N. Isaacs, Physical Organic Chemistry, Longman, London, 1995.
 - 35 R. W. Hoffmann, Chem. Rev., 1989, 1841.
 - 36 D. Seebach, B. Lamatsch, R. Amstutz, A. K. Beck, M. Dobler, M. Egli, R. Fitzi, M. Gautschi, B. Herradon, P. C. Hidber, J. J. Irwin, R. Locher, M. Maestro, T. Maetzke, A. Mourino, E. Pfammatter, D. A. Plattner, C. Schlickli, W. B. Schweizer, P. Seiler, G. Stucky, W. Petter, J. Escalante, E. Juaristi, D. Quintana, C. Miravitlles and E. Molins, *Helv. Chim. Acta*, 1992, **75**, 913.
 - 37 K. Ando, N. S. Green, Y. Li and K. N. Houk, J. Am. Chem. Soc., 1999, 121, 5334.
 - 38 G. D. Rose, L. M. Gierasch and J. A. Smith, Adv. Protein Chem., 1985, 37, 1.
 - 39 N. J. Ede, I. D. Rae and M. T. W. Hearn, *Tetrahedron Lett.*, 1990, **31**, 6071.
 - 40 L. Halab and W. D. Lubell, J. Org. Chem., 1999, 64, 3312.
 - 41 R. Zhang and J. S. Madelengoitia, J. Org. Chem., 1999, 64, 547.
 - 42 R. Zhang, F. Brownewell and J. S. Madalengoitia, J. Am. Chem. Soc., 1998, **120**, 3894.
 - 43 E. Beausoleil, R. Sharma, S. W. Michnick and W. D. Lubell, J. Org. Chem., 1998, 63, 6572.
 - 44 P. Dumy, M. Keller, D. E. Ryan, B. Rohwedder, T. Wohr and M. Mutter, J. Am. Chem. Soc., 1997, **119**, 918.
 - 45 D. A. Fletcher, R. F. McMeeking and D. Parkin, J. Chem. Inf. Comput. Sci., 1996, 36, 746.

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