

Bioorganic & Medicinal Chemistry Letters 10 (2000) 403-406

The Discovery and Synthesis of Highly Potent, A_{2a} Receptor Agonists

Suzanne E. Keeling, ^{a,*} F. David Albinson, ^b Barry E. Ayres, ^a Peter R. Butchers, ^a
C. Lynn Chambers, ^a Peter C. Cherry, ^a Frank Ellis, ^a George B. Ewan, ^a
Michael Gregson, ^a John Knight, ^a Keith Mills, ^b Paul Ravenscroft, ^b
Linda H. Reynolds, ^a Shahin Sanjar^c and Michael J. Sheehan^a

^aMedicinal Sciences, Glaxo Wellcome Medicines Research Centre, Gunnels Wood Road, Stevenage, Herts, SG1 2NY, UK ^bChemical Development, Glaxo Wellcome Medicines Research Centre, Gunnels Wood Road, Stevenage, Herts SG1 2NY, UK ^cDisease Sciences, Glaxo Wellcome Medicines Research Centre, Gunnels Wood Road, Stevenage, Herts SG1 2NY, UK

Received 16 November 1999; accepted 5 January 2000

Abstract—A series of N6,2-disubstituted adenosine analogues have been synthesized and their functional activity measured against A_{2a} and A_1 receptors. Examples of compounds with both a lipophilic N6-substituent and amino-functionalized 2-position were highly active at the A_{2a} receptor on the human neutrophil. © 2000 Published by Elsevier Science Ltd. All rights reserved.

Introduction

The actions of adenosine, and the family of four Gprotein coupled 7-trans-membrane receptors (A_1 , A_{2a} , A_{2b} , A_3) which facilitate its physiological effects, have been of great interest for a number of years.^{1–3} Adenosine is used clinically for the treatment of supraventricular tachycardia,⁴ but its lack of selectivity coupled with its rapid metabolism precludes its therapeutic use to any great degree. Consequently much effort has been focused on the search for metabolically stable and selective purinergic receptor agonists.⁵ In this communication we will describe the discovery and synthesis of a series of highly potent A_{2a} purinergic agonists.

Results and Discussion

The structure–activity relationships of adenosine analogues using binding assays have been well-documented.^{5,6} As part of our own general investigation into the structure–activity relationships of substituted adenosines using functional assays, we have investigated the effects on A_{2a} and A_1 activity of substitution at the 2 and N6 positions of the adenine ring, as shown in Table 1. We noted that substitution at the 6-position of adenine ribosides generally conferred A_1 agonist activity whereas 2-substitution improved activity at the A_{2a} receptor, although there were many subtleties. Two pairs of compounds illustrate the marked influence of the position of substitution on activity: moving the cyclopentyl substituent from N6 in *N*-cyclopentyl-2chloroadenosine **1** to the 2 position in 2-cyclopentylaminoadenosine **2** reduced the A_1 activity by three orders of magnitude and improved the A_{2a} activity 3-fold. The related hydroxylated compounds **3** and **4** exhibit a similar relationship: in this case, the A_{2a} activity increased by two orders of magnitude and the A_1 activity is decreased ~40-fold.

The ability to obtain potent agonists at the adenosine A_{2a} receptor which inhibited human neutrophil function^{7,8} offered us the opportunity to investigate further the potential therapeutic application of A_{2a} agonists in the treatment of inflammatory diseases. Selective and potent A_{2a} agonists were required and our medicinal chemistry strategy explored three main areas: 2-alkylaminoadenosines, 2-alkylamino-NECAs and 2-alkylamino-N6-arylalkyl-NECAs.

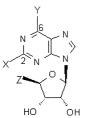
We found that compounds with cycloalkyl substituents at the 2 position e.g. **2** and **6**, were more potent at the A_{2a} receptor than the corresponding acyclic-alkylamino compound **7**, and the incorporation of a hydroxyl group

^{*}Corresponding author. Fax: +44-1438-763438; e-mail: ser0079@ glaxowellcome.co.uk

⁰⁹⁶⁰⁻⁸⁹⁴X/00/\$ - see front matter O 2000 Published by Elsevier Science Ltd. All rights reserved. PII: S0960-894X(00)00017-2

into the 2-cyclopentyl ring yielded A_{2a} agonists more potent than NECA **5**, and with significantly reduced A_1 activity. This is illustrated by the pair of compounds **8** and **9**, and notably the 4'-*N*-ethylcarboxamide substituent present in compound **9** confers several fold improvement in activity as compared to the 4'-hydroxymethyl substituent in **8**. Work investigating N6- substituted analogues in the 4'-hydroxymethyl series had resulted in the discovery of moderately active A_{2a} agonists (compounds **10–12**), but with comparatively lower

Table 1. Relationship between substituent position and activity^c



		10	Off		
	Х	Y	Z	$A_{2a}{}^{a}\left(HN\right)$	A1 ^b (GPI)
1	Cl	NH-	HOCH ₂ -	17	1
2	MH-	H ₂ N-	HOCH ₂ -	5.1	549
3	Cl	OHNH-	HOCH ₂ -	171	2
4	OH OH	H ₂ N-	HOCH ₂ -	1.8	84
5 6	H	H ₂ N- H ₂ N-	EtNH.CO- EtNH.CO-	1 2.2	1 80
7	NH-	H ₂ N-	EtNH.CO-	85	No data
8		H ₂ N-	HOCH ₂ -	0.8	1470
9	OH NH-	H ₂ N-	EtNH.CO-	0.1	260
10	NH-	1-Naphthyl-CH ₂ NH-	HOCH ₂ -	25	No data
11	NH-	3-ClPhEtNH-	HOCH ₂ -	39	>1000
12	NH-	Ph ₂ CHCH ₂ NH-	HOCH ₂ -	36	>612
13	NH-	Ph ₂ CHCH ₂ NH-	EtNH.CO-	6.2	>5000
14	OH NH-	Ph ₂ CHCH ₂ NH-	EtNH.CO-	0.16	390
15	O _{NH-}	1-Naphthyl-CH ₂ NH-	HOCH ₂ -	No data	No data

^aAgonist activities were measured in vitro using inhibition of human neutrophil activation^{9,10} (HN).

^bInhibition of guinea pig Ileum^{9,10} twitch (GPI).

°NECA (5'-*N*-ethylcarboxamidoadenosine) **5** was used as an internal standard^{9–11} and all agonist potencies are quoted as an equieffective concentration ratio (ECR) relative to NECA. ECR values were determined by dividing the EC_{50} of the test compound by the EC_{50} of NECA.

A₁ activity than the unsubstituted compound **2**. Replacement of the 4'- hydroxymethyl group in **12** by the 4'-*N*-ethylcarboxamide substituent again increased the A_{2a} activity several fold (compound **13**), as did hydroxylation of the 2-cyclopentylamino ring (**14**). Interestingly, rat binding data was reported in the literature¹² at this time on N6- arylalkyl compounds, such as **15**. This compound has a K_i = 10300 nM at A₂ receptors on rat striatal membranes and K_i = 45000 nM at A₁ on rat whole-brain membranes, compared to its 2-H analogue which had figures of 9.4 and 24 nM respectively. The observed weakened binding affinities when a 2-cyclohexylamine substituent is present is contrasted by our functional results on compound **10** where agonist activity is retained at the A_{2a} receptor on the human neutrophil.

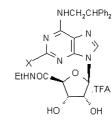
At this point, we had identified several potent compounds, but their solubilities were generally poor and potentially a hindrance to their utility as biological tools. We focused upon analogues of **13** and **14** and sought to improve solubility within the series by incorporating a basic amino group in the 2 substituent. Additionally, it had been reported¹³ that NECA analogues with large 2 substituents possessed significant A_2 selectivity. The results are shown in Table 2.

Compounds 16–18 are examples of analogues which showed significantly improved aqueous solubility compared to that of 13, in addition to activity at the A_{2a} receptor on the human neutrophil at least 10 times more potent than NECA 5 itself.

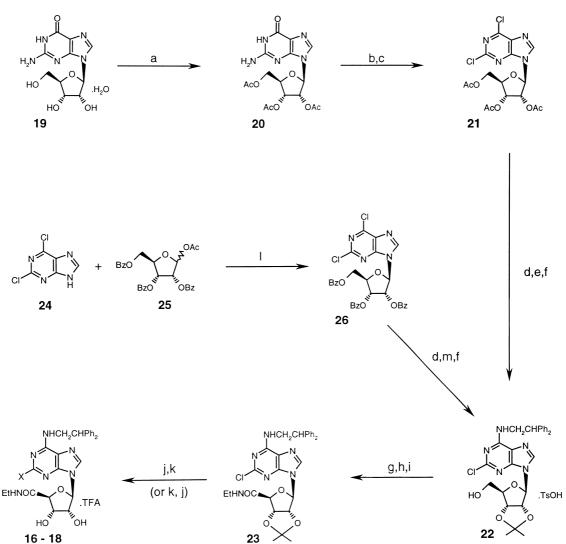
Synthesis of A2a Agonists with Improved Solubility

The analogues 16–18, were prepared from the common intermediate 23, synthesised using one of the two procedures shown in Scheme 1. Compound 22 was reached either from guanosine hydrate 19 in six steps, or in four steps from 2,6-dichloropurine 24 and a fully-protected ribofuranose sugar 25. Tri-acetylation of 19 gave 20,





	Х	$A_{2a}\left(HN\right)$	A1 (GPI)	Solubility pH 2.6 (mg/ml)
13	NH - (free base)	6.2	>5000	0.024
16	NM-	0.08	263	>=1.87
17	H ₂ N [•] NH-	0.05	369	1.57
18	HN NH-	0.03	672	>=1.82



Scheme 1. Synthesis of A_{2a} agonists 16–18. X is defined as in Table 2. Reagents: (a) Ac₂O, Et₃N, MeCN, DMAP; (b) POCl₃, PhNMe₂, Et₄N⁺Cl⁻, MeCN, reflux; (c) *t*-BuONO, Pyridine.HCl, CuCl, DCM, 0–20 °C; (d) Ph₂CHCH₂NH₂, *i*-Pr₂NEt, *i*-PrOH, reflux; (e) Na₂CO₃, MeOH, H₂O; (f) Me₂C(OMe)₂, TsOH, Me₂CO; (g) KMnO₄, KOH, Dioxan; (h) SOCl₂, reflux; (i) EtNH₂, DCM; (j) XH, DMSO, 120 °C; (k) TFA, H₂O; (l) 140 °C, neat; (m) K₂CO₃, MeOH.

which was converted to 21 using a two-step procedure via the 6-chloro derivative followed by diazotization and a modified Sandmeyer reaction. Substitution of the 6-Cl by 2,2-diphenylethylamine in the presence of diisopropylethylamine in propan-2-ol was followed by removal of the acetate protecting groups and reprotection of the 2',3'-diol as the acetonide to yield **22** as its p-toluenesulfonic acid salt. Alternatively, 26 was synthesised directly by heating a mixture of 24 and 25, and yielded 22 following steps similar to the conversion of 21. Oxidation of the free hydroxyl group yielded the intermediate acid which was elaborated to the ethyl amide 23. Substitution of the 2-Cl with a range of amines at 120 °C in DMSO and deprotection using aqueous TFA, or vice versa, followed by purification using preparative HPLC yielded the series of compounds 16–18 as their trifluoroacetic acid salts.

Conclusions

N6,2-disubstituted adenosine analogues have been synthesized and shown to be potent A_{2a} agonists at receptors on the human neutrophil with low activity at the A_1 receptors in the system studied. The improved physical properties will allow their pharmacological effects to be further evaluated for use as potential agents in the treatment of inflammatory diseases.

Acknowledgements

The authors gratefully acknowledge the support of Physical Sciences in this programme, and for helpful discussions with Brian Cox and Richard P. C. Cousins in the preparation of the manuscript.

References and Notes

- 1. Daly, J. W. J. Med. Chem. 1982, 25, 197.
- 2. Jacobsen, K. A.; van Galen, P. J. M.; Williams, M. J. Med. Chem. 1992, 35, 407.
- 3. Poulsen, S.-A.; Quinn, R. J. Bioorg. Med. Chem. 1998, 6, 619.
- 4. Pantely, G. A.; Bristow, J. D. Circulation 1990, 82, 1854.
- 5. Trivedi, B. K.; Bridges, A. J.; Bruns, R. F. In Adenosine and Adenosine Receptors; Williams, M., Ed.; Humana: Clifton, NJ,
- **1990**; pp 57–103.
- 6. Siddiqi, S. M.; Jacobsen, K. A.; Esker, J. L.; Olah, M. E.; Ji,
- X.; Melman, N.; Tiwari, K. N.; Secrist, III, J. A.; Schneller, S.
- W.; Cristalli, G.; Stiles, G. L.; Johnson, C. R.; Ijzerman, A. P. *J. Med. Chem.* **1995**, *38*, 1174 and pp 627–628 and references therein.
- 7. Marone, G.; Rosaria, P.; Sergio, V. Int. Arch. Allergy Appl. Immunol. 1985, 77, 259.

- 8. Cronstein, B. N.; Rosenstein, E. D.; Kramer, S. B.; Weissmann, G.; Hirschhorn, R. J. Immunol. 1985, 135, 1366.
- 9. Gurden, M. F.; Coates, J.; Ellis, F.; Evans, B.; Foster, M.; Hornby, E.; Kennedy, I.; Martin, D. P.; Strong, P.; Vardey, C. J.; Wheeldon, A. *Br. J. Pharmacol.* **1993**, *109*, 693.
- 10. Kennedy, I.; Gurden, M.; Strong, P. Gen. Pharmacol. 1992, 23, 303.
- 11. The assays are fully described in ref 7. NECA **5** has measured EC_{50} of 5nM (4.1–6.1; n=46) on HN and 52 nM (4.4–6.1; n=20) on GPI: EC_{50} figures for NECA are geometric mean values, with 95% confidence limits shown in parentheses where n= number of observations. Where ECR figures are based on n is >=2, arithmetic mean values are given.
- 12. Trivedi, B. K.; Bruns, R. F. J. Med. Chem. **1989**, 32, 1667. 13. Francis, J. E.; Webb, R. L.; Ghai, G. R.; Hutchison, A. J.; Moskal, M. A.; de Jesus, R.; Yokoyama, R.; Rovinski, S. L.; Contrado, N.; Dotson, R.; Barclay, B.; Stone, G. A.; Jarvis, M. F. J. Med. Chem. **1991**, 34, 2570.