# **Communications**



#### H<sub>2</sub> Production

F. Wang, W.-J. Liang, J.-X. Jian, C.-B. Li, B. Chen, C.-H. Tung, L.-Z. Wu\* \_\_\_\_\_

Exceptional Poly(acrylic acid)-Based Artificial [FeFe]-Hydrogenases for Photocatalytic  $H_2$  Production in Water



**Light, polymer, action**: A set of watersoluble poly(acrylic acid) catalysts PAA-g-Fe<sub>2</sub>S<sub>2</sub> containing {Fe<sub>2</sub>S<sub>2</sub>}, an [FeFe]hydrogenase active-site mimic, is synthesized. This system, combined with CdSe quantum dots and ascorbic acid, has an exceptional turnover number and initial turnover frequency (27135 and 3.6 s<sup>-1</sup>) for the photocatalytic production of H<sub>2</sub> in water, which is the highest efficiency to date for [FeFe]-hydrogenase mimics.

### H<sub>2</sub> Production

# **Exceptional Poly(acrylic acid)-Based Artificial [FeFe]-Hydrogenases** for Photocatalytic H<sub>2</sub> Production in Water\*\*

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Generation of hydrogen (H<sub>2</sub>) from water by solar-energy conversion is considered a promising way to deal with the energy crisis and climate change.<sup>[1,2]</sup> One of the key challenges at this stage is to create catalysts for H<sub>2</sub> production with high efficiency and low cost. [FeFe]-hydrogenase, an enzyme in algae, is the fastest proton reduction catalyst in nature known to date.<sup>[3]</sup> The H<sub>2</sub> production rate of the active site of [FeFe]hydrogenase, a Fe<sub>2</sub>S<sub>2</sub> subunit coordinated by CO and CN ligands, achieves turnovers as high as 6000–9000 molecular H<sub>2</sub> per active site per second.<sup>[4]</sup> Such an efficient catalyst with its noble-metal-free structure has aroused much interest in the last decade.<sup>[5,6]</sup> With reference to the crystal structure of natural [FeFe]-hydrogenase,<sup>[7,8]</sup> a large number of [FeFe]hydrogenase mimics have been synthesized on the basis of the Fe<sub>2</sub>S<sub>2</sub> cluster.<sup>[9-19]</sup> From a photochemical point of view, molecular dyads and triads,<sup>[20-25]</sup> multi-component systems,<sup>[26-33]</sup> and assembled hybrid systems,<sup>[34-37]</sup> have been developed. Although two systems with [FeFe]-hydrogenase mimics as catalysts performed H<sub>2</sub> photo-production with a turnover number (TON) of over 200,<sup>[27,28]</sup> most of analogous systems finished their photochemical H<sub>2</sub> production with low turnover numbers (TON < 5) in organic solutions or a mixture of organic solvents and water.<sup>[12,38,39]</sup> In 2011, we designed a robust water-soluble [FeFe]-hydrogenase mimic by linking three hydrophilic ether chains to the  $Fe_2S_2$  active site, and achieved for the first time photocatalytic H<sub>2</sub> production in water.<sup>[29]</sup> With this water-soluble [FeFe]-hydrogenase mimic as a catalyst, CdTe quantum dots (QDs) as a photosensitizer (PS), and ascorbic acid (H<sub>2</sub>A) as a proton source and sacrificial electron donor, the system exhibited a high efficiency for photocatalytic  $H_2$  production (TON = 505). Since then, incorporation of the  $Fe_2S_2$  active site to a water-soluble group,<sup>[29,30]</sup> protein,<sup>[31]</sup> and peptide,<sup>[32]</sup> even the hydrophobic cavity of cyclodextrins,<sup>[33]</sup> has been developed to realize photocatalytic H<sub>2</sub> production in water because water is

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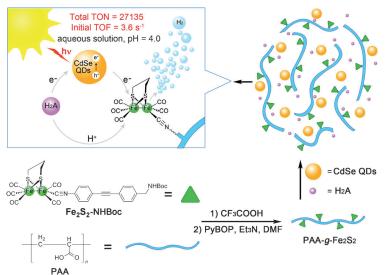
believed to be an ideal solvent for proton mobility and has non-toxic as well as economic advantages.<sup>[40,41]</sup> Indeed, the TON of these water-soluble systems is enhanced in a range of 75 to 84.<sup>[30–32]</sup> Very recently, a systematic comparison suggested that the efficiency and stability of photocatalytic [FeFe]-hydrogenase systems in water is much better than in organic solvents.<sup>[30]</sup> Nevertheless, the efficiency for H<sub>2</sub> evolution is far less than the natural [FeFe]-hydrogenase (turnover frequency (TOF) 6000–9000 molecule H<sub>2</sub> per active site per second).

Poly(acrylic acid) (PAA) is a common hydrophilic polymer, widely used in drug-delivery, self-assembly, nanoparticle modification, and bioimaging.<sup>[42-45]</sup> The carboxy group in the PAA chain not only enhances the water-solubility of the polymer, but also provides modification sites for functionalization. Herein we report that the Fe<sub>2</sub>S<sub>2</sub> active site can be anchored on the side chain of water-soluble PAA. With this grafted polymer as a catalyst, CdSe QDs as a photosensitizer, ascorbic acid as proton source and sacrificial electron donor, we have successfully constructed the first set of polymerbased [FeFe]-hydrogenase mimics for photocatalytic H<sub>2</sub> production. The system shows exceptional TON (based on the Fe<sub>2</sub>S<sub>2</sub> subunit) and initial TOF of up to 27135 and 3.6 s<sup>-1</sup>, respectively, for photocatalytic H<sub>2</sub> production in water.

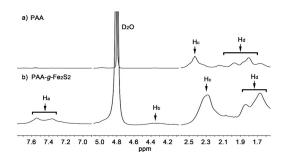
The water-soluble polymer catalyst PAA-g-Fe<sub>2</sub>S<sub>2</sub> was synthesized by stirring the amine-modified Fe<sub>2</sub>S<sub>2</sub> precursor Fe<sub>2</sub>S<sub>2</sub>-NH<sub>2</sub>, which is a deprotected product of Fe<sub>2</sub>S<sub>2</sub>-NHBoc, in a mixed solution of  $CF_3COOH$  and  $CH_2Cl_2$  (v/v = 1/2), and PAA ( $M_w = 1800$ ) in the presence of PyBOP ((benzotriazol-1vloxy)tripyrrolidinophosphonium hexafluorophosphate) in a Et<sub>3</sub>N/DMF mixed solution at room temperature (Scheme 1).<sup>[42,43]</sup> The crude polymer product was precipitated in diethyl ether and dissolved in methanol three times and finally obtained as red to yellow solids depending on the grafting amount of  $Fe_2S_2$  active site. The as-prepared product was characterized by <sup>1</sup>H NMR, IR, UV/Vis spectroscopy and GPC (Supporting Information, Figure S1). The <sup>1</sup>H NMR spectra of PAA-g-Fe<sub>2</sub>S<sub>2</sub> polymer and PAA in D<sub>2</sub>O are shown in Figure 1. The signals at  $\delta = 7.56 - 7.36$  ppm (H<sub>a</sub>) and  $\delta = 4.33 \text{ ppm}$  (H<sub>b</sub>) of the PAA-g-Fe<sub>2</sub>S<sub>2</sub> polymer are attributed to the aromatic protons and methylene protons next to amide of the Fe<sub>2</sub>S<sub>2</sub> moiety, respectively. The IR spectrum of PAA-g-Fe<sub>2</sub>S<sub>2</sub> exhibits a signal for cyanide (CN) at 2125 cm<sup>-1</sup> and three characteristic CO signals at 2040, 1998, and 1975 cm<sup>-1</sup> of the Fe<sub>2</sub>S<sub>2</sub> active site, which are the same as those in the precursor Fe<sub>2</sub>S<sub>2</sub>-NHBoc (Figure 2b).<sup>[22]</sup> The UV/ Vis absorption spectra of PAA-g-Fe<sub>2</sub>S<sub>2</sub> shows a strong characteristic absorption of Fe<sub>2</sub>S<sub>2</sub> moiety at 309 nm in water (Figure 2a), which is a blue-shift of 5 nm compared to the

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 $\textit{Scheme 1.}\ The synthetic route to PAA-g-Fe_2S_2$  for the photocatalytic  $H_2$  production.



*Figure 1.* The <sup>1</sup>H NMR spectra of a) PAA and b) PAA-g-Fe<sub>2</sub>S<sub>2</sub> in D<sub>2</sub>O.

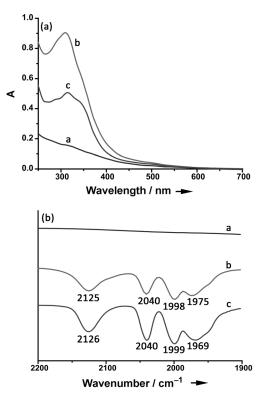
absorption at 314 nm of precursor  $Fe_2S_2$ -NHBoc in CH<sub>3</sub>CN. All of these results confirmed the  $Fe_2S_2$  moiety was chemically grafted on the PAA chain.

Using different initial amount of precursor Fe<sub>2</sub>S<sub>2</sub>-NHBoc, three PAA-g-Fe<sub>2</sub>S<sub>2</sub> polymer catalysts (PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1, PAAg-Fe<sub>2</sub>S<sub>2</sub>-2 and PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3) with different amounts of grafted Fe<sub>2</sub>S<sub>2</sub> active site were prepared. All three polymer catalysts are quite soluble in water. The grafting amount of Fe<sub>2</sub>S<sub>2</sub> active site for PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1, PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2, and PAAg-Fe<sub>2</sub>S<sub>2</sub>-3 is  $3.86 \times 10^{-4}$ ,  $7.07 \times 10^{-5}$ , and  $4.07 \times 10^{-6}$  mol g<sup>-1</sup>, respectively, which was determined by inductively coupled plasma-atomic emission spectrometry (ICP-AES) for the relative amount of iron in samples.

The photocatalytic  $H_2$  production system was constructed by using the PAA-*g*-Fe<sub>2</sub>S<sub>2</sub> polymer as a catalyst, 3-mercaptopropionic acid (MPA) stabilized CdSe QDs (MPA-CdSe QDs) as the photosensitizer, and ascorbic acid (H<sub>2</sub>A) as the proton source and sacrificial electron donor. The CdSe QDs was selected as the photosensitizer not only for its broad visible-light absorption, aqueous dispersion, and economical advantage over precious metal photosensitizers, but also for its simpler preparation than its counterpart CdTe QDs.

We investigated the H<sub>2</sub> production activity of the system using moderate Fe<sub>2</sub>S<sub>2</sub> grafting amount polymer PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2. The system, containing PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 (0.25 mg mL<sup>-1</sup>), MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>), and H<sub>2</sub>A (0.01m) at initial pH 4.0, evolved H<sub>2</sub> immediately with TON of 392 for 3 h of irradiation (blue LED,  $\lambda = 450$  nm). The system showed high activity in the first 3 h; further irradiation led to no obvious improvement in the amount of H<sub>2</sub> production. Control experiments demonstrated that the presence of all of the components in the system, catalyst, photosensitizer, H<sub>2</sub>A, and light is necessary for photocatalytic H<sub>2</sub> evolution.

The degradation of catalyst and consumption of  $H_2A$  influenced the rate of  $H_2$  production greatly. Increasing or decreasing the pH value of the solution to either 5 or 2 resulted in the rate of  $H_2$  evolution dropping dramatically (Figure 3 a), consistent with the reported systems using ascorbic acid as a proton source and sacrificial electron donor.<sup>[29-31]</sup> The  $H_2$  production rate also depends on the concentration of photosensitizer. At fixed concentration of catalyst PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 and H<sub>2</sub>A,

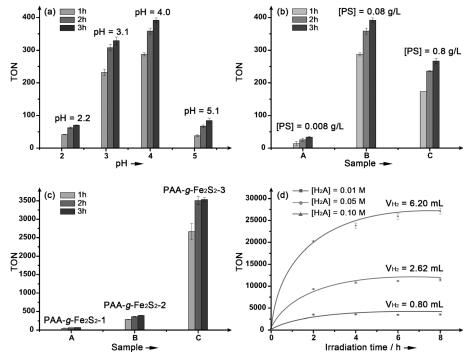


**Figure 2.** UV/Vis absorption (a) and IR (b) spectra of PAA (line a), PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 (line b), and Fe<sub>2</sub>S<sub>2</sub>-NHBoc (Line c); the concentration of samples for UV/Vis absorption is 0.067 mg mL<sup>-1</sup> for PAA and for PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 in aqueous solution, and  $1.0 \times 10^{-5}$  m for Fe<sub>2</sub>S<sub>2</sub>-NHBoc in CH<sub>3</sub>CN.

the optimal concentration of photosensitizer was found to be  $0.08 \text{ mg} \text{mL}^{-1}$ , the reaction was accelerated by increasing the concentration of MPA-CdSe QDs from  $0.008 \text{ mg} \text{mL}^{-1}$  to  $0.08 \text{ mg} \text{mL}^{-1}$  in solution (Figure 3b). Further increasing the concentration of CdSe QDs from  $0.08 \text{ mg} \text{mL}^{-1}$  to  $0.8 \text{ mg} \text{mL}^{-1}$ , however, resulted in lower rate of H<sub>2</sub> production and aggregation of CdSe QDs. This change is probably due to

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**Figure 3.** a) Photocatalytic H<sub>2</sub> production in H<sub>2</sub>O at different pH values; sample concentration: PAAg-Fe<sub>2</sub>S<sub>2</sub>-2 (0.25 mg mL<sup>-1</sup>), MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>), H<sub>2</sub>A (0.01 M); b) Photocatalytic H<sub>2</sub> production at initial pH 4.0 in H<sub>2</sub>O as a function of concentration of the photosensitizer (PS) MPA-CdSe QDs; sample concentration: MPA-CdSe QDs (0.008 mg mL<sup>-1</sup> for sample A, 0.08 mg mL<sup>-1</sup> for sample B, 0.8 mg mL<sup>-1</sup> for sample C), PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 (0.25 mg mL<sup>-1</sup>), H<sub>2</sub>A (0.01 M); c) Photocatalytic H<sub>2</sub> production at initial pH 4.0 in H<sub>2</sub>O as a function of catalysts with different loadings of Fe<sub>2</sub>S<sub>2</sub> active site; PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 to PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3 for sample A–C (0.25 mg mL<sup>-1</sup>), MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>), H<sub>2</sub>A (0.01 M); d) Photocatalytic H<sub>2</sub> production at initial pH 4.0 in H<sub>2</sub>O as a function of ascorbic acid concentration; sample concentration: MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>), PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3 (0.25 mg mL<sup>-1</sup>), H<sub>2</sub>A (0.01 M for line A, 0.05 M for line B, 0.1 M for line C). All samples were irradiated by blue LED lamp ( $\lambda$  = 450 nm).

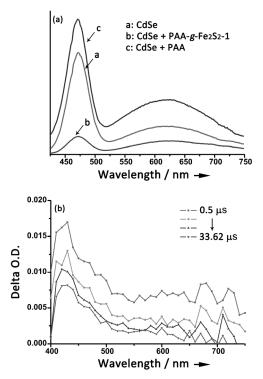
the light-filter effect at high concentration of photosensitizer. Next, the H<sub>2</sub> production activity of three PAA-g-Fe<sub>2</sub>S<sub>2</sub> polymers, PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1, PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2, and PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3, was examined under the identical concentration of photosensitizer and H<sub>2</sub>A at pH 4.0. As shown in Figure 3c, PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3, the one with the smallest Fe<sub>2</sub>S<sub>2</sub> grafted amount, gives rise to the highest  $H_2$  production efficiency (TON = 3536) of the three PAA-g-Fe<sub>2</sub>S<sub>2</sub> polymers for 3 h of irradiation. Finally, the photocatalytic  $H_2$  production with the samples containing PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3  $(0.25 \text{ mgmL}^{-1}),$ MPA-CdSe QDs  $(0.08 \text{ mgmL}^{-1})$  at different concentrations of H<sub>2</sub>A were studied at initial pH 4.0. The rate of photocatalytic  $H_2$ production relies on the concentration of  $H_2A$  (Figure 3d), and the initial rate of H<sub>2</sub> production obeys a first-order dependence on the concentration of H<sub>2</sub>A (Supporting Information, Figure S2). Surprisingly, the TON reached 27135 for  $8\,h$  of irradiation at 0.1m of  $H_2A$  and the rate was as fast as 3.6 molecular  $H_2$  per Fe<sub>2</sub>S<sub>2</sub> active site per second in the initial 30 min (Supporting Information). All of these results confirm that the rate of photocatalytic H<sub>2</sub> production depends on pH value of solution, [photosensitizer] ([PS]), [HA<sup>-</sup>], and the optical power of light source (Supporting Information, Figure S3). The quantum yield (QY) of the system for  $H_2$ production is 5.07%, which was obtained by the sample of PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 as catalyst under optimized conditions (Supporting Information). The TON (based on Fe<sub>2</sub>S<sub>2</sub> subunit) 27135 and initial TOF 3.6 s<sup>-1</sup>, to our knowledge, are the highest values reported to date for photocatalytic H<sub>2</sub> production from [FeFe]-hydrogenase mimics.

The photophysical properties of the polymer-based systems were studied to understand the mechanism of the reaction. The average diameter of MPA-CdSe QDs used in our systems is 1.8 nm as determined by TEM. As shown in Figure 4a, the emission spectrum of MPA-CdSe QDs shows a strong excitonic emission at 470 nm and a relative weak and broad surface trap emission ranged from 520-770 nm.[46] With the addition of PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 or PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 (1.0 mg) into the aqueous solution of MPA-CdSe QDs ( $0.08 \text{ mgmL}^{-1}$ , 3 mL), the excitonic and trap emissions of CdSe QDs were both quenched. The quenching efficiency estimated at excitonic emission for PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 and PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 is 81% and 34%, respectively. Because of the small spectroscopic overlap of absorption of PAA-g-Fe<sub>2</sub>S<sub>2</sub> and the emission of MPA-CdSe QDs (Supporting Information, Figure S5), the energy transfer

between excited MPA-CdSe QDs to PAA-g-Fe<sub>2</sub>S<sub>2</sub> would be negligible if it occurs, so the emission quenching observed in the cases of PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 and PAA-g-Fe<sub>2</sub>S<sub>2</sub>-2 is attributed to the photoinduced electron transfer (PET) from the MPA-CdSe QDs to Fe<sub>2</sub>S<sub>2</sub> core of PAA-g-Fe<sub>2</sub>S<sub>2</sub>. The rate constant of  $6.15 \times 10^{12} \text{ m}^{-1} \text{ s}^{-1}$  for PET was estimated at excitonic emission by the Stern-Volmer equation (Supporting Information, Figure S7). It was worth noting that when the same amount of PAA-g-Fe<sub>2</sub>S<sub>2</sub>-3 or PAA was introduced in the system, the emission intensity of MPA-CdSe QDs increased by 35% or 46%, respectively. The enhancement was also observed by Yang and Gao et al. with the addition of PAA into the aqueous solution of CdTe QDs. They explained and experimentally demonstrated that the coordination between carboxy groups of PAA and cadmium ions on the surface of QDs is responsible for this emission enhancement.<sup>[47,48]</sup> We believe this coordination works the same way in our case. The PAA chain wraps round the nanoparticles by coordination between the carboxy groups and cadmium ions of CdSe QDs and thus suppresses to some extent the non-radiative decay of MPA-CdSe QDs. Actually, the PAA-g-Fe<sub>2</sub>S<sub>2</sub> plays two roles for the emission of MPA-CdSe QDs. On the one hand, the  $Fe_2S_2$ active site in PAA-g-Fe<sub>2</sub>S<sub>2</sub> quenches the emission of MPA-CdSe QDs; on the other hand, the PAA chain suppresses the

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**Figure 4.** a) The emission spectra of MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>) in the absence and presence of PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-1 (0.33 mg mL<sup>-1</sup>) or PAA (0.33 mg mL<sup>-1</sup>) in water; excitation wavelength: 400 nm. b) Transient absorption spectrum of PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-1 (1.0 mg mL<sup>-1</sup>) and MPA-CdSe QDs (0.08 mg mL<sup>-1</sup>) in H<sub>2</sub>O at pH 7.4 upon laser-pulsed by 355 nm light.

mobility of the nanoparticles and enhances the emission of MPA-CdSe QDs. In the cases of PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-1 and PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-2, the quenching process dominates the overall emission of MPA-CdSe QDs. In contrast, with PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-3 the emission of MPA-CdSe QDs is enhanced because of the low amount of Fe<sub>2</sub>S<sub>2</sub> active site in PAA-*g*-Fe<sub>2</sub>S<sub>2</sub>-3.

The electron transfer from excited CdSe QDs to the  $Fe_2S_2$ active site of PAA-g-Fe<sub>2</sub>S<sub>2</sub> was evidenced by a flash photolysis study at room temperature. Compared with signal silence of a sample of MPA-CdSe QDs with PAA, the solution of CdSe QDs and PAA-g-Fe<sub>2</sub>S<sub>2</sub>-1 at pH 7.4 gave a strong absorption at 430 nm, a broad absorption at the range of 550-650 nm, and a relative strong absorption at 680 nm after excitation by laser light at 355 nm. These absorptions are similar to those reported for  $Fe^{I}Fe^{0}$  species generated by reduction of [FeFe]-hydrogenase mimics.<sup>[20,49]</sup> The decay monitored at 430 nm was mono-exponential with a lifetime of 449 µs (Figure S11), which was quenched completely by adjusting the pH to 5.2 by adding HCl to the solution. This quenching is probably due to the formation of protonated Fe<sup>I</sup>Fe<sup>II</sup>H species in acidic solution.<sup>[39,50-52]</sup> The free energy change of electron transfer from excited MPA-CdSe QDs to the Fe<sub>2</sub>S<sub>2</sub> core was estimated by the Rehm-Weller equation. The reductive potential  $(E_{1/2})$  of -0.43 V (all potentials are versus the normal hydrogen electrode (NHE)) of the Fe<sub>2</sub>S<sub>2</sub> active site was taken from cyclic voltammograms obtained using PAA-g- $Fe_2S_2-1$  in aqueous solution (Figure S12). The valence band

energy level  $(E_{vb})$  of CdSe QDs is 0.57 eV,<sup>[53]</sup> and the excitedstate energy  $(E_{00})$  of MPA-CdSe QDs is 2.64 eV. Therefore, the free-energy change  $(\Delta G^0)$  was determined to be -1.64 eV, indicating that the electron transfer from excited MPA-CdSe QDs to the Fe<sub>2</sub>S<sub>2</sub> of PAA-g-Fe<sub>2</sub>S<sub>2</sub> is exothermic.

From the results described above, we could speculate about the general mechanism of the system. The photoexcited electron in the conduction band of CdSe QDs transfers to the  $Fe_2S_2$  core of PAA-g-Fe<sub>2</sub>S<sub>2</sub> and generates a one-electron reduced Fe<sup>I</sup>Fe<sup>0</sup> species. This active species further reacts with a proton in catalytic cycle to produce  $H_2$ .<sup>[39,50-52]</sup> As the oxidative potential of H<sub>2</sub>A is negative enough to reduce the CdSe QDs but too positive to reduce the Fe<sub>2</sub>S<sub>2</sub> core of PAA-g-Fe<sub>2</sub>S<sub>2</sub> directly,<sup>[54]</sup> the oxidative CdSe QDs regenerates by accepting an electron from the sacrificial electron donor HA-. As reported in the literature and observed in our experiments,[55] the photo-corrosion of CdSe QDs would lead to aggregation of CdSe QDs in aqueous systems. But in this case, the absorption and emission spectra of CdSe QDs  $(0.08 \text{ mg mL}^{-1})$  were almost unchanged for 4 h of irradiation, and no obvious aggregation was observed in the presence of PAA or PAA-g-Fe<sub>2</sub>S<sub>2</sub> in aqueous solution (Figures S15 and S16). Evidently, the PAA moiety in PAA-g-Fe<sub>2</sub>S<sub>2</sub> not only functions as a water-soluble framework to bring the  $Fe_2S_2$ core into water, but also plays a role in protecting MPA-CdSe QDs from photo-corrosion and aggregation. Therefore, it is reasonable to consider that the polymer chain of PAA wraps around the MPA-CdSe QDs by coordination and thus narrows distance between the photosensitizer and the  $Fe_2S_2$ active site in aqueous solution. This unique structure prefers to facilitate electron transfer from the excited MPA-CdSe QDs to PAA-g-Fe<sub>2</sub>S<sub>2</sub> and consequently improves the rate of photocatalytic H<sub>2</sub> production.

In summary, a new set of water-soluble polymer catalysts PAA-g-Fe<sub>2</sub>S<sub>2</sub> has been designed and successfully synthesized. A system, containing PAA-g-Fe<sub>2</sub>S<sub>2</sub> as the catalyst, CdSe QDs as the photosensitizer, and ascorbic acid as proton source and sacrificial electron donor, shows high efficiency for the photocatalytic H<sub>2</sub> production in water. The TON of 27135, initial TOF of 3.6 s<sup>-1</sup>, are the highest known to date for [FeFe]-hydrogenase mimics, competitive with those from current state-of-the-art catalytic systems for H<sub>2</sub> production. The PAA chain of PAA-g-Fe<sub>2</sub>S<sub>2</sub> plays three roles in the system: 1) it is a framework to bring the  $Fe_2S_2$  active site into aqueous solution; 2) it is a good stabilizer protecting the MPA-CdSe QDs from aggregation and enhancing the emission quantum yield of the CdSe QDs; 3) it narrows the distance between the photosensitizer and the Fe<sub>2</sub>S<sub>2</sub> core for more efficient electron transfer ( $k_{\rm ET} = 6.15 \times 10^{12} \,\mathrm{M}^{-1} \,\mathrm{s}^{-1}$ ) from the excited MPA-CdSe QDs to the Fe<sub>2</sub>S<sub>2</sub> active site. The three functions jointly contribute to a highly efficient photocatalytic H<sub>2</sub> production. Static-state and time-resolved spectroscopic studies demonstrate that the electron transfer from the excited MPA-CdSe QDs to the  $Fe_2S_2$  catalyst center is the key step to trigger the catalytic cycle. The unique performance of the polymer-based [FeFe]-hydrogenase system indicates that this approach is a promising strategy to improve the photocatalytic efficiency of H<sub>2</sub> production in water. Extension of the present systems is ongoing in our laboratory.

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