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Enzymatic multicomponent reaction for simultaneous synthesis of two important scaffolds, pyridin-2-ones and α -alkylated nitriles

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 \mathbb{R}^{1} \mathbb{R}^{1} \mathbb{R}^{2} \mathbb{R}^{3} \mathbb{R}^{3} \mathbb{R}^{2} 4 Enzymatic Multicomponent Reaction Hydrogen Transfer \mathbb{R}^3 \mathbb{R}^2 5 5' 5NH NC Ő

Enzymatic multicomponent reaction for simultaneous synthesis of two important scaffolds, pyridin-2-ones and α -alkylated nitriles

Zhi-Qiang Liu, Yu-Jing Hu, Xiao-Yang Chen, Qi Wu,* Xian-Fu Lin* Department of Chemistry, Zhejiang University, Hangzhou 310027, People's Republic of China, Fax: + 86 571 87952618; E-mail: <u>llc123@zju.edu.cn</u>

Abstract

An efficient approach for the synthesis of highly substituted pyridin-2-one derivatives and α -alkylated nitriles through enzymatic multicomponent reaction (MCR) was developed. This MCR involved bio-mimetic reduction between 3,4-dihydropyridin-2-one and activated olefin, both of which were *in situ* generated in the Acylase Amano (AA)-catalyzed domino reaction starting from benzaldehyde, cyanoacetamide and ketone. A wide range of substituted benzaldehydes and ketones were accepted by this reaction. Both final products (pyridin-2-one derivatives and α -alkylated nitriles) were important skeletons and synthetic intermediates. The synthetic application of prepared α -alkylated nitrile was demonstrated by converting it into the corresponding α -alkyl- β^2 -amino acid with high yield.

Keywords

Enzyme catalysis; Multicomponent reactions; Transfer hydrogenation; Pyridin-2-one; Alkylated nitrile

1. Introduction

As one representative of substituted six-membered lactams, compounds with pyridine-2-one scaffold have shown significant pharmaceutical activities such as antifungal¹, antibacterial² and antitumor³. Specifically, amrinone (**a**) and milrinone (**b**) in Fig. 1 have been found to display effective activities on therapy of myocardial infarction, and compound (**c**) has the activity of antihypertensive. Meanwhile, the pyridin-2-one ring system has successfully been applied in the natural synthesis as a building block⁴. Thus, development of convenient, efficient and mild strategy to establish these lactam cores is still interesting in the realm of synthetic organic chemistry. Although several synthetic methodologies have been reported⁵, most of them have their own drawbacks such as harsh reaction conditions, low yields or complicated multistep routes. Similarly, α -alkylated nitriles are also important building blocks for synthesizing ketones and biologically active agents⁶, the main methods for constructing α -alkylated nitriles are reactions of nitriles with alkyl halides (or alcohols or carbonyl compounds) using homogeneous inorganic bases such as NaH and NaNH₂⁷ as catalysts. Toxicity of halogenated substrates, the use of homogeneous strong bases and metals are general drawbacks. With ever-increasing environmental concerns, developing new method to construct pyridine-2-one and α -alkylated nitrile derivatives still remains a significant challenge.



Fig. 1. Compounds with pyridin-2-one core

Since enzymes were discovered to maintain activity in organic solvents by Klibanov in the early 1980s⁸, enzymes have been proved as efficient and green biocatalysts in organic synthesis due to their intrinsic advantages of high selectivity, mild reaction conditions, simple operation and environmentally benign behavior. In the past decade, a growing number of enzymes were found to be capable of catalyzing not only their "natural" reactions but also one or more alternative reactions, and this feature is called catalytic promiscuity⁹. Our group has found that the Michael and Markovnikov addition could be catalyzed by alkaline protease and acylase¹⁰. And recently we have discovered many enzymatic multicomponent reactions to synthesize heterocyclic compounds¹¹. More and more new cases of enzymatically promiscuous reactions were reported by other groups¹², for example, Zhang^{12b} provided a new strategy to synthesize spirooxindole derivatives via hydrolase, then Trypsin was founded to catalyze the formation of 4-thiazolidinones and 3,4-dihydropyrimidin-2-ones through three-component reaction^{12e,12f}, and Guan reported the first enzymatic aza-Diels-Alder Reaction^{12a}. Though many works involved single-enzyme catalyzed multistep transformations were found, more explorations concerning new enzymatic domino reaction are still urgent in terms of the development of enzymology and industrial importance. Herein, we discovered an interesting reaction to simultaneously synthesize important pyridine-2-ones and α -alkylated nitriles via enzymatic multicomponent reaction involving in situ hydrogen transfer.

2. Results and discussion

Our initial idea was the synthesis of 3,4-dihydropyridin-2-one derivatives (5') via a multicomponent reaction starting from benzaldehyde (1), cyanoacetamide (2) and monoketone (3) catalyzed by enzyme (Scheme 1). Surprisingly, compounds 4 and 5 were obtained after flash chromatograph separation, which were very similar to the direct Knoevenagel condensation product (4') and expected product of three components (5'). It seemed that hydrogen transfer process happened between compounds 4' and 5'.



Scheme 1. enzymatic one-pot multicomponent reaction involving in situ hydrogen transfer

Considering the efficiency of this amusing phenomenon, we chose 4-chlorobenzaldehyde (1a),

cyanoacetamide (2) and butanone (3a) as the starting materials, catalyst screening was firstly performed in ethylene glycol (EG) at 50 °C. Among the selected commercial enzymes, Acylase Amano (AA) exhibited the best catalytic activity with 50% yield (Table 1, entry 1), 33% yield was obtained by D-aminoacylase (DA) (Table 1, entry 2). The moderate yields were mainly due to the left substrates. Butanone could form two pyridine-2-one products due to its different active positions. While other hydrolases, such as CAL-B (Table 1, entry 3), Lipase from Candida rugosa, Lipozyme from Mucor miehei and Amano Lipase PS (immobilized on diatomite), showed no activity toward the reaction (data not shown). Next, we investigated the effect of different solvents on the model reaction, probably because cyanoacetamide had low solubility in dioxane, toluene and acetonitrile, and no products were observed in these generally used solvents (data not shown). Fortunately, the reaction in diethylene glycol (Di-EG) gave the best yield of 71% at 50 °C after 48 h (Table 1, entry 9), and 50% yield in EG was gotten (Table 1, entry 1), while both DMF and DMSO provided the yields less than 20% (Table 1, entries 7 and 8). Determining Di-EG as the optimum solvent, we performed some control experiments to verify the specific catalytic effect of the enzyme on the initial three component reaction. No product was detected when the reaction underwent in the absence of enzyme (Table 1, entry 4). In addition, since AA is a zinc-dependent enzyme, EDTA (ethylene diamine tetraacetic acid) was used to denature the enzyme. We found that no reaction happened with EDTA-denatured AA or bovine serum albumin (BSA) (Table 1, entries 6 and 5), ruling out the possibility that amino acid of the protein surface or other impurities in AA could promote the initial three component reaction. On the other hand, these results also validated that the specific active site and the tertiary structure of AA was essential for the three component reaction.

It is noteworthy that the ratio of three starting materials has important influence on the output of the multicomponent reaction. When the ratio of compounds 1a:2:3a was 0.5:0.75:0.5, the total yield reached to 80%. Further changing the ratio, no improvement of yield was obtained. Thus, we chose the molar ratio of substrate (1:2:3=0.5:0.75:0.5) for subsequent studies. Actually in our previous report, the interesting *in situ* hydrogen transfer process to form α -alkylated nitriles (4) and pyridine-2-ones (5) was not observed because of the much lower amount of aromatic aldehyde in the optimized feeding ratio of three starting materials (1:2:3=0.125:1:0.75), thus the hydrogen-accepting Knoevenagel product (4') was not enough and only 3,4-dihydropyridin-2-ones (5') was obtained^{11b}.

Ia	DIE	1 9	
~	. •		

	CHO H ₂ N + C	$=0 + =0 \frac{ca}{sc}$	talyst lvent	VH_2 H_2 N_2		CN
	1a 2	3a	4a	CI 5a	ı G) ja
Entry	Catalyst	Solent	4a $(\mu mol)^{b}$	$5a (\mu mol)^b$	6a $(\mu mol)^{b}$	Total yield(%) ^d
1	AA	EG	131	95	26	50
2	DA	EG	84	65	18	33
3	CAL-B	EG	n.d.	n.d.	n.d.	n.d.

Optimization of reaction conditions^a

4	-	Di-EG	n.d.	n.d.	n.d.	n.d.
5	BSA	Di-EG	Trace	trace	Trace	trace
6	AA ^c	Di-EG	Trace	trace	Trace	trace
7	AA	DMF	Trace	trace	Trace	trace
8	AA	DMSO	41	33	5	16
9	AA	Di-EG	175	124	58	71

^{*a*} Reactions were carried out in EG (1 mL) with equimolar substrate (0.5 mmol) in the presence of 20 mg enzyme at 50 $^{\circ}$ C for 48h.

^b Determined by HPLC.

^c Denatured by EDTA in water for 12h at 100 °C.

^d Total yields of 4+5+6 based on the amount of aromatic aldehyde, calculating equation is Yield =[(Amount

4+**5**+**6**) / 500] × 100%.

n.d.: Not determined.

With the optimal conditions in hand, we further examined the scope and limitation of the new enzymatic multicomponent reaction. It can be seen from Table 2 that a wide range of substituted benzaldehydes could successfully participate in the reaction. Generally, the enzyme exhibited better activity for benzaldehydes with electron-withdrawing substitutions than with electron-donating ones. For example, 4-(trifluoromethyl)benzaldehyde gave an excellent yield of 94% (Table 2, entry 1), while 4-isopropyl and 2-methoxybenzaldehyde only could get products in 70% yields (Table 2, entries 2 and 4). 3-Substituted benzaldehyde could also be accepted in the reaction (Table 2, entry 5).

Table 2

Examination of the substituent effect of the aromatic aldehyde ^a



Entry R	Product	Amount	Due du et	Amount	Product	Amount	Total	
		(µmol) ^b	Product	(µmol) ^b		$(\mu mol)^b$	yield(%) ^c	
1	4-CF ₃	4b	233	5b	156	6b	81	94
2	4-iPr	4c	172	5c	122	6c	56	70
3	4-F	4d	212	5d	155	6d	51	84
4	2-OCH ₃	4e	171	5e	135	6e	37	69
5	3-Cl	4f	205	5f	155	6f	51	82

^{*a*} Reactions were carried out in Di-EG (1 mL) with **1** (0.5 mmol), **2** (0.75 mmol) and **3** (0.5 mmol) in the presence of 20 mg AA at 50 $^{\circ}$ C for 48h.

^b Determined by HPLC.

^c Total yields of 4+5+6 based on the amount of aromatic aldehyde, calculating equation is Yield =[(Amount

4+5+6) / 500] ×100%

Then reactions of various ketones with different aldehydes were further tested. Both acetone and 3-pentanone could successfully participate in the reaction to form α -alkylated nitriles and pyridine-2-ones. But the yield declined sharply with the increase of chain length, from 70% to 32% (Table 3, entries 1 and 2), possibly because the activity of α -hydrogen in carbonyl compound reduced when increasing the chain length of ketones. Theoretically, 1,3-dicarbonyl compounds like methyl acetoacetate could also give the corresponding products. In our previous report,^{11b} the amount of cyanoacetamide and methyl acetoacetate was excessive compared to aromatic aldehyde, and these substrates were simultaneously added to the reaction flask, giving the product of 3,4-dihydropyridin-2-one. Herein we first mixed aromatic aldehyde and cyanoacetamide together, when they fully changed to the Knoevenagel condensation product, half amount of methyl acetoacetate was introduced to the system. Finally, the expected α -alkylated nitrile and pyridine-2-one were obtained through slow transformation after 4 days (Table 3, entry 4).

Table 3

Variation of aldehyde and ketone ^a

$\begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} CHO \\ + \end{array} \\ R^{1} \end{array} \\ \end{array} \\ \begin{array}{c} NH_{2} \end{array} \\ \end{array} \\ \begin{array}{c} + \end{array} \\ R^{2} \end{array} \\ \end{array} \\ \begin{array}{c} P \\ R^{3} \end{array} \\ \begin{array}{c} AA \\ Di-EG \end{array} \\ \begin{array}{c} P \\ R^{1} \end{array} \\ \begin{array}{c} O \\ R^{1} \end{array} \\ \begin{array}{c} NH_{2} \end{array} \\ \begin{array}{c} + \end{array} \\ \begin{array}{c} R^{3} \\ H \\ H \end{array} \\ \begin{array}{c} NH \\ H \\ H \end{array} \\ \begin{array}{c} O \\ R^{1} \end{array} \\ \begin{array}{c} O \\ R^{1} \end{array} \\ \begin{array}{c} NH_{2} \end{array} \\ \begin{array}{c} + \end{array} \\ \begin{array}{c} R^{3} \\ H \\ H \\ H \end{array} \\ \begin{array}{c} O \\ H \end{array} \\ \begin{array}{c} O \\ H \\ H \end{array} \\ \begin{array}{c} O \\ H \\ H \end{array} \\ \begin{array}{c} O \\ H \\ H \\ H \\ H \\ H \end{array} \\ \begin{array}{c} O \\ H \\$								
	1	2		3	Y	4	Ŕ	5
Enter	\mathbf{p}^1	P ²	D ³	Droduct	Amount	Droduct	Amount	Total yield
Entry F	ĸ	ĸ	ĸ	Ploduct	$(\mu mol)^{b}$	Product	$(\mu mol)^b$	(%) ^c
1	4-iPr	Н	Н	4c	175	5g	176	70
2	4-iPr	CH ₃	CH ₃	4 c	79	5h	83	32
3 ^d	4-iPr	CH ₃	CH ₃	4c	84	5h	89	35
4 ^e	p-Cl	Н	COOCH ₃	4 a	158	5i	154	62

D2

^{*a*} Reactions were carried out in Di-EG (1 mL) with **1** (0.5 mmol), **2** (0.75 mmol) and **3** (0.5 mmol) in the presence of 20 mg AA at 50 $^{\circ}$ C for 48h.

^b Determined by HPLC.

^c Total yields of 4+5+6 based on the amount of aromatic aldehyde, calculating equation is Yield =[(Amount 4+5) / 500] ×100%.

^d Protected under nitrogen atmosphere.

^{*e*} After 6h the reaction of **1** (0.5 mmol) and **2** (0.5 mmol) in the presence of 20 mg AA at 50 °C, 1 mL Di-EG, **3** (0.25 mmol) was added for 4 days

Basing on the above successful synthesis of two series of important compounds, i.e. pyridine-2-one derivatives and α -alkylated nitriles, we hope to confirm the existence of hydrogen transfer process. Firstly, we should determine whether the oxygen acts as oxidant in the system. Therefore, controlled reaction under the protection of nitrogen was performed, the yield was in accordance with that in air

(Table 3, entries 2 and 3). This result excluded the role of oxygen as oxidant in the reaction. Secondly, can the solvent be the hydrogen donor? When we only put Knoevenagel product and the enzyme in the solvent (Di-EG), no hydrogenation product was observed after two days, which excluded the possibility that the solvent acted as hydrogen donor. The amounts of pyridine-2-one derivatives should be equal to that of α -alkylated nitriles if hydrogen transfer process existed, which were confirmed by their respective amount in Table 1-3. In order to further verify the internal transfer hydrogenation process quantitatively, we tried to prepare two supposed intermediates, 3,4-dihydropyridin-2-one (Scheme 2, 5i' and Scheme 3, 5h') and Knoevenagel condensation products (4a'). When we mixed 5i' with 4a' in Di-EG containing AA, two compounds (5i and 4a) were successfully detected after chromatography as we expected. To have a better understanding for the reaction process, we monitored the transforming course of 5i' and 4a increased as the reaction proceeded. Correspondingly, the substrates (5i' and 4a') gradually reduced. Same results were observed for 5h' and 4a', which further confirmed the *in situ* hydrogen transfer process.



Scheme 2. Quantitatively monitoring the transforming process between two intermediates (5i' and 4a')

Another question needs to be considered is whether the *in situ* hydrogen transfer process happened in the active site of this enzyme. We mixed 0.2 mmol **5h'** and **4a'** in Di-EG for one day under the catalyst-free condition, or in the presence of BSA or AA (Scheme 3). No reaction occurred under the catalyst-free condition, and the catalysis of AA could provide the products with 90% yields (**5h** and **4a**). Interestingly the control reaction under the catalysis of BSA also gave the products in 70% yields. Then could amino acid catalyze this process? Aspartate (acidic), lysine (basic) and proline (neutral) as biocatalyst were investigated respectively, only lysine could promote the transformation with 71% yield, indicating the *in situ* hydrogen transfer process was possibly catalyzed by some surface basic amino acids of the enzyme.



Scheme 3. Controlled experiments to explore the hydrogen transfer process

Thus, a tentatively proposed mechanism was illustrated on the basis of these data in Scheme 4. First, AA catalyzed the fast formation of Knoevenagel condensation reaction of aldehyde and cyanoacetamide to compound 1. Then the zinc ion would coordinate with carbonyl group of cyanoacetamide and ketone to increase the nucleophilicity, Asp 366 of AA deprived one α -H of ketone, which occurred the Michael addition with compound 1 to produce intermediate 2. Next, the electron-rich N-H performed the nucleophilic addition to the carbonyl group of ketone, at the same time, one molecular water was lost from intermediate 2 to form product 3. Lysine catalyzed the transfer hydrogenation process as a hydrogen carrier, obtained proton from compound 3 and transferred to compound 1. The final products pyridin-2-one derivatives 4 and α -alkylated nitriles 5 were obtained.



Scheme 4. The proposed mechanism of this multicomponent reaction.

In order to validate the utility of this strategy, firstly, we examined the reaction on tenfold amount of entry 3 in Table 2, total isolated yield was 69%. Then we synthesized the α -substituted β^2 -amino acid from one product, α -alkylated nitrile *via* simple conversions. β -amino acids are key building blocks of natural products, β -peptides and pharmaceuticals¹³. Compounds with this skeleton usually exhibit antivirus and antibacterial activity¹⁴. So developing new method to construct β -amino acids is still of urgency. Our protocol is outlined in Scheme 5. Initially, Raney Ni was used as the catalyst for the hydrogenation of cyano group in the reductive product 4d under hydrogen atmosphere, 4da was obtained in high yield of 92%. Further simple treatment of 4da with concentrated hydrochloric acid at room temperature for 3 h provided the target α -substituted β^2 -amino acid (4db) in excellent yield of 96%. This successful demonstration of converting prepared α -alkylated nitrile into the corresponding α -alkyl- β^2 -amino acid with high yield displayed the great potential of this enzymatic multicomponent reaction in organic synthesis.



Scheme 5. Reagents and conditions: (i) Raney Ni (10% w/w), H₂ (1 atm), 2 M NH₃/MeOH, 50 °C, 20 h; (ii) aq HCl, rt, 3h

3. Conclusions

In summary, we have discovered an interesting process involving transfer hydrogenation between 3,4-dihydropyridin-2-one and activated olefin, both of them were *in situ* generated in an Acylase Amano (AA)-catalyzed multicomponent domino reaction starting from benzaldehyde, cyanoacetamide and ketone. A wide range of substituted benzaldehydes and ketones were accepted by this reaction. The resultant two final products (pyridin-2-one derivatives and α -alkylated nitriles) were both building blocks of a variety of biologically active agents. As a demonstration, these α -alkylated nitriles could be transformed to synthetically important α -substituted β^2 -amino acids via simple conversions. The reaction process was verified in detail by some controlled experiments, separating intermediates and monitoring the transfer hydrogenation course between 3,4-dihydropyridin-2-one and activated olefin quantitatively. This novel single enzyme-catalyzed multicomponent reaction is attractive in terms of high atom economy, easy workup procedure, tolerance of various functional groups and mild reaction conditions, and will widen the application of enzyme in organic synthesis.

4. Material and methods

4.1 General information for reagents and analytical methods

D-aminoacylase from *E. coli* (10000 U/mg, 1 U is defined as enzyme quantity, which produces 1 mmol of D-Amino acid per 30 min) and Acylase 'Amano' (AA) from *Aspergillus oryzae* (\geq 30000 U/g, 1 U is defined as enzyme quantity, which produces 1 mmol of L-Amino acid per 30 min) were purchased from Amano Enzyme Inc (Japan). Lipase immobilized on acrylic resin from *C. antarctica* (10000 U/g, recombinant, expressed in *Aspergillus oryzae*) was purchased from Sigma (Steinheim, Germany). The ¹H and ¹³C NMR spectra were recorded with TMS as internal standard using a Bruker AMX-400 or 500 MHz spectrometer. Chemical shifts were expressed in parts per million and coupling constants (*J*) in hertz. Analytical HPLC was performed using an Agilent TC-C18 series (250×4.6 mm) and a UV detector (210 nm). The mobile phase was methanol and water. IR spectra were measured with a Nicolet Nexus FTIR 670 spectrophotometer. Melting points were determined using XT-4 apparatus and were not corrected. All the known products were characterized by comparing the ¹H NMR, ¹³C NMR, and HRMS.

4.2 General procedure for the enzymatic multicomponent reaction

A mixture of aromatic aldehyde (0.5 mmol), cyanoacetamide (0.75 mmol), monoketone (0.5 mmol), and 20 mg AA in 1 mL Di-EG was shaken at 50 °C and 200 rpm for 48h. After completion of the reaction as monitored by TLC, the detecting solvent was petroleum ether/ethyl acetate mixture (1/2 v/v), then the mixture was loaded directly onto a silica gel column using petroleum ether/ethyl acetate mixture (1/1 v/v) as eluent to afford the desired product of α -alkylated nitriles and pyridin-2-ones, the α -alkylated nitriles was first obtained, importantly, it had very slight UV absorption at 254 nm, but it can be confirmed in the iodine jar.

4.3 Procedure for the scale-up synthesis of α-alkylated nitrile and pyridin-2-one

A mixture of 4-fluorobenzaldehyde (5 mmol), cyanoacetamide (7.5 mmol), butanone (5 mmol), and 150 mg AA in 10 mL Di-EG was shaken at 50 $^{\circ}$ C and 200 rpm for 72h. The mixture was loaded directly onto a silica gel column using petroleum ether/ethyl acetate mixture (1/1 v/v) as eluent to afford the desired product of **4d** (337 mg), **5d** (350mg) and **6d** (61 mg), the total isolated yield was 69 %.

4.4 Procedure for the synthesis of α -alkyl- β^2 -amino acid

0.3 mmol compound **4d** and Raney Ni (10% w/w) were put into 10 mL NH₃/MeOH solvent (2 M), the mixture was stirred at 50 °C for 20 h under hydrogen atmosphere. After completion of the reaction, Raney Ni was filtered and the solvent was removed under reduced pressure, then the residual solid was washed with 2 mL petroleum ether/ethyl acetate mixture (1/1 v/v) twice. The obtained compound **4da** was put into 10 mL concentrated hydrochloric acid for 3 h at room temperature, the final product was gotten after removing solvent.

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Table 1

4

5

6

7

Optimization of reaction conditions ^a



n.d.

trace

trace

trace

n.d.

Trace

Trace

Trace

n.d.

trace

trace

trace

8AADMSO41335169AADi-EG1751245871a Reactions were carried out in EG (1 mL) with equimolar substrate (0.5 mmol) in the presence of 20

n.d.

Trace

Trace

Trace

mg enzyme at 50 °C for 48h.

^b Determined by HPLC.

_

BSA

 AA^{c}

AA

^c Denatured by EDTA in water for 12h at 100 °C.

Di-EG

Di-EG

Di-EG

DMF

^d Total yields of 4+5+6 based on the amount of aromatic aldehyde, calculating equation is Yield

=[(Amount 4+5+6) / 500] × 100%.

n.d.: Not determined.

Table 2

Examination of the substituent effect of the aromatic aldehyde ^a



^a Reactions were carried out in Di-EG (1 mL) with 1 (0.5 mmol), 2 (0.75 mmol) and 3 (0.5 mmol) in the

presence of 20 mg AA at 50 $^{\rm o}{\rm C}$ for 48h.

^b Determined by HPLC.

^c Total yields of 4+5+6 based on the amount of aromatic aldehyde, calculating equation is Yield =[(Amount 4+5+6) / 500] ×100%

Table 3

Variation of aldehyde and ketone ^a

$\begin{array}{c} \begin{array}{c} & CHO \\ & + \end{array} \\ R^{1} \end{array} \\ O \end{array} \\ \begin{array}{c} & O \\ & + \end{array} \\ R^{2} \end{array} \\ \begin{array}{c} & O \\ & AA \\ \hline Di-EG \end{array} \\ \begin{array}{c} & O \\ & R^{1} \end{array} \\ \begin{array}{c} & O \\ & NH_{2} \end{array} \\ \begin{array}{c} & + \end{array} \\ \begin{array}{c} & R^{3} \\ & NH \\ & O \end{array} \\ \begin{array}{c} & R^{2} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & R^{3} \end{array} \\ \begin{array}{c} & R^{2} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & R^{3} \end{array} \\ \begin{array}{c} & R^{2} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & R^{3} \end{array} \\ \begin{array}{c} & R^{2} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & O \end{array} \\ \begin{array}{c} & R^{3} \\ & R^{3} \end{array} \\ \begin{array}{c} & R^{2} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & O \end{array} \\ \begin{array}{c} & R^{3} \\ & NH \end{array} \\ \begin{array}{c} & O \\ & O \end{array} \\ \end{array} $								
1		2		3		4		[≫] 1 CN R ¹ 5
Entry	R^1	R ²	R ³	Product	Amount (µmol) ^b	Product	Amount (µmol) ^b	Total yield (%) ^c
1	4-iPr	Н	Н	4c	175	5g	176	70
2	4-iPr	CH_3	CH ₃	4 c	79	5h	83	32
3 ^d	4-iPr	CH_3	CH ₃	4 c	84	5h	89	35
4 ^e	p-Cl	Н	$COOCH_3$	4a	158	5i	154	62

^{*a*} Reactions were carried out in Di-EG (1 mL) with **1** (0.5 mmol), **2** (0.75 mmol) and **3** (0.5 mmol) in the presence of 20 mg AA at 50 °C for 48h.

^b Determined by HPLC.

^c Total yields of **4+5+6** based on the amount of aromatic aldehyde, calculating equation is Yield

=[(Amount 4+5) / 500] \times 100%.

^d Protected under nitrogen atmosphere.

^{*e*} After 6h the reaction of **1** (0.5 mmol) and **2** (0.5 mmol) in the presence of 20 mg AA at 50 °C, 1 mL Di-EG, **3** (0.25 mmol) was added for 4 days















Enzymatic multicomponent reaction for simultaneous synthesis of two

important scaffolds, pyridin-2-ones and α-alkylated nitriles

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Supplementary Data

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Supporting Information

Characterizations for Compounds

4a: 3-(4-chlorophenyl)-2-cyanopropanamide

Cl[•] m.p. 180-181 °C; IR (neat): 3423, 3315, 3206, 2252, 1667, 1611; ¹H NMR (400 MHz, DMSO- d_6) δ 7.78 (s, 1H), 7.49 (s, 1H), 7.41-7.31 (m, 4H), 3.96-3.93 (m, 1H), 3.14 (dd, J = 13.6, 6.8 Hz, 1H), 3.16 (dd, J = 13.6, 8.8 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ 171.3, 141.1, 137.0, 136.2, 136.2, 133.6, 133.6, 123.5, 39.6; HRMS (EI) calcd. for C₁₀H₉N₂OCl [M⁺]: 208.0403, found: 208.0403

6a: 4-(4-chlorophenyl)-6-ethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 260-261 °C; IR (neat): 2220, 1645, 1614; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.65 (s, 1H), 7.66-7.60 (m, 4H), 6.34 (s, 1H), 2.58 (q, J = 7.6 Hz, 2H), 1.19 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.4, 159.1, 157.5, 135.2, 134.9, 129.9, 128.9, 116.4, 104.9, 97.7, 26.1, 12.7; HRMS (EI) calcd. for C₁₄H₁₁N₂OCl [M⁺]: 258.0560, found: 258.0562

5a: 4-(4-chlorophenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



CI m.p. 278-279 °C; IR (neat): 2223, 1652, 1604; ¹H NMR (400 MHz, DMSO- d_6) δ 12.64 (s, 1H), 7.60-7.33 (m, 4H), 2.30 (s, 3H), 1.68 (s, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 159.9, 150.7, 150.7, 135.1, 133.9, 129.7, 128.8, 116.2, 111.3, 99.7, 18.0, 13.9; HRMS (EI) calcd. for C₁₄H₁₁N₂OCl [M⁺]: 258.0560, found: 258.0561

4b: 2-cyano-3-(4-(trifluoromethyl)phenyl)propanamide



NMR (400 MHz, DMSO- d_6) δ 7.79 (s, 1H), 7.71-7.51 (m, 5H), 4.01 (t, J = 7.6 Hz, 1H), 3.25 (dd, J = 13.8, 6.6 Hz, 1H), 3.16 (dd, J = 13.6, 8.4 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ 171.2, 147.0, 135.2, 135.1, 133.1, 132.9, 130.6, 130.5, 128.4, 123.4, 40.0; HRMS (EI) calcd. for C₁₁H₉N₂OF₃ [M⁺]: 242.0667, found: 242.0667

6b: 6-ethyl-2-oxo-4-(4-(trifluoromethyl)phenyl)-1,2-dihydropyridine-3-carbonitrile



m.p. 200-201 °C; IR (neat): 2272, 1694, 1617; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.74 (s, 1H), 7.92-7.81 (m, 4H), 6.37 (s, 1H), 2.58 (q, J = 7.6 Hz, 2H), 1.18 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.3, 158.9, 157.9, 140.2, 130.4, 130.2, 129.0, 126.5, 125.7, 125.7, 125.0, 122.8, 116.2, 105.0, 98.1, 26.1, 12.6; HRMS (EI) calcd. for C₁₅H₁₁N₂OF₃ [M⁺]: 292.0823, found: 292.0827

5b: 5,6-dimethyl-2-oxo-4-(4-(trifluoromethyl)phenyl)-1,2-dihydropyridine-3-carbonitrile



 F_3C m.p. 264-265 °C; IR (neat): 2227, 1652, 1617; ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.73 (s, 1H), 7.92-7.56 (m, 4H), 2.33 (s, 3H), 1.68 (s, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆) δ 160.2, 159.8, 151.1, 140.4, 129.5, 129.3, 128.7, 125.7, 125.7, 125.0, 122.9, 116.1, 111.1, 99.5, 18.0, 13.9; HRMS (EI) calcd. for C₁₅H₁₁N₂OF₃ [M⁺]: 292.0823, found: 292.0826

4c: 2-cyano-3-(4-isopropylphenyl)propanamide



m.p. 150-151 °C; IR (neat): 3433, 3367, 3194, 2253, 1659, 1630;

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.78 (s, 1H), 7,48 (s, 1H), 7.20 (s, 4H), 3.93-3.89 (m, 1H), 3.11 (dd, *J* = 13.8, 6.6 Hz, 1H), 2.99 (dd, *J* = 13.6, 8.8 Hz, 1H), 2.89-2.83 (m, 1H), 1.19 (d, *J* = 6.8 Hz, 6H); ¹³C NMR (125 MHz, DMSO-*d*₆) δ 171.6, 152.3, 139.4, 134.1, 131.6, 123.7, 40.1, 38.3, 29.1; HRMS (EI) calcd. for C₁₃H₁₆N₂O [M⁺]: 216.1263, found: 216.1266

6c: 6-ethyl-4-(4-isopropylphenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 280-281 °C; IR (neat): 2219, 1652, 1605; ¹H NMR (400 MHz, DMSO- d_6) δ 12.54 (s, 1H), 7.57-7.41 (m, 4H), 6.33 (s, 1H), 3.00-2.93 (m, 1H), 2.58 (q, J = 7.4 Hz, 2H), 1.25-1.17 (m, 9H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.6, 160.2, 157.0, 151.0, 133.6, 128.1, 126.7, 116.7, 105.0, 97.2, 33.3, 26.1, 23.6, 12.6; HRMS (EI) calcd. for C₁₇H₁₈N₂O [M⁺]: 266.1419, found: 266.1417

5c: 4-(4-isopropylphenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 290-291 °C; IR (neat): 2219, 1648, 1605; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.56 (s, 1H), 7.40-7.20 (m, 4H), 2.99-2.92 (m, 1H), 2.30 (s, 3H), 1.70 (s, 3H), 1.24 (d, J = 6.8 Hz, 6H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.8, 160.1, 150.3, 149.2, 133.7, 127.7, 126.5, 116.4, 111.4, 99.7, 33.2, 23.7, 18.0, 14.0; HRMS (EI) calcd. for C₁₇H₁₈N₂O [M⁺]: 266.1419, found: 266.1424

4d: 2-cyano-3-(4-fluorophenyl)propanamide



F m.p. 149-150 °C; IR (neat): 3468, 3323, 3201, 2250, 1676, 1614; ¹H NMR (400 MHz, DMSO- d_6) δ 7.77 (s, 1H), 7.49 (s, 1H), 7.35-7.14 (m, 4H), 3.95-3.91 (m, 1H), 3.14 (dd, J = 13.8, 7.0 Hz, 1H), 3.05 (dd, J = 13.8, 8.6 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ 166.7, 162.8, 160.9, 133.6, 133.6, 131.5, 131.4, 118.8, 115.8, 115.6, 34.8; HRMS (EI) calcd. for C₁₀H₉N₂OF [M⁺]: 192.0699, found: 192.0703

6d: 6-ethyl-4-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile



F m.p. 128-129 °C; IR (neat): 2272, 1695, 1618; ¹H NMR (400 MHz, DMSO- d_6) δ 12.58 (s, 1H), 7.68-7.33 (m, 4H), 6.30 (s, 1H), 2.55 (q, J = 7.4 Hz, 2H), 1.16 (t,

J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 164.2, 162.2, 161.4, 159.3, 157.3, 132.5, 132.5, 130.6, 130.5, 116.5, 115.9, 115.7, 105.0, 97.7, 26.1, 12.6; HRMS (EI) calcd. for C₁₄H₁₁N₂OF [M⁺]: 242.0855, found: 242.0856

5d: 4-(4-fluorophenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



F m.p. 253-254 °C; IR (neat): 2224, 1652, 1603; ¹H NMR (400 MHz, DMSO- d_6) δ 12.59 (s, 1H), 7.37-7.33 (m, 4H), 2.30 (s, 3H), 1.69 (s, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 164.0, 162.0, 161.6, 160.7, 151.3, 133.3, 133.0, 130.8, 130.7, 117.0, 116.5, 116.3, 112.2, 100.6, 18.7, 14.6; HRMS (EI) calcd. for C₁₄H₁₁N₂OF [M⁺]: 242.0855, found: 242.0856

4e: 2-cyano-3-(2-methoxyphenyl)propanamide



COCH₃ m.p. 116-117 °C; IR (neat): 3334, 3199, 2245, 1683, 1602; ¹H NMR (400 MHz, DMSO- d_6) δ 7.79 (s, 1H), 7.47 (s, 1H), 7.29-6.88 (m, 4H), 3.92-3.88 (m, 1H), 3.08 (dd, J = 13.6, 6.8 Hz, 1H), 3.02 (dd, J = 13.6, 9.2 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ 166.3, 157.2, 130.4, 128.7, 124.5, 120.3, 118.5, 110.8, 55.4, 37.3, 30.2; HRMS (EI) calcd. for C₁₁H₁₂N₂O₂ [M⁺]: 204.0899, found: 204.0901

6e: 6-ethyl-4-(2-methoxyphenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile



 OCH_3^{v} m.p. 249-250 °C; IR (neat): 2223, 1644, 1617; ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.50 (s, 1H), 7.48-7.03 (m, 4H), 6.18 (s, 1H), 3.78 (s, 3H), 2.55 (q, *J* = 7.4 Hz, 2H), 1.16 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆) δ 161.7, 159.1, 157.0, 156.2, 131.9, 129.8, 125.8, 121.1, 116.7, 112.4, 106.6, 100.6, 56.1, 26.5, 13.1; HRMS (EI) calcd. for C₁₅H₁₄N₂O₂ [M⁺]: 254.1055, found: 254.1057

5e: 4-(2-methoxyphenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



OCH₃ m.p. 262-263 °C; IR (neat): 2223, 1652, 1605; ¹H NMR (400 MHz, DMSO- d_6) δ 12.55 (s, 1H), 7.49-7.06 (m, 4H), 3.76 (s, 3H), 2.30 (s, 3H), 1.63 (s, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 160.1, 159.7, 155.2, 149.7, 130.8, 128.8, 124.9, 120.7, 116.3, 112.2, 111.7, 100.4, 55.6, 17.9, 13.5; HRMS (EI) calcd. for C₁₅H₁₄N₂O₂ [M⁺]: 254.1055, found: 254.1052

4f: 3-(3-chlorophenyl)-2-cyanopropanamide



Cl m.p. 115-116 °C; IR (neat): 3366, 3200, 2250, 1675, 1624; ¹H NMR (400 MHz, DMSO- d_6) δ 7.78 (s, 1H), 7.51 (s, 1H), 7.39-7.25 (m, 4H), 4.00-3.96 (m, 1H), 3.16 (dd, J = 13.6, 6.8 Hz, 1H), 3.07 (dd, J = 14.0, 8.8 Hz, 1H); ¹³C NMR (125 MHz, DMSO- d_6) δ 166.0, 139.4, 132.9, 130.2, 128.8, 127.8, 127.1, 118.2, 34.6; HRMS (EI) calcd. for C₁₀H₉N₂OCl [M⁺]: 208.0403, found: 208.0402

6f: 4-(3-chlorophenyl)-6-ethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 253-254 °C; IR (neat): 2220, 1640, 1616; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.68 (s, 1H), 7.68-7.58 (m, 4H), 6.37 (s, 1H), 2.58 (q, J = 7.6 Hz, 2H), 1.19 (t, J = 7.6 Hz, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.3, 158.7, 157.6, 138.1, 133.4, 130.7, 130.1, 127.7, 126.8, 116.3, 105.0, 97.9, 26.1, 12.7; HRMS (EI) calcd. for C₁₄H₁₁N₂OCl [M⁺]: 258.0560, found: 258.0563

5f: 4-(3-chlorophenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 249-250 °C; IR (neat): 2223, 1652, 1591; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.65 (s, 1H), 7.56-7.27 (m, 4H), 2.32 (s, 3H), 1.70 (s, 3H); ¹³C NMR (125 MHz, DMSO- d_6) δ 160.1, 159.9, 150.8, 138.3, 133.3, 130.7, 129.0, 127.4, 126.4, 116.1, 111.3, 99.7, 18.0, 13.9; HRMS (EI) calcd. for C₁₄H₁₁N₂OCl [M⁺]: 258.0560, found: 258.0564

5g: 4-(4-isopropylphenyl)-6-methyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 265-266 °C; IR (neat): 2224, 1652, 1615; ¹H NMR (400 MHz,

DMSO- d_6) δ 12.59 (s, 1H), 7.55-7.40 (m, 4H), 6.33 (s, 1H), 2.99-2.92 (m, 1H), 2.29 (s, 3H), 1.22 (d, J = 6.8 Hz, 6H); ¹³C NMR (125 MHz, DMSO- d_6) δ 161.6, 160.0, 151.9, 151.0, 133.5, 128.0, 126.7, 116.8, 106.5, 96.9, 33.3, 23.6, 19.1; HRMS (EI) calcd. for C₁₆H₁₆N₂O [M⁺]: 252.1263, found: 252.1263

5h: 6-ethyl-4-(4-isopropylphenyl)-5-methyl-2-oxo-1,2-dihydropyridine-3-carbonitrile



m.p. 254-255 °C; IR (neat): 2223, 1652, 1598; ¹H NMR (400 MHz, DMSO-*d*₆) δ 12.53 (s, 1H), 7.40-7.22 (m, 4H), 2.99-2.92 (m, 1H), 2.60 (q, *J* = 7.6 Hz, 2H), 1.74 (s, 3H), 1.24 (d, *J* = 6.8 Hz, 6H), 1.14 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (125 MHz, DMSO-*d*₆) δ 162.5, 160.2, 154.6, 149.1, 133.7, 127.7, 126.5, 116.3, 110.5, 100.3, 33.2, 24.6, 23.7, 13.5, 12.7; HRMS (EI) calcd. for C₁₈H₂₀N₂O [M⁺]: 280.1576, found: 280.1579

6-ethyl-4-(4-isopropylphenyl)-5-methyl-2-oxo-1,2,3,4-tetrahydropyridine-3-carbonitril e



5h':

m.p. 184-185 °C; IR (neat): 2253, 1699, 1682; ¹H NMR (400 MHz, CDCl₃) δ 8.21, 8.13 (s, 1H), 7.27-7.08 (m, 4H), 4.14-3.72 (m, 1H), 3.60-3.56 (m, 1H), 2.93-2.86 (m, 1H), 2.33-2.21 (m, 2H), 1.70 (s, 3H), 1.30-1.23 (m, 6H), 1.18-1.11 (m, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 162.6, 162.0, 149.0, 148.9, 134.6, 133.1, 132.7, 132.6, 128.1,

127.4, 127.4, 127.1, 116.5, 115.1, 110.4, 108.4, 48.0, 46.7, 41.6, 40.9, 33.8, 23.9, 23.9, 23.9, 23.4, 23.2, 16.2, 16.0, 12.3, 12.2; HRMS (EI) calcd. for $C_{18}H_{22}N_2O$ [M⁺]: 282.1732, found: 282.1734

5i: Methyl 4-(4-chlorophenyl)-5-cyano-2-methyl-6-oxo-1,6-dihydropyridine-3carboxylate



m.p. 265-266 °C; IR (neat): 2231, 1734, 1651; ¹H NMR (400 MHz,

CDCl₃) δ 13.01 (bs, 1H), 7.58-7.34 (m, 4H), 3.40 (s, 3H), 2.40 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 165.2, 159.7, 158.3, 153.6, 134.6, 134.3, 129.1, 128.6, 115.3, 111.6, 100.7, 52.1, 18.4; HRMS (EI) calcd. for C₁₅H₁₁N₂O₃Cl [M⁺]: 302.0458, found: 302.0459

4da: 3-amino-2-(4-fluorobenzyl)propanamide



F NH₂ m.p. 121-122 °C; IR (neat): 3355, 3298, 1670, 1606, 1510; ¹H NMR (400 MHz, DMSO- d_6) δ 7.31 (s, 1H), 7.21-7.07 (m, 4H), 6.75 (s, 1H), 2.74-2.42 (m, 5H); ¹³C NMR (100 MHz, DMSO- d_6) δ 175.6, 161.8, 159.4, 136.4, 130.5, 130.4, 114.7, 114.6, 50.7, 43.8, 34.5; HRMS (EI) calcd. for C₁₀H₁₃FN₂O [M⁺]: 196.1012, found: 196.1012

4db: 3-amino-2-(4-fluorobenzyl)propanoic acid



F NH₂ m.p. 242-243 °C; IR (neat): 3362, 3184, 1665, 1637, 1511; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.10 (s, 2H), 7.75 (s, 1H), 7.38-7.10 (m, 4H), 2.91-2.67 (m, 5H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ173.4, 162.1, 159.7, 134.6, 134.6, 130.9, 130.8, 115.0, 114.8, 44.4, 34.8; HRMS (EI) calcd. for C₁₀H₁₂FNO₂ [M⁺]: 197.0852, found: 197.0849

Copies of ¹H NMR, ¹³C NMR and HRMS of Compounds

4a: 3-(4-chlorophenyl)-2-cyanopropanamide





6a: 4-(4-chlorophenyl)-6-ethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





5a: 4-(4-chlorophenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile







6b: 6-ethyl-2-oxo-4-(4-(trifluoromethyl)phenyl)-1,2-dihydropyridine-3-carbonitrile







5,6-dimethyl-2-oxo-4-(4-(trifluoromethyl)phenyl)-1,2-dihydropyridine-3-carbonitrile





4c: 2-cyano-3-(4-isopropylphenyl)propanamide





6c: 6-ethyl-4-(4-isopropylphenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile





5c: 4-(4-isopropylphenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile











6d: 6-ethyl-4-(4-fluorophenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile











4e: 2-cyano-3-(2-methoxyphenyl)propanamide





6e: 6-ethyl-4-(2-methoxyphenyl)-2-oxo-1,2-dihydropyridine-3-carbonitrile





5e: 4-(2-methoxyphenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





4f: 3-(3-chlorophenyl)-2-cyanopropanamide





6f: 4-(3-chlorophenyl)-6-ethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





5f: 4-(3-chlorophenyl)-5,6-dimethyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





5g: 4-(4-isopropylphenyl)-6-methyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





5h: 6-ethyl-4-(4-isopropylphenyl)-5-methyl-2-oxo-1,2-dihydropyridine-3-carbonitrile





5h':

6-ethyl-4-(4-isopropylphenyl)-5-methyl-2-oxo-1,2,3,4-tetrahydropyridine-3-carbonitril





5i: Methyl 4-(4-chlorophenyl)-5-cyano-2-methyl-6-oxo-1,6-dihydropyridine-3carboxylate







4db: 3-amino-2-(4-fluorobenzyl)propanoic acid



