Contents lists available at ScienceDirect

European Journal of Medicinal Chemistry

journal homepage: http://www.elsevier.com/locate/ejmech



Synthesis and evaluation of bibenzyl glycosides as potent tyrosinase inhibitors

Reiko Tajima^a, Hiromi Oozeki^a, Seiichi Muraoka^a, Saori Tanaka^a, Yukari Motegi^a, Hiroyuki Nihei^a, Yoichi Yamada^b, Noriyoshi Masuoka^c, Ken-ichi Nihei^{a,*}

^a Department of Applied Biochemistry, Faculty of Agriculture, Utsunomiya University, Utsunomiya, Tochigi 321-0943, Japan ^b Department of Chemistry, Faculty of Education, Utsunomiya University, Utsunomiya, Tochigi 321-0943, Japan

^c Department of Life Science, Okayama University of Science, Okayama 700–0005, Japan

ARTICLE INFO

Article history: Received 18 October 2010 Received in revised form 27 January 2011 Accepted 27 January 2011 Available online 3 February 2011

Keywords: Tyrosinase inhibitor Bibenzyl glycoside Wittig reaction Glycosylation

1. Introduction

Tyrosinase, a widely available copper-containing oxidoreductase, catalyzes the oxidation of monophenol and o-diphenol to o-quinone [1]. In mammals, L-tyrosine serves as a typical monophenol substrate. Numerous phenolics having monophenol and o-diphenol structures found in plants and insects are similarly oxidized by tyrosinase [2]. This enzymatic oxidation is the ratelimiting step in several phenomena of colour development and cuticle sclerotization; the remainder of the reaction sequence proceeds spontaneously at physiological conditions [3].

The presence of the oxidation products of L-tyrosine has recently been linked to the demise of neurons in several neurodegenerative disorders such as Parkinson's and Huntington's diseases [4.5]. Thus, the study of tyrosinase inhibition is clearly important for the development of therapeutic agents, and also for that of antioxidants, functional cosmetics and insecticides [6].

An effective tyrosinase inhibitor should optimally be both safe and potent. For maximal safety, a tyrosinase inhibitor should contain a glycoside moiety. For example, the cosmetics industry, which uses tyrosinase inhibitors as whitening agents, has adopted the use of the two glycosides: arbutin, a hydroquinone glycoside, and aloesin, a C-glycosylated chromone, primarily because other

ABSTRACT

Bibenzyl glycosides 1-6 were synthesized from 2,4-dihydoxybenzaldehyde and xylose, glucose, cellobiose or maltose. The key steps in the synthesis were the Wittig reaction and trichloroacetimidate glycosylation. Tests for tyrosinase inhibitory activity showed that all were significantly active, indicating that they are unique hydrophilic tyrosinase inhibitors. Bibenzyl xyloside 2 is a particularly potent inhibitor ($IC_{50} = 0.43 \ \mu$ M, 17 times higher than that of kojic acid). These results suggest that the hydrophilic cavity of tyrosinase might accommodate the bulky carbohydrate on the bibenzyl scaffold. © 2011 Elsevier Masson SAS. All rights reserved.

> tyrosinase inhibitors such as linoleic acid, hinokitiol, kojic acid, naturally occurring hydroquinones and catechols are known to cause side effects [7]. On the other hand, 4-substituted resorcinol moiety would be requisite to the potent tyrosinase inhibitor [8,9]. For example, 4-hexylresorcinol is effective for preventing shrimp melanosis and for controlling browning in fresh and dried fruit slices [10,11].

> Bibenzyl xyloside 1 was tentatively identified in 1993 in the methanol extract of Chlorophytum arundinaceum (liliaceae) [12]. Its structure, shown in Fig. 1, contains both the glycoside and 4resorcinol moieties. Thus, it shows potential as a key molecule to design novel tyrosinase inhibitors that are safe for use and exhibit potency and hydrophilicity. In this study, we describe the concise



Fig. 1. Structure of bibenzyl derivatives 1-7.



Corresponding author. Tel.: +81 28 649 5412; fax: +81 28 649 5401. E-mail address: nihei98@cc.utsunomiya-u.ac.jp (K. Nihei).

^{0223-5234/\$ -} see front matter © 2011 Elsevier Masson SAS. All rights reserved. doi:10.1016/j.ejmech.2011.01.065



Fig. 2. Synthetic plan for bibenzyl glycosides.

synthesis and tyrosinase inhibitory activity of bibenzyl **1** and its derivatives **2**–**7** [13].

2. Results and discussion

2.1. Synthesis of bibenzyl glycoside 1

Bibenzyl glycosides were synthesized by the strategic plan shown in Fig. 2. The bibenzyl structure can be assumed to be consisting of two benzaldehydes and a sugar moiety. Leading reactions in the strategic plan were Wittig olefination to form a stilbene scaffold and trichloroacetimidate etherification to introduce a sugar moiety on the phenolic hydroxyl of the bibenzyl. Hence, commercially available 2,4-dihydroxybenzaldehyde and xylose were selected as starting materials.

According to the previous report [14], aldehyde **8**, the left part of the bibenzyl structure, was prepared in two steps from 2,4-dihydroxybenzaldehyde. Reaction of 2,4-dihydroxybenzaldehyde by *O*-benzylation, NaBH₄ reduction and chlorination gave chloride **9** [15]. Reaction of chloride **9** with excess triphenylphospine (TPP) furnished phosphonium salt **10** that was transformed to a phosphorus ylide in the presence of lithium hexamethyldisilazide (LiHMDS) as a base (Scheme 1). The ylide was coupled with aldehyde **8** to build the stilbene framework and give stilbene **11** (90%; *cis:trans* = 2:1, as determined by ¹H NMR). Selective reduction at the olefinic position of stilbene **11** by H₂–Pd/C-ethylenediamine complex (H₂–Pd(en)/C) furnished bibenzyl **12** (85%) [16]. The removal of the methoxymethyl (MOM) moiety from the structure of bibenzyl **12** under acidic conditions yielded the corresponding phenol **13** (88%).

Glycosylation of bibenzyl **13** with imidate **14** prepared from p-xylose was quantitatively accomplished using the catalytic amount of trimethylsilyl trifluoromethanesulfonate (TMSOTf). Note that the by-product α -glycoside of xyloside **15** was not detected in the chromatographic separation. The use of BF₃·Et₂O as a Lewis acid decreased reaction yield to approximately 80%. The removal of the benzyl (Bn) protective groups from xyloside **15** by

 Table 1

 Tyrosinase inhibitory activities of bibenzyl derivatives 1–7 and kojic acid.

Compounds tested	IC ₅₀ (μM)
1	$1.6\pm0.43^{\text{a}}$
2	0.43 ± 0.18
3	0.73 ± 0.11
4	0.77 ± 0.04
5	0.68 ± 0.05
6	0.83 ± 0.06
7	0.37 ± 0.06
Kojic acid	$\textbf{7.4} \pm \textbf{1.4}$

 a The IC_{50} values represent means \pm SE of three different experiments.

hydrogenolysis with H_2 –Pd(OH)₂ on carbon, followed by ester exchange on the acetyl moieties by using NaOMe, gave bibenzyl glycoside **1** in excellent yield (94%, two steps). The overall yield of this synthetic scheme from aldehyde **8** was 63% (six steps).

2.2. Tyrosinase inhibitory activity of bibenzyl glycoside 1

The tyrosinase inhibitory activities of the synthesized bibenzyl glycosides evaluated against L-DOPA oxidation catalyzed by purified tyrosinase, are listed in Table 1. Kojic acid was used as the reference standard. The potent inhibitory activity, $IC_{50} = 1.6 \mu M$, was observed for 2-xylosylated bibenzyl **1**, that was 5 times higher than that of kojic acid.

A number of bibenzyls that exhibit various biological activities, such as antifungal, phytotoxic and anti-HIV effects, have been isolated from the extracts of several organisms including liverworts, algae and ferns [17–20]. However, significant tyrosinase inhibitory activity exhibited on the part of a bibenzyl glycoside is a novel observation. This finding prompted us to further investigate derivatives 2-7 by a chemical approach.

2.3. Synthesis of bibenzyls 2-7

Bibenzyl glycosides **2** and **3** were synthesized by a procedure similar to that of bibenzyl glycoside **1** (Scheme 2). The reaction of 2,4-dihydroxybenzaldehyde with MOMCl gave benzaldehyde **16**. In the presence of LiHMDS, the Wittig reaction of benzaldehyde **16** with phosphonium salt **10** furnished stilbene **17** (84%; *cis:trans* = 3:2, as determined by ¹H NMR). Pd(en)/C-catalyzed hydrogenation at the olefinic position of stilbene **17** gave bibenzyl **18** (72%). The removal of the MOM moieties from bibenzyl **18** under acidic conditions quantitatively yielded diphenol **19**.



Scheme 1. Synthesis of bibenzyl xyloside 1. Reagents and conditions: (a) TPP, toluene, reflux, 0.5 h, 59%; (b) LiHMDS, THF, 0 °C to rt, 1 h, 90%; (c) H₂–Pd(en)/C, THF, rt, 12 h, 85%; (d) TsOH, THF, MeOH, reflux, 2 h, 88%; (e) TMSOTF, CH₂Cl₂, 0 °C, 5 min, 100%; (f) H₂–Pd(OH)₂/C, EtOAc, rt, 12 h; (g) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 94% (2 steps).



Scheme 2. Syntheses of bibenzyl xylosides **2** and **3**. Reagents and conditions: (a) **10**, LiHMDS, THF, 0 °C to rt, 1 h, 84%; (b) H₂–Pd(en)/C, THF, rt, 12 h, 72%; (c) TsOH, THF, MeOH, reflux, 2 h, 100%; (d) **14**, TMSOTF, CH₂Cl₂, 0 °C, 5 min, 71%; (e) H₂–Pd(OH)₂/C, EtOAc, rt, 12 h; (f) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 85% (2 steps); (g) **14** (excess), TMSOTF, CH₂Cl₂, 0 °C, 5 min, 77%; (h) H₂–Pd(OH)₂/C, EtOAc, rt, 12 h; (i) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 85% (2 steps).

We speculated that the synthesis of bibenzyl glycosides **2** and **3** should be possible by selective glycosylation of diphenol **19**, with chemoselectivity governed by the molar ratio of diphenol **19** to imidate **14**. The reaction of diphenol **19** with 1.5 equivalent of glycosyl donor **14** selectively furnished mono-glycoside **20** (71%). A cross peak between an oxygenated aryl carbon at C-4 and an anomeric proton was observed in the HMBC spectrum, so the glycosylated site on the benzene ring was determined to be as shown in Scheme 2. A mono-glycosylated product at C-2 and diglycoside **21** were not detected by TLC analysis under these conditions, presumably because of steric crowding from alkyl substitution at C-1. The reaction of mono-glycoside **20** by hydrogenolysis at the O-benzyl position and the removal of the acetyl group by NaOMe gave the desired bibenzyl glycoside **2** (85%, two steps).

In contrast, the reaction of diphenol **19** with 4.0 equivalent of glycosyl donor **14** gave diglycoside **21** (77%). The successive removal of the benzyl and acetyl groups of diglycoside **21** furnished bibenzyl glycoside **3** (85%, two steps). The overall yields of bibenzyl glycosides **2** and **3** from aldehyde **16** were 37 and 40%, respectively (five steps).

The successful synthesis of mono-glycoside **2** indicated that several bibenzyl derivatives could be obtained by the selective glycosylation of diphenol **19**. The use of imidates **22–24**, prepared respectively from D-glucose, D-cellobiose and D-maltose, as glycosyl donors gave glycosides **25–27**, respectively by Scheme 3 (74%, 75% and 57%, respectively). Hydrogenolysis followed by the removal of the acetyl groups from the bibenzyls yielded glycosides **4–6** (63%, 72% and 56%, respectively). The overall yields of glycosides **4–6** from aldehyde **16** were 28%, 33% and 19%, respectively. Symmetric bibenzyl **7** was readily prepared by a method previously reported [21].

2.4. Tyrosinase inhibitory activity of bibenzyls 2-7

Xylosylated bibenzyl **2** exhibited a tyrosinase inhibitory activity of $IC_{50} = 0.43 \mu$ M, 17 times higher than that of kojic acid (Table 1).



Scheme 3. Syntheses of bibenzyl glycosides **4**–**6.** Reagents and conditions: (a) **19**, TMSOTF, CH_2Cl_2 , 0 °C, 5 min, 74%; (b) H_2 –Pd(OH)₂/C, EtOAc, rt, 12 h; (c) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 63% (2 steps); (d) **19**, TMSOTF, CH_2Cl_2 , 0 °C, 5 min, 75%; (e) H_2 –Pd (OH)₂/C, EtOAc, rt, 12 h; (f) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 72% (2 steps); (g) **19**, TMSOTF, CH_2Cl_2 , 0 °C, 1 h, 57%; (h) H_2 –Pd(OH)₂/C, EtOAc, rt, 12 h; (i) NaOMe, MeOH, 0 °C, 0.5 h, then H⁺, 56% (2 steps).

Aglycone **7** and xyloside **2** exhibited similar inhibitory activities, indicating that glycosylation at the C-4 position of the bibenzyl skeleton has minimal effect on activity. In contrast, C-2-xylosylated bibenzyls **1** and **3** exhibited relatively low inhibitory activities, possibly because the resorcinol moiety could be weakly bound on the active site of tyrosinase for bulky xylosyl substitution at C-2.

Glycosylation at the C-4 position with a large sugar moiety appears to influence inhibitory activity. For example, glucoside **4**, cellobioside **5** and maltoside **6** exhibited relatively low activities (IC₅₀ = 0.77, 0.68 and 0.83 μ M, respectively). Noted that all the synthesized bibenzyls did not act as substrates, as confirmed by incubated experiments with tyrosinase observed by UV spectrophotometry (data not shown) [22–24]. Although the natural occurrence of xyloside **1** is questionable because of inconsistencies in the ¹H and ¹³C NMR data for natural **1** and synthetic **1** [12,13], it is appropriate to newly categorize the bibenzyl glycosides as tyrosinase inhibitors that are both potent and hydrophilic.

Hydrophobic substitutions on tyrosinase inhibitors tend to enhance inhibitory activity [25,26]. The isopropyl ester moiety of 4-resorcinol inhibitor was recently reported to interact significantly with the hydrophobic site on tyrosinase [27]. However, hydrophilic interactions between an inhibitor and tyrosinase have not yet been investigated in detail. The high activities of our synthesized bibenzyl glycosides suggest that tyrosinase might possess a hydrophilic cavity that can accommodate a carbohydrate moiety. Taking advantage of this cavity might enable the design of novel tyrosinase inhibitors for practical use.

3. Conclusions

A series of bibenzyl glycosides was synthesized from 2,4dihydroxybenzaldehyde and D-xylose, D-glucose, D-cellobiose, or D-maltose via the Wittig reaction and trichloroacetimidate glycosylation. In evaluations of tyrosinase inhibitory activity against L-DOPA oxidation catalyzed by the purified enzyme, all of the synthesized bibenzyl glycosides showed greater activity than the common inhibitor kojic acid, used as a reference standard. Bibenzyl xyloside **2** is particularly potent inhibitor (IC₅₀ = 0.43 μM, 17 times higher than that of kojic acid). The high activities of the bulky bibenzyl glycosides suggest that tyrosinase might possess a hydrophilic cavity at its catalytic center. This hydrophilic interaction as well as the known hydrophobic interaction of the development of new and effective tyrosinase inhibitors.

4. Experimental

4.1. Chemistry

4.1.1. General

Optical rotations were recorded by a Horiba SEPA-300. IR spectra were measured with a Horiba FT-720 spectrometer. NMR spectra were recorded with a JEOL EX-400 spectrometer operating at 400 MHz for the ¹H NMR spectra and at 100 MHz for the ¹³C NMR spectra. Chemical shifts were recorded as ppm relative to the TMS signal for CDCl₃ or the solvent signal (¹H NMR; CH₃OH: 3.30 ppm or pyridine: 8.71 ppm, ¹³C NMR; CH₃OH: 49.0 ppm or pyridine: 149.2 ppm). HRMS spectra were measured on a JEOL AccuTOF mass spectrometer fitted with an electrospray ion source in positive or negative ionization mode. Preparative HPLC was carried out by using 1.0 mL/min of flow rate with Wakosil-II 5C18 AR Prep as a column and Jasco 880PU as a pump.

4.1.2. 2',4,4'-Tribenzyloxy-2-methoxymethoxystilbene (11)

TPP (2.5 g, 9.5 mmol) and 2,4-di(benzyloxy)benzyl chloride (**9**) [15] (2.0 g, 5.9 mmol) were dissolved in toluene (10 mL) and the resultant solution was refluxed for 0.5 h. The reaction mixture was allowed to cool to room temperature and diluted with hexane (50 mL). Yellowish precipitate was collected by filtration and washed with Et_2O (2 mL) to give compound **10** as a white powder (2.1 g, 59% from **9**), which was used in the next step without further purification.

To a suspension of 10 (0.78 g, 1.3 mmol) in THF (5 mL), 0.84 mL of LiHMDS (1.6 M in THF, 1.3 mmol) was added at 0 °C. The resultant solution was stirred at room temperature for 0.5 h. The solution was then cooled to 0 °C, and a THF (5 mL) solution of 4-benzyloxy-2methoxymethoxybenzaldehyde (8) [14] (0.20 g, 0.73 mmol) was slowly added. After being stirred for 1 h at room temperature, the reaction mixture was poured into Et₂O (60 mL) and washed with saturated aqueous NH₄Cl solution (15 mL \times 3) and brine (15 mL \times 3). The aqueous layers were extracted with Et₂O (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (2.5-5% EtOAc in hexane) gave the title compound 11 (0.37 g; *cis:trans* = 2:1, as determined by ¹H NMR; 90% from **8**) as colorless oil. IR (film) v_{max} 3031, 2871, 1604, 1498, 1257, 1026, 735 cm⁻¹. *Cis*-**11**; ¹H NMR (400 MHz, CDCl₃) δ 7.39 (15H, m, ArH), 7.13 (2H, d, J = 8.3 Hz, ArH), 6.77 (1H, d, J = 2.4 Hz, ArH), 6.70 (1H, d, J = 12.2 Hz, CH), 6.60 (1H, d, J = 12.2 Hz, CH), 6.57 (1H, d, J = 2.4 Hz, ArH), 6.43 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.37 (1H, dd, J = 2.4, 8.3 Hz, ArH), 5.13 (2H, s, CH₂), 5.05 (2H, s, CH₂), 5.01 (2H, s, CH₂), 4.99 (2H, s, CH₂), 3.45 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 159.0 (C), 158.9 (C), 157.3 (C), 155.9 (C), 137.1 (C), 136.9 (C), 136.8 (C), 130.5 (CH), 130.4 (CH), 128.59 (CH), 128.55 (CH), 128.49 (CH), 128.0 (CH), 127.9 (CH), 127.6 (CH), 127.2 (CH), 124.0 (C), 123.7 (C), 120.6 (CH), 120.1 (CH), 107.4 (CH), 105.8 (CH), 102.5 (CH), 100.7 (CH), 94.9 (CH₂),

70.3 (CH₂), 70.08 (CH₂), 70.05 (CH₂), 56.1 (CH₃). *Trans*-**11**; ¹H NMR (400 MHz, CDCl₃) δ 7.54 (1H, d, *J* = 8.8 Hz, ArH), 7.48 (1H, d, *J* = 8.8 Hz, ArH), 7.39 (17H, m, ArH and CH), 6.78 (1H, d, *J* = 2.4 Hz, ArH), 6.63 (1H, dd, *J* = 2.4, 8.8 Hz, ArH), 6.58 (2H, m, ArH), 5.15 (2H, s, CH₂), 5.08 (2H, s, CH₂), 5.05 (4H, s, CH₂), 3.43 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 159.1 (C), 158.9 (C), 156.9 (C), 155.4 (C), 136.9 (C), 128.60 (CH), 128.55 (CH), 128.02 (CH), 127.97 (CH), 127.9 (CH), 127.6 (CH), 127.5 (CH), 127.4 (CH), 127.1 (CH), 121.9 (C), 121.7 (C), 121.4 (CH), 121.1 (CH), 108.0 (CH), 106.5 (CH), 102.6 (CH), 100.9 (CH), 94.8 (CH₂), 70.4 (CH₂), 70.2 (CH₂), 70.1 (CH₂), 56.0 (CH₃). ESIHRMS *m*/*z* 581.2282 [M + Na]⁺ (calcd for C₃₇H₃₄NaO, 581.2304).

4.1.3. 2',4,4'-Tribenzyloxy-2-methoxymethoxybibenzyl (12)

A THF (3 mL) solution of **11** (0.11 g, 0.20 mmol) was hydrogenated over 5% Pd(en) on carbon (33 mg) for 12 h under atmospheric pressure [16]. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (2-10% EtOAc in hexane) gave the title compound 12 (95 mg, 85%) as a white solid. IR (film) ν_{max} 3031, 1604, 1498, 1257, 1026, 735 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.37 (15H, m, ArH), 7.01 (1H, d, *J* = 8.8 Hz, ArH), 6.94 (1H, d, J = 8.8 Hz, ArH), 6.74 (1H, d, J = 2.4 Hz, ArH), 6.59 (1H, d, J = 2.4 Hz, ArH), 6.50 (2H, m, ArH), 5.03 (2H, s, CH₂), 5.02 (4H, s, CH₂), 5.01 (2H, s, CH₂), 3.39 (3H, s, CH₃), 2.84 (4H, s, CH₂). ¹³C NMR (100 MHz, CDCl₃) δ 158.2 (C), 158.1 (C), 157.4 (C), 155.9 (C), 137.3 (C), 137.2 (C), 130.3 (CH), 130.2 (CH), 128.6 (CH), 128.5 (CH), 127.93 (CH), 127.88 (CH), 127.76 (CH), 127.5 (CH), 127.3 (CH), 124.1(C), 123.8 (C), 106.9 (CH), 105.3 (CH), 102.2 (CH), 100.5 (CH), 94.4 (CH₂), 70.2 (CH₂), 70.1 (CH₂), 69.9 (CH₂), 55.8 (CH₃), 30.7 (CH₂), 30.5 (CH₂). ESIHRMS m/z 583.2457 [M + Na]⁺ (calcd for C₃₇H₃₆NaO₅, 583.2460).

4.1.4. 2',4,4'-Tribenzyloxy-2-hydroxybibenzyl (13)

TsOH·H₂O (39 mg, 0.21 mmol) was added to a 50% THF/MeOH solution (6 mL) of bibenzyl 12 (95 mg, 0.17 mmol) at room temperature. After refluxed for 2 h, the reaction mixture was poured into EtOAc (60 mL) and washed with H_2O (15 mL \times 3) and brine (15 mL \times 3). The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (10-20% EtOAc in hexane) gave the title compound 13 (78 mg, 88%) as a white solid. IR (film) *v*_{max} 3400, 3031, 2866, 1612, 1504, 1290, 1026, 735 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.37 (15H, m, ArH), 7.02 (1H, d, J = 8.3 Hz, ArH), 6.94 (1H, d, J = 8.3 Hz, ArH), 6.61 (1H, d, *J* = 2.4 Hz, ArH), 6.50 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 6.49 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.47 (1H, bs, OH), 5.11 (2H, s, CH₂), 5.00 (2H, s, CH₂), 4.99 (2H, s, CH₂), 2.76 (4H, m, CH₂). ¹³C NMR (100 MHz, CDCl₃) & 158.5 (C), 157.1 (C), 154.9 (C), 137.1 (C), 137.0 (C), 136.6 (C), 130.4 (CH), 130.3 (CH), 128.7 (CH), 128.6 (CH), 128.5 (CH), 128.1 (CH), 128.0 (CH), 127.9 (CH), 127.5 (CH), 127.4 (CH), 123.0 (C), 120.1 (C), 106.9 (CH), 105.7 (CH), 102.6 (CH), 100.9 (CH), 70.3 (CH₂), 70.2 (CH₂), 70.0 (CH₂), 31.4 (CH₂), 30.8 (CH₂). ESIHRMS m/z 515.2224 $[M - H]^{-}$ (calcd for C₃₅H₃₁O₄, 515.2222).

4.1.5. 2-(2",3",4"-Tri-O-acetyl-β-*D*-xylopyranosyl)-2',4,4'-tribenzyloxybibenzyl (**15**)

To a cold (0 °C) and stirred solution of bibenzyl **13** (29 mg, 56 μ mol) and 2,3,4-tri-O-acetyl- α -D-xylopyranosyl trichloroacetimidate (**14**) (47 mg, 0.11 mmol) in CH₂Cl₂ (1 mL) was added 5 μ L of TMSOTf solution (0.11 M in CH₂Cl₂, 0.55 μ mol). After being stirred for 5 min at 0 °C, the resultant solution was poured into EtOAc (60 mL) and washed with saturated aqueous NaHCO₃ solution (15 mL \times 3) and brine (15 mL \times 3). The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (15–25% EtOAc in hexane) gave the title compound **15** (43 mg, 100% from **13**) as a white solid. $[\alpha]_D^{19}$ -11.8 (c 1.27, CHCl₃). IR (film) v_{max} 3031, 1754, 1506, 1220, 1039, 739 m⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.37 (15H, m, ArH), 6.99 (1H, d, J = 8.3 Hz, ArH), 6.86 (1H, d, J = 8.3 Hz, ArH), 6.67 (1H, d, *J* = 2.0 Hz, ArH), 6.57 (1H, d, *J* = 2.0 Hz, ArH), 6.52 (1H, dd, *J* = 2.0, 8.3 Hz, ArH), 6.48 (1H, dd, J = 2.0, 8.3 Hz, ArH), 5.17 (1H, d, *J* = 6.4 Hz, CH), 5.15 (1H, t, *J* = 8.4 Hz, CH), 5.01 (8H, m, CH), 4.14 (1H, dd, J = 4.9, 12.2 Hz, CH₂), 3.43 (1H, dd, J = 8.4, 12.2 Hz, CH₂), 2.77 (4H, m, CH₂), 2.06 (3H, s, CH₃), 2.05 (3H, s, CH₃), 1.97 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 170.0 (C), 169.8 (C), 169.3 (C), 158.1 (C), 157.9 (C), 157.3 (C), 155.0 (C), 137.3 (C), 137.1 (C), 130.5 (CH), 130.3 (CH), 128.54 (CH), 128.46 (CH), 128.0 (CH), 127.9 (CH), 127.7 (CH), 127.5 (CH), 127.2 (CH), 124.2 (C), 123.3 (C), 108.1 (CH), 105.2 (CH), 103.0 (CH), 100.4 (CH), 98.2 (CH), 70.6 (CH), 70.14 (CH₂), 70.07 (CH₂), 70.0 (CH), 69.7 (CH₂), 68.3 (CH), 61.6 (CH₂), 30.0 (CH₂), 20.8 (CH₃), 20.6 (CH₃). ESIHRMS m/z 797.2918 [M + Na]⁺ (calcd for C46H46NaO11, 797.2938).

4.1.6. 2', 4,4'-Trihydroxy-2-(β -D-xylopyranosyl)bibenzyl (**1**)

Bibenzyl **15** (14 mg, 18 μ mol) in EtOAc (1 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (3 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (EtOAc) gave the phenol as a white solid, which was used in the next step without further purification.

To cold $(0 \circ C)$ and stirred solution of the phenol in MeOH (5 mL) was added 0.10 mL of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H₂O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (20% MeCN in H₂O, $t_{\rm R}$ = 10.1 min). The title compound **1** (6.5 mg, 94%) was obtained as a colorless solid. $[\alpha]_{D}^{20}$ -10.6 (*c* 0.09, MeOH). IR (Nujol) ν_{max} 3350, 1603, 1508 cm⁻¹. ¹H NMR (pyridine- d_5 , 400 MHz) δ 7.42 (1H, d, J = 2.0 Hz, ArH), 7.33 (1H, d, J = 7.8 Hz, ArH), 7.32 (1H, d, J = 8.0 Hz, ArH), 7.01 (1H, d, J = 2.0 Hz, ArH), 6.90 (1H, dd, J = 2.0, 8.0 Hz, ArH), 6.73 (1H, dd, *J* = 2.0, 7.8 Hz, ArH), 5.43 (1H, d, *J* = 7.8 Hz, CH), 4.34 (1H, dd, *J* = 7.8, 8.3 Hz, CH), 4.28 (1H, dd, J = 4.9, 10.7 Hz, CH₂), 4.26 (1H, m, CH), 4.20 (1H, dd, J = 8.3, 8.8 Hz, CH), 3.66 (1H, m, CH₂), 3.50 (2H, m, CH₂), 3.35 (2H, m, CH₂). 13 C NMR (pyridine- d_5 , 100 MHz) δ 157.7 (C), 157.6 (C), 157.1 (C), 156.8 (C), 130.9 (CH), 130.3 (CH), 122.6 (C), 120.1 (C), 109.3 (CH), 106.7 (CH), 104.0 (CH), 103.2 (CH), 77.7 (CH), 74.3 (CH), 70.2 (CH), 66.5 (CH₂), 31.2 (CH₂), 31.0 (CH₂). HRESIMS m/z $377.1240 [M - H]^{-}$ (calcd for C₁₉H₂₁O₈, 377.1236).

4.1.7. 2',4'-Dibenzyloxy-2,4-di(methoxymethoxy)stilbene (17)

At 0 °C, LiHMDS (1.3 mL, 1.6 M solution in THF, 2.1 mmol) was added to a suspension of 10 (1.3 g, 2.1 mmol) in THF (5 mL). The resultant solution was stirred at room temperature for 0.5 h. The solution was then cooled to 0 °C, and a THF (5 mL) solution of 2,4-di (methoxymethoxy)benzaldehyde (16) (0.3 g, 1.3 mmol) was slowly added. After being stirred for 1 h at room temperature, the reaction mixture was poured into EtOAc (100 mL) and washed with saturated aqueous NH₄Cl solution (20 mL \times 3) and brine (20 mL \times 3). The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (20-30% EtOAc in hexane) gave the title compound **17** (0.57 g; *cis:trans* = 3:2, as determined by ¹H NMR; 84% from **16**) as colorless oil. IR (film) ν_{max} 3031, 2871, 1604, 1498, 1257, 1026, 735 cm⁻¹. *Cis*-**17**; ¹H NMR (400 MHz, CDCl₃) δ 7.37 (10H, m, ArH), 7.13 (1H, d, J = 8.3 Hz, ArH), 7.12 (1H, d, J = 8.3 Hz, ArH), 6.79 (1H, d, J = 2.4 Hz, ArH), 6.71 (1H, d, *J* = 12.2 Hz, CH), 6.60 (1H, d, *J* = 12.2 Hz, CH), 6.58 (1H, d, *J* = 2.4 Hz, ArH), 6.49 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 6.37 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 5.14 (2H, s, CH₂), 5.13 (2H, s, CH₂), 5.06 (2H, s, CH₂), 4.99 (2H, s, CH₂), 3.47 (3H, s, CH₃), 3.46 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 159.1 (C), 157.32 (C), 157.28 (C), 155.7 (C), 137.1 (C), 137.0 (C), 130.5 (CH), 130.4 (CH), 128.6 (CH), 128.5 (CH), 128.0 (CH), 127.8 (CH), 127.6 (CH), 127.2 (CH), 124.2 (CH), 123.7 (CH), 120.0 (C), 119.8 (C), 109.0 (CH), 105.8 (CH), 103.8 (CH), 100.7 (CH), 94.9 (CH₂), 94.5 (CH₂), 70.3 (CH₂), 70.1 (CH₂), 56.1 (CH₃). Trans-17; ¹H NMR (400 MHz, CDCl₃) δ 7.56 (1H, d, I = 8.8 Hz, ArH), 7.48 (1H, d, I = 8.8 Hz, ArH), 7.39 (10H, m, ArH), 7.35 (1H, d, *I* = 16.6 Hz, CH), 7.29 (1H, d, *I* = 16.6 Hz, CH), 6.80 (1H, d, J = 2.4 Hz, ArH), 6.70 (1H, dd, J = 2.4, 8.8 Hz, ArH), 6.60 (2H, m, ArH), 5.16 (4H, s, CH₂), 5.08 (2H, s, CH₂), 5.06 (2H, s, CH₂), 3.48 (3H, s, CH₃), 3.45 (3H, s, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 159.2 (C), 157.3 (C), 157.0 (C), 155.3 (C), 137.0 (C), 136.9 (C), 128.61 (CH), 128.55 (CH), 128.0 (CH), 127.9 (CH), 127.5 (CH), 127.4 (CH), 127.10 (CH), 127.05 (CH), 122.32 (CH), 122.26 (C), 121.6 (C), 109.5 (CH), 106.5 (CH), 103.9 (CH), 100.8 (CH), 94.8 (CH₂), 94.5 (CH₂), 70.4 (CH₂), 70.2 (CH₂), 56.13 (CH₃), 56.08 (CH₃), HRESIMS m/z 532.2003 [M + Na]⁺ (calcd for C32H32NaO6, 532.2097).

4.1.8. 2',4'-Dibenzyloxy-2,4-di(methoxymethoxy)bibenzyl (18)

A THF (5 mL) solution of 17 (0.20 g, 0.40 mmol) was hydrogenated over 5% Pd(en) on carbon (62 mg) for 12 h [16]. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (15% EtOAc in hexane) gave the title compound 18 (0.15 g, 72%) as a white solid. IR (film) v_{max} 3031, 1604, 1498, 1257, 1026, 735 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.37 (m, 10H, ArH), 7.03 (1H, d, J = 7.8, Hz, ArH), 6.95 (1H, d, J = 8.3 Hz, ArH), 6.75 (1H, d, *J* = 2.0 Hz, ArH), 6.58 (2H, m, ArH), 6.50 (1H, dd, *J* = 1.4, 7.8 Hz, ArH), 5.12 (2H, s, CH₂), 5.04 (2H, s, CH₂), 5.02 (4H, s, CH₂), 3.47 (3H, s, CH₃), 3.40 (3H, s, CH₃), 2.84 (4H, bs, CH₂). ¹³C NMR (100 MHz, CDCl₃) δ 158.2 (C), 157.4 (C), 156.4 (C), 155.8 (C), 137.3 (C), 137.1 (C), 130.3 (CH), 130.1 (CH), 128.6 (CH), 128.5 (CH), 127.9 (CH), 127.8 (CH), 127.5 (CH), 127.2 (CH), 125.1 (C), 123.8 (C), 108.6 (CH), 105.3 (CH), 103.4 (CH), 100.5 (CH), 94.7 (CH₂), 94.3 (CH₂), 70.2 (CH₂), 69.9 (CH₂), 56.0 (CH₃), 55.9 (CH₃), 30.6 (CH₂), 30.5 (CH₂). ESIHRMS m/z 537.2247 $[M + Na]^+$ (calcd for C₃₂H₃₄NaO₆, 537.2253).

4.1.9. 2',4'-Dibenzyloxy-2,4-dihydroxystilbene (19)

TsOH·H₂O (0.19 g, 0.38 mmol) was added to a 50% THF/MeOH solution (8 mL) of bibenzyl 18 (0.29 g, 1.5 mmol) at room temperature. After refluxed for 2 h, the reaction mixture was poured into EtOAc (100 mL) and washed with H₂O (20 mL \times 3) and brine (20 mL \times 3). The aqueous layers were extracted with EtOAc $(20 \text{ mL} \times 3)$, and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (30–35% EtOAc in hexane) gave the title compound **19** (0.16 g, 100%) as a white solid. IR (film) *v*_{max} 3400, 3031, 2866, 1612, 1504, 1290, 1026, 735 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.40 (10H, m, ArH), 7.03 (1H, d, J = 8.3 Hz, ArH), 6.90 (1H, d, J = 8.3 Hz, ArH), 6.62 (1H, bs, ArH), 6.51 (1H, d, J = 8.3 Hz, ArH), 6.30 (2H, m, ArH), 5.46 (1H, bs, OH), 5.12 (2H, s, CH₂), 5.00 (2H, s, CH₂), 4.73 (1H, bs, OH), 2.75 (4H, m, CH₂). ¹³C NMR (100 MHz, CDCl₃) δ 158.5 (C), 157.0 (C), 155.0 (C), 136.9 (C), 136.5 (C), 130.6 (CH), 130.3 (CH), 128.7 (CH), 128.6 (CH), 128.1 (CH), 128.0 (CH), 127.5 (CH), 127.4 (CH), 122.9 (C), 119.9 (C), 107.2 (CH), 105.6 (CH), 103.0 (CH), 100.9 (CH), 70.3 (CH₂), 70.2 (CH₂), 31.4 (CH₂), 30.8 (CH₂). ESIHRMS m/z 425.1661 $[M - H]^-$ (calcd for C₂₈H₂₅O₄, 425.1753).

4.1.10. 4-(2",3",4"-Tri-O-acetyl-β-D-xylopyranosyl)-2',4'-dibenzyloxy-2-hydroxybibenzyl (**20**)

To a cold (0 °C) and stirred solution of bibenzyl **19** (103 mg, 0.24 mmol) and imidate **14** (149 mg, 0.37 mmol) in CH₂Cl₂ (2 mL) was added 21 μ L of TMSOTf solution (0.11 M in CH₂Cl₂). After being stirred for 5 min at 0 °C, the resultant solution was poured into EtOAc (100 mL) and washed with saturated aqueous NaHCO₃ solution (20 mL \times 3) and brine (20 mL \times 3). The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic

layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (45% EtOAc in hexane) gave the title compound 20 (117 mg, 71% from 19) as a white solid. IR (film) $\nu_{\rm max}$ 3032, 1753, 1506, 1223, 1039, 737 m⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.40 (10H, m, ArH), 7.04 (1H, d, J = 8.3 Hz, ArH), 6.96 (1H, d, *J* = 8.8 Hz, ArH), 6.62 (1H, d, *J* = 2.4 Hz, ArH), 6.51 (1H, dd, *J* = 2.4, 7.3 Hz, ArH), 6.48 (2H, m, ArH), 5.52 (1H, bs, OH), 5.22 (1H, t, I = 7.8 Hz, CH), 5.15 (1H, dd, I = 6.4, 7.8 Hz, CH), 5.13 (2H, s, CH₂), 5.09 (1H, d, J = 6.4 Hz, CH), 5.01 (2H, s, CH₂), 4.99 (1H, m, CH), 4.20 (1H, dd, *J* = 4.9, 12.2 Hz, CH₂), 3.49 (1H, dd, *J* = 7.8, 12.2 Hz, CH₂), 2.76 (4H, s, CH₂), 2.08 (9H, m, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 170.1 (C), 169.9 (C), 169.5 (C), 158.5 (C), 157.0 (C), 156.1 (C), 154.9 (C), 136.9 (C), 136.5 (C), 130.5 (C), 130.2 (C), 128.7 (CH), 128.6 (CH), 128.1 (CH), 128.0 (CH), 127.5 (CH), 127.4 (CH), 122.8 (C), 122.4 (C), 108.6 (CH), 105.6 (CH), 104.6 (CH), 100.8 (CH), 98.6 (CH), 70.8 (CH), 70.1 (CH₂), 68.5 (CH), 61.8 (CH₂), 31.2 (CH₂), 30.8 (CH₂), 20.7 (CH₃). ESIHRMS m/z 683.2473 $[M - H]^-$ (calcd for C₃₉H₃₉O₁₁, 683.2492).

4.1.11. 2,2',4'-Trihydroxy-4- β -D-xylopyranosylbibenzyl (**2**)

Bibenzyl **20** (96 mg, 0.12 mmol) in EtOAc (2 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (24 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (60–70% EtOAc in hexane) gave the phenol as a white solid, which was used in the next step without further purification.

To cold $(0 \circ C)$ and stirred solution of the phenol in MeOH (5 mL) was added 110 µL of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H_2O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (20% MeCN in H₂O, $t_R = 7.5$ min). The title compound **2** (40 mg, 85%) was obtained as a colorless solid. $[\alpha]_{D}^{20}$ –5.6 (*c* 0.17, MeOH). IR (Nujol) *v*_{max} 3520, 3350, 1595, 1510 cm⁻¹. ¹H NMR (pyridine- d_5 , 400 MHz) δ 7.35 (1H, d, J = 8.3 Hz, ArH), 7.33 (1H, d, J = 8.3 Hz, ArH), 7.22 (1H, d, J = 2.4 Hz, ArH), 7.08 (1H, d, J = 2.4 Hz, ArH), 6.90 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.80 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 5.44 (1H, d, *J* = 6.8 Hz, CH), 4.31 (1H, m, CH), 4.28 (1H, m, CH₂), 4.26 (1H, m, CH), 4.23 (1H, m, CH), 3.66 (1H, dd, J = 9.3, 10.2 Hz, CH₂), 3.44 (4H, m, CH₂). ¹³C NMR (pyridine- d_5 , 100 MHz) δ 157.7 (C), 157.3 (C), 157.1 (C), 157.0 (C), 130.6 (CH), 130.4 (CH), 123.5 (C), 120.1 (C), 107.2 (CH), 106.6 (CH), 104.4 (CH), 103.3 (CH), 102.8 (CH), 77.8 (CH), 74.2 (CH), 70.3 (CH), 66.6 (CH₂), 30.8 (CH₂), 30.6(CH₂). HRESIMS m/z 377.1234 [M - H]⁻ (calcd for C₁₉H₂₁O₈, 377.1236).

4.1.12. 2,4-Di(2",3",4"-tri-O-acetyl-β-D-xylopyranosyl)-2',4'-dibenzyloxybibenzyl (**21**)

To a cold (0 °C) and stirred solution of bibenzyl 19 (87 mg, 0.20 mmol) and imidate 14 (387 mg, 9.2 mmol) in CH₂Cl₂ (5 mL) was added 36 µL of TMSOTf solution (0.11 M in CH₂Cl₂). After being stirred for 5 min at 0°C, the resultant solution was poured into EtOAc (60 mL) and washed with saturated aqueous NaHCO₃ solution $(15 \text{ mL} \times 3)$ and brine $(15 \text{ mL} \times 3)$. The aqueous layers were extracted with EtOAc (15 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (40-50% EtOAc in hexane) gave the title compound **21** (149 mg, 77% from **19**) as a white solid. $[\alpha]_D^{19}$ –18.5 (*c* 0.30, CHCl₃). IR (film) ν_{max} 1755, 1610, 1506, 1221, 1041, 737 m⁻¹. ¹H NMR $(100 \text{ MHz}, \text{CDCl}_3) \delta$ 7.38 (10H, m, ArH), 6.99 (1H, d, J = 8.3 Hz, ArH),6.86 (1H, d, J = 8.3 Hz, ArH), 6.68 (1H, d, J = 2.4 Hz, ArH), 6.57 (1H, d, J = 2.4 Hz, ArH), 6.54 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.50 (1H, dd, J = 2.4, 8.3 Hz, ArH), 5.15 (6H, m, CH and CH₂), 5.02 (4H, m, CH), 4.99 (2H, m, CH), 4.20 (1H, dd, J = 4.7, 12.1 Hz, CH₂), 4.16 (1H, dd, J = 4.7, 12.1 Hz, CH₂), 3.50 (1H, dd, J = 7.8, 11.7 Hz, CH₂), 3.47 (1H, dd, J = 7.8, 11.7 Hz, CH₂), 2.80 (4H, m, CH₂), 2.04 (18H, m, CH₃). ^{13}C NMR (100 MHz. CDCl₃) δ 170.0 (C), 169.88 (C), 169.86 (C), 169.8 (C), 169.4 (C), 169.3 (C), 158.2 (C), 157.3 (C), 155.5 (C), 154.8 (C), 137.2 (C), 137.0 (C), 130.6 (CH), 130.2 (CH), 128.53 (CH), 128.48 (CH), 127.9 (CH), 127.8 (CH), 127.5 (CH), 127.3 (CH), 126.5 (C), 123.1 (C), 110.4 (CH), 105.2 (CH), 104.6 (CH), 100.3 (CH), 98.5 (CH), 97.9 (CH), 70.7 (CH), 70.2 (CH₂), 70.1 (CH₂), 69.8 (CH), 69.7 (CH), 69.1 (CH), 68.5 (CH₂), 68.2 (CH₂), 61.8 (CH₂), 61.4 (CH₂), 30.1 (CH₂), 29.9 (CH₂), 20.8 (CH₃). ESIHRMS *m*/*z* 965.3307 [M + Na]⁺ (calcd for C₅₀H₅₄NaO₁₈, 965.3208).

4.1.13. 2', 4'-Dihydroxy-2, 4-di(β -D-xylopyranosyl)bibenzyl (**3**)

Bibenzyl **21** (19 mg, 20 μ mol) in EtOAc (2 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (3 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (70–80% EtOAc in hexane) gave the phenol as a white solid, which was used in the next step without further purification.

To cold $(0 \,^{\circ}C)$ and stirred solution of the phenol in MeOH (4 mL) was added 100 µL of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H₂O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (15% MeCN in H₂O, $t_{\rm R}$ = 12.5 min). The title compound **3** (8.7 mg, 85%) was obtained as a colorless solid. $[\alpha]_{D}^{20}$ -48.0 (*c* 0.05, MeOH). IR (Nujol) ν_{max} 3350, 1508 cm⁻¹. ¹H NMR (pyridine- d_5 , 400 MHz) δ 7.49 (1H, bs, ArH), 7.31 (1H, d, J = 8.3 Hz, ArH), 7.28 (1H, d, J = 8.3 Hz, ArH), 7.04 (1H, d, J = 8.3 Hz, ArH), 7.02 (1H, bs, ArH), 6.72 (1H, d, *J* = 8.3 Hz, ArH), 5.52 (1H, d, *J* = 5.4 Hz, CH), 5.38 (1H, d, J = 7.3 Hz, CH), 4.28 (8H, m, CH and CH₂), 3.85 (1H, t, *I* = 9.8 Hz, CH₂), 3.67 (1H, dd, *I* = 8.8, 11.2 Hz, CH₂), 3.43 (2H, m, CH₂), 3.30 (2H, m, CH₂). ¹³C NMR (pyridine- d_5 , 100 MHz) δ 157.6 (C), 157.2 (C), 156.8 (C), 156.7 (C), 130.8 (CH), 130.1 (CH), 126.0 (C), 119.9 (C), 109.6 (CH), 106.7 (CH), 105.3 (CH), 103.2 (CH), 103.1 (CH), 102.7 (CH), 77.8 (CH), 77.6 (CH), 74.2 (CH), 70.3 (CH), 70.2 (CH), 66.6 (CH₂), 66.5 (CH₂), 30.90 (CH₂), 30.86 (CH₂). HRESIMS m/z 509.1649 $[M - H]^{-}$ (calcd for C₂₄H₂₉O₁₂, 509.1659).

4.1.14. 4-(2",3",4",6"-Tetra-O-acetyl-β-D-glucopyranosyl)-2',4'-dibenzyloxy-2-hydroxybibenzyl (**25**)

To a cold (0 °C) and stirred solution of bibenzyl 19 (50 mg, 0.12 mmol) and 2,3,4,6-tetra-O-acetyl-α-D-glucopyranosyl trichloroacetimidate (22) (140 mg, 0.30 mmol) in CH₂Cl₂ (2 mL) was added 10 µL of TMSOTf solution (0.11 M in CH₂Cl₂). After being stirred for 5 min at 0 °C, the resultant solution was poured into EtOAc (100 mL) and washed with saturated aqueous NaHCO₃ solution $(20 \text{ mL} \times 3)$ and brine $(20 \text{ mL} \times 3)$. The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (50-55% EtOAc in hexane) gave the title compound **25** (66 mg, 74% from **19**) as a white solid. $[\alpha]_D^{23}$ – 4.48 (*c* 1.46, CHCl₃). IR (film) v_{max} 3033, 1750, 1503, 1220, 1040, 736 m⁻¹. ¹H NMR (400 MHz, $CDCl_3$) δ 7.39 (10H, m, ArH), 7.02 (1H, d, I = 7.8 Hz, ArH), 6.95 (1H, d, I = 7.8 Hz, ArH), 6.62 (1H, d, I = 2.4 Hz, ArH), 6.50 (1H, dd, I = 2.4, 7.8 Hz, ArH), 6.47 (1H, d, J = 2.4 Hz, ArH), 6.45 (1H, dd, J = 2.4, 7.8 Hz, ArH), 5.57 (1H, bs, OH), 5.27 (1H, t, J = 8.0 Hz, CH), 5.23 (1H, t, J = 8.0 Hz, CH), 5.13 (1H, m, CH), 5.11 (2H, s, CH₂), 5.00 (2H, s, CH₂), 4.99 (1H, d, J = 6.8 Hz, CH), 4.27 (1H, dd, J = 5.4, 12.2 Hz, CH₂), 4.15 (1H, dd, J = 2.4, 12.2 Hz, CH₂), 3.82 (1H, ddd, *J* = 2.4, 5.4, 8.0 Hz, CH), 2.75 (4H, m, CH₂), 2.07 (3H, s, CH₃), 2.043 (3H, s, CH₃), 2.037 (3H, s, CH₃), 2.02 (3H, s, CH₃). ^{13}C NMR (100 MHz, CDCl₃) δ 171.0 (C), 170.6 (C), 169.73 (C), 169.65 (C), 158.9 (C), 157.3 (C), 156.8 (C), 155.3 (C), 137.2 (C), 136.8 (C), 130.7 (CH), 130.6 (CH), 129.0 (CH), 128.9 (CH), 128.5 (CH), 128.3 (CH), 127.84 (CH), 127.76 (CH), 123.0 (C), 122.8 (C), 109.2 (CH), 106.1 (CH), 105.1 (CH), 101.3 (CH), 99.6 (CH), 73.1 (CH), 72.3 (CH), 71.5 (CH), 70.6 (CH₂), 70.5 (CH₂), 68.7 (CH), 62.3 (CH₂), 31.6 (CH₂), 31.2 (CH₂), 20.9 (CH₃). ESIHRMS m/z 755.2700 [M – H]⁻ (calcd for C₄₂H₄₃O₁₃, 755.2704).

4.1.15. $4-(\beta-D-Glucopyranosyl)-2,2',4'-trihydroxybibenzyl$ (**4**)

Bibenzyl **25** (35 mg, 46 µmol) in EtOAc (2 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (24 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (60–70% EtOAc in hexane) gave the phenol as a white solid, which was used in the next step without further purification.

To cold $(0 \circ C)$ and stirred solution of the phenol in MeOH (5 mL) was added 43 uL of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H₂O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (20% MeCN in H₂O, $t_{\rm R}$ = 6.8 min). The title compound **4** (12 mg, 63%) was obtained as a colorless solid. $[\alpha]_D^{21}$ -33.4 (*c* 0.2, MeOH). IR (Nujol) ν_{max} 3522, 3350, 1595, 1510 cm⁻¹. ¹H NMR (CD₃OD, 400 MHz) δ 6.89 (1H, d, J = 8.3 Hz, ArH), 6.77 (1H, d, *J* = 8.3 Hz, ArH), 6.56 (1H, d, *J* = 2.4 Hz, ArH), 6.47 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 6.27 (1H, d, J = 2.4 Hz, ArH), 6.16 (1H, dd, J = 2.4, 8.3 Hz, ArH), 4.82 (1H, d, J = 7.3 Hz, CH), 3.89 (1H, dd, J = 2.0, 12.2 Hz, CH₂), 3.70 (1H, dd, *J* = 4.9, 12.2 Hz, CH₂), 3.39 (4H, m, CH), 2.70 (4H, m, CH₂). ¹³C NMR (CD₃OD, 100 MHz) δ 158.2 (C), 157.3 (C), 157.0 (C), 156.9 (C), 131.44 (CH), 131.39 (CH), 124.3 (C), 121.2 (C), 108.5 (CH), 107.3 (CH), 104.9 (CH), 103.4 (CH), 102.4 (CH), 78.1 (CH), 74.9 (CH), 71.4 (CH), 62.6 (CH₂), 31.4 (CH₂), 31.2 (CH₂). HRESIMS m/z 431.1376 $[M + Na]^+$ (calcd for C₂₀H₂₄NaO₉, 431.1318).

4.1.16. 4-(2",2"',3",3",4",4"',6",6""-Octa-O-acetyl-β-*D*-cellobiopyranosyl)-2',4'-dibenzyloxy-2-hydroxybibenzyl (**26**)

To a cold (0 °C) and stirred solution of bibenzyl **19** (50 mg. 0.12 mmol) and 2,2',3,3',4,4',6,6'-octa-O-acetyl-α-D-cellobiopyranosyl trichloroacetimidate (23) (185 mg, 0.24 mmol) in CH₂Cl₂ (2 mL) was added 10 µL of TMSOTf solution (0.11 M in CH₂Cl₂). After being stirred for 5 min at 0 °C, the resultant solution was poured into EtOAc (100 mL) and washed with saturated aqueous NaHCO₃ solution (20 mL \times 3) and brine (20 mL \times 3). The aqueous layers were extracted with EtOAc (20 mL \times 3), and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (70% EtOAc in hexane) gave the title compound **26** (92 mg, 75% from **19**) as a white solid. $[\alpha]_D^{22}$ -14.56 (c 1.40, CHCl₃). IR (film) v_{max} 3035, 1750, 1503, 1220, 1040, 734 m⁻¹.¹H NMR (400 MHz, CDCl₃) δ 7.38 (10H, m, ArH), 7.01 (1H, d, *J* = 8.3 Hz, ArH), 6.92 (1H, d, *J* = 8.3 Hz, ArH), 6.60 (1H, d, *J* = 2.4 Hz, ArH), 6.48 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 6.44 (1H, d, *J* = 2.4 Hz, ArH), 6.42 (1H, dd, J = 2.4, 8.3 Hz, ArH), 5.64 (1H, bs, OH), 5.22 (1H, t, J = 9.3 Hz, CH), 5.11 (3H, m, CH), 5.09 (2H, s, CH₂), 5.05 (1H, t, J = 9.8 Hz, CH), 4.99 (2H, s, CH₂), 4.93 (2H, m, CH), 4.49 (2H, m, CH), 4.36 (1H, dd, J = 4.4, 12.7 Hz, CH₂), 4.12 (1H, dd, J = 5.9, 12.7 Hz, CH₂), 4.04 (1H, dd, J = 2.0, 12.7 Hz, CH₂), 3.82 (1H, m, CH), 3.69 (2H, m, CH and CH₂), 2.73 (4H, m, CH₂), 2.02 (21H, m, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 170.5 (C), 170.4 (C), 170.2 (C), 169.8 (C), 169.6 (C), 169.3 (C), 169.1 (C), 158.5 (C), 157.0 (C), 156.4 (C), 155.0 (C), 136.9 (C), 136.5 (C), 130.4 (CH), 130.2 (CH), 128.7 (CH), 128.6 (CH), 128.1 (CH), 128.0 (CH), 127.5 (CH), 127.4 (CH), 122.7 (C), 122.5 (C), 108.8 (CH), 105.7 (CH), 104.7 (CH), 100.9 (CH), 100.8 (CH), 99.0 (CH), 72.9 (CH), 72.8 (CH), 72.5 (CH), 72.0 (CH), 71.6 (CH), 71.4 (CH), 70.3 (CH₂), 70.2 (CH₂), 67.7 (CH), 61.9 (CH₂), 61.5 (CH₂), 31.2 (CH₂), 30.8 (CH₂), 20.7 (CH₃), 20.6 (CH₃), 20.5 (CH₃). ESIHRMS m/z 1043.3528 [M - H]⁻ (calcd for C₅₄H₅₉O₂₁, 1043.3549).

4.1.17. $4-(\beta$ -D-Cellobiopyranosyl)-2,2',4'-trihydroxybibenzyl (5)

Bibenzyl **26** (46 mg, 44 μ mol) in EtOAc (2 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (24 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (80–85% EtOAc in hexane) gave the phenol as a white solid, which was used in the next step without further purification.

To cold $(0 \,^{\circ}C)$ and stirred solution of the phenol in MeOH (5 mL) was added 52 µL of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H₂O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (20% MeCN in H₂O, $t_{\rm R}$ = 6.4 min). The title compound **5** (18 mg, 72%) was obtained as a colorless solid. $[\alpha]_D^{21}$ -66.0 (*c* 0.04, MeOH). IR (Nujol) *v*_{max} 3520, 3350, 1595, 1510 cm⁻ ¹H NMR (CD₃OD, 400 MHz) δ 6.89 (1H, d, I = 8.3 Hz, ArH), 6.77 (1H, d, J = 8.3 Hz, ArH), 6.56 (1H, d, J = 2.4 Hz, ArH), 6.47 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.27 (1H, d, *J* = 2.4 Hz, ArH), 6.17 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 4.93 (1H, d, *J* = 7.2 Hz, CH), 4.44 (1H, d, *J* = 7.8 Hz, CH), 3.91 (3H, m, CH and CH₂), 3.66 (2H, m, CH₂), 3.61 (1H, dd, J = 8.3, 8.8 Hz, CH), 3.54 (1H, m, CH), 3.48 (1H, dd, *J* = 7.8, 8.3 Hz, CH), 3.32 (3H, m, CH), 3.24 (1H, t, J = 8.3 Hz, CH), 2.70 (4H, m, CH₂). ¹³C NMR (CD₃OD, 100 MHz) & 158.1 (C), 157.3 (C), 156.93 (C), 156.88 (C), 131.44 (CH), 131.39 (CH), 124.4 (C), 121.2 (C), 108.5 (CH), 107.3 (CH), 104.9 (CH), 104.6 (CH), 103.4 (CH), 102.2 (CH), 80.3 (CH), 78.1 (CH), 77.9 (CH), 76.6 (CH), 76.4 (CH), 74.9 (CH), 74.7 (CH), 71.4 (CH), 62.4 (CH₂), 61.7 (CH₂), 31.3 (CH₂), 31.2 (CH₂). HRESIMS *m*/*z* 569.1876 $[M - H]^{-}$ (calcd for C₂₆H₃₃O₁₄, 569.1870).

4.1.18. 4-(2",2"',3",3"',4",4"',6",6"'-octa-O-acetyl-β-Dmaltopyranosyl)-2',4'-dibenzyloxy-2-hydroxybibenzyl (**27**)

To a cold (0 °C) and stirred solution of bibenzyl 19 (64 mg, 0.15 mmol) and 2,2',3,3',4,4',6,6'-octa-O-acetyl- α -D-maltopyranosyl trichloroacetimidate (24) (218 mg, 0.28 mmol) in CH₂Cl₂ (2 mL) was added 14 uL of TMSOTf solution (0.11 M in CH₂Cl₂). After being stirred for 1 h at 0 °C, the resultant solution was poured into EtOAc (100 mL) and washed with saturated aqueous NaHCO₃ solution ($20 \text{ mL} \times 3$) and brine (20 mL \times 3). The aqueous layers were extracted with EtOAc $(20 \text{ mL} \times 3)$, and the combined organic layers were dried over Na₂SO₄. Filtration and concentration followed by silica gel chromatography (60% EtOAc in hexane) gave the title compound 27 (89 mg, 57% from **19**) as a white solid. $[\alpha]_{D}^{22}$ +42.38 (*c* 0.68, CHCl₃). IR (film) ν_{max} 3040, 1735, 1503, 1220, 1040, 735 m⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.39 (10H, m, ArH), 7.04 (1H, d, J = 8.3 Hz, ArH), 6.96 (1H, d, J = 8.3 Hz, ArH), 6.62 (1H, d, J = 2.4 Hz, ArH), 6.51 (1H, dd, J = 2.4, 8.3 Hz, ArH), 6.47 (1H, d, J = 2.4 Hz, ArH), 6.44 (1H, dd, J = 2.4, 8.3 Hz, ArH), 5.69 (1H, bs, OH), 5.43 (1H, d, J = 3.9 Hz, CH), 5.37 (1H, t, J = 9.8 Hz, CH), 5.31 (1H, t, J = 8.8 Hz, CH), 5.12 (2H, s, CH₂), 5.05 (3H, m, CH), 5.01 (2H, s, CH₂), 4.87 (1H, dd, J = 3.9, 10.8 Hz, CH₂), 4.47 (1H, dd, J = 2.4, 11.7 Hz, CH₂), 4.26 (2H, m, CH and CH₂), 4.07 (2H, m, CH and CH₂), 3.97 (1H, m, CH), 3.84 (1H, m, CH), 2.76 (4H, m, CH₂), 2.06 (21H, m, CH₃). ¹³C NMR (100 MHz, CDCl₃) § 170.6 (C), 170.5 (C), 170.2 (C), 170.0 (C), 169.7 (C), 169.5 (C), 158.5 (C), 157.0 (C), 156.2 (C), 155.0 (C), 136.9 (C), 136.5 (C), 130.4 (CH), 130.2 (CH), 128.7 (CH), 128.6 (CH), 128.1 (CH), 128.0 (CH), 127.53 (CH), 127.46 (CH), 122.7 (C), 122.5 (C), 108.8 (CH), 105.7 (CH), 104.8 (CH), 100.9 (CH), 98.6 (CH), 95.6 (CH), 75.3 (CH), 72.7 (CH), 72.2 (CH), 72.0 (CH), 70.3 (CH₂), 70.2 (CH₂), 70.0 (CH), 69.3 (CH), 68.5 (CH), 68.0 (CH), 62.8 (CH₂), 61.6 (CH₂), 31.3 (CH₂), 30.8 (CH₂), 20.9 (CH₃), 20.73 (CH₃), 20.68 (CH₃), 20.62 (CH₃), 20.61 (CH₃), 20.58 (CH₃). ESIHRMS m/z $1067.3465 [M + Na]^+$ (calcd for C₅₄H₆₀NaO₂₁, 1067.3525).

4.1.19. 2,2',4'-Trihydroxy-4-(β -D-maltopyranosyl)bibenzyl (**6**)

Bibenzyl **27** (49 mg, 46 μ mol) in EtOAc (2 mL) was hydrogenated over 20% Pd(OH)₂ on carbon (24 mg) for 12 h. Filtration through Celite 535 and concentration in vacuo followed by silica gel chromatography (70–80% EtOAc in hexane) gave the phenol as a white solid, which was used in the next step without further purification.

To cold (0 °C) and stirred solution of the phenol in MeOH (5 mL) was added 28 μ L of NaOMe (5.2 M in MeOH) slowly. After being stirred for 0.5 h at 0 °C, the resultant solution was neutralized with Amberlite IR-120 (H⁺). Filtration and concentration followed by

solid phase extraction with Sep-Pak Plus C-18 cartridge (0-40% MeCN in H₂O) gave the crude bibenzyl that was further purified by preparative RP-HPLC (17.5% MeCN in H₂O, $t_{\rm R}$ = 6.0 min). The title compound **6** (15 mg, 56%) was obtained as a colorless solid. $[\alpha]_D^{21}$ +21.7 (*c* 0.09, MeOH). IR (Nujol) *v*_{max} 3520, 3350, 1600, 1510 cm⁻¹ ¹H NMR (CD₃OD, 400 MHz) δ 6.88 (1H, d, I = 8.3 Hz, ArH), 6.76 (1H, d, *J* = 8.3 Hz, ArH), 6.55 (1H, d, *J* = 2.4 Hz, ArH), 6.46 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 6.26 (1H, d, *J* = 2.4 Hz, ArH), 6.15 (1H, dd, *J* = 2.4, 8.3 Hz, ArH), 5.19 (1H, d, *J* = 3.9 Hz, CH), 4.84 (1H, d, *J* = 7.8 Hz, CH), 3.87 (3H, m, CH and CH₂), 3.66 (5H, m, CH and CH₂), 3.48 (4H, m, CH), 2.69 (4H, m, CH₂). ¹³C NMR (CD₃OD, 100 MHz) δ 158.1 (C), 157.3 (C), 157.0 (C), 156.9 (C), 131.44 (CH), 131.38 (CH), 124.3 (C), 121.2 (C), 108.4 (CH), 107.2 (CH), 104.9 (CH), 103.4 (CH), 102.9 (CH), 102.3 (CH), 80.9 (CH), 77.8 (CH), 76.7 (CH), 75.1 (CH), 74.8 (CH), 74.5 (CH), 74.4 (CH), 71.5 (CH), 62.8 (CH₂), 62.0 (CH₂), 31.78 (CH₂), 31.68 (CH₂). HRESIMS m/z 569.1801 [M – H]⁻ (calcd for C₂₆H₃₃O₁₄, 569.1870).

4.2. Tyrosinase inhibitory assay

Tyrosinase (E.C.1.14.18.1) purchased from Sigma-Aldrich was purified by the procedure as previously reported [28]. The enzyme concentration was adjusted at 1.0 µg/mL by the Bradford method with bovine serum albumin as a standard [29]. All inhibitors were first dissolved in DMSO and used for the experiment by appropriate dilution with DMSO. First, 0.3 mL of a 5.0 mM of L-DOPA aqueous solution was mixed with 0.6 mL of 0.25 M sodium phosphate buffer (pH 6.8) and 1.9 mL of water, incubated at 30 °C for 10 min. Then, 0.1 mL of the DMSO solution of the inhibitor and 0.1 mL of enzyme solution were added, in this order, to the mixture. This solution was immediately monitored for the formation of dopachrome by measuring the linear increase in optical density at 475 nm. The reaction was carried out under a constant temperature of 30 °C. Absorption measurements were recorded using a Jasco V-630 spectrophotometer. The experimental data were analyzed by using Sigma Plot 9.0 to estimate IC₅₀ values.

Acknowledgments

We are grateful to Dr. Michinori Karikomi (Utsunomiya University) for assistance to obtain optical rotations and, Drs. Tadashi Yanagisawa and Masayuki ligo for their invaluable discussion. This study was supported in part by a Grant of Utsunomiya University Exploratory Research for Young Scientists and a Grant-in-Aid of Research for Promoting Technological Seeds from Japan Science and Technology Agency (JST).

References

- [1] E.I. Solomon, P. Chen, M. Metz, S. Lee, A.E. Palmer, Angew. Chem. Int. Ed. 40 (2001) 4570 - 4590.
- T.S. Chang, Int. J. Mol. Sci. 10 (2009) 2440-2475.
- R. Halaban, R.S. Patton, E. Cheng, S. Svedine, E.S. Trombetta, M.L. Wahl, [3] S. Ariyan, D.N. Hebert, J. Biol. Chem. 277 (2002) 14821–14828.
- [4] A. De Iuliis, G. Arrigoni, L. Anderson, A. Zambenedetti, A. Burlina, P. James, P. Arslan, F. Vianello, Biochim. Biophys. Acta 1784 (2008) 1687-1693.
- [5] M. Asanuma, I. Miyazaki, N. Ogawa, Neurotox. Res. 5 (2003) 165-176.
- S. Parvez, M. Kang, H.S. Chung, H. Bae, Phytother. Res. 21 (2007) 805-816. [6]
- S.Y. Seo, V. Sharma, N. Sharma, J. Agric. Food Chem. 51 (2003) 2837-2853.
- [8] K. Shimizu, R. Kondo, K. Sakai, Planta Med. 66 (2000) 11-15.
- [9] N. Jun, G. Hong, K. Jun, Bioorg. Med. Chem. 15 (2007) 2396-2402.
- [10] R. Iyengar, C. Boumont, A. McEvily, J. Food Compos. Anal. 4 (1991) 148-157. V.H. Frankos, D.F. Schmitt, L.C. Haws, A. McEvily, R. Iyengar, S.A. Miller, Regul. Toxicol. Pharmacol. 14 (1991) 202–212. [11]
- [12] M. Tandon, Y.N. Shukla, Phytochemistry 32 (1993) 1624-1625.
- [13] H. Oozeki, R. Tajima, K. Nihei, Bioorg. Med. Chem. 18 (2008) 5252-5254.
- [14] H. Kogen, N. Toda, K. Tago, S. Marumoto, K. Takami, M. Ori, N. Yamada, K. Koyama, S. Naruto, K. Abe, R. Yamazaki, T. Hara, A. Aoyagi, Y. Abe, T. Kaneko, Org. Lett. 4 (2002) 3359-3362.
- M. Matsushita, T. Kanemura, S. Hatakeyama, H. Irie, T. Toki, M. Miyashita, [15] Tetrahedron 51 (1995) 10687-10698.
- [16] H. Sajili, K. Hattori, K. Hirota, J. Org. Chem. 63 (1998) 7990-7992.
- Y. Asakawa, Phytochemistry 56 (2001) 297-312. [17]
- [18] S.D. Lorimer, N.B. Perry, J. Nat. Prod. 56 (1993) 1444-1450.
- [19] Y. Hernández-Romero, L. Acevedo, M.L.Á. Sánchez, W.T. Shier, H.K. Abbas, R. Mata, J. Agric. Food Chem. 53 (2005) 6276-6280.
- [20] K.P. Manfredi, V. Vallurupalli, M. Demidova, K. Kindscher, L.K. Pannell, Phytochemistry 58 (2001) 153-157.
- E.R. Silcoff, T. Sheradsky, New J. Chem. 23 (1999) 1187-1192. [21]
- [22] K. Nihei, I. Kubo, Bioorg. Med. Chem. Lett. 13 (2003) 2409-2412.
- [23] I. Kubo, K. Nihei, K. Shimizu, Bioorg. Med. Chem. 12 (2004) 5343-5347.
- I. Kubo, K. Nihei, K. Tsujimoto, Bioorg. Med. Chem. 12 (2004) 5349-5354. [24]
- [25] I. Kubo, I. Kinst-Hori, J. Agric. Food Chem. 46 (1998) 5338-5341
- [26] T. Matsuda, Y. Odaka, N. Ogawa, K. Nakamoto, H. Kuninaga, J. Agric. Food
- Chem. 56 (2008) 597-601. [27] S. Khatib, O. Nerya, R. Musa, S. Tamir, T. Peter, J. Vaya, J. Med. Chem. 50 (2007) 2676-2681.
- [28] J.C. Espín, H.J. Wichers, J. Agric. Food Chem. 47 (1999) 2638-2644.
- [29] M.A. Bradford, Anal. Biochem. 72 (1976) 248-254.