

L-Ribonucleosides from L-Xylose

Elisabeth Moyroud and Peter Strazewski*

Institute of Organic Chemistry, University of Basel, St. Johannis-Ring 19, CH-4056 Basel, Switzerland.

Received 22 October 1998; accepted 30 November 1998

Abstract: L-Xylose was converted into a L-ribose derivative *via* an oxidation/reduction procedure. The L-ribosyl donor was submitted to a glycosidation reaction according to Vorbrüggen's conditions to give L-ribonucleosides (L-uridine, L-cytidine, L-adenosine and L-guanosine) in high yield. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: L-ribonucleosides; glycosidation; L-xylose; L-ribose

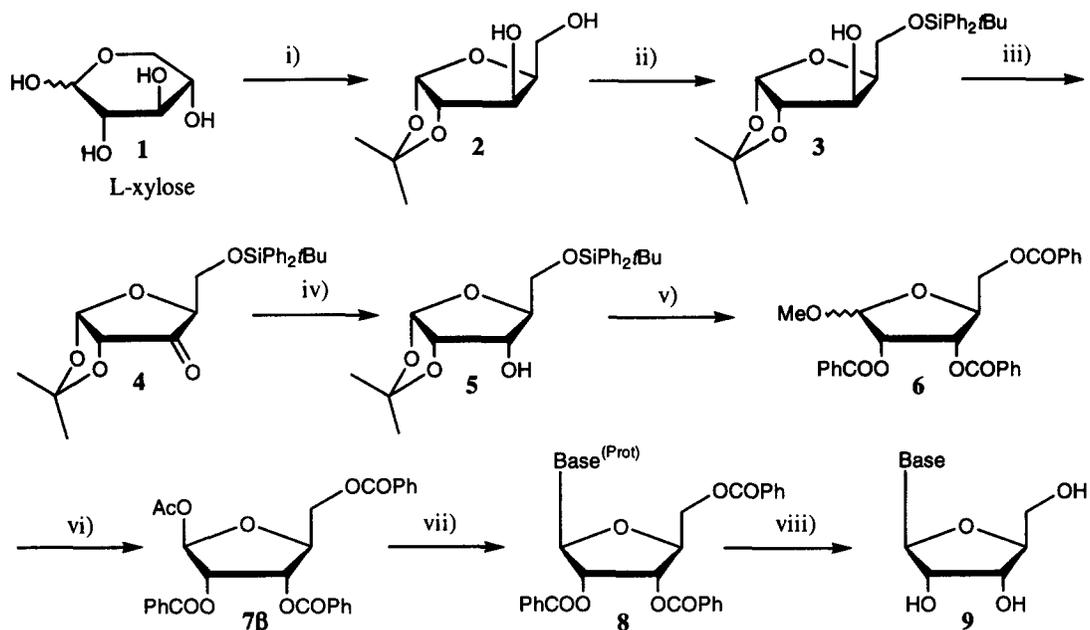
Introduction

The use of L-carbohydrates and the corresponding nucleosides in medicinal applications has greatly increased. In particular several modified nucleosides derived from L-sugars have shown a high potential as useful antiviral agents.¹ Also, due to the stereoselectivity of enzymes, L-ribose-modified oligoribonucleotides become attractive candidates for diagnostic and therapeutic uses because L-RNA ligands remain uncleaved in biological fluids.² The preparation of L-ribonucleosides requires the availability of suitably protected L-ribofuranosides which can be coupled with purine or pyrimidine bases. The mirror-image nucleosides thus obtained can be further processed to give properly protected L-RNA building blocks. The conversion of L-arabinose,²⁻⁵ D-glucose⁶ or D-ribose⁷ into L-ribose have been described. In our synthetic approach the key step was the conversion of L-xylose (**1**) into a ribo-configured derivative.

Results and discussion

We proceeded from commercially available L-xylose (**1**, Fig. 1). The first step was the acetonation of the sugar to form **2** with an overall yield of 80% after treatment with 0.12 M HCl solution for 30 min.⁸ Compound **2** was then silylated with *tert*-butyldiphenylsilyl chloride at the primary hydroxyl group under standard conditions to give **3** as colorless crystals in 97% yield after crystallisation from petrol ether. The inversion of configuration at C(3) was accomplished *via* an oxidation/reduction procedure. Ketone **4** was obtained by treatment with CrO₃/pyr complex in CH₂Cl₂ in 97% yield. The reduction step was performed with LiAlH₄ at -78 °C for 1 hour and led to **5** in 96% yield. The L-ribosyl derivative **5** was deprotected and O-methylated at C(1) in one pot followed by O-benzoylation to obtain **6** in 86% yield (α - and β -anomer) which could be used for glycosidation.⁹

Figure 1



8a: Base : uracil: CH₃CN, o/n, 60-65°C: **89%**

8b: Base^(prot) : 4-N-acetylcytosine: CH₃CN, o/n, 60-65°C: **86%**

8c: Base^(prot) : 6-N-benzoyladenine: CH₂ClCH₂Cl, o/n, 80°C: **84%**

8d: Base^(prot) : 2-N-acetyl-6-O-diphenylcarbamoylguanine: toluene, 1h, 80°C: **70%**

9a: L-uridine: **95%**

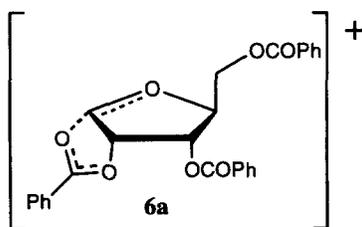
9b: L-cytidine: **89%**

9c: L-adenosine: **92%**

9d: L-guanosine: **90%**

i) Acetone, H₂SO₄: **80%**; ii) *t*BuPh₂SiCl, imidazole, DMF: **97%**; iii) CrO₃, pyr, CH₂Cl₂: **97%**; iv) LiAlH₄, CH₂Cl₂: **96%**; v) MeOH, H₂SO₄; BzCl, pyr: **86%**; vi) Ac₂O, AcOH, H₂SO₄: **77%** (α : β 1:1); vii) B^{TMS}, TMSOTf, solvents; viii) NaOMe/MeOH or NH₃/H₂O.

However, according to Vorbrüggen¹⁰ the best leaving group for the formation of intermediate **6a** is acetyl at C(1) and in β -position. The conversion of **6** led to the standard ribosyl donor 1-O-acetyl-2,3,4-tri-O-benzoyl-L-ribofuranoside^{5,11} (**7**) as an α / β mixture (1:1). The pure β -anomer was obtained *via* crystallisation from isopropyl alcohol.



We tried to perform the isomerisation of the α - to the β -anomer by heating **7** (mainly α -anomer) at 50 °C under acidic conditions for several hours but we observed only decom-

position of the starting material. To increase the overall yield, a regeneration of **6** (α - and β -anomer) from **7** (α -anomer) was feasible *via* total deprotection under basic conditions (0.1 M NaOMe in MeOH), followed by the same reaction sequence *v*) as before. We observed reaction yields (**7a** to **6a/b**) between 55% and 87% (α + β). We obtained L-ribosyl donor **7b** in an overall yield of 24% starting from L-xylose (**1**), regeneration of **7a** to **6a/b** not included. The introduction of the β -N-glycosidic bond was realised through the well-established procedure reported by Vorbrüggen *et al.*¹² Using the standard protected bases (uracil, 4-N-acetylcytosine and 6-N-benzoyladenine), we obtained **8(a-c)** in good yields (89%, 86%, 84% for **8a**, **8b**, **8c**, respectively). Only in the case of glycosidation with 6-N-benzoyladenine we observed traces of the undesired N7 isomer which we separated by column chromatography. Glycosidation with guanine-type heterocycles produces N7/N9 isomeric mixtures¹³ that are frequently difficult to separate. We applied to our L-ribosyl donor **7** the glycosidation conditions using 2-N-acetyl-6-O-diphenylcarbonylguanidine developed by Robins *et al.*,¹³ a convenient route to guanine N9 derivatives. Compound **8d** was obtained as a pure N9 isomer in high yield (70%), no traces of N7 product were observed. The four protected L-ribonucleosides **8(a-d)** were deprotected with NH₃/H₂O/MeOH at 60°C or NaOMe (0.1 M in MeOH) at room temperature overnight to furnish compounds **9(a-d)** in high yields (89%–95%).

Experimental

¹H NMR spectra were obtained at 300 MHz on a VARIAN Gemini 300 spectrometer using tetramethylsilane as an internal standard. ¹³C NMR spectra were obtained at 75 MHz using the same internal standard. Mass spectra were obtained on a MAT 312 mass spectrometer using Fast Atomic Bombardment (FAB) ionisation method (in *p*-nitrobenzyl alcohol, if not stated otherwise) and positive ion detection. $[\alpha]$ values were obtained on a Perkin-Elmer 141 polarimeter. Melting points were determined by visual observation on a Kofler block and are corrected. TLC was performed on a pre-coated silica gel F₂₅₄ plates with fluorescent indicator. Glycosides and nucleosides were visualised on tlc plates by subsequent spraying with naphthoresorcin and conc. H₂SO₄ solutions in ethanol, respectively, followed by heating. Column chromatography was performed with flash silica gel (35–70 μ m) from Uetikon. Dry pyridine was obtained by distillation over CaH₂. All other solvent were used as purchased. Abbreviations: PE: petrol ether, TBME: *tert*-butylmethyl ether, DMAP: 4-dimethylamino-pyridine, BSA: N,O-bis-trimethylsilylacetamide, TMSOTf: trimethylsilyltrifluoromethanesulfonate, MFSTA: N-methyl-N-trimethylsilyltrifluoroacetamide, TBDPSCl: *tert*-butyldiphenylsilyl chloride.

5-O-*tert*-Butyldiphenylsilyl-1,2-O-isopropylidene- α -L-xylofuranose (3). To a magnetically stirred solution of 1,2-O-isopropylidene-L-xylofuranose⁸ (**2**) (5.0 g, 26.29 mmol) and imidazole (7.16 g, 105.16 mmol) in DMF (40 ml) at 0°C, was added dropwise TBDPSCl (7.20 ml, 28.13 mmol) over 10 min. The mixture was allowed to reach room temperature and stirred for 1 hour. TBME (100 ml) and water (15 ml) were added to the mixture, and the organic layer was separated and washed with water (20 ml \times 2) and brine (20 ml). After drying over Na₂SO₄ and evaporation, crystallisation from PE gave pure **3** (9.02 g, 21.05 mmol, 97%) as

colorless crystals. R_f (PE/TBME 6/4): 0.60; mp: 92-94°C; $[\alpha]_D^{25} = +3.7 \pm 0.5$ ($c = 1.65$, CHCl_3). $^1\text{H NMR}$ (CDCl_3): 1.06 (s, 9H, *t*Bu); 1.33 (s, 3H, CH_3), 1.47 (s, 3H, CH_3); 4.04 (s, 1H, HOC(3)); 4.14 (m, 2H, $\text{H}_2\text{C}(5)$); 4.20 (m, 1H, HC(4)); 4.39 (s, 1H, HC(3)); 4.57 (d, 1H, $^3J = 3.6$, HC(2)); 6.03 (d, 1H, $^3J = 3.8$, HC(1)); 7.40-7.79 (m, 10H, Ph). $^{13}\text{C NMR}$ (CDCl_3): 19.12 ($\underline{\text{C}}(\text{CH}_3)_3$); 26.22, 26.84 ($\underline{\text{C}}(\text{H}_3)_2\text{C}$); 26.74 ($\underline{\text{C}}(\text{H}_3)_3\text{C}$); 62.83 (C(5)); 76.90 (C(4)); 78.48 (C(3)); 85.50 (C(2)); 105.05 (C(1)); 111.55 ($\underline{\text{C}}(\text{CH}_3)_2$); 127.96-135.74 (Ph). **MS(FAB)**: 4.67 ($[\text{M}+\text{K}]^+$, 19.0%); 429 ($[\text{M}+\text{H}]^+$, 1.8%); 173 ($[\text{M}-\text{OSiPh}_2\text{tBu}]^+$, 20.6%); 135 (100%)

(2S,3R,5R)-5-O-(tert-Butyldiphenylsilyl)oxymethyl-2,3-O-isopropylidene-2,3-dioxytetrahydrofuran-4-one (4). CrO_3 (6.91 g, 69.06 mmol) was added to dry pyridine (11.11 ml, 138.13 mmol) in dichloromethane (120 ml) and the resulting mixture was stirred for 15 min at 20°C. A solution of **3** (7.40 g, 17.26 mmol) in CH_2Cl_2 (40 ml) was added, immediately followed by Ac_2O (6.53 ml, 69.06 mmol). After stirring for 30 min, EtOAc (70 ml) and toluene (70 ml) were added. The mixture was decanted and filtered through a short SiO_2 column. The eluate was evaporated and co-evaporated with toluene to give **4** (7.14 g, 16.74 mmol, 97%) as a yellowish oil. R_f (PE/TBME 6/4): 0.82. $^1\text{H NMR}$ (CDCl_3): 1.07 (s, 9H, *t*Bu); 1.48 (s, 3H, CH_3); 1.49 (s, 3H, CH_3); 3.87 (dd, 1H, $^2J_{5a,5b} = 11.0$, $^3J_{5a,4} = 2.2$, HaC(5)); 3.92 (dd, 1H, $^2J_{5b,5a} = 11.0$, $^3J_{5b,4} = 1.9$, HbC(5)); 4.40 (m, 1H, $^3J = 2.0$, HC(4)); 4.43 (dd, 1H, $^3J = 4.5$, HC(2)); 6.27 (d, 1H, $^3J = 4.5$, HC(1)); 7.40-7.70 (m, 10H, Ph). $^{13}\text{C NMR}$ (CDCl_3): 19.07 ($\underline{\text{C}}(\text{CH}_3)_3$); 26.67 ($\underline{\text{C}}(\text{H}_3)_3\text{C}$); 27.20, 27.71 ($\underline{\text{C}}(\text{H}_3)_2\text{C}$); 64.46 (C(5)); 77.12 (C(4)); 81.50 (C(2)); 103.77 (C(1)); 114.20 ($\underline{\text{C}}(\text{CH}_3)_2$); 127.69-135.51 (Ph); 210.85 (C(3)). **MS(FAB)**: 465 ($[\text{M}+\text{K}]^+$, 12.8%); 427 ($[\text{M}+\text{H}]^+$, 1.5%); 187 ($[\text{M}-\text{SiPh}_2\text{tBu}]^+$, 2.8%); 171 ($[\text{M}-\text{OSiPh}_2\text{tBu}]^+$, 2.4%); 135 (100%)

5-O-tert-Butyldiphenylsilyl-1,2-O-isopropylidene- α -L-ribofuranose (5). Compound **4** (10 g, 23.44 mmol) in anhydrous THF (300 ml) was cooled at -78°C and LiAlH_4 (889 mg, 23.44 mmol) was added. The mixture was stirred for 1 hour at -78°C until the disappearance of **4** was observed on tlc. The reaction was quenched with water (1 ml) and then EtOAc (50 ml) was added. The mixture was filtered over *Celite* and the organic layer was dried over Na_2SO_4 . Evaporation and crystallisation from PE gave **5** (9.65 g, 22.50 mmol, 96%) as colorless crystals. R_f (PE/TBME 6/4): 0.56; mp: 59-61°C; $[\alpha]_D^{25} = -31 \pm 1$ ($c = 1.6$, CHCl_3). $^1\text{H NMR}$ (CDCl_3): 1.08 (s, 9H, *t*Bu); 1.39 (s, 3H, CH_3); 1.58 (s, 3H, CH_3); 2.41 (d, 1H, $^3J = 9.9$, HOC(3)); 3.82-3.88 (m, 3H, HC(4), $\text{H}_2\text{C}(5)$); 4.15 (m, 1H, HC(3)); 4.58 (t, 1H, $^3J = 3.9$, HC(2)); 5.84 (d, 1H, $^3J = 3.8$, HC(1)); 7.34-7.72 (2m, 10H, Ph). $^{13}\text{C NMR}$ (CDCl_3): 19.24 ($\underline{\text{C}}(\text{CH}_3)_3$); 26.57 ($\underline{\text{C}}(\text{H}_3)_2\text{CH}$); 26.77 ($\underline{\text{C}}(\text{H}_3)_3\text{C}$); 62.36 (C(5)); 71.22 (C(3)); 78.76 (C(2)); 81.24 (C(4)); 104.16 (C(1)); 112.50 ($\underline{\text{C}}(\text{H})(\text{CH}_3)_2$); 127.63-135.62 (Ph). **MS(FAB)**: 467 ($[\text{M}+\text{K}]^+$, 20.6%); 173 ($[\text{M}-\text{OSiPh}_2\text{tBu}]^+$, 11.7%); 135 (100%)

1-O-Methyl-2,3,5-tri-O-benzoyl-L-ribofuranose (6). To a solution of **5** (5.81 g, 13.56 mmol) in anhydrous methanol (250 ml) was added H_2SO_4 (0.13 ml) at 4°C. The mixture was stirred during 2-3 days at 4°C then neutralised with *Amberlite IRA 68* (OH^- form), filtered and evaporated. The crude product was coevaporated three times with anhydrous pyridine, dissolved in pyridine (230 ml) containing DMAP (828 mg, 6.78 mmol). Benzoyl chloride

(6.30 ml, 54.22 mmol) was added dropwise at 0°C. The solution was stirred overnight at room temperature. Ice (150 g) was added and stirred for 1 hour. The mixture was extracted with CH₂Cl₂ (200 ml×3). The organic layers were extracted with water, sat. NaHCO₃ solution, dried over Na₂SO₄ and evaporated. Purification by chromatography (PE/TBME 8/2) gave **6** (5.55 g, 11.66 mmol, 86%) as a yellow oil. R_f (PE/EtOAc 8/2): 0.58 (α- and β-anomers). ¹H NMR (CDCl₃): 3.42 (s, 2.25H, CH₃, β-anomer); 3.49 (s, 0.75H, CH₃, α-anomer); 4.49–4.67 (m, 0.75H, HC(4), H₂C(5), α-an.); 4.69–4.77 (m, 2.25H, HC(4), H₂C(5), β-an.); 5.16 (s, 0.75H, HC(1), β-an.); 5.33 (dd, 0.25H, ³J=7.1, 4.4, HC(1), α-an.); 5.68 (d, 0.75H, ³J=4.9, HC(2), β-an.); 5.73 (dd, 0.25H, ³J=7.0, 3.4, HC(3), α-an.); 5.88 (dd, 0.75H, ³J=6.6, 4.9, HC(3), β-an.); 7.26–8.10 (m, 22H, Ph). ¹³C NMR (CDCl₃): 55.37 (CH₃, β-anomer); 55.75 (CH₃, α-anomer); 64.07 (C(5), α-an.); 64.78 (C(5), β-an.); 70.80 (C(3), α-an.); 71.81 (C(2), α-an.); 72.45 (C(3), β-an.); 75.47 (C(2), β-an.); 79.02 (C(4), β-an.); 79.44 (C(4), α-an.); 101.94 (C(1), α-an.); 106.44 (C(1), β-an.); 128.32–133.44 (Ph); 165.28, 165.38, 165.24, 166.24 (PhCO). MS(FAB): 515 ([M+K]⁺, 10.9%); 477 ([M+H]⁺, 2.8%); 445 ([M-OCH₃]⁺, 24.2%); 105 ([PhCO]⁺, 100%)

1-O-Acetyl-2,3,5-tri-O-benzoyl-β-L-ribofuranose (7β). Glacial acetic acid (0.94 ml) and AcO₂ (2.11 ml) were added to **6** (1.78 g, 3.73 mmol). The mixture was stirred for 5 min at 0°C, then H₂SO₄ (0.30 ml) was added dropwise during 5 min. The mixture was stirred at 0°C during 10 min until the precipitation occurred. The mixture was kept over night at 4°C. Ice (50 g) was added, and stirred for 30 min, then the mixture was extracted with CH₂Cl₂ (150 ml×3). The organic layers were washed with a cold sat. NaHCO₃ solution (200 ml×5). After drying over Na₂SO₄ and evaporation, crystallisation of the β-anomer from isopropyl alcohol afforded **7β** (712 mg, 1.411 mmol, 39%) as white crystals, (crude yield: 77% (1:1 α:β)). R_f (PE/EtOAc 8/2): α: 0.30, β: 0.43; mp: 129–131°C; [α]_D²⁵ = -43 ± 1 (c= 1.30, CHCl₃). ¹H NMR (CDCl₃): 1.99 (s, 3H, CH₃); 4.50–4.52 (m, 1H, HaC(5)); 4.75–4.80 (m, 2H, HbC(5), HC(4)); 5.79 (d, 1H, ³J=4.7, HC(2)); 5.92 (dd, 1H, ³J=7.9, 4.6, HC(3)); 6.44 (s, 1H, HC(1)); 7.30–8.10 (m, 15H, Ph). ¹³C NMR (CDCl₃): 20.85 (CH₃); 63.68 (C(5)); 71.35 (C(3)); 74.98 (C(2)); 79.95 (C(4)); 98.38 (C(1)); 128.38–133.64 (Ph); 165.00, 165.34, 165.95 (PhCO); 169.64 (CH₃CO). MS(FAB): 543 ([M+K]⁺, 13.5%); 445 ([M-OCOCH₃]⁺, 21.4%); 105 ([PhCO]⁺, 100%)

2',3',5'-tri-O-Benzoyl-β-L-uridine (8a). Compound **7β** (250 mg, 0.50 mmol) and uracil (167 mg, 1.49 mmol) were co-evaporated three times with anhydrous CH₃CN under argon. CH₃CN abs. (6 ml) and BSA (0.37 ml, 1.49 mmol) were added at room temperature. The mixture was stirred at 60–65°C until the base dissolved. TMSOTf (87 μl, 0.74 mmol) was added, the mixture stirred over night at 65°C. CH₂Cl₂ (50 ml) was added and the mixture was washed with sat. NaHCO₃ solution (10 ml). After drying over Na₂SO₄ and evaporation, purification by chromatography (PE/EtOAc 7/3 and 5/5) afforded **8a** (246 mg, 0.44 mmol, 89%) as a white foam. R_f (PE/EtOAc 2/8): 0.37. ¹H NMR (CDCl₃): 4.65–4.87 (m, 3H, HC(4'), H₂C(5')); 5.61 (d, 1H, ³J=8.0, HC(5)); 5.77 (t, 1H, ³J=5.7, HC(2')); 5.90 (t, 1H, ³J=3.6, HC(3')); 6.32 (d, 1H, ³J=5.7, HC(1')); 7.33–8.08 (m, 15H, Ph); 8.11(d, 1H, ³J=8.4, HC(6)); 9.11 (s, 1H, NH). ¹³C NMR (CDCl₃): 63.70 (C(5')); 71.10 (C(3')); 73.73 (C(2')); 80.46 (C(4')); 88.12 (C(1'));

103.37 (C(5)); 128.31–133.76 (Ph); 139.63 (C(6)); 150.12 (C(2)); 162.91 (C(4)); 165.25, 165.28, 166.01 (PhCO). **MS(FAB)**: 595 ([M+K]⁺, 5.7%); 557 ([M+H]⁺, 2.6%); 445 ([M-base]⁺, 13.4%); 105 ([PhCO]⁺, 100%)

4-N-Acetyl-2',3',5'-tri-O-benzoyl-β-L-cytidine (8b). Compound **7β** (250 mg, 0.50 mmol) and 4-N-acetylcytosine (228 mg, 1.49 mmol) were treated as described above. Purification by chromatography (PE/EtOAc 5/5 and 2/8) furnished **8b** (255 mg, 0.43 mmol, 86%) as a white foam. R_f (PE/EtOAc 2/8): 0.46. ¹H NMR (CDCl₃): 2.20 (s, 3H, CH₃); 4.69–4.86 (m, 3H, HC(4'), H₂C(5')); 5.84 (dd, 1H, ³J=5.9, 4.4, HC(2')); 5.90 (t, 1H, ³J=5.4, HC(3')); 6.40 (d, 1H, ³J=4.2, HC(1')); 7.32–7.95 (m, 16H, Ph, HC(5)); 8.10 (d, 1H, ³J=7.8, HC(6)); 9.98 (br s, 1H, NH). ¹³C NMR (CDCl₃): 24.82 (CH₃); 63.53 (C(5')); 70.90 (C(3')); 74.75 (C(2')); 80.59 (C(4')); 90.07 (C(1')); 97.33 (C(5)); 128.47–133.64 (Ph); 144.56 (C(6)); 154.60 (C(2)); 163.23 (C(4)); 165.15, 165.22, 166.09 (PhCO); 170.89 (CH₃CO). **MS(FAB)**: 636 ([M+K]⁺, 6.4%); 598 ([M+H]⁺, 2.9%); 445 ([M-base]⁺, 10.8%); 105 ([PhCO]⁺, 100%)

6-N-Benzoyl-2',3',5'-tri-O-benzoyl-β-L-adenosine (8c). Compound **7β** (200 mg, 0.40 mmol) and 6-N-benzoyladenine (284 mg, 1.19 mmol) were co-evaporated three times with anhydrous 1,2-dichloroethane under argon. Anhydrous 1,2-dichloroethane (5 ml) and MFSTA (0.44 ml, 2.38 mmol) were added at room temperature. The mixture was stirred at 80°C until the base dissolved. TMSOTf (0.11 ml, 0.59 mmol) was added, the mixture stirred over night at 80°C. CH₂Cl₂ (40 ml) was added, the mixture was washed with sat. NaHCO₃ solution (10 ml). After drying over Na₂SO₄ and evaporation, purification by chromatography (PE/EtOAc 5/5 to 2/8) afforded **8c** (228 mg, 0.33 mmol, 84%) as a white foam. R_f (PE/EtOAc 2/8): 0.57. ¹H NMR (CDCl₃): 4.72 (dd, 1H, ²J_{5'a,5'b}=12.0, ³J_{5'a,4'}=3.1, HaC(5')); 4.82–4.86 (m, 1H, HC(4')); 4.88 (dd, 1H, ²J_{5'b,5'a}=11.9, ³J_{5'b,4'}=4.1, HbC(5')); 6.30 (t, 1H, ³J=5.2, HC(2')); 6.46 (d, 1H, ³J=5.4, HC(3')); 6.53 (d, 1H, ³J=5.0, HC(1')); 7.32–8.10 (m, 20H, Ph); 8.26 (s, 1H, HC(2)); 8.65 (s, 1H, HC(8)); 9.64 (br s, 1H, NH). ¹³C NMR (CDCl₃): 63.38 (C(5')); 71.34 (C(3')); 73.77 (C(2')); 80.66 (C(4')); 86.96 (C(1')); 123.54 (C(5)); 127.85–133.60 (Ph); 141.71 (C(8)); 149.80 (C(4)); 151.60 (C(2)); 152.58 (C(6)); 164.95, 165.12, 165.93 (PhCO, PhCONH). **MS(FAB)**: 722 ([M+K]⁺, 8.4%); 684 ([M+H]⁺, 7.0%); 445 ([M-base]⁺, 13.6%); 105 ([PhCO]⁺, 100%)

2-N-Acetyl-6-O-diphenylcarbamoyl-2',3',5'-tri-O-benzoyl-β-L-guanosine (8d). 2-N-Acetyl-6-O-diphenylcarbamoylguanidine¹³ (168 mg, 0.45 mmol) was co-evaporated three times with anhydrous 1,2-dichloroethane under argon. Absolute 1,2-dichloroethane (3 ml) and BSA (0.15 ml, 0.60 mmol) were added at room temperature. The mixture was stirred at 80°C until the base dissolved, evaporated in vacuo, then dissolved in anhydrous toluene (1 ml). Compound **7β** (150 mg, 0.30 mmol) was coevaporated three times with anhydrous toluene, then dissolved in toluene (4 ml), then added to the base soln. under argon. TMSOTf (81 μl, 0.45 mmol) was added, the mixture stirred for one hour at 80°C. CH₂Cl₂ (50 ml) was added, the mixture was washed with sat. NaHCO₃ solution (20 ml). After drying over Na₂SO₄ and evaporation, purification by chromatography (PE/EtOAc 8/2 and 5/5) furnished **8d** (174 mg, 0.21 mmol, 70%) as a white foam. R_f (PE/EtOAc 2/8): 0.82. ¹H NMR (CDCl₃): 2.42 (s, 3H,

CH₃); 4.72–4.95 (m, 3H, HC(4'), H₂C(5)); 6.28–6.38 (m, 3H, HC(3'), HC(2'), HC(1')); 7.20–8.00 (m, 25H, Ph); 8.06 (s, 1H, HC(8)); 8.42 (br s, 1H, NH). ¹³C NMR (CDCl₃): 24.99 (CH₃); 63.63 (C(5')); 71.46 (C(3')); 74.22 (C(2')); 80.55 (C(4')); 87.53 (C(1')); 121.26 (C(5)); 126.64–142.45 (Ph); 141.67 (C(8)); 150.13 (CONPh₂); 152.27 (C(4)); 154.25 (C(2)); 156.32 (C(6)); 165.04, 165.15, 166.11 (PhCO); 169.83 (CH₃CO). MS(FAB): 871 ([M+K]⁺, 3.7%); 833 ([M+H]⁺, 3.0%); 445 ([M-base]⁺, 18.3%); 105 ([PhCO]⁺, 100%)

L-Uridine (9a). Compound **8a** (1.36 g, 2.44 mmol) was dissolved in methanol (10 ml) then NH₃ (35% in water, 20 ml) was added. The mixture was stirred over night at 60°C in a sealed vessel. After evaporation to dryness, water (50 ml) was added and extracted with EtOAc (50 ml×3). The water layer was evaporated to give **9a** (566 mg, 2.32 mmol, 95%) which was recrystallised from H₂O/EtOH as white crystals. R_f (MeOH/EtOAc 1/9): 0.23; mp: 159–163°C; [α]_D²⁵ = -16 ± 1 (c = 0.55, H₂O). ¹H NMR (CD₃OD): 3.73 (dd, 1H, ²J_{5'a,5'b} = 12.1, ³J_{5'a,4'} = 3.1, HaC(5')); 3.83 (dd, 1H, ²J_{5'b,5'a} = 12.2, ³J_{5'b,4'} = 2.6, HbC(5')); 3.99–4.02 (m, 1H, HC(4')); 4.13–4.19 (m, 2H, HC(2'), HC(3')); 5.69 (d, 1H, ³J = 8.0, HC(5)); 5.90 (d, 1H, ³J = 4.3, HC(1')); 8.01 (d, 1H, ³J = 8.0, HC(6)). ¹³C NMR (CD₃OD): 62.28 (C(5')); 71.31 (C(3')); 75.74 (C(2')); 86.36 (C(4')); 90.73 (C(1')); 102.65 (C(5)); 142.74 (C(6)); 152.46 (C(2)); 166.20 (C(4)). MS(FAB): 283 ([M+K]⁺, 35.3%); 245 ([M+H]⁺, 74.6%); 133 ([M-base]⁺, 18.9%); 113 ([base+H₂]⁺, 100%)

L-Cytidine (9b). Compound **8b** (1.27 g, 2.13 mmol) was dissolved in 0.1M NaOMe/MeOH (50 ml). The mixture was stirred over night at room temperature. A minimal amount of Dowex (H⁺ form) was added to neutralise the mixture. After filtration and evaporation to dryness, water (50 ml) was added and extracted with EtOAc (50 ml×3). The water layer was evaporated to give **9b** (462 mg, 1.90 mmol, 89%) which was recrystallised from H₂O/EtOH as white crystals. R_f (MeOH/EtOAc/H₂O 1/4/0.7): 0.34; mp: 200–205°C; [α]_D²⁵ = -33 ± 1 (c = 0.82, H₂O). ¹H NMR (DMSO-d₆): 3.52–3.70 (m, 2H, H₂C(5')); 3.83–3.85 (m, 1H, HC(4')); 3.92–3.96 (m, 2H, HC(2'), HC(3')); 5.04–5.10 (m, 2H, HOC(3'), HOC(5')); 5.35 (d, 1H, ³J = 4.5, HOC(2')); 5.74 (d, 1H, ³J = 7.5, HC(5)); 5.79 (d, 1H, ³J = 3.6, HC(1')); 7.24 (br s, 2H, NH₂); 7.87 (d, 1H, ³J = 7.5, HC(6)). ¹³C NMR (DMSO-d₆): 60.70 (C(5')); 69.51 (C(3')); 74.07 (C(2')); 84.14 (C(4')); 89.23 (C(1')); 94.03 (C(5)); 141.57 (C(6)); 155.54 (C(2)); 165.63 (C(4)). MS(FAB): 282 ([M+K]⁺, 26.2%); 244 ([M+H]⁺, 48.8%); 112 ([base+H₂]⁺, 100%)

L-Adenosine (9c). Compound **8c** (1.23 g, 1.80 mmol) was dissolved in methanol/THF (5/2, v:v, 14 ml) then NH₃ (35% in water, 20 ml) was added. The mixture was stirred over night at 60°C in a sealed vessel. After evaporation to dryness, water (50 ml) was added and extracted with EtOAc (50 ml×3). The water layer was evaporated to give **9c** (443 mg, 1.66 mmol, 92%) which was recrystallised from H₂O/EtOH as a white powder. R_f (MeOH/EtOAc/H₂O 1/4/0.7): 0.59; mp: 235–238°C; [α]_D²⁵ = +87 ± 1 (c = 0.45, 1.25M NaOH). ¹H NMR (DMSO-d₆): 3.53–3.72 (m, 2H, H₂C(5')); 3.96–3.98 (m, 1H, HC(4')); 4.14–4.18 (m, 1H, HC(3')); 4.62 (dd, 1H, ³J = 11.4, 6.0, HC(2')); 5.22 (d, 1H, ³J = 4.4, HOC(3')); 5.45–5.49 (m, 2H, HOC(2'), HOC(5')); 5.89 (d, 1H, ³J = 6.0, HC(1')); 7.39 (br s, 2H, NH₂); 8.15 (s, 1H, HC(2)); 8.37 (s, 1H, HC(8)). ¹³C NMR (DMSO-d₆): 61.72 (C(5')); 70.71 (C(3')); 73.47 (C(2')); 85.94 (C(4')); 87.94

(C(1')); 119.40 (C(5)); 136.98 (C(8)); 149.08 (C(4)); 152.42 (C(2)); 156.20 (C(6)).
MS(FAB): 306 ([M+K]⁺, 1.5%); 268 ([M+H]⁺, 1.0%); 267 ([M]⁺, 2.3%); 136 (100%)

L-Guanosine (9d). Compound **8d** (2.43 g, 2.97 mmol) was dissolved in methanol/THF (5/2, v:v, 14 ml) then NH₃ (35% in water, 20 ml) was added. The mixture was stirred over night at 60°C in a sealed vessel. After evaporation to dryness, water (250 ml) was added and extracted with CH₂Cl₂ (100 ml×3). The water layer was evaporated to give **9d** (757 mg, 2.67 mmol, 90%) which was recrystallised from H₂O/EtOH as a pale yellow powder. R_f (MeOH/EtOAc/H₂O 1/4/0.7): 0.28; mp: 239–243°C ; [α]_D²⁵ = +80 ± 1 (c = 0.85, 1.25M NaOH). ¹H NMR (DMSO-d₆): 3.54–3.65 (m, 2H, H₂C(5')); 3.89 (d'd', 1H, ³J = 7.5, 3.8, HC(4')); 4.09 (dd, 1H, ³J = 4.8, 3.8, HC(3')); 4.41 (t, 1H, ³J = 5.5, HC(2')); 5.13 (t, 1H, ³J = 5.5, HOC(5')); 5.19 (d, 1H, ³J = 4.7, HOC(3')); 5.45 (d, 1H, ³J = 6.1, HOC(2')); 5.71 (d, 1H, ³J = 6.0, HC(1')); 6.52 (br s, 2H, NH₂); 7.95 (s, 1H, HC(8)); 10.69 (br s, 1H, NH). ¹³C NMR (DMSO-d₆): 61.40 (C(5')); 70.38 (C(3')); 73.71 (C(2')); 85.20 (C(4')); 86.33 (C(1')); 116.09 (C(5)); 135.58 (C(8)); 151.33 (C(4)); 153.70 (C(2)); 156.77 (C(6)). **MS(FAB):** 322 ([M+K]⁺, 4.3%); 284 ([M+H]⁺, 11.6%); 283 ([M]⁺, 5.5%); 122 (100%)

Acknowledgement: The financial support by the *Swiss National Science Foundation* is gratefully acknowledged.

References and Footnotes

- [1] Abstract book and Congress Proceedings of the 13th *International Round Table: Nucleosides & Nucleotides and their Biological Applications*, 6–10.9.1998, Montpellier, France.
- [2] Nolte A, Klußmann S, Bald R, Erdmann VA, Fürste JP. *Nature Biotechnology* **1996**, *14*, 1116–1119.
- [3] Abe Y, Takizawa T, Kunieda T. *Chem. Pharm. Bull.* **1980**, *28*, 1324–1326.
- [4] Holý A, Sorm F. *Collect. Czech. Chem. Commun.* **1969**, *34*, 3383–3401; *ibid.* **1972**, *37*, 4072–4087; *ibid.* **1973**, *38*, 423–427.
- [5] Visser GM, Van Westrenen J, Van Boeckel CAA, Van Boom JH. *Recl. Trav. Chim. Pays Bas* **1986**, *105*, 528–537.
- [6] Pitsch S. *Helv. Chem. Acta* **1997**, *80*, 2286–2314.
- [7] Jung M, Xu Y. *Tetrahedron Lett.* **1997**, *38*, 4199–4202.
- [8] Poopeiko NE, Kvasnyuk EI, Mikhailopulo IA, Lidaks MJ. *Synthesis* **1985**, 605–609.
- [9] Altmann KH, Dan NM, Fabbro D, Freier SM, Geiger T, Häner R, Hüsken D, Martin P, Monia BP, Müller M, Natt F, Nicklin P, Phillips J, Pielas U, Sasmor H, Moser HE. *Chimia* **1996**, *50*, 168–176. In our hands the α/β mixture of methyl riboside derivative **6** could not be well separated and only the β anomer reacted under the applied glycosidation conditions.
- [10] Vorbrüggen H, Krolkiewicz K, Bennua B. *Chem. Ber.* **1981**, *114*, 1234–1255.
- [11] Redondo EF, Rinderknecht H. *Helv. Chim. Acta* **1959**, *42*, 1171–1173.
- [12] (a) Vorbrüggen H, Höfle G. *Chem. Ber.* **1981**, *114*, 1256–1268. (b) Vorbrüggen H. *Acta Biochim. Polon.* **1996**, *43*, 25–36.
- [13] Zou R, Robins MJ. *Can. J. Chem.* **1987**, *65*, 1436–1437 and ref. cited therein.