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## HIV-1 replication inhibitors of the styrylquinoline class: introduction of an additional carboxyl group at the C-5 position of the quinoline

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Abstract—Novel variants of HIV-1 replication inhibitors of the styrylquinoline class, bearing an additional acid group or a propenoic acid moiety at the C-5 position of the quinoline have been synthesized. Key steps included Heck reaction and palladium catalyzed carbonylation reaction of 5-haloquinaldine derivatives. These compounds exhibited reinforced anti-integrase potency and significant antiviral activities.

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The substantial incidence of resistance observed in therapy-experienced patients and newly acquired HIV-1 infections underscores the need for new antiretroviral agents. All oral agents licensed to treat HIV-1 diseases target two of the three essential, virally-encoded enzymes, reverse transcriptase and protease.<sup>1</sup> The third enzyme, integrase inserts the viral DNA into the cellular genome through a multi-step process that includes two catalytic reactions, 3' endonucleolytic processing of the viral DNA ends, and joining of the viral and cellular DNAs (strand transfer).<sup>2</sup> The availability of in vitro assays using recombinant integrase has allowed the discovery of numerous classes of compounds with inhibitory activity.<sup>3</sup>

We have reported that polyhydroxylated styrylquinolines (SQLs), exemplified by **1**, are potent HIV-1 integrase inhibitors in in vitro experiments, block the replication of HIV-1 in cell culture, and are devoid of cytotoxicity.<sup>4</sup> Although the exact mechanism by which drug **1** and analogs exert their inhibitory potency remains unknown, it has been recently proposed that such drugs might act prior to integration by preventing viral DNA-integrase-binding.<sup>5</sup> Thus, SQLs constitute unique class of integrase inhibitors, distinct from the diketoacid group (illustrated by L 731,988, **2**), which affect viral DNA integration.<sup>3a</sup> Within the carbon framework of SQLs, we have identified the salicylic acid moiety at C-7, C-8 of the quinoline ring, and the 4'-OH on the ancillary aromatic nucleus as critical pharmacophores for antiviral activity.

Since it was recognized that carboxylic acid groups are beneficial to both integrase inhibition potency and cytotoxicity, we envisioned to elaborate new structural variants possessing an additional carboxyl group at C-5 while keeping the salicylic acid system at C-7/C-8. Herein, we report the synthesis and the evaluation of the biological activity of new styrylquinoline derivatives **3a**,**b** bearing a carboxyl group either directly bound to the quinoline ring or through an ethylenic spacer.

From the outset of the work we planned to incorporate the requisite function at an early stage of the synthesis, via a palladium-catalyzed coupling reaction of the 5bromoquinaldine  $\mathbf{6}$ , before to elaborate the arylethenyl

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system through a Perkin-type condensation with aromatic aldehydes 4. Our starting point for the synthesis of compound 3a was the 8-hydroxy-2-methyl-quinoline-7-carboxylic acid 7 available, although in moderate yield, by Kolbe–Schmidt carboxylation of 8-hydroxyquinaldine (Scheme 1).<sup>6</sup>

Bromination of acid 7 using bromine or NBS provided the desired bromo acid 6 in 30–40% yield, along a substantial amount of dibromoquinoline 8.<sup>7</sup> Since these procedures turned out to be quite erratic, we turned to a more reliable method. Thus methylation of 7 produced the corresponding dimethylated derivative, which uneventfully gave bromoester 9 in 72% yield upon treatment with bromine. To our delight, Heck reaction of 9 with ethyl acrylate delivered the desired unsaturated ester 10 in 66% yield. Finally, Perkin-type condensation of quinaldine 9 with 3,4-dihydroxybenzaldehyde (4, R=H), followed by deprotection afforded the desired diacid 3a in 49% overall yield from 10 (Scheme 2).<sup>8</sup>

We next turned our attention to the synthesis of diacid **3b**, bearing the carboxyl group directly bound to the quinoline nucleus. Introduction of a nitrile function as carboxyl group surrogate was first investigated. Thus, when the *n*-butyl ester **11**, easily available from acid 7,<sup>4f</sup> was reacted with sodium iodide in the presence of chloramine-T,<sup>9</sup> the desired 5-iodoquinaldine derivative **12** was obtained in 80% yield. Protection of the phenol group as a pivalate, followed by palladium(0) and copper(I) co-catalyzed cyanation reaction<sup>10</sup> delivered the carbonitrile **13** in 62% overall yield. However the final hydrolysis was disappointing. For example, hydrolysis of **13** with concentrated hydrochloric acid in refluxing acetic acid gave mainly the hydroxy-acid **14** in which the nitrile group was unaffected. On the other hand,







Scheme 2. Reagents and conditions: (a) NBS, DMF, 20 °C, 1 h, 6: 40%; (b) 20 equiv MeI, 20 equiv  $K_2CO_3$ , DMF–acetone, 12 h at 50 °C, 95%; (c) Br<sub>2</sub>, DMF, CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 72%; (d) 5 mol % Pd(OAc)<sub>2</sub>, 20 mol % PPh<sub>3</sub>, 5 equiv H<sub>2</sub>C=CHCO<sub>2</sub>Et, 5 equiv Et<sub>3</sub>N, 110 °C, 2.5 h, 66%; (e) (i) 3,4-dihydroxybenzaldehyde (4, R = H) 4 equiv, Ac<sub>2</sub>O, 140 °C, 12 h; (f) HBr 48%, AcOH, 100 °C, 30 h, 49%.

the use of more drastic conditions (e.g., 50% H<sub>2</sub>SO<sub>4</sub>, 100 °C) led to complete decomposition (Scheme 3).

This problem combined with the modest yield encountered in the Kolbe–Schmidt carboxylation to access the acid 7, prompted us to explore an alternative strategy based on the regioisomeric quinaldine 16.<sup>11</sup> Since this material was originally prepared according to a lengthy sequence, we decided to explore the Doebner and Miller condensation of commercially available 3amino-4-hydroxybenzoic acid 15. In the event treatment 15 with crotonaldehyde in refluxing 6 N hydrochloric acid provides quinoline 16 in 52% yield. Iodination of 16 with the KI/I<sub>2</sub> system as reported<sup>12</sup> provided iodo phenol 17. Benzylation of 17 afforded ester 18, which,



Scheme 3. Reagents and conditions: (a) 1.5 equiv NaI, 1.5 equiv *p*-TolN(Na)Cl, DMF, 20 °C, 1 h, 80%; (b) *t*-BuCOCl, py, DMF, 18 h, 20 °C, 82%; (c) 5 mol % Pd(PPh<sub>3</sub>)<sub>4</sub>, 10 mol % CuI, 2 equiv KCN, refluxing THF, 2 h, 76% (d) HCl concd, AcOH, 110 °C, 16 h, 54%.

upon Stille reaction with tributyl-(1-ethoxy-vinyl)-stannane  $19^{13}$  gave the enol ether 20 (82% yield). The later product was ozonolyzed to give rise to diester 21, but with a disappointing low yield.<sup>14</sup> Notwithstanding this drawback we continued our synthesis. Thus saponification of the ethyl ester followed by conventional hydrogenolysis of the two benzyl groups furnished the expected diacid 22 in 38% overall yield from 21 (Scheme 4).

Although this route was successful, it suffered from the tedious procedure for the introduction of the C-7 carboxylic group. Clearly a direct hydroxycarbonylation reaction would be a more suitable method. In the event, treatment iodoquinaldine **18** with palladium acetate and dppp in wet DMSO under 1 atm of CO directly afforded the expected acid **23** in 41% yield.<sup>15</sup>

To our surprise, the debenzylation of **23** turned out to be ineffective probably due to the presence in this polar material of phosphine traces poisoning the palladium catalyst. We thus have recourse to a variant of the carbonylation reaction using benzyl alcohol as nucleophilic partner.

Heating a DMSO solution of **18** in the presence of benzyl alcohol and palladium acetate under CO pressure led to diester **24** with a 55% yield after chromatographic purification. By contrast with the reaction of **23**, deprotection of this material now proceeded smoothly to give diacid **22** in 70% yield. With an efficient route to this material, we next proceeded to the implementation of the arylethenyl appendage. Thus condensation of **22** with 3,4-dihydroxy-5-methoxybenzaldehyde **25**, followed by hydrolysis of the intermediate acetylated



Scheme 4. Reagents and conditions: (a) HCl 6 N, CH<sub>3</sub>CH=CHCHO, 100 °C, 1 h, 52%; (b)  $I_2/KI$ , aq NaOH, 0.5 h, 15 °C, 70% (c) 3 equiv BnBr, 3 equiv  $K_2CO_3$ , DMF, 24 h, 50 °C, 80%; (d) 1.8 equiv 19, 5 mol % Pd<sub>2</sub>dba<sub>3</sub>·CHCl<sub>3</sub>, DMF, 100 °C, 3 h, 82%; (e) (i) O<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, (ii) Me<sub>2</sub>S, 20 °C, 8 h 20%; (f) 2 N NaOH, MeOH, THF, 18 h; (g) Pd(OH)<sub>2</sub>, DMF, AcOH, 1 atm H<sub>2</sub>, 20 °C, 8 h, 38% from 21.

compound produced the targeted diacid **3b** in 73% overall yield from **22** (Scheme 5).<sup>16</sup>

Diacids **3a.b** were assayed in vitro for inhibitory activity against HIV-1 integrase, and for antiviral activity against HIV-1 replication in cells. In vitro IC<sub>50</sub> was determined as the drug concentration that inhibits 50% of the recombinant integrase activity in a standard 3'processing and strand transfer assays. Cellular antiviral  $IC_{50}$  was determined as the drug concentration that inhibits 50% of viral particles production in de novo infection assay of HeLa P4 cells.<sup>5b,17</sup> TC<sub>50</sub> was the drug concentration that corresponds to 50% of cells survival as determined by a standard MTT assay.<sup>18,19</sup> Results are listed in comparison with reference compound 1 in Table 1.<sup>4b</sup> Diacids **3a**,**b** were found to be highly potent inhibitors of integrase on both 3'-processing and strand transfer assays (Fig. 1). This result clearly established that carboxyl groups on the quinoline ring are beneficial to the inhibitor/integrase interaction as initially suspected. When compared to 1, a marked decline in antiviral activity was observed for **3b**, on the other hand **3a** kept a good activity on cell culture, but with a slightly increased cytotoxicity.

In short we have devised efficient syntheses of two new styrylquinoline derivatives with additional carboxylic residues on the quinoline ring. Although improved anti-integrase activities were observed, no decisive



Scheme 5. Reagents and conditions: (a) 15 mol % Pd(OAc)<sub>2</sub>, 20 mol % dppp, 10 equiv Et<sub>3</sub>N, DMSO, H<sub>2</sub>O, 2 atm CO, 70 °C, 20 °C 1 h then 70 °C 2 h, 41%; (b) 15 mol % Pd(OAc)<sub>2</sub>, 20 mol % dppp, 10 equiv Et<sub>3</sub>N, DMSO, 10 equiv BnOH, 2 atm CO, 70 °C, 20 °C 1 h then 70 °C 2 h, 55%; (c) Pd(OH)<sub>2</sub>, 1 atm H<sub>2</sub>, AcOEt, EtOH, AcOH, 20 °C, 24 h, 70%; (d) (i) 2.5 equiv 25, Ac<sub>2</sub>O, 4 d, (ii) py, H<sub>2</sub>O, 110 °C, 3 h, 73%.

Table 1. Biological activities of 3a,b

Compound	1	3a	3b
3' Processing <sup>a</sup>	1.0	0.3	0.2
Integration <sup>a</sup>	1.7	0.06	0.07
Antiviral activity <sup>a</sup>	0.3	35	1.6
Cytotoxicity <sup>b</sup>	>100	>100	60

<sup>a</sup> IC<sub>50</sub>, μM.

<sup>b</sup>TC<sub>50</sub>, μM.



Figure 1. Phosphorimager picture of HIV IN integration activity in presence of increasing concentration of compounds 3a,b. Integration assays were performed at 100 nM of IN, 12.5 nM of U5-2, and 7.5 mM of MgCl<sub>2</sub>.

breakthrough resulted from this substitution pattern on the antiviral potency. Further studies aimed at exploring modulations of the ancillary rings of compounds **3a**,**b** are ongoing.

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## **References and notes**

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- 8. Compound **3a**: Brick red solid; mp > 300 °C (dec); IR (neat, cm<sup>-1</sup>) v 3400–2300 (broad)  $\delta$  1677, 1626, 1587, 1518; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz)  $\delta$  9.04 (d, J = 9.1 Hz, 1H), 8.42 (d, J = 9.1 Hz, 1H), 8.41 (s, 1H), 8.27 (d, J = 15.7 Hz, 1H), 8.08 (d, J = 16.4 Hz, 1H), 7.73 (d, J = 16.4 Hz, 1H), 7.22 (broad s, 1H), 7.14 (d, J = 8.1 Hz, 1H), 6.90 (d, J = 8.1 Hz, 1H), 6.49 (d, J = 15.7 Hz, 1H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz)  $\delta$  169.5 (C), 167.9 (C), 163.1 (C), 152.3 (C), 149.1 (C), 146.2 (C), 142.1 (CH), 139.0 (CH), 138.2 (CH), 134.3 (C), 128.8 (C), 127.9 (CH), 127.5 (C), 121.9 (CH), 121.4 (CH), 118.7 (CH), 118.3 (CH), 116.5 (CH), 115.8 (C), 114.9 (CH), 114.0 (C).
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- 14. An improved yield was observed using methanol as cosolvent, but extensive trans-esterification occurred in such conditions.
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- 16. Compound **3b**: Brick red solid; mp > 260 °C (dec); IR (neat, cm<sup>-1</sup>)  $\nu$  3500–2500 (broad), 1693, 1690, 1584; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 200 MHz)  $\delta$  9.77 (d, *J* = 9.4 Hz, 1H), 8.73 (s, 1H), 8.47 (d, *J* = 9.4 Hz, 1H), 7.99 (d, *J* = 16.1 Hz, 1H), 7.78 (d, *J* = 16.1 Hz, 1H), 6.92 (s, 1H), 6.89 (s, 1H), 3.90 (s, 3H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 50 MHz)  $\delta$  169.4 (C), 168.3 (C), 167.4 (C), 151.5 (C), 149.5 (C), 146.9 (C), 143.1 (CH), 141.6 (CH), 138.6 (C), 136.0 (CH), 135.0 (C), 130.4 (C), 126.7 (C), 122.2 (CH), 118.5 (CH), 113.4 (C), 110.7 (CH), 109.0 (C), 104.5 (CH), 56.8 (CH<sub>3</sub>); Anal. Calcd for C<sub>20</sub>H<sub>15</sub>NO<sub>8</sub>·2/3H<sub>2</sub>O: C, 58.68; H, 4.02; N, 3.42. Found: C, 58.53; H, 4.21; N, 3.36.
- 17. HeLa P4 cells were infected with viruses and grown in the presence of increasing concentrations of drugs. After 72 h, the infectivity of viruses was determined by measuring beta-galactosidase (β-Gal) production by the chlorophenol red-β-D-galactopyranoside (CPRG) assay.<sup>18</sup> The 50%

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