

Tetrahedron 54 (1998) 2827--2832

TETRAHEDRON

Stereoselective Synthesis of the C-Linked Analogue of β-D-Galactopyranosyl-L-serine

Alessandro Dondoni,* Alberto Marra, and Alessandro Massi

Dipartimento di Chimica, Laboratorio di Chimica Organica, Università, 44100 Ferrara, Italy.

Received 5 December 1997; accepted 8 January 1998

Abstract: The coupling of the D-serinal derivative 7 with the D-galactopyranosylmethylene phosphorane generated from the phosphonium salt 6 and reduction of the resulting alkene led to the C-glycosylated amino alcohol 9 that in turn was oxidized to the title amino acid in 44% overall yield. © 1998 Elsevier Science Ltd. All rights reserved:

The synthesis of the so called C-glycosyl amino acids possessing an anomeric C-C bond instead of the C-N or C-O bond between the sugar and the amino acid moieties is an issue that is currently addressed in various laboratories.¹ These synthetic amino acids can be used for the modification of bioactive glycopeptides by the attachment of carbohydrates through chemical and enzymatic resistant carbon-carbon bond.² Among the few genuine isosteres so far reported wherein the glycosidic oxygen atom has been replaced by a methylene group, lb,d,h,k,n the C-analogue 2 of β-D-galactopyranosyl-L-serine (1) has been prepared^{1d} and incorporated into a 17-amino acid α -helical peptide for both biological and conformational studies.³ The amino acid 2 was also employed^{1e} for the synthesis of water-soluble carbon-linked galactosphingolipid analogues that proved to bind specifically to HIV-1 gp120 and therefore represented potential inhibitors of the first step in the infection process causing the AIDS. In both cases tetra-O-benzylated N-Fmoc and N-Boc derivatives of 2 were prepared by Wittig condensation of the C-glycosyl aldehyde 3 (see Scheme 1) with a suitable phosphorane serving as a β -alaninol anion equivalent. Therefore the B-D-linkage at the anomeric centre of the sugar and the S-configuration at the carbon bearing the amino group were already in place in the reagents employed. We would like to describe the application of the same concept in a reversed manner and report below an improved synthesis of 2 by condensation of a β -linked D-galactose phosphorous ylide with a D-serine derived aldehyde as the key coupling step. We considered an alternative synthetic approach to the amino acid 2 because the Wittig olefination of the high value sugar aldehyde 3 was reported^{1d} to occur in low yield (34%) whilst we needed substantial amounts of 2 for the synthesis of glycopeptide and sphingosine mimetics.



The formyl C-glycoside 3 was prepared in gram quantities (1-5 g) by our thiazole-based method starting from tetra-O-benzyl-D-galactonolactone.⁴ The crude product obtained by the improved thiazole-to-formyl unmasking protocol⁵ was reduced (NaBH₄) to the alcohol 4 in almost quantitative yield (Scheme 1). Pure compound 4 was readily transformed into the iodomethyl derivative 5 under standard iodination conditions⁶ and the latter was efficiently converted into the corresponding phosphonium iodide 6 by coupling with neat triphenylphosphine at 120 °C. When the same reaction was carried out in the presence of various solvents, the salt 6 was obtained in much lower yield. Compound 6 proved to be a non-hygroscopic material, storable for long period without appreciable decomposition.



Scheme 1. Reagents and conditions: a) NaBH₄, Et₂O-MeOH, 0 °C, 10 min; b) 4₂, Ph₃P, Imidazole,toluene, reflux, 2 h; c) Ph₃P, 120 °C, 2 h.

With an efficient entry to the sugar phosphonium salt 6 at hand, suitable conditions were searched for an efficient coupling with the readily accessible⁷ N-Boc-N, O-isopropylidene-D-serinal 7 (Scheme 2). The sugar phosphorane, generated from 6 by treatment with *n*BuLi (1 equiv) in THF-HMPA at -40 °C, was reacted with a solution of the aldehyde 7 (1 equiv) in THF and the mixture allowed to warm up to -10 °C. Suitable workup and flash chromatography on silica gel afforded pure (Z)-8 (54%, $J_{3,4} = 11.3$ Hz) and (E)-8 (8%, $J_{3,4} = 16.0$ Hz) slightly contaminated by uncharacterized byproducts.



Scheme 2. Reagents and conditions: a) 6, *n*BuLi, 4:1 THF-HMPA, 4 Å MS, -40 to -10 °C, 2 h; b) TsNHNH₂ , AcONa , 4:1 DME-H₂O, reflux, 5 h; c) Jones reagent, acetone, 0 °C to r.t., 3 h; d) CH₂N₂, Et₂O-MeOH, 0 °C, 5 min.

In order to preserve the O-benzyl protective groups of the sugar moiety, the double bond of the Z- and Ealkene 8 was reduced by the use of diimide⁸ generated in situ from p-toluenesulfonhydrazide and sodium acetate.^{1e} In this way each individual olefin afforded the same C-alkyl galactoside 9 in comparable yields (82-84%). That the original β -D-linkage at the anomeric carbon of 3 was maintained in 9 was demonstrated by the coupling constant value of 9.0 Hz between the trans-diaxial protons at C-5 and C-6. To complete the synthesis, deacetonation and oxidation of 9 with the Jones reagent afforded in a single step the crude tetra-O-benzylgalactosyl-N-Boc- α -amino-acid 2a in 94% yield, contaminated (~5%) by the corresponding α -amino-alcohol derivative⁹ 11. Moreover, the amino acid 2a was fully characterized as the methyl ester 10. The overall yield of isolated 2a from the sugar phosphonium salt 6 was 44%.

In conclusion, a practical alternative procedure has been developed for the preparation of the O-benzyl N-Boc protected sugar amino acid 2a. This kind of protection and the free carboxylic group provide the arrangement required for the incorporation of 2a into a peptide chain. Both the sugar and the amino acid building blocks 6 and 7 can be easily prepared in gram quantities. In respect to the earlier synthesis, ^{1d} this procedure involves a higher yield (almost double) Wittig condensation and a simpler elaboration of the resulting alkene. The synthesis of 2a illustrates a new approach that may be extended to other C-linked glycosyl serines starting from suitable sugar phosphoranes.

Acknowledgement. Financial support from the Ministero dell' Università e della Ricerca Scientifica e Tecnologica (Italy) is gratefully acknowledged. We thank Mr. P. Formaglio (University of Ferrara, Italy) for NMR measurements.

EXPERIMENTAL

All moisture-sensitive reactions were performed under a nitrogen atmosphere using oven-dried glassware. All solvents were dried over standard drying agents¹⁰ and freshly distilled prior to use. Commercially available powdered 4-Å molecular sieves (50 μ m average particle size) were used without further activation. Flash column chromatography¹¹ was performed on silica gel 60 (230-400 mesh). Reactions were monitored by TLC on silica gel 60 F₂₅₄ with detection by charring with sulfuric acid. Melting points were determined with a capillary apparatus and are uncorrected. Optical rotations were measured at 20 ± 2 °C in the stated solvent. ¹H (300 MHz) and ¹³C (75 MHz) NMR were recorded at r. t. for CDCl₃ solutions, unless otherwise specified. Assignments were aided by decoupling and/or homo- and heteronuclear two-dimensional experiments. MALDI-TOF mass spectra were acquired using α -cyano-4-hydroxycinnamic acid as the matrix.

2,6-Anhydro-3,4,5,7-tetra-*O***-benzyl-D***-glycero-L-manno***-heptitol** (4). To a stirred, cooled (0 °C) solution of aldehyde 3 (2.76 g, 5.0 mmol; >95% pure by ¹H NMR analysis at 140 °C in DMSO-d₆) in Et₂O (10 mL) and MeOH (10 mL) was added sodium borohydride (189 mg, 5.0 mmol). The mixture was stirred at 0 °C for 10 min, then diluted with acetone (2 mL) and concentrated. The residue was eluted from a short column of silica gel with 3:1 cyclohexane-AcOEt to give 4 (2.58 g, 93%) as a syrup; $[\alpha]_D = +2.2$ (*c* 1, CHCl₃). ¹H NMR: δ 7.41-7.23 (m, 20 H, 4 Ph), 4.97 and 4.61 (2 d, 2 H, J = 11.5 Hz, PhCH₂), 4.94 and 4.66 (2 d, 2 H, J = 10.7 Hz, PhCH₂), 4.78 and 4.71 (2 d, 2 H, J = 11.7 Hz, PhCH₂), 4.49 and 4.43 (2 d, 2 H, J = 12.0 Hz, PhCH₂), 3.95 (dd, 1 H, $J_{4.5} = 2.8$, $J_{5.6} = -0.5$ Hz, H-5), 3.95 (dd, 1 H, $J_{2.3} = 9.5$, $J_{3.4} = 9.6$ Hz, H-3), 3.87 (ddd, 1 H, $J_{1a,1b} = 11.5$, $J_{1a,2} = 2.8$, $J_{1a,OH} = 5.5$ Hz, H-1a), 3.72 (ddd, 1 H, $J_{1b,2} = 5.2$, $J_{1b,OH} = 7.5$ Hz, H-1b),

3.65 (dd, 1 H, H-4), 3.62-3.50 (m, 3 H), 3.36 (ddd, 1 H, H-2), 2.03 (dd, 1 H, OH). Anal. Calcd for C₃₅H₃₈O₆: C, 75.79; H, 6.91. Found: C, 76.10; H, 7.03.

2,6-Anhydro-3,4,5,7-tetra-*O*-benzyl-1-deoxy-1-iodo-D-glycero-L-manno-heptitol (5). A mixture of alcohol 4 (2.22 g, 4.0 mmol), triphenylphosphine (3.15 g, 12.0 mmol), imidazole (0.82 g, 12.0 mmol), iodine (2.03 g, 8.0 mmol), and anhydrous toluene (40 mL) was refluxed for 2 h, then cooled to r. t., diluted with Et₂O (50 mL), washed with 5% aqueous Na₂S₂O₃ (2 x 20 mL), and concentrated. The brown solid was triturated with Et₂O (50 mL) and filtered through a pad of Celite to remove most of crystalline triphenylphosphine oxide. The solution was concentrated and the residue was eluted from a column of silica gel with 7:1 cyclohexane-AcOEt to give 5 (2.05 g, 77%) as a syrup; $[\alpha]_D = -14.2$ (c 1, CHCl₃). ¹H NMR: δ 7.41-7.23 (m, 20 H, 4 Ph), 4.99 and 4.71 (2 d, 2 H, J = 11.0 Hz, PhCH₂), 4.97 and 4.67 (2 d, 2 H, J = 11.8 Hz, PhCH₂), 4.77 and 4.63 (2 d, 2 H, J = 11.5 Hz, PhCH₂), 4.54 and 4.45 (2 d, 2 H, J = 11.7 Hz, PhCH₂), 4.01 (dd, 1 H, $J_{4.5} = 2.5$, $J_{5.6} = -0.5$ Hz, H-5), 3.80 (dd, 1 H, $J_{2.3} = 9.2$, $J_{3.4} = 9.4$ Hz, H-3), 3.66-3.58 (m, 4 H), 3.54 (dd, 1 H, $J_{1a,1b} = 10.5$, $J_{1a,2} = 2.3$ Hz, H-1a), 3.30 (dd, 1 H, $J_{1b,2} = 7.0$ Hz, H-1b), 3.19 (ddd, 1 H, H-2). Anal. Calcd for C₃₅H₃₇IO₅: C, 63.26; H, 5.61. Found: C, 63.55; H, 5.46.

(2,6-Anhydro-3,4,5,7-tetra-O-benzyl-1-deoxy-D-glycero-L-manno-heptitol-1yl)triphenylphosphonium iodide (6). A mixture of iodide 5 (1.99 g, 3.0 mmol) and triphenylphosphine (3.93 g, 15.0 mmol) was heated with stirring at 120 °C under a nitrogen atmosphere for 2 h, then cooled to r. t., triturated with toluene (3 x 10 mL) and Et₂O (2 x 10 mL), and dried to give 6 (2.56 g, 92%) as a white amorphous solid; $[\alpha]_D = -37.7$ (c 1, CHCl₃). ¹H NMR: δ 7.75-7.50, 7.41-7.28, 7.14-7.10 (3 m, 35 H, 7 Ph), 5.03 and 4.95 (2 d, 2 H, J = 11.4 Hz, PhCH₂), 4.92 and 4.55 (2 d, 2 H, J = 11.5 Hz, PhCH₂), 4.75 and 4.69 (2 d, 2 H, J = 11.6 Hz, PhCH₂), 4.18 and 4.10 (2 d, 2 H, J = 11.5 Hz, PhCH₂), 3.85 (dd, 1 H, $J_{4,5} = 2.8$, $J_{5,6} = 0.8$ Hz, H-5), 3.70-3.57 (m, 1 H), 3.54 (dd, 1 H, $J_{3,4} = 9.3$ Hz, H-4), 3.45-3.28 (m, 3 H), 3.25 (ddd, 1 H, $J_{6,7a} = 4.8$, $J_{6,7b} = 6.8$ Hz, H-6), 3.12 (dd, 1 H, $J_{7a,7b} = 9.7$ Hz, H-7a), 3.05 (dd, 1 H, H-7b). Anal. Calcd for C₃₅H₃₇IO₅P: C, 68.68; H, 5.65. Found: C, 68.96; H, 5.67. The use of refluxing toluene, DMF at 120 °C or sulfolane at 200 °C as the solvent led to very low yield of phosphonium salt 6.

(Z/E)-5,9-Anhydro-6,7,8,10-tetra-O-benzyl-2,3,4-trideoxy-1,2-N,O-isopropylidene-2-(tert-butoxycarbonylamino)-D-threo-L-galacto-dec-3-enitol (8). To a stirred, cooled (-40 °C) mixture of phosphonium salt 6 (923 mg, 1.00 mmol), powdered 4-Å molecular sieves (1.00 g), anhydrous hexamethylphosphoramide (2 mL), and anhydrous THF (6 mL) was slowly added *n*butyllithium (400 μ L, 1.00 mmol, of a 2.5 solution in hexanes). After 5 min, to the resulting red-coloured suspension was slowly added a solution of the aldehyde 7 (228 mg, 1.00 mmol) in anhydrous THF (2 mL). The mixture was allowed to warm up to -10 °C in 2 h, then diluted with E₂O (100 mL) and filtered through a pad of Celite. The solution was washed with 1 M phosphate buffer at pH = 7 (30 mL), dried (MgSO₄), and concentrated. The residue was eluted from a column of silica gel with 6:1 cyclohexane-AcOEt to afford first (Z)-8 (405 mg, 54%) as a syrup; $[\alpha]_D =$ -22.3 (c 1, CHCl₃). ¹H NMR (C₂D₂Cl₄, 120 °C) selected data: δ 5.73 (dd, 1 H, J = 8.2, 11.3 Hz, CH=), 5.63 (dd, 1 H, J = 5.2, 11.3 Hz, CH=). MALDI-TOF MS: 773.4 (M⁺+Na), 789.4 (M⁺+K). Anal. Calcd for C₄₆H₅₅O₈N: C, 73.67; H, 7.39; N, 1.87. Found: C, 74.01; H, 7.48; N, 1.68. Eluted second was syrupy (E)-8 (60 mg, ~8%) contaminated by small amounts of uncharacterised byproducts. ¹H NMR (DMSO-d₆, 160 °C) selected data: δ 5.77 (dd, 1 H, J = 6.0, 16.0 Hz, CH=), 5.70 (dd, 1 H, J = 5.1, 16.0 Hz, CH=), 4.84 and 4.58 $(2 d, 2 H, J = 11.6 Hz, PhCH_2)$, 4.77 and 4.68 $(2 d, 2 H, J = 12.0 Hz, PhCH_2)$, 4.74 and 4.63 $(2 d, 2 H, J = 11.5 Hz, PhCH_2)$, 4.53 and 4.48 $(2 d, 2 H, J = 12.2 Hz, PhCH_2)$. MALDI-TOF MS: 773.2 (M⁺+Na), 789.2 (M⁺+K).

5,9-Anhydro-6,7,8,10-tetra-O-benzyl-2,3,4-trideoxy-1,2-N,O-isopropylidene-2-(tertbutoxycarbonylamino)-D-threo-L-galacto-decitol (9). To a stirred, warmed (85 °C) solution of alkene (Z)-8 (375 mg, 0.50 mmol) and freshly recrystallized p-toluenesulfonhydrazide (186 mg, 1.00 mmol) in dimethoxyethane (5 mL) was added a 1 M aqueous solution of sodium acetate (1.00 mL) in four portions during 2 h. After an additional 3 h at 85 °C the reaction mixture was diluted with H₂O (5 mL) and extracted with CH₂Cl₂ (2 x 30 mL). The organic phase was dried (MgSO₄) and concentrated. The residue was eluted from a column of silica gel with 6:1 cyclohexane-AcOEt to give 9 (316 mg, 84%) as a syrup; $[\alpha]_D = +4.8$ (c 1, CHCl₃). ¹H NMR (DMSO-d₆, 160 °C): δ 7.40-7.20 (m, 20 H, 4 Ph), 4.84 and 4.63 (2 d, 2 H, J = 11.5 Hz, PhCH₂), 4.84 and 4.57 (2 d, 2 H, J = 11.9 Hz, PhCH₂), 4.78 and 4.67 (2 d, 2 H, J = 12.0 Hz, PhCH₂), 4.53 and 4.48 (2 d, 2 H, J = 12.0 Hz, PhCH₂), 4.04 (dd, 1 H, $J_{7,8} = 2.7$, $J_{8,9} = -0.5$ Hz, H-8), 3.88 (dd, 1 H, $J_{1a,2} = 6.0$, $J_{1a,1b} = 8.5$ Hz, H-1a), 3.84-3.77 (m, 1 H, H-2), 3.70 (dd, 1 H, $J_{6,7} = 9.5$ Hz, H-7), 3.66-3.54 (m, 5 H), 3.22 (ddd, 1 H, $J_{4a,5} = J_{5,6} = 9.0$, $J_{4b,5} = 2.8$ Hz, H-5), 1.88-1.67 and 1.62-1.42 (2 m, 2 H-3, 2 H-4), 1.43 and 1.39 (2 s, 6 H, 2 Me), 1.40 (s, 9 H, tBu). Anal. Calcd for $C_{46}H_{57}O_8N$: C, 73.47; H, 7.64; N, 1.86. Found: C, 73.62; H, 7.74; N, 1.73. When the same reaction was performed using (E)-8 instead of (Z)-8 as the starting material, similar results were obtained.

5,9-Anhydro-6,7,8,10-tetra-O-benzyl-2,3,4-trideoxy-2-(*tert*-butoxycarbonylamino)-Dthreo-L-galacto-deconic acid (2a). To a stirred, cooled (0 °C) solution of 9 (301 mg, 0.40 mmol) in acetone (8 mL) was added freshly prepared 1 M Jones reagent (1.20 mL, 1.20 mmol). The mixture was allowed to warm to r. t. in 30 min, stirred at r. t. for an additional 2.5 h and then diluted with isopropanol (-0.5 mL). The suspension was neutralized with saturated aqueous NaHCO₃, diluted with Et₂O (100 mL) and washed with brine (2 x 20 mL). The organic phase was dried (MgSO₄) and concentrated to afford 2a (273 mg, -94%) contaminated by the amino alcohol 11 and other minor byproducts. Compound 2a showed ¹H and ¹³C NMR spectra consistent with those reported.^{1e} Prolonged reaction time or larger excess of Jones reagent led to lower yields of 2a due to acidic cleavage of the benzyl groups.

Methyl 5,9-Anhydro-6,7,8,10-tetra-O-benzyl-2,3,4-trideoxy-2-(tert-butoxycarbonylamino)-D-threo-L-galacto-deconate (10). Treatment of a solution of crude acid 2a in 1:1 Et₂O-MeOH with ethereal diazomethane at 0 °C for 5 min gave, after column chromatography on silica gel (4:1 cyclohexane-AcOEt), the ester 10 as a syrup, $[\alpha]_D = -4.3$ (c 1, CHCl₃). ¹H NMR: δ 7.40-7.25 (m, 20 H, 4 Ph), 5.04 (d, 1 H, J = 8.0 Hz, NH), 4.93 and 4.63 (2 d, 2 H, J = 11.6 Hz, PhCH₂), 4.93 and 4.62 (2 d, 2 H, J = 10.8 Hz, PhCH₂), 4.75 and 4.67 (2 d, 2 H, J = 11.7 Hz, PhCH₂), 4.47 and 4.41 (2 d, 2 H, J = 11.8 Hz, PhCH₂), 4.27-4.20 (m, 1 H, H-2), 3.98 (dd, 1 H, J_{7,8} = 2.6, J_{8,9} = ~0.5 Hz, H-8), 3.68 (s, 3 H, OMe), 3.66-3.47 (m, 5 H), 3.18 (ddd, 1 H, J_{4a,5} = J_{5,6} = 9.0, J_{4b,5} = 2.3 Hz, H-5), 1.95-1.75 (m, 3 H), 1.58-1.48 (m, 1 H), 1.41 (s, 9 H, *t*Bu). Anal. Calcd for C₄₄H₅₃O₉N: C, 71.42; H, 7.22; N, 1.89. Found: C, 71.20; H, 7.33; N, 1.78.

REFERENCES AND NOTES

- (a) Colombo, L.; Casiraghi, G.; Pittalis, A. J. Org. Chem. 1991, 56, 3987. (b) Petrus, L.; BeMiller, J. N. Carbohydr. Res. 1992, 230, 197. (c) Kessler, H.; Wittmann, V.; Köck, M.; Kottenhahn, M.Angew. Chem. Int. Ed. Engl. 1992, 31, 902. (d) Bertozzi, C. R.; Hoeprich, P. D.; Bednarski, M. D. J. Org. Chem. 1992, 57, 6092. (e) Bertozzi, C. R.; Cook, D. G.; Kobertz, W. R.; Gonzales-Scarano, F.; Bednarski, M. D. J. Am. Chem. Soc. 1992, 114, 10639. (f) Gurjar, M. K.; Mainkar, A. S.; Syamala, M. Tetrahedron: Asymmetry 1993, 4, 2343. (g) Axon, J. R.; Beckwith, A. L. J. J. Chem. Soc., Chem. Commun. 1995, 549. (h) Dorgan, B. J.; Jackson, R. F. W. Synlett 1996, 859. (i) Herpin, T. F.; Motherwell, W. B.; Weibel, J.-M. Chem. Commun. 1997, 923. (j) Burkhart, F.; Hoffmann, M.; Kessler, H. Angew. Chem. Int. Ed. Engl. 1997, 36, 1191. (k) Lay, L.; Meldal, M.; Nicotra, F.; Panza, L.; Russo, G. Chem. Commun. 1997, 1469. (l) Dondoni, A.; Junquera, F.; Merchan, F. L.; Merino, P.; Scherrmann, M.-C.; Tejero, T. J. Org. Chem. 1997, 62, 5484. (m) López-Herrera, F. J.; Sarabia-García, F.; Heras-López, A.; Pino-González, M. S. J. Org. Chem. 1997, 62, 6056. (n) Debenham, S. D.; Debenham, J. S.; Burk, M. J.; Toone, E. J. J. Am. Chem. Soc. 1997, 119, 9897.
- (a) Kunz, H. Angew. Chem. Int. Ed. Engl. 1987, 26, 294. (b) Kunz, H. Pure Appl. Chem. 1993, 65, 1223. (c) Meldal, M.; Bock, K. Glycoconjugate J. 1994, 11, 59. (d) Lee, Y. C.; Lee, R. T. Acc. Chem. Res. 1995, 28, 321. (e) Dwek, R. A. Chem. Rev. 1996, 96, 683. (f) Kunz, H. In Preparative Carbohydrate Chemistry, Hanessian, S., Ed., M. Dekker: New York, 1997, p 265. (g) G. Arsequell, G. Valencia, Tetrahedron Asymm. 1997, 8, 2839.
- It is worth recalling that the O-linked galactosyl-serine 1 is present in collagens and other associated structural glycopeptides. See: Lamport, D. T. A.; Katona, L.; Roering, S. Biochem. J. 1973, 133, 125. Muir, L.; Lee, Y. C. J. Biol. Chem. 1970, 245, 502.
- 4. Dondoni, A.; Scherrmann, M.-C. J. Org. Chem. 1994, 59, 6404.
- 5. Dondoni, A.; Marra, A.; Perrone, D. J. Org. Chem. 1993, 58, 275.
- 6. Garegg, P. J.; Samuelsson, B. J. Chem. Soc., Perkin I 1980, 2866.
- 7. For an improved synthesis of this α -amino aldehyde, see: Dondoni, A.; Perrone, D. Synthesis 1997, 527.
- 8. Dewey, R. S.; van Tamelen, E. E. J. Am. Chem. Soc. 1961, 83, 3729.
- We prepared a pure sample of this alcohol by acid hydrolysis of 9 (4:1 AcOH-H₂O, 80 °C, 15 min) in 70% yield after column chromatography (3:1 Et₂O-cyclohexane); mp 87-89 °C (from hexane), lit.^{1e} mp 80-81 °C;
 [α]_D = -11.6 (c 0.8, CHCl₃), previously^{1e} unreported. Anal. Calcd for C₄₃H₅₃O₈N: C, 72.55; H, 7.50; N, 1.97. Found: C, 72.40; H, 7.62; N, 1.90. The alcohol 11 proved to be identical by NMR analysis with compound 7 described in ref. le.



- Perrin, D. D.; Armarego, W. L. F. Purification of Laboratory Chemicals, 3rd ed.; Pergamon Press: Oxford, 1988.
- 11. Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.