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Vitamin D₃: Synthesis of *seco* C-9,11,21-*trishnor*-17-Methyl-1 α , 25-dihydroxyvitamin D₃ Analogues

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Abstract—The synthesis and biological activities of *seco* C-9,11,21-*trishnor*-17-methyl-1 α ,25-dihydroxyvitamin D₃ analogues (D-ring analogues) are described. © 2002 Elsevier Science Ltd. All rights reserved.

The observation that 1 α ,25-dihydroxy vitamin D₃ (**1**; calcitriol) is active in the regulation of cell proliferation and differentiation, next to the classical role in calcium-bone homeostasis, has led in recent years to the development of analogues capable of dissociating cell differentiating effects from calcemic effects.^{1,2} Among the three fragments of the vitamin D skeleton especially structural modifications of the side chain and of the A-ring have been studied in the past.³

Some years ago, we embarked on an extensive study of the structure–function relationship with the focus on the least studied part of the molecule, that is the central CD-ring regio.⁴ In this respect we decided stripping the molecule to its five-carbon backbone (C-8–C-20) and resubstituting it again in various ways. Presently we want to describe 21-*nor*, 17-methyl D-ring analogues lacking the six-membered C-ring, that is with general formula **3** and **4**. We decided to select a ‘D-ring’ carrying a *gem*-dimethyl group at C-13 (steroid numbering) as these substituents mimic respectively the angular C-18 methyl group and C-12 in the parent steroid **1**, which are known to have an influence on restricting the side chain orientations.^{3,5} It is generally assumed that the relative position in space of the 1 α - and 25-hydroxy groups is important for the biological activity and that the side chain occupies a very restricted topology at the binding site of the vitamin D receptor (VDR).^{3,5,6} This is also influenced by the 21-methyl group; it is of interest

to see if its absence can be compensated by a 17-methyl substituent. Also some 19-*nor* analogues **4** are described, as this structural feature lowers the toxic calcemic effect (**1** vs **2**).⁷

(1*S*,3*R*)-Camphoric acid **5** is an ideal template for the central fragment (D-ring). Construction of analogues will involve (i) chemoselective manipulation of the carboxylic functions, (ii) formation of side chains and (iii) producing the C-8 aldehyde function needed for coupling with A-ring precursors **6a**,⁸ and **6b** (Fig. 1).^{7a}

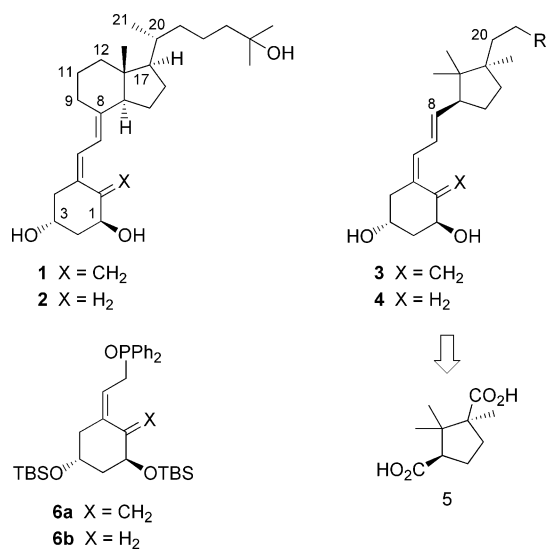


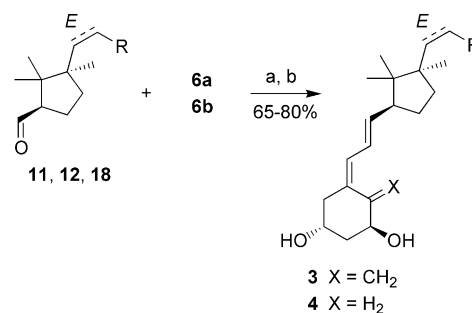
Figure 1.

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A highly efficient two-step differentiation of the carboxylic functions in **5** was attained via silylation of the corresponding diol **7** affording **8a** (87%) next to the di-TBDPS ether (5%) (Scheme 1). This method was far superior to an approach involving the C-8 mono-methyl ester (97%; MeOH, concd H₂SO₄, 30 °C) of **5** and selective reduction of both carboxylic functions. Introduction of the side chain via tosylate **8b** was impossible because all attempted substitutions at the sterically hindered C-20, failed. We therefore turned our attention to addition reactions on aldehyde precursor **9**. Julia coupling⁹ with the appropriate phenylsulfone (PhSO₂CH₂R) and reductive elimination¹⁰ led in good yields to the E-double bond isomers **10** (for respective side chain moieties R, see Scheme 2). The 25-hydroxy group in R is protected as a TES ether. Subsequent deprotection and oxidation gave the required C-8-formyl precursors **11** (for analogues WY 621, WY 624 and WY 903). For the synthesis of aldehyde **12** (synthesis of saturated side chain analogues KS 699, KS 703, WY 619, WY 625, WY 838 and WY 836) **10** was first hydrogenated.

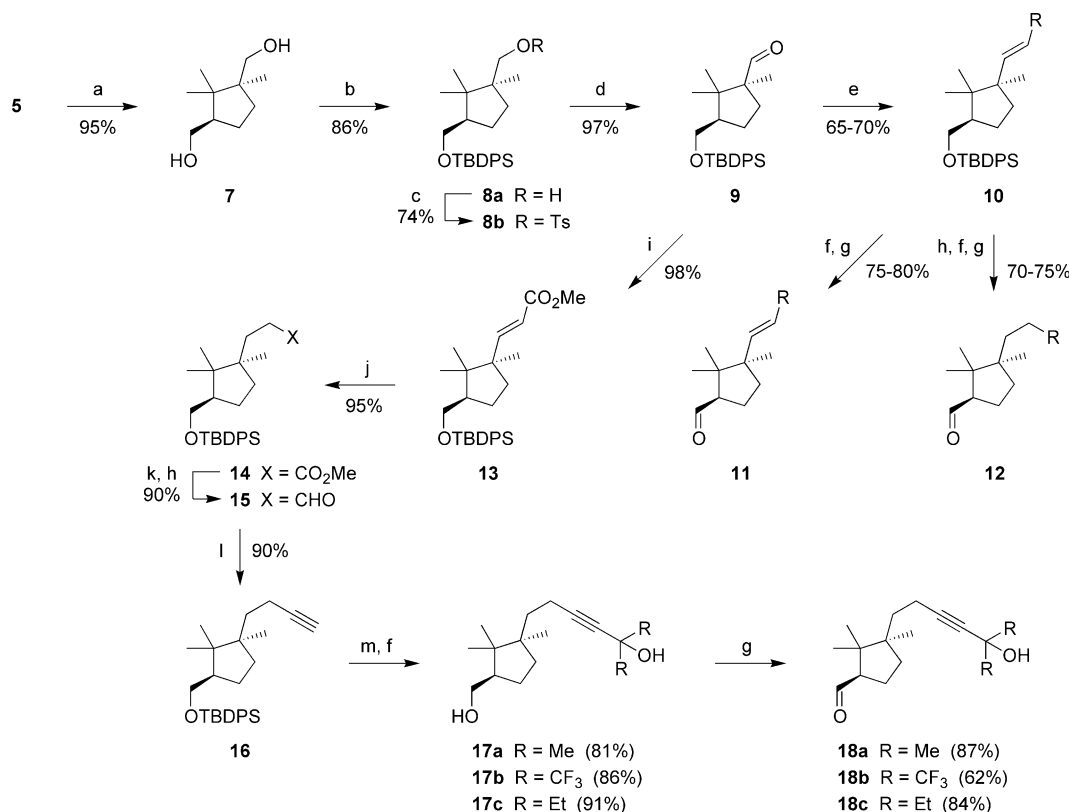
Of special interest are 23-yne vitamin D₃ analogues with a conformationally restricted side chain; they frequently display enhanced biological properties.³ In the present context their synthesis was performed via an initial Horner–Emmons reaction¹¹ on **9**. After catalytic hydrogenation of resulting **13**, the ester function in **14** was transformed into an aldehyde function in a two-

step sequence (reduction to the corresponding alcohol and Swern oxidation¹²). On large scale the direct Dibal-H reduction was troublesome as it invariably led to mixtures of **14**, **15** and the primary alcohol. Then,



4 (X = H ₂)	3 (X = CH ₂)	C ₂₀ -C ₂₂	R
WY 838	KS 699	single	(CH ₂) ₂ C(Me) ₂ OH
WY 836	KS 703	single	(CH ₂) ₂ C(Et) ₂ OH
-	WY 619	single	(CH ₂) ₂ C(CF ₃) ₂ OH
-	WY 625	single	CH ₂ -CF ₂ C(Me) ₂ OH
-	WY 621	double (E)	(CH ₂) ₂ C(CF ₃) ₂ OH
WY 903	WY 624	double (E)	CH ₂ -CF ₂ C(Me) ₂ OH
WY 906	WY 722	single	≡C-(Me) ₂ OH
CD 578	WY 718		≡C-(CF ₃) ₂ OH
-	WY 727	single	≡C-(Et) ₂ OH

Scheme 2. (a) *n*-BuLi, THF, -78 → -20 °C, 3 h; (b) TBAF, THF, rt, 12 h.



Scheme 1. (a) LiAlH₄, THF-Et₂O, rt, 5 h; (b) TBDPSCI, imidazole, DMF, 0 °C → rt, 5 h; (c) TsCl, Et₃N, DMAP, CH₂Cl₂, Δ, 12 h; (d) (COCl)₂, DMSO, CH₂Cl₂, -78 °C, then Et₃N, -78 → 0 °C; (e) (i) LDA, THF, -78 °C, 2 h; (ii) Ac₂O, Et₃N, DMAP, -78 °C → rt, 2 h; (iii) Na-Hg, (3.1 or 4.8%), Na₂HPO₄, MeOH, -40 → rt, 5 h; (f) TBAF, THF, rt, 4 h; (g) (COCl)₂, DMSO, CH₂Cl₂, then Et₃N, -78 °C → -10 °C, 4 h; (h) H₂, 5% Rh/Al₂O₃, H₂, EtOAc, rt, 2–4 h; (i) (EtO)₂P(O)CH₂CO₂CH₃, NaH, THF, 0 °C, 2 h; (j) 4 atm. H₂, 10% Pd/C, 3% EtOAc in hexane, rt, 15 h; (k) DIBALH, CH₂Cl₂, -78 °C, 3 h; (l) (MeO)₂P(O)CHN₂, *t*-BuOK, THF, -78 → 0 °C, 15 h; (m) LDA, R₂CO, THF, -40 → 10 °C, 3 h.

treatment of **15** with dimethyl diazophosphonate afforded alkyne **16**.¹³ The remaining carbon atoms of the side chain were introduced upon reaction of lithiated **16** with a ketone (Me₂CO for WY 722 and WY 906, (CF₃)₂CO for WY 718 and CD 578 and Et₂CO for WY 727). Deprotection and Swern oxidation then led to C-8 aldehydes **17**.

Finally, construction of the title compounds **3** and **4** involved Lythgoe coupling of aldehydes **11**, **12** and **17** with A-ring phosphine oxides **6a**⁸ and **6b**^{7a} followed by deprotection of the hydroxy functions (Scheme 2).

The coupling was performed on D-ring side chain fragments possessing a free 25-hydroxy function therefore an excess of **6a,b** (4–5 equiv) was used; the A-ring phosphine oxide could be recuperated. The combined yield of this two-step sequence was higher (65–80%) than for the alternative approach involving TES protection (40–50%).

The affinity to the pig intestinal mucosa vitamin D receptor (VDR) was evaluated as described previously.¹⁴ The relative affinity of the analogues was calculated from their concentration needed to displace 50% of [³H] 1 α ,25(OH)₂D₃ from its receptor compared with the activity of **1** (assigned a value of 100%). The affinity for VDR varied between 40 and 80% compared to **1**.

The in vivo calcemic effects were tested in vitamin D-replete normal NMRI mice by daily intraperitoneal injections of 1 α ,25(OH)₂D₃ **1**, the analogues or the solvent during 7 consecutive days, using serum calcium concentration as parameter. The biological evaluation was determined in vitro on different cell lines (HL 60, MCF-7, MG 63, keratinocytes). All results are the mean of at least 3 experiments and are expressed as percentage activity (at 50% dose response) in comparison with **1** (= 100% activity). All analogues were 10 to more than 100-fold less calcemic than 1 α ,25(OH)₂D₃. The antiproliferative activity was comparable to **1**, except

for the more potent fluorinated analogues; especially WY 718 and CD 578 with a 26,27-hexafluoro-23-yne side chain. The latter, a 19-*nor*-analogue, displays high ratios of cell antiproliferation activities versus calcemic effect. Further details of the biological activities and considerations obtained from comparison with other D-ring analogues will be published elsewhere (Table 1).

Acknowledgements

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References and Notes

- Bouillon, R.; Van Baelen, H. *Saudi Med. J.* **1989**, *10*, 260.
- DeLuca, H. F.; Burmester, J.; Darwish, H.; Krisinger, J. In *Comprehensive Medicinal Chemistry*; Pergamon: New York, 1990; vol. 3, p 1129.
- Bouillon, R.; Okamura, W. H.; Norman, A. W. *Endocr. Rev.* **1995**, *16*, 200.
- (a) Bouillon, R.; De Clercq, P.; Pirson, P.; Vandewalle, M. Novel structural analogues of vitamin D; patent PCT/EP 93.202037.3; priority date 09-07-1993 (b) Sabbe, K.; D'Halleweyn, C.; De Clercq, P.; Vandewalle, M.; Bouillon, R.; Verstuyf, A. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 1697. (c) Guidong, Z.; Yongjun, C.; Xiaoming, Z.; Vandewalle, M.; De Clercq, P.; Bouillon, R.; Verstuyf, A. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 1703. (d) Yong-Yung, C.; De Clercq, P.; Vandewalle, M. *Tetrahedron Lett.* **1996**, *52*, 9361. (e) Wu, Y.; Shi, L.; D'Halleweyn, C.; Van Haver, D.; De Clercq, P.; Vandewalle, M. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 928. (f) Linclau, B.; De Clercq, P.; Bouillon, R.; Verstuyf, A.; Vandewalle, M. *Bioorg. Med. Chem. Lett.* **1997**, *11*, 1461 and 1465. (g) Sas, B.; De Clercq, P.; Vandewalle, M. *Synlett* **1997**, *10*, 1167. (h) Verstuyf, A.; Verlinden, L.; Van Baelen, H.; Sabbe, K.; D'Halleweyn, C.; De Clercq, P.; Vandewalle, M.; Bouillon, R. *J. Bone Miner. Res.* **1998**, *13*, 549. (i) Zhou, X.; Zhu, G.-D.; Van Haver, D.; Vandewalle, M.; De Clercq, P. J.; Verstuyf, A.; Bouillon, R. *J. Med. Chem.* **1999**, *42*, 3539. (j) Verstuyf, A.; Verlinden, L.; Van Etten, E.; Shi, L.; Wu, Y.; D'Halleweyn, C.; Van Haver, D.; Zhu, G.-D.; Chen, Y.-J.; Zhou, X.; Haussler, M. R.; De Clercq, P.; Vandewalle, M.; Van Baelen, H.; Mathieu, C.; Bouillon, R. *J. Bone Miner. Res.* **2000**, *2*, 237; for an analogous approach, see: Kutner, A.; Zhao, H.; Fitak, H.; Wilson, S. R. *Bioorg. Chem.* **1995**, *23*, 22. Hilpert, H.; Wirz, B. *Tetrahedron* **2001**, *57*, 681.
- (a) Okamura, W. H.; Midland, M. M.; Hammond, M. W.; Rahman, N. A.; Dormanen, M. C.; Nemere, I.; Norman, A. W. *J. Steroid Biochem. Mol. Biol.* **1995**, *53*, 603. (b) Van Haver, D.; De Clercq, P. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 1029.
- Rochel, N.; Wurtz, J. M.; Mitscher, A.; Klaholz, B.; Moras, D. *Mol. Cell* **2000**, 173.
- (a) Perlman, K. L.; Sicinski, H. K.; DeLuca, H. F. *Tetrahedron Lett.* **1990**, *31*, 1823. (b) Bouillon, R.; Sarandeses, L. A.; Allewaert, K.; Zhao, J.; Mascarenas, J. L.; Mourino, A.; Vrielynck, S.; De Clercq, P.; Vandewalle, M. *Bone Miner. Res.* **1993**, *8*, 1009.
- Baggiolini, E. G.; Iacobelli, J. A.; Hennessy, B. M.; Batcho, A. D.; Sereno, J. F.; Uskokovic, M. R. *J. Org. Chem.* **3098**, 51.
- Julia, M.; Paris, J. M. *Tetrahedron Lett.* **1973**, *49*, 4833.
- Roush, W. R.; Russo-Rodrigues, S. *J. Org. Chem.* **1985**, *50*, 5465.
- Tanner, D.; Somfai, P. *Tetrahedron* **1987**, *43*, 4395.

Table 1. Biological activities

Compd	In vitro studies					In vivo studies
	VDR	HL60	MG-63	MCF-7	Keratinocytes	Ca serum
1	100	100	100	100	100	100
KS 699	30	40	60	80	40	<1
WY 838	40	9	40	30	10	0.25
KS 703	80	100	200	250	90	5
WY 836	75	85	80	100	40	0.25
WY 619	80	150	200	1250	750	10
WY 625	75	80	50	90	1350	3
WY 621	80	50	50	375	300	13
WY 624	40	20	12	70	200	<0.1
WY 903	70	20		10	60	0.5
WY 722	9	100	50	150	400	0.1
WY 906	60	60		70	85	0.25
WY 718	80	215	350	3500	3500	1
CD 578	100	300		2000	4500	1
WY 727	70	90	20	150	90	1

12. Omura, K.; Swern, D. *Tetrahedron* **1978**, *34*, 1651.
13. (a) Seyferth, D.; Marmor, R. S.; Hilbert, P. *J. Org. Chem.* **1971**, *36*, 1379. (b) Gilbert, J. C.; Weeracoriva, U. *J. Org. Chem.* **1979**, *44*, 4997.
14. Bouillon, R.; Allewaert, K.; Van Leeuwen, J. P. T. M.; Tan, B. K.; Xiang, D. Z.; De Clercq, P.; Vandewalle, M.; Pols, H. A. P.; Bos, M. P.; Van Baelen, H.; Birkenhäger, J. C. *J. Biol. Chem.* **1992**, *267*, 3044.