## Mesoionic Xanthine Analogues: Antagonists of Adenosine Receptors

Richard A. Glennon,\*<sup>†</sup> Shanaz M. Tejani-Butt,<sup>†</sup> William Padgett,<sup>‡</sup> and John W. Daly\*<sup>‡</sup>

Department of Medicinal Chemistry, Medical College of Virginia, Virginia Commonwealth University, Richmond, Virginia 23298, and Laboratory of Bioorganic Chemistry, Bldg. 4, Room 212, National Institute of Arthritis, Diabetes, and Digestive and Kidney Diseases, National Institutes of Health, Bethesda, Maryland 20205. Received February 21, 1984

A variety of mesoionic xanthines including mesoionic thiazolo[3,2- $\alpha$ ]pyrimidines, benzothiazolopyrimidines, and 1,3,4-thiadiazolo[3,2- $\alpha$ ]pyrimidines were antagonists of A<sub>1</sub>-adenosine receptors (inhibition of binding of [<sup>3</sup>H]-cyclohexyladenosine) and A<sub>2</sub>-adenosine receptors (inhibition of 2-chloroadenosine-elicited accumulations of cyclic AMP) in brain tissue. Most of the compounds were less potent than theophylline and none were remarkably selective for A<sub>1</sub>- or A<sub>2</sub>-adenosine receptors. However, members of the thiadiazolopyrimidine class of mesoionics exhibited very low or no activity as antagonists of A<sub>2</sub>-adenosine receptors while exhibiting activity only 2-4-fold lower than that of theophylline at A<sub>1</sub>-adenosine receptors. Unlike the case for theophylline, the presence of a phenyl substituent in the five-membered ring did not enhance the potency of a mesoionic thiadiazolopyrimidine. The nature of the substituents on the mesoionic ring did not appear to have marked effects on potency unlike the marked effect of the nature of 1,3-substituents on activity of nonmesoionic xanthines. The benzothiazolo[3,2- $\alpha$ ]pyrimidines were the most potent antagonists, being nearly as potent as theophylline at A<sub>1</sub>-adenosine receptors and somewhat more potent than theophylline at A<sub>2</sub>-adenosine receptors.

Theophylline and other alkylxanthines were considered for decades to owe their pharmacological effects primarily to inhibition of cyclic nucleotide phosphodiesterases and a resultant elevation of cyclic AMP and/or cyclic GMP levels. It is now realized that theophylline and certain other xanthines are more potent as antagonists of adenosine receptors than they are as inhibitors of phosphodiesterases.<sup>1</sup> One class of adenosine receptors  $(A_1)$  is inhibitory to adenylate cyclase; blockade of this receptor is probably involved in the lipolytic<sup>2</sup> and central stimulation<sup>3</sup> activity of theophylline and other xanthines. The other class of adenosine receptors  $(A_2)$  is stimulatory to adenylate cyclase. Blockade of this receptor by xanthines could, under certain conditions, increase heart rate<sup>4</sup> and blood pressure,<sup>4b</sup> reduce coronary blood flow,<sup>5</sup> and reduce hormone production.<sup>6,7</sup> Antagonism of adenosine receptors might also be involved in the diuretic effects produced by xanthines.<sup>8</sup> At present, the xanthines are the only major class of adenosine receptor antagonists. In a search for more potent and/or selective antagonists, a series of mesoionic xanthine analogues was investigated. Such mesoionic analogues do, like the xanthines, inhibit phosphodiesterases. $^{9-12}$  In the current study, inhibition of binding of [<sup>3</sup>H]cyclohexyladenosine to rat cerebral cortical membranes<sup>13,14</sup> was used to assess the potency of mesoionic analogues at an A<sub>1</sub>-adenosine receptor. Inhibition of 2chloroadenosine-elicited accumulations of cyclic AMP in guinea pig cerebral cortical slices<sup>15</sup> was used to assess potency of mesoionic analogues as antagonists of an A<sub>2</sub>adenosine receptor. A potent phosphodiesterase inhibitor (rolipram)<sup>16</sup> was included in the latter protocol to eliminate any possible effects of phosphodiesterase inhibition by the mesoionic analogues.

**Chemistry.** The synthesis of most of the mesoionic xanthines analogues has been previously reported;<sup>9-12</sup> for the most part, they were prepared by condensation of the appropriately substituted alkylamino heterocycle with bis(2,4,6-trichlorophenyl) malonate or bis(2,4,6-trichlorophenyl) alkylmalonate. The condensation is described for the preparation of the novel derivative **2b**. The thiadia-zolopyrimidine derivative **3a** was prepared by condensation of 2-(ethylamino)-1,3,4-thiadiazole with carbon suboxide.

<sup>‡</sup>National Institute of Arthritis, Diabetes, and Digestive and Kidney Diseases.

The synthesis of **3d** required 2-(ethylamino)-5-phenyl-1,3,4-thiadiazole (8); 4-ethyl-3-thiosemicarbazide was allowed to react with trimethyl orthobenzoate in the presence of an acid catalyst to afford a white crystalline material (mp 173–175 °C). Although the spectral data obtained of this product were consistent with what might be anticipated for 8, Chandra et al.<sup>17</sup> had previously reported that treatment of benzal thiosemicarbazone with ferric chloride affords 8 with a melting point of 238–240 °C. Ortho ester cyclizations of alkylthiosemicarbazides can yield mixtures of thiadiazoles and mercaptotriazoles,<sup>18</sup> and although our product was distinctly different from 3phenyl-5-mercapto-1,2,4-triazole (mp 141–142 °C),<sup>19</sup> the

- (2) Londos, C.; Cooper, D. M. F.; Schlegel, W.; Rodbell, M. Proc. Natl. Acad. Sci. U.S.A. 1978, 75, 5362.
- (3) Snyder, S. H.; Katims, J. J.; Annau, Z.; Bruns, R. F.; Daly, J. W. Proc. Natl. Acad. Sci. U.S.A. 1981, 78, 3260.
- (4) (a) Evans, D. B.; Schenden, J. A.; Bristol, J. A. Life Sci. 1982, 31, 2425. (b) Von Borstal, R. W.; Wurtman, R. J.; Conlay, L. A. Ibid. 1983, 32, 1151.
- (5) Alfonso, S. Circ. Res. 1970, 26, 743.
- (6) Londos, C.; Cooper, D. M. F.; Wolff, J. Proc. Natl. Acad. Sci. U.S.A. 1980, 77, 2551.
- (7) Petrack, B.; Czernik, A. J.; Ansell, J.; Cassidy, J. Life Sci. 1981, 28, 2611.
- (8) Osswald, H. Nauyn-Schmiedeberg's Arch. Pharmacol. 1975, 288, 79.
- (9) Glennon, R. A.; Rogers, M. E.; Bass, R. G.; Ryan, S. B. J. Pharm. Sci. 1978, 67, 1762.
- (10) Glennon, R. A.; Rogers, M. E.; Smith, J. D.; El-Said, M. K.; Egle, J. L. J. Med. Chem. 1981, 24, 658.
- (11) Glennon, R. A.; Gaines, J. J.; Rogers, M. E. J. Med. Chem. 1981, 24, 766.
- (12) Rogers, M. E.; Glennon, R. A.; Smith, J. D.; Boots, M. R.; Nanavati, N.; Maconoughey, J. E.; Aub, D.; Thomas, S.; Bass, R. G.; Mbagwu, G. J. Med. Chem. 1981, 24, 1284.
- (13) Bruns, R. F.; Daly, J. W.; Snyder, S. H. Proc. Natl. Acad. Sci. U.S.A. 1980, 77, 5547.
- (14) Daly, J. W.; Butts-Lamb, P.; Padgett, W. Cell. Mol. Neurobiol. 1983, 3, 69.
- (15) Smellie, F. W.; Davis, C. W.; Daly, J. W.; Wells, J. N. Life Sci. 1979, 24, 2475.
- (16) Schwabe, U.; Miyake, M.; Ohga, Y.; Daly, J. W. Mol. Pharmacol. 1976, 12, 900.
- (17) Chandra, D. S.; Choudhury, R. S. K. Chem. Abstr. 1928, 22, 4123.
- (18) Coburn, R. A.; Bhooshan, B.; Glennon, R. A. J. Org. Chem. 1973, 38, 3947.
- (19) Shah, M. H.; Mhasalkar, M. Y.; Patki, V. M.; Deliwala, C. V.; Sheth, U. K. J. Pharm. Sci. 1969, 58, 1398.

0022-2623/84/1827-1364\$01.50/0 © 1984 American Chemical Society

<sup>&</sup>lt;sup>†</sup>Virginia Commonwealth University.

<sup>(1)</sup> Daly, J. W. J. Med. Chem. 1982, 25, 197.

					$\mathrm{IC}_{50}$ , <sup>a</sup> $\mu\mathrm{M}$		
no.	R	R′	R″	R‴	A <sub>1</sub> receptor	A <sub>2</sub> receptor	
			Xanthine	s			
1a 1b 1c 1d 1e	${}^{CH_3}_{C_2H_5}_{n-C_3H_7}_{CH_2C_6H_5}_{CH_3}_{CH_4}$	$\begin{array}{c} \mathrm{CH}_3\\ \mathrm{C}_2\mathrm{H}_5\\ n\text{-}\mathrm{C}_3\mathrm{H}_7\\ \mathrm{CH}_2\mathrm{C}_6\mathrm{H}_5\\ \mathrm{CH}_3\\ \mathrm{CH} \end{array}$	H H H CH <sub>3</sub>	H H H H C.H.	$28 \pm 6  6.4 \oplus 0.4  1.5 \pm 6  7 \pm 2  110 \pm 22  0.8 \pm 0.3 $	$42 \pm 6 \\ 12 \pm 2 \\ 8.1 \pm 2.4 \\ 35 \pm 5 \\ 150 \pm 15 \\ 1.65 \pm 0.12$	
	0113	City	Thiezolopyrim	uidines	0.0 - 0.0		
			I mazotopyrm	numes			
2a 2b 2c 2d 2e 2f 2g	$C_2H_5 \\ n-C_3H_7 \\ C_2H_5 \\ C_2H_5 \\ CH_2C_6H_5 \\ C_2H_5 \\ C_2H$	$C_{2}H_{5}$ $n-C_{3}H_{7}$ $CH_{2}C_{3}H_{5}$ $CH_{2}C_{6}H_{5}$ $CH_{2}C_{3}H_{5}$ $CH_{2}C_{3}H_{5}$ b	Н Н Н С¢Н₅ Н	H H H H H H	$120 \\ 54 \pm 6 \\ 100 \\ 160 \\ 160 \\ 120 \\ 95$	150 250 120 150 200 200 250	
			Thiadiazolopyri	midines			
3a 3b 3c 3d 3e 3f 3g 3h	$\begin{array}{c} {\rm H} \\ {\rm CH}_3 \\ {\rm C}_2 {\rm H}_5 \\ {\rm C}_2 {\rm H}_5 \\ {\rm CH}_2 {\rm C}_6 {\rm H}_5 \\ {\rm CH}_2 {\rm C}_6 {\rm H}_4 (\rm 4{\ Cl}) \\ {\rm CH}_3 \\ {\rm C}_2 {\rm H}_5 \end{array}$	$C_{2}H_{5}$ $C_{2}H_{5}$ $C_{2}H_{5}$ $C_{2}H_{5}$ $C_{2}H_{5}$ $C_{2}H_{5}$ $n-C_{5}H_{11}$ $C_{2}H_{5}$		H H C <sub>6</sub> H₅ H H H CH₃	$110 \pm 10 \\ 60 \pm 30 \\ 72 \pm 20 \\ 80 \pm 40 \\ 58 \pm 2 \\ 45 \bullet 7 \\ 120 \pm 30 \\ 500$	$\begin{array}{c} \gg 250 \ (0\%) \\ \gg 250 \ (0\%) \\ \gg 250 \ (15\%) \\ 250 \\ \gg 200 \ (10\%) \\ > 250 \ (35\%) \\ \gg 250 \ (5\%) \\ \gg 250 \ (0\%) \end{array}$	
		I	Benzothiazolopy	rimidines			
4a 4b 4c 4d	$\begin{array}{c} \mathbf{C_2H_5}\\ \mathbf{C_2H_5}\\ \textbf{\textit{n-C_3H_7}}\\ \mathbf{C_2H_5} \end{array}$	CH2C3H5 CH2C3H5 i-C4H9 i-C4H9	H OC <sub>2</sub> H <sub>5</sub> H H		$37 \pm 3$ $20 \pm 5$ 30 $41 \pm 4$	$18 \pm 5$ 54 20 15 \pm 5	
			Triazolopyrin	nidine			
5	n-C <sub>3</sub> H <sub>7</sub>	CH3			≫250 (20%)	≫250 (5%)	
			Imidazothia	zine			
6	$C_2H_5$				$290 \pm 20$	≫250 (25%)	
			Isoquinopyrin	nidine			
 7	$C_2H_5$	$C_2H_5$			27	30	

Table I. Effects of Xanthines and Various Mesoionic Xanthine Analogues on A1- and A2-Adenosine Receptors

<sup>a</sup> IC<sub>50</sub> values for A<sub>1</sub> receptors were obtained from antagonism of binding of 1 nM [<sup>3</sup>H]cyclohexyladenosine to rat cerebral cortical membranes. IC<sub>50</sub> values for A<sub>2</sub>-adenosine receptors were obtained from antagonism of accumulations of cyclic AMP elicited by 15  $\mu$ M 2chloroadenosine in guinea pig cerebral cortical slices. Values are from single determination or are means  $\bullet$  SEM for two to three determinations. Percentages in parentheses indicate the percent inhibition at the highest concentration tested. <sup>b</sup>R' = 3,5-dimethoxybenzyl.

identity of 8 was verified by another synthetic route. 2-Amino-5-phenyl-1,3,4-thiadiazole was prepared by a literature procedure,<sup>20</sup> and acylated with acetic anhydride<sup>21</sup> and the resultant amide reduced with LiAlH<sub>4</sub> to afford 8 with a melting point of 173–174 °C. Compound 8 was cyclized to 3d by condensation with bis(2,4,6-trichlorophenyl) ethylmalonate.

## **Results and Discussion**

**Xanthines.** Theophylline (1a) is nearly equipotent as an antagonist at  $A_1$ - and  $A_2$ -adenosine receptors<sup>14</sup> (Table I). Addition of a methyl group at the 7-position (R") yields caffeine (1e) and reduces potency about 3-fold at both  $A_1$ - and  $A_2$ -adenosine receptors. A phenyl group at the 8-position (R") of theophylline (i.e., 1f) greatly increases potency at  $A_1$ - and  $A_2$ -receptors<sup>13,15,22-24</sup> (Table I).

- (21) Kubota, S.; Yasufumi, U.; Kazuichi, F.; Toyooka, K.; Shibuya, M. J. Org. Chem. 1980, 45, 1473.
- (22) Bruns, R. F. Biochem. Pharmacol. 1981, 30, 325.
- (23) Fredholm, B. B.; Persson, C. G. Eur. J. Pharmacol. 1982, 81, 673.
- (24) Bruns, R. F.; Daly, J. W.; Snyder, S. H. Proc. Natl. Acad. Sci. U.S.A. 1983, 80, 2077.

The nature of substituents at the 1- and 3-positions (R, R') of xanthines has significant effects on potency and selectivity at  $A_1$ - and  $A_2$ - adenosine receptors. The 1,3-diethyl derivative 1b is 5-fold more potent than theophylline at both  $A_1$  and  $A_2$  receptors (Table I). The 1,3-dipropyl analogue of theophylline (i.e., 1c) is 20-fold more potent at  $A_1$  receptors but only 5-fold more potent at  $A_2$  receptors than theophylline. The 1,3-dibenzyl analogue 1d is 4-fold more potent at  $A_1$  receptors than theophylline while being equipotent with theophylline at  $A_2$  receptors. Further investigation of structure-activity relations for xanthine antagonists is in progress.

**Mesoionic Analogues.** Most of the mesoionic analogues are less potent than theophylline as adenosine antagonists. Whether or not this reflects the effect of the mesoionic ring or the effect of alterations in the fivemembered ring on receptor affinity is unknown. There have been only limited studies on effects of replacement of N<sub>7</sub> and N<sub>9</sub> nitrogen atoms of xanthines with sulfur or carbon on potencies at adenosine receptors. 8-Azatheophylline, for example, was virtually inactive as an antagonist at A<sub>2</sub>-adenosine receptors of fibroblasts, while 1ethyl-3-propyl-7-thiaxanthine was 10-fold less potent than 1,3-diethylxanthine.<sup>22</sup> 9-Oxa-8-phenyltheophylline was at

<sup>(20)</sup> Hoggarth, E. J. Chem. Soc. 1949, 1163.



least 500 times less active than 8-phenyltheophylline. One direct comparison of the effect of structural modification of the five-membered ring on activity can be made for a xanthine and a mesoionic xanthine analogue; in the case of the mesoionic triazolopyrimidine 5, a methyl substituent is present at what would be the xanthine 9-position. Compound 5 is virtually inactive at adenosine receptors (Table I) as are the corresponding 9-methylxanthines isocaffeine and 1,9-dimethylxanthine.<sup>3,22</sup> The weakly active mesoionic imidazothiazine 6 also possesses an alkyl group at the xanthine 9-position.

A series of seven mesoionic thiazolopyrimidines, 2, with various substituents were evaluated (Table I). Compounds 2a-2g were considerably weaker than theophylline, and, in contrast to the marked effects of substituents on activity in the 1 series, substituent effects in series 2 were less pronounced. Nevertheless, dipropyl derivative 2b was, as in the 1 series, more active than its diethyl derivative 2a at A<sub>1</sub> sites. Compound 2b also displays selectivity for A<sub>1</sub> sites.

Benz-fusion of the series 2 compounds affords mesoionic benzothiazolopyrimidines 4. These compounds are as potent or are slightly more potent than theophylline (1a) as adenosine antagonists and, with the exception of the ethoxy-substituted derivative 4b, exhibit some selectivity for  $A_2$  sites. There was little effect of substituent variation on activity, although, admittedly, substituent selection was not very large.

Several of the mesoionic thiadiazolopyrimidines 3 were quite selective for A<sub>1</sub>-adenosine receptors, showing nearly no activity at A<sub>2</sub>-adenosine receptors at the highest concentration tested (250  $\mu$ M). Remarkably, the thiadiazolopyrimidine 3a was as active at  $A_1$  receptors as was 3b; in a corresponding nonmesoionic xanthine pair, 3methylxanthine is about 120-fold less active than theophylline.<sup>24</sup> Increasing the size of the xanthine 3-position substituent from ethyl (3b) to *n*-pentyl (3g) had only a slight effect on activity. The presence of the 4-chlorobenzyl substituent of 3f resulted in a relatively potent  $A_1$ adenosine antagonist, which now has significant, albeit weak, activity as an A<sub>2</sub> antagonist. The presence of a methyl group in the five-membered ring of 3c (i.e., 3h) is not tolerated, 3h being virtually inactive. This is remarkable, since in the case of theophylline, the presence of a methyl substituent in an equivalent position (i.e., 8-methyltheophylline) has little effect on the potency at either  $A_1$  (unpublished results) or  $A_2$  receptors.<sup>22</sup> An 8phenyl group in theophylline (i.e., 1f) increases potency by 35-fold, and again the lack of corresponding effect when the potency of 3c is compared with that of 3d is remarkable. Possibly, the sharper bond angle of the C-S-C portion of the thiadiazole ring of **3h** and **3d**, imparted by the presence of the sulfur atom, might locate the methyl or phenyl group in a somewhat different orientation than that in 8-methyltheophylline or 1f, respectively.

The mesoionic thiazine analogue 6 was essentially devoid of activity, while the mesoionic isoquinopyrimidine 7 was equipotent with theophylline.

Nearly all of the mesoionic compounds discussed herein have been previously examined as inhibitors of cyclic-AMP phosphodiesterase (PDE).<sup>9-12</sup> In general, the benzothiazolopyrimidines and thiadiazolopyrimidines are more active than their thiazolopyrimidine counterparts, while the mesoionic triazolopyrimidine 5 and the methyl-substituted thiadiazolopyrimidine 3h are inactive. There does not appear to exist, for these few compounds, a significant relationship between the data in Table I and the ability of these agents to inhibit cAMP PDE.<sup>9-12</sup>

The data indicate some analogies and some differences between the structure-activity requirements for mesoionics and the structure-activity requirements for xanthines as adenosine antagonists. Further comparisons are required, and at that time speculations as to the nature of interactions of the receptor surface with the planar heterocyclic ring of adenosine antagonists (xanthines, mesoionics, pteridines,<sup>22</sup> pyrazolopyridines,<sup>25-27</sup> pyrazolopyrimidines,<sup>28</sup> etc.) may be possible.

## **Experimental Section**

Proton magnetic resonance (<sup>1</sup>H NMR) spectra were recorded on a Perkin-Elmer R-24 high-resolution spectrometer and chemical shifts are reported relative to Me<sub>4</sub>Si as an internal standard. Infrared spectra were obtained on a Perkin-Elmer 257 spectrophotometer and mass spectra were determined on a Finnigan 4000 series GC/MS. Spectral data were consistent with the assigned structures. Melting points were determined on a Thomas-Hoover melting point apparatus and are uncorrected. Elemental analyses were performed by Atlantic Microlab, Atlanta, GA, and determined values were within 0.4% of theoretical. Most of the compounds used in this study were those previously reported, or they were synthesized according to literature procedures.<sup>9-12</sup>

Anhydro-6,8-di-*n*-propyl-5-hydroxy-7-oxothiazolo[3,2-*a*]pyrimidinium Hydroxide (2b). 2-(*n*-Propylamino)thiazole (0.1 g, 0.7 mmol) and bis(2,4,6-trichlorophenyl) *n*-propylmalonate (0.35 g, 0.7 mmol) were heated, neat, at 160 °C, under a stream of N<sub>2</sub>, until a clear melt resulted (ca. 3 min). When cool, the resultant yellow oil was triturated with anhydrous Et<sub>2</sub>O (20 mL) and the crude solid product was collected by filtration. Recrystallization from EtOAc afforded 0.08 g (45%) of **2b** as white crystals, mp 128-130 °C. Anal. (C<sub>12</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub>S) C, H, N.

An hydro-8-ethyl-5-hydroxy-7-oxo-1,3,4-thiadiazolo[3,2a]pyrimidinium Hydroxide (3a). Dibromomalonyl dichloride<sup>29</sup> (3.0 g, 10 mmol) in anhydrous  $Et_2O$  (30 mL) was added dropwise to zinc shavings (2.0 g), under a slow stream of N<sub>2</sub>, at such a rate that the stirred reaction mixture boiled gently. The carbon suboxide that was formed was bubbled into a solution of 2-(ethylamino)-1,3,4-thiadiazole (0.05 g, 0.38 mmol) in anhydrous  $Et_2O$  (5 mL) to yield an almost instantaneous precipitate of crude 3a. The solid was collected by filtration and recrystallized from MeCN to afford 0.07 g (92%) of 3 as white crystals, mp 208-210 °C (lit.<sup>30</sup> 208-209 °C).

**2-(Ethylamino)-5-phenyl-1,3,4-thiadiazole (8).** Method A. Concentrated HCl (0.05 mL) was added to a solution of 4-ethyl-3-thiosemicarbazide (1.19 g, 10 mmol) and trimethyl orthobenzoate (3.6 g, 20 mmol) in 95% EtOH (15 mL). The reaction mixture was stirred at room temperature for 1.5 h, heated at reflux for 1.5 h, and allowed to cool. The solvent was evaporated under

- (25) Psychoyos, S.; Ford, C. J.; Phillips, M. A. Biochem. Pharmacol. 1982, 31, 1441.
- (26) Williams, M.; Risley, E. A.; Huff, J. R. Can. J. Physiol. Pharmacol. 1981, 59, 897.
- (27) Murphy, K. M. M.; Snyder, S. H. Life Sci. 1981, 28, 917.
- (28) Davies, L. P.; Chow, S. C.; Skerritt, J. H.; Brown, D. J.; Johnston, G. A. R. Life Sci. 1984, 34, 2117.
- (29) Staudinger, H.; Bereza, S. Chem. Ber. 1908, 41, 4461.
- (30) Coburn, R. A.; Glennon, R. A. J. Heterocycl. Chem. 1973, 10, 487.

reduced pressure to near dryness to yield a crude white product; this product was collected by filtration, washed well with petroleum ether, and recrystallized from 95% EtOH to afford 0.65 g (30%) of 8: mp 173-175 °C; IR (KBr) 3200 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>) § 7.9-7.5 (m, 5, arom H), 7.6 (br s, 1, NH), 3.5 (q, 2, CH<sub>2</sub>), 1.3 (t, 3, CH<sub>3</sub>).

Method B. A solution of 2-acetamido-5-phenyl-1,3,4-thiadiazole<sup>21</sup> (0.2 g, 1 mmol) in THF (15 mL) was added dropwise to a stirred suspension of LiAlH<sub>4</sub> (0.08 g, 2 mmol) in THF (15 mL) at 0 °C. The reaction mixture was heated at reflux for 3 h and cooled to 0 °C and the excess LiAlH<sub>4</sub> destroyed by the successive dropwise addition of H<sub>2</sub>O (1 mL), 15% aqueous NaOH (1.5 mL), and  $H_2O$  (3 mL). The mixture was filtered, the filtrate was dried  $(MgSO_4)$  and evaporated to dryness to afford 0.09 g (40%) of 8, mp 173–174 °C after recrystallization from 95% EtOH. Anal. (C<sub>10</sub>H<sub>11</sub>N<sub>3</sub>S) C, H, N.

Anhydro-2-phenyl-6,8-diethyl-5-hydroxy-7-oxo-1,3,4-thiadiazolo[3,2-a]pyrimidinium Hydroxide (3d). Bis(2,4,6-trichlorophenyl)ethylmalonate (0.39 g, 0.8 mmol) and 8 (0.2 g, 0.8 mmol) were heated, neat, at 160 °C until a clear melt resulted (ca. 5 min). The cooled product was triturated with anhydrous  $Et_2O$  (20 mL) and collected by filtration. Recrystallization from *i*-PrOH yielded 0.17 g (98%) of **3d** as pale yellow crystals: mp 233–235 °C; IR (KBr) 1685, 1645 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.0–7.6 (m, 5, arom H), 4.25 (q, 2, CH<sub>2</sub>), 2.6 (q, 2, NCH<sub>2</sub>), 1.6 (t, 3 CH<sub>3</sub>), 1.1 (t, 3, NCH<sub>2</sub>CH<sub>3</sub>). Anal. ( $C_{15}H_{15}N_3SO_2$ ) C, H, N.

Biochemical Assay. Inhibition of binding of 1 nM [<sup>3</sup>H]cyclohexyladenosine (New England Nuclear Corp.) to A1adenosine receptors in rat cerebral cortical membranes was assayed as described.<sup>14</sup> The  $K_D$  for [<sup>3</sup>H]cyclohexyladenosine was about 1 nM. Inhibition of binding by a range of concentrations of each compound was assessed in triplicate in one to three separate experiments. Inhibition of 2-chloroadenosine-stimulated accumulation of cAMP in [<sup>3</sup>H]adenine-labeled guinea pig cerebral cortical slices was determined essentially as described.<sup>14</sup> In addition, 10 µg/mL of adenosine deaminase was present in final incubations to eliminate contributions from endogenous adenosine and 30 µM rolipram [4-[3-(cyclopentyloxy)-4-methoxyphenyl]-2-pyrrolidone, ZK 62711, Schering AG, West Berlin] was present in final incubations to inhibit phosphodiesterases. The  $EC_{50}$  of 2-chloroadenosine was approximately 7  $\mu$ M. Inhibition of the response to 15  $\mu$ M 2-chloroadenosine by a range of concentrations of each compound was determined in triplicate in one to two separate experiments.  $K_i$  values can be calculated from the observed IC<sub>50</sub> values (Table I) by using the equation:  $K_i = IC_{50}/1$ + [adenosine analogue]/ $K_{\rm D}$  or EC<sub>50</sub> of the adenosine analogue.

Acknowledgment. This work was supported in part by an A. H. Robins Graduate Fellowship to S.M.T.

Registry No. 1a, 58-55-9; 1b, 5169-95-9; 1c, 31542-62-8; 1d, 31542-68-4; 1e, 58-08-2; 1f, 961-45-5; 2a, 91265-82-6; 2b, 91265-80-4; 2c, 91265-83-7; 2d, 91265-84-8; 2e, 91280-59-0; 2f, 91265-85-9; 2g, 91265-86-0; 3a, 39456-06-9; 3b, 91265-87-1; 3c, 53528-96-4; 3d, 91265-81-5; 3e, 53528-87-3; 3f, 91265-88-2; 3g, 53528-93-1; 3h, 91265-89-3; 4a, 91265-90-6; 4b, 91265-91-7; 4c, 91265-92-8; 4d, 91265-93-9; 5, 91265-76-8; 6, 91265-77-9; 7, 91265-78-0; 8, 91265-79-1; 2-(n-propylamino)thiazole, 78508-32-4; bis(2,4,6-trichlorophenyl) n-propylmalonate, 77427-41-9; carbon suboxide, 12795-06-1; 2-(ethylamino)-1,3,4-thiadiazole, 13275-68-8; 4ethyl-3-thiosemicarbizide, 13431-34-0; trimethyl orthobenzoate, 707-07-3; 2-acetamido-5-phenyl-1,3,4-thiadiazole, 28898-88-6; bis(2,4,6-trichlorophenyl) ethylmalonate, 15781-72-3.

## Synthesis of a Tricyclic Aphidicolin Analogue That Inhibits DNA Synthesis in Vitro

John E. McMurry<sup>\*1</sup> and Thomas R. Webb

Department of Chemistry, University of California, Santa Cruz, California 95064. Received March 1, 1984

We have hypothesized that the biological activity of the antiviral antitumor diterpene aphidicolin requires a specific stereochemical relationship between two rigidly held hydroxyl groups on the  $\alpha$  face of the molecule. The complex tetracyclic carbon skeleton is not necessary but appears to serve only as a framework on which to hold the hydroxyls. In support of this theory, we have prepared a simple tricyclic triol analogue (7) whose activity approaches that of the natural product in inhibiting in vitro DNA synthesis.

Aphidicolin (1), a diterpenoid tetrol produced by the mold Cephalosporium aphidicola Petch,<sup>2</sup> has provoked wide interest in recent years owing to its striking biological activity; more than 100 publications have appeared in the last 5 years reporting its use in biochemical studies. For example, aphidicolin shows marked activity against herpes virus, both in vitro and in the rabbit eye.<sup>3,4</sup> In addition, aphidicolin possesses considerable antitumor activity in the C6 mouse colon and B16 mouse melanosarcoma screens<sup>5</sup> and has been shown to inhibit the growth of

(4)Ikegami, S.; Taguchi, T.; Ohashi, M.; Oguro, M.; Nagano, H.; Mano, Y. Nature (London) 1978, 275, 458.

leukemic T- and B-lymphocytes.<sup>6</sup> These biological properties, together with the unusual structure of aphidicolin, have also occasioned much activity among synthetic organic chemists. Six different total syntheses of the natural product have been recorded,<sup>7-12</sup> but there has

- Douros, J.; Suffness, M. "New Anticancer Drugs"; Carter, S. (5)K., Sakurai, Y., Eds.; Springer-Verlag: Berlin, 1980; p 29.
- Pedrali-Noy, G.; Belvedere, M.; Crepaldi, T.; Focher, F.; Spa-(6)dari, S. Cancer Res. 1982, 42, 3810.
- (7)(a) McMurry, J. E.; Andrus, A.; Ksander, G. M.; Musser, J. H.; Johnson, M. A. J. Am. Chem. Soc. 1979, 101, 1330. McMurry, J. E.; Andrus, A.; Ksander, G. M.; Musser, J. H.; Johnson, M. A. Tetrahedron, Suppl. 1981, 37, 319. Trost, B. M.; Nishimura, Y.; Yamamoto, K. J. Am. Chem. Soc.
- (8)1979, 101, 1328.
- (9)Corey, E. J.; Tius, M. A.; Das, J. J. Am. Chem. Soc. 1980, 102, 1742.
- (10) Ireland, R. E.; Godfrey, J. D.; Thaisrivongs, S. J. Am. Chem. Soc. 1981, 103, 2446.
- Van Tamelen, E. E.; Zawacky, S. R.; Russell, R. K.; Carlson, (11)J. G. J. Am. Chem. Soc. 1983, 105, 142.

<sup>(1)</sup> Author to whom inquiries should be addressed: Department of Chemistry, Baker Laboratory, Cornell University, Ithaca, NY 14853.

<sup>(2)</sup> (a) Brundret, K. M.; Dalziel, W.; Hesp, B.; Jarvis, J. A. J.; Neidle, S. J. Chem. Soc., Chem. Commun. 1972, 1027. (b) Dalziel, W.; Hesp, B.; Stevenson, K. M.; Jarvis, J. A. J. J. Chem. Soc., Perkin Trans. 1 1973, 2841. (3) Bucknall, R. A.; Moores, J.; Simms, R.; Hesp, B. Antimicrob.

Agents Chemother. 1973, 4, 294.