A Convenient New Synthesis of 3-Substituted β-Lactams Formally Derived from 1-(Aminomethyl)cyclopropanecarboxylic Acids^[‡]

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1,3-Dipolar cycloaddition of *N*-benzyl-*C*-(methoxycarbonyl)nitrone (3a), N-benzyl-C-phenylnitrone (3b), N-benzyl-C-cyanonitrone (3c), N-(p-methoxybenzyl)-C-cyanonitrone (3d), N-phenyl- (3e) and N-(2-pyridyl)-C-methylnitrones (3f) to bicyclopropylidene (2) gave the corresponding cycloadducts 5a-f in 100, 95, 94, 100, 93 and 71% yields, respectively. Treatment of these bisspirocyclopropanated isoxazolidines with trifluoroacetic acid in acetonitrile furnished the corresponding 3-spirocyclopropanated β -lactams **7a**-**f** in 78, 75, 75, 94, 96 and 96% yields, respectively. The structures of the cycloadduct **5b** and of the β -lactam **9a** were proved by X-ray crystal structure analyses. Thus, this new method furnishes

Introduction

Since a number of β -lactams play an important role as powerful anti-infectants,^[1] we were pleased to find that 1,3dipolar cycloadducts 4 of nitrones 3 with methylenecyclopropanes 1 underwent facile and clean fragmentation upon treatment with trifluoroacetic acid to vield β -lactams $6^{[2]}$ (for a mechanistic interpretation of this transformation see ref.^[2b]). As a logical extension of this method, we immediately conceived the possibility of subjecting the readily available 1,3-dipolar cycloadducts of nitrones 3 with bicyclopropylidene 2,^[3] the bisspirocyclopropanated isoxazol-

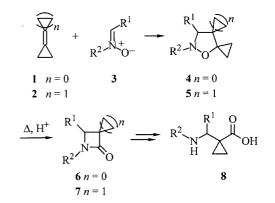
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compounds with a 5-azaspiro[2.3]hexan-4-one skeleton in 68-94% overall yield in two simple steps. β -Lactams **7a**, **7b**, and 7d were converted into their N-acyl derivatives 9a, 9b, 9d and 11 in 44, 28, 39 and 78% yields, respectively. Heating of the β -lactams **9b** and **11** with *tert*-butyl glycinate (**12**) or with tert-butyl (S)-phenylalaninate (14) in DMF led to ringopening of the β -lactam moiety to give β -dipeptides 15, 16 and 17 in 61, 84 and 79% yields, respectively, while β -lactam 9a gave the amide 13 (51% yield). β -Lactams 7e and 7f turned out not to be transformable into such peptide products

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idines 5, to these fragmentation conditions. By the same reaction mode, these compounds would lead to 2-spirocyclopropanated β -lactams 7 with variable substituents at the 3-position (Scheme 1). This would open up a convenient route to substituted 1-(aminomethyl)cyclopropanecarboxylic acids 8, a subclass of the cyclopropyl-containing β-alanine analogues,^[4] which have recently been reviewed exhaustively.^[5] The parent 1-(aminomethyl)cyclopropanecarboxylic acid (8, $R^{1} = R^{2} = H$) has been used by Seebach et al. to build interesting oligo-β-peptides that form ribbontype arrangements of eight-membered hydrogen-bonded



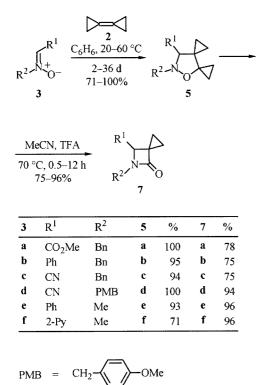
Scheme 1. 1,3-Dipolar cycloadditions of nitrones 3 to methylenecyclopropane (1) and bicyclopropylidene (2), and subsequent possible transformations of the cycloadducts 4 and 5

^[‡] Cyclopropyl Building Blocks for Organic Synthesis, 102. For Part 101 see: A. de Meijere, R. R. Kostikov, A. I. Savchenko, S. I. Kozhushkov, Eur. J. Org. Chem. 2004, in press. Part 100: M. Limbach, S. Dalai, A. de Meijere, Adv. Synth. Catal. 2004, 346, 760-766.

rings.^[6a] Thus, a new general route to β -alanine analogues of type **8** would facilitate new studies towards β -peptide analogues incorporating these building blocks. Herein, we report our first successes along these avenues.

Results and Discussion

As could be expected from literature precedents,^[7] 1,3dipolar cycloadditions of *N*-benzyl-*C*-(methoxycarbonyl)nitrone (**3a**),^[8] *N*-benzyl-*C*-phenylnitrone (**3b**),^[9] *N*-benzyl-*C*-cyanonitrone (**3c**),^[10] *N*-(*p*-methoxybenzyl)-*C*-cyanonitrone (**3d**), *N*-methyl-*C*-phenyl- (**3e**) and *N*-methyl-*C*-(2pyridyl)nitrones (**3f**)^[11] to bicyclopropylidene (**2**) in benzene at ambient or elevated temperature gave the corresponding cycloadducts **5a**-**f** in 100, 95, 94, 100, 93 and 71% yields, respectively (Scheme 2). As in previous cases, for those cycloadditions that were carried out at moderate temperatures in order to avoid the thermal rearrangement of the cycloadducts **5** at elevated temperature, very long reaction times were necessary (cf. ref.^[7]).



Scheme 2. Preparation of cycloadducts 5a-f from nitrones 3a-f and bicyclopropylidene (2), and their conversion into β -lactams 7a-f

The structures of 5a-f were assigned on the basis of their NMR spectra. However, no signals for the benzylic CH₂ group or the nitrile and quaternary spirocyclopropane carbon atoms in position 4 were seen in the ¹³C NMR spectra of 5c and 5d at room temperature under standard conditions; at 25 °C these signals are close to coalescence, due to a dynamic conformational exchange of the molecules with a rate close to the NMR time-scale. Upon raising the

temperature, these signals became sharp only at 100 °C, and, after cooling, they returned to their initial broadened state (see Exp. Sect.). The structural features of one of the cycloadducts - 8-benzyl-9-phenyl-7-oxa-8-azadispiro-[2.0.2.3]nonane (**5b**) – were established by an X-ray crystal structure analysis (Figure 1).

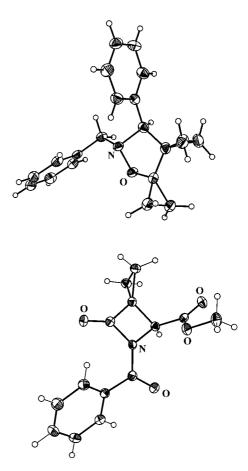
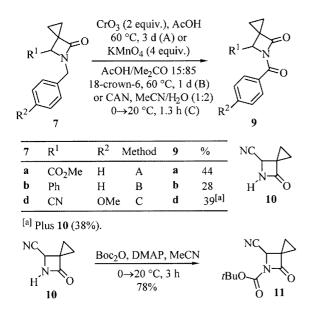


Figure 1. Crystal structures of 8-benzyl-9-phenyl-7-oxa-8-azadispiro[2.0.2.3]nonane (**5b**) and methyl 5-benzoyl-6-oxo-5-azaspiro[2.3]hexane-4-carboxylate (**9a**)^[12]

Treatment of compounds 5a-f with trifluoroacetic acid (TFA) in acetonitrile at 70 °C indeed furnished the corresponding 3-spirocyclopropanated β -lactams 7a-f in 78, 75, 75, 94, 96 and 96% yields, respectively, after purification by column chromatography (Scheme 2). Thus, the overall yields of this new approach to compounds with a 5-azaspiro[2.3]hexan-4-one skeleton in two simple steps range from 68-94% (cf. ref.^[13]).

An interesting application of the readily available spirocyclopropanated β -lactams **7a**-**f** would be their conversion into small oligopeptides in which the cyclopropanated β alanines derived from **7a**-**f** would act as a conformational lock.^[6] Nucleophilic ring opening of these β -lactams **7** with an appropriately protected glycine derivative should directly lead to simple dipeptides. Such transformations are known, however, to require an additional acyl substituent on the nitrogen.^[14] In the present case, the preparation of such activated β -lactams might be achieved by two routes. The first one would be by N-debenzylation of β -lactams 7a-d followed by N-acylation. Unfortunately, attempted debenzylation of these compounds by hydrogenation under palladium catalysis led to hydrogenolytic opening of the $N-C(R^1)$ bond of the lactam ring. In the case of **7b**, this bond is also benzylic, and its cleavage is facilitated by ring strain.^[15] Attempted debenzylations of 7c under Birch conditions (sodium in ammonia),^[16a] or of 7d by buffered sodium persulfate,^[16b] only led to decomposition of the starting material or no reaction, respectively. However, the alternative approach, in which the benzylic methylene is oxidized to a carbonyl group, turned out to be more successful (Scheme 3). This type of oxidation was achieved for compounds 7a.b with chromium trioxide in acetic acid (method A)^[17a] or potassium permanganate in an acetic acid/acetone mixture (method B),^[17b] which furnished the corresponding *N*-benzoyl-β-lactams **9a**,**b** in 44 and 28% yields, respectively. The structure of the *N*-benzoyl- β -lactam **9a** was rigorously proved by an X-ray crystal structure analysis (Figure 1).

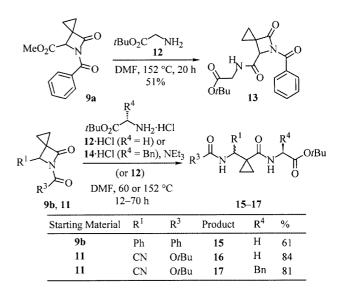


Scheme 3. Oxidation of the benzylic methylene group in $\beta\text{-lactams}\ \textbf{7a,b,d}$

The methylene group in the more labile *p*-methoxybenzyl group of the β -lactam **7d** can be oxidized using cerium ammonium nitrate (CAN) in aqueous acetonitrile (method C, cf. ref.^[17c]). However, this oxidation was accompanied by oxidative *N*-deprotection of **7d**, giving *N*-(*p*-methoxybenzoyl)- β -lactam **9d** and *N*-deprotected β -lactam **10** in almost equal yields (39 and 38%, respectively). The latter was *tert*-butoxycarbonyl protected using an established procedure^[17d] (Scheme 3).

The β -lactams **9a**, **9b** and **11** did not react with *tert*-butyl glycinate (**12**) in DMF at ambient temperature. Upon heating of **9a** with **12** in DMF under reflux (152 °C), the ester group was transformed into a *tert*-butoxycarbonylmethy-lenamido group to give the new lactam **13** (Scheme 4) without ring opening, even after prolonged heating with an ex-

cess of 12. However, the β -lactams **9b** and **11** reacted with **12** in the desired manner under these conditions to give the corresponding β -dipeptides **15** and **16** in 61 and 84% yields, respectively (Scheme 4), as pure colorless solids, after column chromatography (**15**) or without any additional purification (**16**).



Scheme 4. Reactions of *N*-acylated β -lactams **9a,b** and **11** with *tert*butyl glycinate (**12**) and *tert*-butyl (*S*)-phenylalaninate (**14**)

Under the same conditions, a 1.1:1 mixture of the diastereomeric dipeptides (2S,2'S)-17 and (2S,2'R)-17 was obtained from the β -lactam 11 and *tert*-butyl (*S*)-phenylalaninate (14). These diastereomeric dipeptides (2S,2'S)-17 and (2S,2'R)-17 could easily be separated by column chromatography (see Exp. Sect.), and the structure of the latter was established by an X-ray crystal structure analysis (Figure 2). The absolute configuration of the dipeptide (2S,2'R)-17 was assigned on the basis of the known (*S*)-configuration of the *tert*-butyl (*S*)-phenylalaninate (14) used in the synthesis of 17.

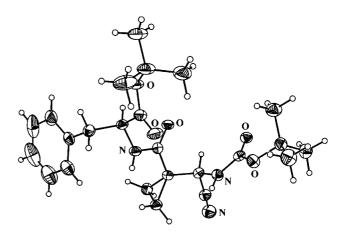


Figure 2. Crystal structure of *tert*-butyl (2S,2'R)-2-{[1-(*tert*-butoxycarbonylaminocyanomethyl)cyclopropylcarbonyl]amino}-3-phenylpropionate [(2S,2'R)-17]^[12]

In summary, a new, original and simple two step approach to cyclopropanated β -alanines in their β -lactam form has been developed. In addition, the potential use of these β -lactams in the construction of simple dipeptides incorporating cyclopropanated β -alanines by ring opening with protected amino acids has been demonstrated.

Experimental Section

General: NMR spectra were recorded on Bruker AM 250 (250 MHz for ¹H and 62.9 MHz for ¹³C NMR), Varian Mercury 200 (200 MHz for ¹H and 50.3 MHz for ¹³C NMR), Unity 300 (300 MHz for ¹H and 75.5 MHz for ¹³C NMR) or Inova 600 (150 MHz for ¹³C NMR) instruments in CDCl₃ unless otherwise specified. Multiplicities were determined by DEPT (Distortionless Enhancement by Polarization Transfer) or APT (Attached Proton Test) measurements. Chemical shifts refer to $\delta_{TMS} = 0.00$ according to the chemical shifts of residual CHCl₃ signals. IR: Bruker IFS 66 (FT-IR) spectrophotometer, measured as KBr pellets or oils between KBr plates. MS (EI, 70 eV): Finnigan MAT 95 spectrometer. High resolution mass data (HRMS) were obtained by preselectedion peak matching at R 10000 to be within ± 2 ppm of the exact mass. M.p.: Büchi 510 capillary melting point apparatus, uncorrected values. TLC: Macherey-Nagel precoated sheets, 0.25 mm Sil G/UV₂₅₄. Column chromatography: Merck silica gel, grade 60, 230-400 mesh. Starting materials: bicyclopropylidene (2),^[18] Nbenzyl-C-(methoxycarbonyl)nitrone (3a),^[8] N-benzyl-C-phenylnitrone (3b),^[9] N-benzyl-C-cyanonitrone (3c),^[10] N-phenyl- (3e) and N-(2-pyridyl)-C-methylnitrones (3f),^[11] and tert-butyl glycinate (12)^[19] were prepared according to published procedures. All operations in anhydrous solvents were performed under an argon atmosphere in flame-dried glassware. Diethyl ether was dried by distillation from sodium benzophenone ketyl, DMF and triethylamine from calcium hydride, CH2Cl2 and acetonitrile from P2O5. mCPBA was enriched according to a published procedure.^[20] All other chemicals were used as commercially available. Organic extracts were dried over MgSO₄ or Na_2SO_4 (for 13, 15–17).

N-(*p*-Methoxybenzyl)-*C*-cyanonitrone (3d): Chloroacetonitrile (22.6 g, 18.9 mL, 0.30 mol) and K₂CO₃ (55.3 g, 0.40 mol) were added to a vigorously stirred solution of p-methoxybenzylamine (27.4 g, 26.1 mL, 0.20 mol) in acetonitrile (2 L). After additional stirring for 12 h at 60 °C, the suspension was filtered through a pad of Celite, and the solvent was evaporated under reduced pressure. The residue was purified by column chromatography ($R_{\rm f} = 0.20$, 660 g of silica gel, 7.5×20 cm column, hexane/Et₂O, 1:2) to give 28.0 g (80%) of (4-methoxybenzylamino)acetonitrile as a dark oil. This oil (28.0 g, 0.160 mol) was dissolved in anhydrous CH₂Cl₂ (760 mL), and m-CPBA (60.7 g, 0.352 mol) was added in small portions at 0 °C. After additional stirring for 30 min at 0 °C and for 1 h at ambient temperature, a 10% aq. solution of Na₂S₂O₃ (300 mL) and sat. aq. NaHCO₃ solution (300 mL) were added, and the mixture was stirred for an additional 1 h. The aqueous phase was extracted with CH_2Cl_2 (2 × 200 mL); the combined organic layers were washed with brine (2 \times 200 mL), dried, and concentrated under reduced pressure. Nitrone 3d (30.4 g, 100%, 1:4 mixture of E- and Z-isomers) was obtained as a yellow solid and used without purification. An analytical sample was obtained by column chromatography ($R_{\rm f} = 0.24, 20$ g of silica gel, 2×13 cm column, hexane/Et₂O, 1:3) and had m.p. 74-75 °C. IR (KBr): $\tilde{v} = 3102$ cm⁻¹, 2994, 2964, 2937, 2838, 2222, 1616, 1587, 1544, 1520, 1462, 1414. ¹H NMR (250 MHz): $\delta = 3.82$ (s, 3 H, OCH₃, *E*-isomer), 3.84 (s, 3 H, OCH₃, Z-isomer), 4.94 (s, 2 H, CH₂, Z), 5.24 (s, 2 H, CH₂, E), 6.53 (s, 1 H, =CH, Z), 6.62 (s, 1 H, =CH, E), 6.90–6.99 (m, 4 H, Ar–H, Z- and E-isomers), 7.29–7.33 (m, 2 H, Ar–H, Z), 7.44–7.48 (m, 2 H, Ar–H, E) ppm. ¹³C NMR (50.3 MHz): $\delta = 55.3$ (CH₃, E), 55.4 (CH₃, Z), 69.4 (CH₂, E), 71.1 (CH₂, Z), 106.8 (CH, Z), 107.0 (CH, E), 112.1 (C, Z), 114.3 (2 CH, E), 114.8 (2 CH, Z), 115.3 (C, E), 122.4 (C, Z), 123.5 (C, E), 131.0 (2 CH, E), 131.5 (2 CH, Z), 160.7 (C, E), 161.0 (C, Z) ppm. MS (EI): m/ z (%) = 190 (2) [M⁺], 121 (100), 77 (12), 51 (8). C₁₀H₁₀N₂O₂ (190.2): calcd. C 63.14, H 5.30, N 14.73; found C 62.88, H 5.49, N 14.93.

Cycloaddition of Nitrones 3c-f to Bicyclopropylidene (2). General Procedure (GP) 1: A solution of the respective nitrone (5 mmol) and bicyclopropylidene (2) (0.85 g, 1.0 mL, 10.6 mmol) in benzene (3 mL) was stirred in a hermetically closed tube at the indicated temperature for the indicated time. After cooling to ambient temperature, the solution was concentrated under reduced pressure, and the crude product was purified by column chromatography on silica gel.

Methyl 8-Benzyl-7-oxa-8-azadispiro[2.0.2.3]nonane-9-carboxylate (5a): Column chromatography ($R_f = 0.14$, 115 g of silica gel, 4.5 \times 15 cm column, hexane/Et₂O, 5:1) of the residue obtained from nitrone 3a (1.0 g, 5.18 mmol) and bicyclopropylidene (2) (832 mg, 0.97 mL, 10.4 mmol) according to GP1 (45 °C, 2 d) gave the cycloadduct **5a** (1.40 g, 100%) as a yellow oil. IR (film): $\tilde{v} = 3064 \text{ cm}^{-1}$, 3031, 3006, 2953, 1766, 1456, 1437. ¹H NMR (250 MHz): $\delta =$ 0.27-0.36 (m, 2 H, cPr-H), 0.40-0.46 (m, 1 H, cPr-H), 0.61-0.78 (m, 3 H, cPr-H), 0.91-0.94 (m, 2 H, cPr-H), 3.65 (s, 3 H, OCH₃), 3.68 (s, 1 H, CH), 4.14 (d, J = 12.5 Hz, 1 H, CH₂), 4.37 (d, J = 12.5 Hz, 1 H, CH₂), 7.23–7.41 (m, 5 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 5.5$ (CH₂), 6.8 (CH₂), 8.6 (CH₂), 10.7 (CH₂), 30.0 (C), 52.1 (CH₃), 62.8 (CH₂), 66.4 (C), 72.7 (CH), 127.7 (CH), 128.4 (2 CH), 129.4 (2 CH), 136.0 (C), 170.6 (C) ppm. MS (EI): m/z (%) = 273 (10) [M⁺], 214 (90), 105 (19), 91 (100). C₁₆H₁₉NO₃ (273.3): calcd. C 70.31, H 7.01, N 5.12; found C 70.10, H 6.80, N 5.01.

8-Benzyl-9-phenyl-7-oxa-8-azadispiro[2.0.2.3]nonane (5b): Column chromatography ($R_{\rm f} = 0.44$, 165 g of silica gel, 5 \times 17 cm column, hexane/Et₂O, 10:1) of the residue obtained from nitrone **3b** (3.76 g, 18.0 mmol) and bicyclopropylidene (2) (1.60 g, 1.87 mL, 20.0 mmol) according to GP1 (60 °C, 25 d) gave the cycloadduct **5b** (5.0 g, 95%) as a colorless solid, m.p. 70 °C. IR (KBr): $\tilde{v} = 3067$ cm⁻¹, 2998, 2845, 1653, 1636, 1456, 1437. ¹H NMR (250 MHz): $\delta = 0.12 - 0.35$ (m, 2 H, cPr-H), 0.34-0.53 (m, 4 H, cPr-H), 0.85-0.98 (m, 2 H, cPr-H), 4.14 (s, 1 H, CH), 4.08 (m, 2 H, CH₂), 7.20–7.40 (m, 10 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 6.7$ (CH₂), 7.3 (CH₂), 7.7 (CH₂), 8.6 (CH₂), 33.0 (C), 61.3 (CH₂), 66.5 (C), 76.3 (CH), 126.8 (CH), 127.6 (CH), 127.9 (2 CH), 128.1 (2 CH), 128.5 (4 CH), 137.7 (C), 138.1 (C) ppm. MS (EI): m/z (%) = 291 (10) [M⁺], 262 (5), 235 (5), 129 (30), 115 (18), 91 (100). C₂₀H₂₁NO (291.4): calcd. C 82.44, H 7.27, N 4.81; found C 82.19, H 6.97, N 4.76.

8-Benzyl-9-cyano-7-oxa-8-azadispiro[2.0.2.3]nonane (5c): Column chromatography ($R_{\rm f} = 0.20$, 54 g of silica gel, 4 × 10 cm column, hexane/Et₂O, 2:1) of the residue obtained from the nitrone **3c** (1.46 g, 9.12 mmol) and bicyclopropylidene (**2**) (1.60 g, 1.9 mL, 20 mmol) according to GP1 (20 °C, 8 d) gave the cycloadduct **5c** (2.07 g, 94%) as a colorless oil. IR (film): $\tilde{v} = 3066 \text{ cm}^{-1}$, 3031, 2959, 2863, 2246, 1497, 1454. ¹H NMR (300 MHz): $\delta = 0.30-0.49$ (m, 2 H, *c*Pr-H), 0.51-0.60 (m, 2 H, *c*Pr-H), 0.80-0.97 (m, 4 H, *c*Pr-H), 3.78 (s, 1 H, CH), 4.11-4.26 (m, 2 H, CH₂), 7.20-7.37

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(m, 5 H, Ar-H) ppm. ¹³C NMR (50.3 MHz): δ = 7.3 (CH₂), 7.7 (CH₂), 8.2 (CH₂), 11.2 (CH₂), 62.5 (CH), 66.6 (C), 128.0 (CH), 128.6 (2 CH), 129.2 (2 CH), 135.0 (C), three carbon atom signals were not detectable at this temperature. ¹³C NMR (75.5 MHz, C₂D₂Cl₄, 100 °C): δ = 7.0 (CH₂), 7.4 (CH₂), 7.7 (CH₂), 10.5 (CH₂), 30.6 (C), 60.8 (CH₂), 62.5 (CH), 66.2 (C), 116.1 (C), 127.6 (CH), 128.3 (2 CH), 128.8 (2 CH), 135.3 (C) ppm. MS (EI): *m/z* (%) = 239 (20) [M⁺ - H], 214 (10), 211 (20), 105 (50), 91 (100). MS (DCI): *m/z* (%) = 481 (5) [2 M + H⁺], 258 (8) [M + NH₄⁺], 241 (100) [M + H⁺]. C₁₅H₁₆N₂O (240.3): calcd. C 74.97, H 6.71, N 11.66; found C 74.76, H 6.65, N 11.63.

9-Cyano-8-(4-methoxybenzyl)-7-oxa-8-azadispiro[2.0.2.3]nonane (5d): Column chromatography ($R_{\rm f}$ = 0.34, 55 g of silica gel, 3 \times 17 cm column, hexane/Et₂O, 10:1) of the residue obtained from the nitrone 3d (2.5 g, 13 mmol) and bicyclopropylidene (2) (2.10 g, 2.46 mL, 26.2 mmol) according to GP1 (20 °C, 8 d) gave the cycloadduct 5d (3.50 g, 100%) as a colorless solid, m.p. 70-71 °C. IR (KBr): $\tilde{v} = 3075 \text{ cm}^{-1}$, 3010, 2934, 2868, 2838, 2246, 1611, 1585, 1512, 1468. ¹H NMR (250 MHz): $\delta = 0.36 - 0.70$ (m, 4 H, *c*Pr-H), 0.82-1.04 (m, 4 H, cPr-H), 3.80 (s, 3 H, OCH₃), 3.81 (s, 1 H, CH), 4.04-4.28 (m, 2 H, CH₂), 6.85-6.91 (m, 2 H, Ar-H), 7.30–7.37 (m, 2 H, Ar-H) ppm. ¹³C NMR (50.3 MHz): $\delta = 7.2$ (CH₂), 7.7 (CH₂), 8.2 (CH₂), 11.2 (CH₂), 55.2 (CH₃), 62.3 (CH), 66.5 (C), 114.0 (2 CH), 126.9 (C), 130.5 (2 CH), 159.4 (C), three carbon atom signals were not detectable at this temperature. ¹³C NMR (75.5 MHz, $C_2D_2Cl_4$, 100 °C): $\delta = 6.9$ (CH₂), 7.4 (CH₂), 7.7 (CH₂), 10.5 (CH₂), 30.5 (C), 55.2 (CH₃), 60.2 (CH₂), 62.3 (CH), 66.2 (C), 114.1 (2 CH), 116.2 (C), 127.3 (C), 130.1 (2 CH), 159.4 (C) ppm. MS (EI): m/z (%) = 270 (20) [M⁺], 241 (8), 135 (25), 121 (100). C₁₆H₁₈N₂O₂ (270.3): calcd. C 71.09, H 6.71, N 10.36; found C 70.97, H 6.58, N 10.12.

8-Methyl-9-(pyridin-2-yl)-7-oxa-8-azadispiro[2.0.2.3]nonane (5f): Column chromatography ($R_{\rm f} = 0.20, 54$ g of silica gel, 4×10 cm column, hexane/Et₂O, 2:1) of the residue obtained from the nitrone 3f (3.0 g, 22.0 mmol) and bicyclopropylidene (2) (2.86 g, 3.35 mL, 36 mmol) according to GP1 (60 °C, 36 d) gave the cycloadduct 5f (3.39 g, 71%) as a colorless oil. IR (film): $\tilde{v} = 3073 \text{ cm}^{-1}$, 3001, 2956, 2871, 2777, 1590, 1570, 1472, 1434. ¹H NMR (250 MHz): $\delta = 0.03 - 0.18$ (m, 2 H, cPr-H), 0.33-0.42 (m, 2 H, cPr-H), 0.58-0.62 (ddd, J = 10.0, 5.0, 5.0 Hz, 1 H, cPr-H), 0.90-0.99(m, 3 H, cPr-H), 2.90 (s, 3 H, NCH₃), 4.07 (s, 1 H, CH), 7.17-7.20 (m, 1 H, Ar-H), 7.68-7.71 (m, 2 H, Ar-H), 8.44-8.47 (m, 1 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 5.9$ (CH₂), 6.5 (CH₂), 9.0 (CH₂), 9.6 (CH₂), 33.1 (C), 45.7 (CH), 66.5 (C), 79.7 (CH₃), 122.5 (CH), 122.8 (CH), 136.8 (CH), 148.3 (CH), 159.3 (C) ppm. MS (EI): m/z (%) = 216 (2) [M⁺], 201 (5), 187 (18), 159 (82), 145 (48), 130 (100), 119 (45), 92 (20), 78 (55), 42 (35). HRMS (EI) calcd. for $C_{13}H_{16}N_2O$ 216.1263 [M⁺], found 216.1263. $C_{13}H_{16}N_2O$ (216.3): calcd. C 72.19, H 7.46; found C 72.45, H 7.79.

Preparation of \beta-Lactams 7a-f. General Procedure (GP) 2: Trifluoroacetic acid (TFA) was added to the solution of the respective isoxazolidine 5 in acetonitrile, and the resulting mixture was stirred at 70 °C for the indicated time. After cooling to ambient temperature, the mixture was filtered through a pad of Celite, concentrated under reduced pressure, and the crude product was purified by column chromatography on silica gel.

Methyl 5-Benzyl-6-oxo-5-azaspiro[2.3]hexane-4-carboxylate (7a): Column chromatography ($R_f = 0.21$, 40 g of silica gel, 3×14 cm column, hexane/Et₂O, 1:1) of the residue obtained from the isoxazolidine 5a (1.41 g, 5.16 mmol) and TFA (0.710 g, 0.480 mL, 6.22 mmol) in acetonitrile (32 mL) according to GP2 (12 h) gave the β-lactam **7a** (990 mg, 78%) as a yellow oil. IR (film): $\tilde{v} = 3065$ cm⁻¹, 3029, 3003, 2953, 2849, 1756, 1729, 1455, 1436. ¹H NMR (250 MHz): $\delta = 0.84-0.92$ (m, 1 H, cPr-H), 1.06-1.16 (m, 1 H, cPr-H), 1.18-1.29 (m, 2 H, cPr-H), 3.71 (s, 3 H, OCH₃), 4.02 (s, 1 H, CH₂), 4.24 (d, J = 15.0 Hz, 1 H, CH₂), 4.91 (d, J = 15.0 Hz, 1 H, CH₂), 7.25-7.39 (m, 5 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 6.5$ (CH₂), 8.0 (CH₂), 37.2 (C), 45.5 (CH₂), 52.1 (CH), 57.4 (CH₃), 127.7 (CH), 128.3 (2 CH), 128.7 (2 CH), 135.2 (C), 170.0 (C), 170.9 (C) ppm. MS (EI): m/z (%) = 245 (4) [M⁺], 217 (20), 186 (30), 158 (40), 91 (100). C₁₄H₁₅NO₃ (245.3): calcd. C 68.56, H 6.16, N 5.71; found C 68.45, H 6.08, N 5.49.

5-Benzyl-6-phenyl-5-azaspiro[2.3]hexane-4-one (7b): Column chromatography ($R_{\rm f} = 0.15, 40$ g of silica gel, 3×14 cm column, hexane/Et₂O, 3:1) of the residue obtained from the isoxazolidine **5b** (1.48 g, 5.08 mmol) and TFA (698 mg, 0.472 mL, 6.12 mmol) in acetonitrile (32 mL) according to GP2 (3 h) gave the β -lactam 7b (1.0 g, 75%) as a colorless solid, m.p. 64–65 °C. IR (KBr): $\tilde{\nu}$ = 3029 cm⁻¹, 1745, 1653, 1559, 1494, 1456. ¹H NMR (250 MHz): $\delta = 0.37 - 0.43$ (ddd, J = 10.0, 7.5, 5.0 Hz, 1 H, cPr-H), 1.01 - 1.06(ddd, J = 10.0, 7.5, 5.0 Hz, 1 H, cPr-H), 1.09-1.18 (ddd, J =10.0, 7.5, 5.0 Hz, 1 H, cPr-H), 1.24–1.32 (ddd, J = 10.0, 7.5, 5.0 Hz, 1 H, cPr-H), 3.85 (d, J = 15.0 Hz, 1 H, CH₂), 4.48 (s, 1 H, CH), 4.88 (d, J = 15.0 Hz, 1 H, CH₂), 7.15–7.21 (m, 4 H, Ar-H), 7.24–7.43 (m, 6 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): δ = 7.2 (CH₂), 8.2 (CH₂), 40.3 (C), 44.5 (CH₂), 61.4 (CH), 127.5 (CH), 127.6 (CH), 128.4 (4 CH), 128.6 (2 CH), 128.7 (2 CH), 135.8 (C), 136.3 (C), 172.4 (C) ppm. MS (EI): m/z (%) = 263 (36) [M⁺], 172 (10), 130 (100), 129 (85), 115 (42), 91 (65). $C_{18}H_{17}NO$ (263.3): calcd. C 82.10, H 6.51, N 5.32; found C 82.0, H 6.25, N 5.12.

5-Benzyl-6-oxo-5-azaspiro[2.3]hexane-4-carbonitrile (**7c**): Column chromatography ($R_f = 0.11$, 41 g of silica gel, 3 × 12 cm column, hexane/Et₂O, 2:1) of the residue obtained from the isoxazolidine **5c** (627 mg, 2.61 mmol) and TFA (357 mg, 0.24 mL, 3.13 mmol) in acetonitrile (15 mL) according to GP2 (12 h) gave the β-lactam **7c** (415 mg, 75%) as a colorless oil. IR (film): $\tilde{v} = 3065 \text{ cm}^{-1}$, 3032, 3009, 2923, 2243, 1772, 1496, 1456, 1382, 1355. ¹H NMR (300 MHz): $\delta = 1.17-1.29$ (m, 2 H, *c*Pr–H), 1.32–1.44 (m, 2 H, *c*Pr–H), 4.14 (s, 1 H, CH), 4.25 (d, *J* = 15.1 Hz, 1 H, CH₂), 4.81 (d, *J* = 15.1 Hz, 1 H, CH₂), 7.27–7.42 (m, 5 H, Ar-H) ppm. ¹³C NMR (75.5 MHz): $\delta = 8.2$ (CH₂), 9.1 (CH₂), 38.3 (C), 46.0 (CH₂), 46.7 (CH), 115.6 (C), 128.3 (CH), 128.4 (2 CH), 129.0 (2 CH), 133.9 (C), 169.8 (C) ppm. MS (EI): *m/z* (%) = 212 (85) [M⁺], 183 (32), 122 (32), 91 (100), 80 (45), 69 (50). C₁₃H₁₂N₂O (212.25): calcd. C 73.56, H 5.70, N 13.20; found C 73.74, H 5.77, N 13.08.

5-(4-Methoxybenzyl)-6-oxo-5-azaspiro[2.3]hexane-4-carbonitrile (7d): Column chromatography ($R_{\rm f} = 0.10, 44$ g of silica gel, 3 \times 13 cm column, hexane/Et₂O, 1.5:1) of the residue obtained from the isoxazolidine 5d (2.00 g, 7.40 mmol) and TFA (1.01 g, 0.68 mL, 8.88 mmol) in acetonitrile (45 mL) according to GP2 (12 h) gave the β -lactam 7d (1.68 g, 94%) as a colorless solid, m.p. 54–56 °C. IR (KBr): $\tilde{v} = 3003 \text{ cm}^{-1}$, 2973, 2913, 2867, 2841, 2249, 1754, 1613, 1585, 1515. ¹H NMR (250 MHz): $\delta = 1.14 - 1.27$ (m, 2 H, cPr-H), 1.30-1.44 (m, 2 H, cPr-H), 3.80 (s, 3 H, OCH₃), 4.10 (s, 1 H, CH), 4.18 (d, J = 15.0 Hz, 1 H, CH₂), 4.75 (d, J = 15.0 Hz, 1 H, CH₂), 6.84–6.93 (m, 2 H, Ar-H), 7.20–7.26 (m, 2 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 8.2$ (CH₂), 9.0 (CH₂), 38.2 (C), 45.5 (CH₂), 46.4 (CH), 55.2 (CH₃), 114.3 (2 CH), 115.7 (C), 125.9 (C), 129.8 (2 CH), 159.5 (C), 169.6 (C) ppm. MS (EI): m/z (%) = 242 (40) [M⁺], 213 (15), 121 (100). C₁₄H₁₄N₂O₂ (242.3): calcd. C 69.41, H 5.82, N 11.56; found C 69.14, H 5.61, N 11.37.

5-Methyl-6-phenyl-5-azaspiro[2.3]hexan-4-one (7e): Column chromatography ($R_{\rm f} = 0.66$, 21 g of silica gel, 14 × 2.5 cm column, CH₂Cl₂/MeOH, 30:1) of the residue obtained from the isoxazolidine **5e** (150 mg, 697 µmol) and TFA (96 mg, 0.10 mL, 840 µmol) in acetonitrile (3 mL) according to GP2 (30 min) gave the β-lactam **7e** (125 mg, 96%) as a colorless oil. IR (film): $\tilde{v} = 3001 \text{ cm}^{-1}$, 2902, 1751, 1456, 1386, 1172, 1039. ¹H NMR (250 MHz): $\delta = 0.42$ (dt, J = 7.3, 2.6 Hz, 1 H, *c*Pr-H), 1.08–1.18 (m, 2 H, *c*Pr-H), 1.25–1.35 (m, 1 H, *c*Pr-H), 2.86 (s, 3 H, NCH₃), 4.63 (s, 1 H, CH), 7.20–7.37 (m, 2 H, Ar-H), 7.38–7.43 (m, 3 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 7.6$ (CH₂), 8.6 (CH₂), 27.7 (NCH₃), 40.2 (C), 64.4 (CH), 127.3 (2 CH), 128.9 (3 CH), 135.1 (C), 175.2 (C) ppm. MS (EI): *m*/*z* (%) = 187 (59) [M⁺], 158 (7), 129 (100), 118 (48), 115 (56). HRMS (EI) calcd. for C₁₂H₁₃NO [M⁺] 187.0997, found 187.0997.

5-Methyl-6-(pyrid-2-yl)-5-azaspiro[2.3]hexan-4-one (7f): Column chromatography ($R_{\rm f} = 0.38, 21$ g of silica gel, 14×2.5 cm column, CH₂Cl₂/MeOH, 20:1) of the residue obtained from the isoxazolidine 5f (400 mg, 1.85 mmol) and TFA (254 mg, 0.20 mL, 2.22 mmol) in acetonitrile (5 mL) according to GP2 (40 min) gave the β-lactam **7f** (335 mg, 96%) as a colorless oil. IR (film): $\tilde{v} = 3003 \text{ cm}^{-1}$, 2924, 1751, 1472, 1291. ¹H NMR (250 MHz): $\delta = 0.35$ (ddd, J = 10.0, 7.5, 5.2 Hz, 1 H, cPr-H), 1.03 (ddd, J = 10.2, 7.0, 5.1 Hz, 1 H, *c*Pr-H), 1.18 (ddd, *J* = 10.0, 7.5, 5.1 Hz, 1 H, *c*Pr-H), 1.26 (ddd, J = 10.2, 7.5, 5.2 Hz, 1 H, cPr-H), 2.91 (s, 3 H, NCH₃), 4.72 (s, 1 H, CH), 7.21-7.27 (m, 1 H, Ar-H), 7.33 (d, J = 7.8 Hz, 1 H, Ar-H), 7.75 (dt, J = 7.7, 1.7 Hz, 1 H, Ar-H), 8.53-8.55 (m, 1 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 6.7$ (CH₂), 8.2 (CH₂), 27.8 (NCH₃), 40.7 (C), 64.7 (CH), 120.6 (CH), 123.1 (CH), 137.1 (CH), 149.5 (CH), 157.4 (C), 173.0 (C) ppm. MS (EI): m/z (%) = 188 (9) $[M^+]$, 159 (54), 145 (16), 130 (100), 119 (19), 78 (27). $C_{11}H_{12}N_2O$ (188.20): calcd. C 70.20, H 6.43; found C 70.31, H 6.56.

Methyl 5-Benzoyl-6-oxo-5-azaspiro[2.3]hexane-4-carboxylate (9a): Finely powdered CrO₃ (0.202 g, 0.202 mmol) was added in one portion to a stirred solution of β -lactam 7a (247 mg, 1.01 mmol) in acetic acid (20 mL) at ambient temperature, and the resulting mixture was stirred at 60 °C for an additional 3 days. After cooling to room temperature and dilution with diethyl ether (20 mL), NaHCO₃ (15.0 g, 179 mmol) was added to the solution in several portions, and the reaction mixture was stirred until the carbon dioxide evolution ceased. The organic phase was washed with aq. sat. NaHCO₃ solution in 20 mL portions until the evolution of carbon dioxide ceased, then it was dried, and concentrated under reduced pressure. Column chromatography ($R_{\rm f} = 0.20, 42$ g of silica gel, 3 \times 13 cm column, hexane/Et₂O, 2:1) of the residue gave 9a (113 mg, 44%) as a colorless solid, m.p. 92–93 °C. IR (KBr): $\tilde{v} = 3090$ cm⁻¹, 3077, 2987, 2961, 1796, 1734, 1676, 1601, 1581, 1481, 1441. ¹H NMR (300 MHz): $\delta = 1.07 - 1.33$ (m, 2 H, *c*Pr-H), 1.34-1.56 (m, 2 H, cPr-H), 3.80 (s, 3 H, OCH₃), 4.80 (s, 1 H, CH), 7.44-7.49 (m, 2 H, Ar-H), 7.54-7.68 (m, 1 H, Ar-H), 8.06-8.15 (m, 2 H, Ar-H) ppm. ¹³C NMR (50.3 MHz): $\delta = 9.6$ (CH₂), 11.4 (CH₂), 34.4 (C), 52.6 (CH), 56.0 (CH₃), 128.1 (2 CH), 129.9 (2 CH), 131.2 (C), 133.3 (CH), 164.5 (C), 167.6 (C), 168.7 (C) ppm. MS (EI): m/ z (%) = 259 (2) [M⁺], 228 (5), 200 (97), 105 (100), 77 (45). The structure of this β-lactam 9a was verified by X-ray crystal structure analysis. Some starting material 7a (72 mg, 29%) was also recovered by column chromatography ($R_{\rm f} = 0.12$).

5-Benzoyl-6-phenyl-5-azaspiro[2.3]hexane-4-one (9b): $KMnO_4$ (721 mg, 4.56 mmol) and 18-crown-6 (15.0 mg, 0.057 mmol, 5 mol%) were added to a stirred solution of β -lactam 7b (0.300 g,

1.14 mmol) in a mixture of acetone/acetic acid 85:15 (30 mL). The solution was stirred at 60 °C for 24 h. After cooling to room temperature, the mixture was diluted with diethyl ether (30 mL), then an aq. sat. solution of NaHCO3 (50 mL) was added in small portions under vigorous stirring, and the reaction mixture was stirred until the carbon dioxide evolution ceased. The organic phase was washed with brine (2 \times 20 mL), dried, and concentrated under reduced pressure. Column chromatography ($R_{\rm f} = 0.26$, 12 g of silica gel, 2×10 cm column, hexane/Et₂O, 3:1) of the residue gave **9b** (90.0 mg, 28%) as a colorless solid, m.p. 121–123 °C. IR (KBr): $\tilde{v} = 3065 \text{ cm}^{-1}$, 3010, 2950, 1800, 1675, 1653, 1448. ¹H NMR (250 MHz): $\delta = 0.66 - 0.76$ (m, 1 H, cPr-H), 1.28-1.39 (m, 2 H, cPr-H), 1.43-1.53 (m, 1 H, cPr-H), 5.35 (s, 1 H, CH), 7.29-7.43 (m, 5 H, Ar-H), 7.45-7.51 (m, 2 H, Ar-H), 7.55-7.62 (m, 1 H, Ar-H), 8.04–8.11 (m, 2 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): δ = 10.6 (CH₂), 11.7 (CH₂), 37.9 (C), 59.7 (CH), 126.9 (2 CH), 128.1 (2 CH), 128.5 (CH), 128.7 (2 CH), 129.9 (2 CH), 132.2 (C), 133.1 (CH), 136.4 (C), 164.8 (C), 170.4 (C) ppm. MS (EI): m/z (%) = 277 (36) [M⁺], 249 (25), 129 (32), 105 (100), 77 (50). C₁₈H₁₅NO₂ (277.3): calcd. C 77.96, H 5.45, N 5.05; found C 77.69, H 5.35, N 4.82. Some starting material 7b (200 mg, 67%) was also recovered by column chromatography ($R_{\rm f} = 0.11$).

Deprotection of β-Lactam 7d: A solution of CAN (6.63 g, 12.1 mmol) in water (85 mL) was added to a stirred solution of β lactam 7d (900 mg, 3.72 mmol) in acetonitrile (40 mL) at 0 °C. After an additional stirring for 20 minutes at the same temperature and for 1 h at ambient temperature, the mixture was extracted with ethyl acetate (3 \times 50 mL). The combined organic layers were washed successively with a 10% aq. Na₂SO₃ solution (2×50 mL), a 5% aq. NaHCO₃ solution (2×50 mL) and brine (50 mL), dried, and concentrated under reduced pressure. Column chromatography (hexane/Et₂O, 1:2, 75 g of silica gel, 3.5×16 cm column) of the residue gave 6-oxo-5-azaspiro[2.3]hexane-4-carbonitrile (10) (172 mg, 38%) as a colorless solid, m.p. 72–73 °C, $R_{\rm f} = 0.15$, and 5-(4-methoxybenzoyl)-6-oxo-5-azaspiro[2.3]hexane-4-carbonitrile (9d) (371 mg, 39%) as a colorless solid, m.p. 109–110 °C, $R_{\rm f} =$ 0.38 (hexane/Et₂O, 1:4). **10**: IR (KBr): $\tilde{v} = 3250 \text{ cm}^{-1}$, 3097, 2720, 2249, 1763, 1331. ¹H NMR (250 MHz): $\delta = 1.26 - 1.37$ (m, 2 H, cPr-H), 1.41-1.48 (m, 2 H, cPr-H), 4.44 (s, 1 H, CH), 6.40 (s, 1 H, NH) ppm. ¹³C NMR (62.9 MHz): $\delta = 9.1$ (CH₂), 9.8 (CH₂), 39.8 (C), 43.6 (CH), 116.9 (C), 171.5 (C) ppm. MS (EI): m/z (%) = 122 (12) [M⁺], 79 (50), 52 (100). C₆H₆N₂O (122.1); calcd. C 59.01, H 4.95, N 22.94; found C 59.20, H 5.08, N 22.81. 9d: IR (KBr): $\tilde{v} = 3084 \text{ cm}^{-1}$, 3005, 2977, 2938, 2843, 2251, 1798, 1666, 1603, 1576, 1514. ¹H NMR (250 MHz): $\delta = 1.47 - 1.67$ (m, 4 H, cPr-H), 3.89 (s, 3 H, OCH₃), 4.92 (s, 1 H, CH), 6.95-7.01 (m, 2 H, Ar-H), 8.08–8.14 (m, 2 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): δ = 11.4 (CH₂), 12.2 (CH₂), 34.8 (C), 44.4 (CH), 55.5 (CH₃), 113.8 (2 CH), 115.4 (C), 122.2 (C), 132.6 (2 CH), 163.3 (C), 164.3 (C), 166.3 (C) ppm. MS (EI): m/z (%) = 256 (20) [M⁺], 135 (100), 92 (9), 77 (9). C14H12N2O3 (256.3): calcd. C 65.62, H 4.72, N 10.93; found C 65.75, H 4.61, N 11.15.

tert-Butyl 4-Cyano-6-oxo-5-azaspiro[2.3]hexane-5-carboxylate (11): Di-*tert*-butyl pyrocarbonate (Boc₂O, 873 mg, 4.00 mmol) and DMAP (24.0 mg, 0.196 mmol) were added in one portion to a stirred solution of β -lactam 10 (245 mg, 2.01 mmol) in anhydrous acetonitrile (30 mL) at 0 °C. The reaction mixture was stirred at ambient temperature for an additional 3 h, diluted with dichloromethane (30 mL), washed successively with 10% aq. Na₂SO₃ solution (2 × 20 mL), aq. sat. NaHCO₃ solution (2 × 20 mL) and brine (2 × 20 mL), dried, and concentrated under reduced pressure. Column chromatography ($R_f = 0.10$, 12 g of silica gel, 2 × 10 cm

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column, hexane/Et₂O, 2:1) furnished **11** (346 mg, 78%) as a colorless solid, m.p. 87 °C. IR (KBr): $\tilde{v} = 3012 \text{ cm}^{-1}$, 2992, 2250, 1823, 1717. ¹H NMR (300 MHz): $\delta = 1.35-1.46$ (m, 2 H, cPr-H), 1.48–1.64 (m, 2 H, cPr-H), 1.56 (s, 9 H, 3 CH₃), 4.64 (s, 1 H, CH) ppm. ¹³C NMR (50.3 MHz): $\delta = 10.6$ (CH₂), 11.4 (CH₂), 28.0 (3 CH₃), 37.0 (C), 45.9 (CH), 85.1 (C), 115.0 (C), 145.7 (C), 166.6 (C) ppm. MS (EI): m/z (%) = 223 (1) [M + H⁺], 167 (9), 149 (30), 57 (100). MS (DCI): m/z (%) = 684 (5) [3 M + NH₄⁺], 462 (40) [2 M + NH₄⁺], 257 (72) [M + NH₃ + NH₄⁺], 240 (100) [M + NH₄⁺]. C₁₁H₁₄N₂O₃ (222.2): calcd. C 59.45, H 6.35, N 12.61; found C 59.55, H 6.14, N 12.47.

Reaction of *N*-Acylated β -Lactams 9a,b, 11 with *tert*-Butyl Glycinate (12) and *tert*-Butyl (*S*)-Phenylalaninate (14). General Procedure (GP) 3: *tert*-Butyl amino ester 12 (or its hydrochloride in the presence of 1 equiv. of triethylamine) was added to a solution of the respective β -lactam 9a,b or 11 in DMF, and the resulting mixture was stirred at 152 °C (if not otherwise specified) for the indicated time. After cooling to ambient temperature, diethyl ether (20 mL) was added, and the organic layer was washed with water (10 mL), brine (2 × 10 mL), and dried over Na₂SO₄. After filtration, the solution was concentrated under reduced pressure, and the crude product was purified by column chromatography on silica gel.

[(5-Benzoyl-6-oxo-5-azaspiro[2.3]hexane-4-carbonyl)tert-Butyl amino]acetate (13): Column chromatography ($R_{\rm f} = 0.10, 12 \text{ g of}$ silica gel, 2×10 cm column, hexane/Et₂O, 1:1) of the residue obtained from the β -lactam 9a (113 mg, 0.436 mmol) and glycinate 12 (171 mg, 1.31 mmol) in DMF (10 mL) according to GP3 (20 h) gave the glycinate 13 (80 mg, 51%) as a colorless solid, m.p. 112–115 °C. IR (KBr): $\tilde{v} = 3400 \text{ cm}^{-1}$ (br.), 2982, 2925, 2853, 1790, 1719, 1644, 1604, 1581, 1516, 1487. ¹H NMR (250 MHz): $\delta = 1.08 - 1.19$ (m, 1 H, cPr-H), 1.31 - 1.45 (m, 2 H, cPr-H), 1.48(s, 9 H, 3 CH₃), 1.58-1.66 (m, 1 H, cPr-H), 4.27 (s, 2 H, CH₂), 5.03 (d, J = 7.4 Hz, 1 H, CH), 6.91 (d, J = 7.4 Hz, 1 H, NH), 7.39-7.47 (m, 2 H, Ar-H), 7.5-7.57 (m, 1 H, Ar-H), 7.75-7.81 (m, 2 H, Ar-H) ppm. ¹³C NMR (150 MHz): $\delta = 13.3$ (CH₂), 15.2 (CH₂), 27.2 (C), 28.0 (3 CH₃), 40.7 (CH₂), 53.4 (CH), 83.2 (C), 127.2 (2 CH), 128.7 (2 CH), 132.3 (CH), 132.5 (C), 165.9 (C), 167.8 (C), 174.7 (C), 177.4 (C) ppm. MS (DCI): m/z (%) = 734 (10) [2 $M + NH_4^+$], 376 (100) [M + NH_4^+]. $C_{19}H_{22}N_2O_5$ (358.39) calcd. C 63.67, H 6.19, N 7.82; found C 63.33, H 5.95, N 7.71.

tert-Butyl {[1-(Benzoylaminophenylmethyl)cyclopropylcarbonyl]amino}acetate (15): Column chromatography ($R_{\rm f} = 0.17, 15 \,{\rm g}$ of silica gel, 2×10 cm column, hexane/Et₂O, 1:2) of the residue obtained from the β -lactam 9b (243 mg, 0.877 mmol) and glycinate 12 (354 mg, 2.7 mmol) in DMF (35 mL) according to GP3 (12 h) gave 15 (217 mg, 61%) as a colorless solid, m.p. 133-134 °C. IR (KBr): $\tilde{v} = 3311 \text{ cm}^{-1}$, 3238, 3064, 2974, 1754, 1734, 1656, 1636. ¹H NMR (300 MHz): $\delta = 0.99 - 1.29$ (m, 4 H, cPr-H), 1.36 (s, 9 H, 3 CH₃), 3.71 (dd, J = 18.3, 5.0 Hz, 1 H, CH₂), 3.78 (dd, J =18.3, 5.0 Hz, 1 H, CH₂), 4.76 (d, J = 8.7 Hz, 1 H, CH), 5.81 (t, J = 5.0 Hz, 1 H, NH), 7.12–7.27 (m, 3 H, Ar-H), 7.32–7.47 (m, 5 H, Ar-H), 7.85-7.89 (m, 2 H, Ar-H), 8.66 (d, J = 8.7 Hz, 1 H, NH) ppm. ¹³C NMR (75.5 MHz): $\delta = 13.1$ (CH₂), 14.1 (CH₂), 27.9 (3 CH₃), 29.5 (C), 41.8 (CH₂), 58.3 (CH), 82.5 (C), 126.5 (2 CH), 127.2 (2 CH), 127.4 (CH), 128.5 (4 CH), 131.5 (CH), 134.1 (C), 140.2 (C), 166.5 (C), 168.8 (C), 173.5 (C) ppm. MS (EI): m/z (%) = 408 (2) [M⁺], 335 (10), 277 (85), 250 (15), 210 (15), 105 (100), 77 (30), 57 (22). MS (DCI): m/z (%) = 426 (55) [M + NH₄⁺], 409 (100) $[M + H^+]$. $C_{24}H_{28}N_2O_4$ (408.5): calcd. C 70.57, H 6.91, N 6.86; found C 70.76, H 7.10, N 7.26.

tert-Butyl {[1-(tert-Butoxycarbonylaminocyanomethyl)cyclopropylcarbonyllamino}acetate (16): The colorless solid (0.20 g, 84%) obtained from β-lactam 11 (150 mg, 0.67 mmol), tert-butyl glycinate hydrochloride (12·HCl) (195 mg, 1.16 mmol) and triethylamine (118 mg, 162 µL, 1.16 mmol) in DMF (16 mL) according to GP3 (12 h) after evaporation of the solvent was essentially pure acetate 16. An analytical sample was obtained by column chromatography $(R_{\rm f} = 0.70, 20 \text{ g of silica gel}, 2 \times 16 \text{ cm column}, \text{Et}_2\text{O})$ and had m.p. 86–88 °C. IR (KBr): $\tilde{v} = 3311 \text{ cm}^{-1}$, 3096, 2989, 2939, 1751, 1688, 1638, 1550, 1511. ¹H NMR (300 MHz): $\delta = 1.08 - 1.16$ (m, 1 H, cPr-H), 1.17-1.28 (m, 2 H, cPr-H), 1.30-1.37 (m, 1 H, cPr-H), 1.44 (s, 9 H, 3 CH₃), 1.47 (s, 9 H, 3 CH₃), 3.87 (dd, J =18.4, 4.4 Hz, 1 H, CH₂), 3.94 (dd, J = 18.4, 4.4 Hz, 1 H, CH₂), 4.24 (d, J = 9.2 Hz, 1 H, CH), 5.83 (br. t, J = 4.4 Hz, 1 H, NH), 6.20 (br. d, J = 9.2 Hz, 1 H, NH) ppm. ¹³C NMR (50.3 MHz): $\delta = 13.2 (CH_2), 14.8 (CH_2), 27.9 (C), 28.0 (3 CH_3), 28.2 (3 CH_3),$ 41.9 (CH₂), 47.0 (CH), 81.2 (C), 82.8 (C), 117.4 (C), 168.8 (2 C), 171.2 (C) ppm. MS (EI): m/z (%) = 353 (2) [M⁺], 297 (15), 224 (28), 197 (42), 57 (100). MS (DCI): m/z (%) = 724 (10) [2 M + NH_4^+], 707 (18) [2 M + H⁺], 371 (85) [M + NH_4^+], 354 (100) [M + H⁺]. C₁₇H₂₇N₃O₅ (353.41): calcd. C 57.77, H 7.70, N 11.89; found C 57.49, H 7.58, N 11.81.

tert-Butyl (2*S*,2'*S*)-2-{[1-(*tert*-Butoxycarbonylaminocyanomethyl)cyclopropylcarbonyl]amino}-3-phenylpropionate [(2*S*,2'*S*)-17] and *tert*-Butyl (2*S*,2'*R*)-2-{[1-(*tert*-Butoxycarbonylaminocyanomethyl)cyclopropylcarbonyl]amino}-3-phenylpropionate [(2*S*,2'*R*)-17]: Column chromatography (44 g of silica gel, 3×13 cm column, hexane/ Et₂O, 1:1) of the residue obtained from the β -lactam 11 (184 mg, 0.83 mmol), *tert*-butyl (*S*)-phenylalaninate hydrochloride (14·HCl) (428 mg, 1.66 mmol) and Et₃N (168 mg, 230 µL, 1.66 mmol) in DMF (33 mL) according to GP3 (60 °C, 12 h) gave (2*S*,2'*S*)-17 (155 mg, 42%) and (2*S*,2'*R*)-17 (143 mg, 39%) as colorless solids.

The diastereoisomer (2*S*,2'*S*)-17: $R_{\rm f} = 0.34$, m.p. 101–102 °C, $[\alpha]_{D}^{20} = +53.4 \ (c = 0.5 \text{ in CHCl}_{3}). \text{ IR (KBr): } \tilde{\nu} = 3438 \text{ cm}^{-1}, 3426,$ 3090, 2981, 2935, 2247, 1738, 1719, 1650, 1540, 1524, 1370. ¹H NMR (250 MHz): $\delta = 1.00 - 1.22$ (m, 4 H, *c*Pr-H), 1.41 (s, 9 H, 3 CH_3), 1.43 (s, 9 H, 3 CH₃), 3.06 (dd, $J = 15, 5 \text{ Hz}, 1 \text{ H}, \text{ CH}_2$), $3.15 (dd, J = 15, 5 Hz, 1 H, CH_2), 4.18 (d, J = 8.7 Hz, 1 H, CH),$ 4.65 (dd, J = 12.5, 5 Hz, 1 H, CH), 5.71 (br. d, J = 8.7 Hz, 1 H, NH), 6.12 (br. d, J = 7.6 Hz, 1 H, NH), 7.11–7.14 (m, 2 H, Ar-H), 7.23–7.29 (m, 3 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta =$ 13.2 (CH₂), 14.7 (CH₂), 27.9 (3 CH₃), 28.2 (3 CH₃), 37.4 (CH₂), 46.9 (CH), 53.4 (CH), 81.1 (C), 82.7 (C), 117.3 (C), 127.1 (CH), 128.4 (2 CH), 129.4 (2 CH), 135.6 (C), 170.2 (C), 170.7 (C) ppm; two carbon atom signals were not detectable at this temperature. ¹³C NMR (75.5 MHz, C₂D₂Cl₄, 100 °C): δ = 12.5 (CH₂), 13.9 (CH₂), 27.5 (C), 27.7 (3 CH₃), 28.0 (3 CH₃), 37.4 (CH₂), 47.1 (CH), 53.5 (CH), 81.1 (C), 82.5 (C), 117.0 (C), 126.9 (CH), 128.2 (2 CH), 129.1 (2 CH), 135.7 (C), 154.3 (C), 169.8 (C), 170.2 (C) ppm. MS (EI): m/z (%) = 443 (9) [M⁺], 387 (10), 331 (28), 314 (30), 286 (22), 242 (10), 148 (100), 120 (85), 91 (15), 57 (40). C₂₄H₃₃N₃O₅ (443.5): calcd. C 64.99, H 7.50, N 9.47; found C 64.76, H 7.76, N 9.60. The diastereoisomer (2*S*,2'*R*)-17: $R_{\rm f} = 0.27$, m.p. 142–143 °C, $[\alpha]_{\rm D}^{20} =$ +22.5 (c = 0.16 in CHCl₃). IR (KBr): $\tilde{v} = 3374$ cm⁻¹, 3281, 3036, 2970, 2250, 1739, 1690, 1649, 1535, 1369. ¹H NMR (250 MHz): $\delta = 0.99 - 1.26$ (m, 4 H, cPr-H), 1.43 (s, 9 H, 3 CH₃), 1.46 (s, 9 H, 3 CH₃), 3.04 (dd, J = 12.5, 5 Hz, 1 H, CH₂), 3.12 (dd, J = 12.5, 7.5 Hz, 1 H, CH₂), 4.10 (d, J = 10 Hz, 1 H, CH), 4.71 (dd, J =12.5, 5 Hz, 1 H, CH), 5.67 (br. d, J = 10 Hz, 1 H, NH), 6.15 (br. d, J = 12.5 Hz, 1 H, NH), 7.06-7.10 (m, 2 H, Ar-H), 7.24-7.31 (m, 3 H, Ar-H) ppm. ¹³C NMR (62.9 MHz): $\delta = 13.3$ (CH₂), 14.7 (CH₂), 27.9 (3 CH₃), 28.2 (3 CH₃), 37.7 (CH₂), 47.1 (CH), 53.3

(CH), 81.1 (C), 83.0 (C), 117.5 (C), 127.1 (CH), 128.4 (2 CH), 129.4 (2 CH), 135.8 (C), 170.2 (C), 170.3 (C) ppm, two carbon atom signals were not detectable at this temperature. ¹³C NMR (75.5 MHz, C₂D₂Cl₄, 100 °C): δ = 12.5 (CH₂), 13.9 (CH₂), 27.6 (C), 27.7 (3 CH₃), 28.0 (3 CH₃), 37.7 (CH₂), 47.1 (CH), 53.4 (CH), 81.1 (C), 82.7 (C), 117.0 (C), 126.8 (CH), 128.2 (2 CH), 129.1 (2 CH), 135.8 (C), 154.3 (C), 169.9 (C), 170.1 (C) ppm. MS (EI): *m*/*z* (%) = 443 (38), 387 (35), 331 (75), 314 (65), 286 (68), 240 (25), 184 (22), 148 (100), 120 (55), 57 (50). C₂₄H₃₃N₃O₅ (443.5): calcd. C 64.99, H 7.50, N 9.47; found C 64.77, H 7.62, N 9.42.

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- ^[1] ^[1a] Review: J. Suckling, Angew. Chem. 1988, 100, 555-570; Angew. Chem. Int. Ed. Engl. 1988, 27, 537-552. For selected communications, see also: ^[1b] S. C. M. Fell, M. J. Pearson, G. Burton, J. Bateson, J. Chem. Soc., Perkin Trans. 1 1991, 1361-1363, and references cited therein. ^[1c] J. H. Bateson, A. W. Guest, Tetrahedron Lett. 1993, 34, 1799-1802. ^[1d] J. C. Sheehan, E. Chacko, Y. S. Lo, D. R Ponzi, E. Sato, J. Org. Chem. 1978, 43, 4856-4859.
- ^[2] ^[2a] F. M. Cordero, M. Salvati, F. Pisaneschi, A. Brandi, *Eur. J. Org. Chem.* 2004, 2205–2213. ^[2b] F. M. Cordero, F. Pisaneschi, M. Salvati, V. Paschetta, J. Ollivier, J. Salaün, A. Brandi, *J. Org. Chem.* 2003, 68, 3271–3280. ^[2c] V. Paschetta, F. M. Cordero, R. Paugam, J. Ollivier, A. Brandi, J. Salaün, *Synlett* 2001, 1233–1236. ^[2d] F. M. Cordero, F. Pisaneschi, A. Goti, J. Ollivier, J. Salaün, A. Brandi, *J. Am. Chem. Soc.* 2000, 122, 8075–8076.
- ^[3] The preparation and chemical transformations of methylene-cyclopropane (1) itself^[3a-c] and its close relative bicyclopropylidene (2)^[3d-g] have recently been reviewed extensively: ^[3a] P. Binger, H. M. Büch, *Top. Curr. Chem.* 1987, *135*, 77–151. ^[3b] P. Binger, T. Schmidt, in *Methods of Organic Chemistry (Houben-Weyl)*, *Vol. E 17c* (Ed.: A. de Meijere), Thieme, Stuttgart, 1997, pp. 2217–2294. ^[3c]A. Brandi, A. Goti, *Chem. Rev.* 1998, *98*, 589–635. ^[3d] A. de Meijere, S. I. Kozhushkov, A. F. Khlebnikov, *Zh. Org. Khim.* 1996, *32*, 1607–1626; *Russ. J. Org. Chem. (Engl. Transl.)* 1996, *32*, 1555–1575. ^[3e] A. de Meijere, S. I. Kozhushkov, *Eur. J. Org. Chem.* 2000, *309*–3822. ^[3g] A. de Meijere, S. I. Kozhushkov, *Eur. J. Org. Chem.* 2000, 3809–3822. ^[3g] A. de Meijere, S. I. Kozhushkov, *Eur. J. Org. Chem.* 2000, 3809–3822. ^[3g] A. de Meijere, S. I. Kozhushkov, *Eur. J. Org. Chem.* 2000, 3809–3822. ^[3g] A. de Meijere, S. I. Kozhushkov, *Eur. J. Org. Chem.* 2000, 3809–3822. ^[3g] A. Braid, H. Nüske, T. Pohlmann, M. Es-Sayed, S. Bräse, *Pure Appl. Chem.* 2000, *72*, 1745–1756.
- ^[4] ^[4a] S. Abele, D. Seebach, *Eur. J. Org. Chem.* 2000, 1–15. ^[4b] Y. Yamamoto, N. Assao, N. Tsukada, in *Advances in Asymmetric Synthesis* (Ed.: A. Hassner), JAI Press Inc., Stamford, 1988, vol. 3. ^[4c] G. Cardillo, C. Tomasini, *Chem. Soc. Rev.* 1996, 25, 117–128. ^[4d] *Enantioselective Synthesis of β-Amino Acids* (Ed.: E. Juaristi), Wiley-VCH, New York, 1997. ^[4e] E. Juaristi, D. Quintana, J. Escalante, *Aldrichim. Acta* 1994, 27, 3–11.
- ^[5] F. Gnad, O. Reiser, *Chem. Rev.* **2003**, *103*, 1603–1623, and references cited therein.
- ^[6] ^[6a] S. Abele, P. Seiler, D. Seebach, *Helv. Chim. Acta* **1999**, *82*, 1559–1571. For the conformational properties of β-alanines **8**, see also: ^[6b] J.-F. Tocanne, *Tetrahedron* **1972**, *28*, 389–416. ^[6c]

M. Pelissier, A. Serafini, J. Devanneaux, J.-F. Labarre, J.-F. Tocanne, *Tetrahedron* **1971**, *27*, 3271–3284.

- [7] Reviews: ^[7a] A. Brandi, S. Cicchi, F. M. Cordero, A. Goti, *Chem. Rev.* 2003, 103, 1213–1269. ^[7b] A. Goti, F. M. Cordero, A. Brandi, *Top. Curr. Chem.* 1996, 178, 1–97.
- [8] O. Tamura, T. Okabe, T. Yamaguchi, K. Gotanda, K. Noe, M. Sakamoto, *Tetrahedron* 1995, *51*, 107–118.
- [9] S. Cicchi, M. Corsi, A. Goti, J. Org. Chem. 1999, 64, 7243-7245.
- ^[10] H. Tokuyama, T. Kuboyama, A. Amano, T. Yamashita, T. Fukuyama, *Synthesis* 2000, 1299–1304.
- ^[11] T. Shimizu, M. Ishizaki, N. Nitada, *Chem. Pharm. Bull.* **2002**, 50, 908–921.
- ^[12] Crystals of the compounds were grown by slow evaporation of their solutions in hexane (5b, 9a) or diethyl ether [(2S, 2'R)-17]. The X-ray single crystal data were collected at 133(2) K on a STOE-IPDS 2 Image Plate diffractometer using graphite monochromated Mo- K_{α} radiation. The structure solutions and refinements on F_2 were performed with the Bruker SHELXTL program suite. **5b**: $C_{20}H_{21}NO$ (M = 291.38), monoclinic, a =9.2097(7), b = 5.6716(3), c = 30.290(2) Å, $a = \gamma = 90$, $\beta = 94.307(6)^\circ$, V = 1577.70(19) Å³, Z = 4, space group $P 2_1/n$, $\rho = 1.227 \text{ g cm}^{-3}$, intensities measured: 7117 ($2\theta_{\text{max}} = 46.50^{\circ}$), independent: 2237 ($R_{int} = 0.0327$), 199 parameters refined, $R_1 = 0.0310$ for 2237 reflections with $I > 2\sigma(I)$, wR_2 (all data) = 0.0768, Goof = 1.040, maximum and minimum residual electron density 0.183 and $-0.136 \text{ e}\cdot \text{\AA}^{-3}$. **9a**: $C_{14}H_{13}NO_4$ (M = 259.25), monoclinic, a = 10.7153(6), b =8.9670(4), c = 13.7210(6) Å, $a = \gamma = 90$, $\beta = 109.056(4)^{\circ}$, V =1246.12(10) Å³, Z = 4, space group $P 2_1/c$, T = 133(2) K, ρ = 1.382 g cm⁻³, intensities measured: 24299 ($2\theta_{\text{max}} = 49.42^{\circ}$), independent: 2073 ($R_{int} = 0.0326$), 173 parameters refined, $R_1 =$ 0.0305 for 2073 reflections with $I > 2\sigma(I)$, wR_2 (all data) = 0.0709, Goof = 1.072, maximum and minimum residual electron density 0.258 and $-0.170 \text{ e} \cdot \text{\AA}^{-3}$. (2S,2'R)-17: C₂₄H₃₃N₃O₅ (M = 443.53), monoclinic, a = 11.905(5), b = 9.2517(16), c =23.731(6) Å, $a = \gamma = 90, \beta = 94.25(3)^{\circ}, V = 2672.1(13) Å^3$, Z = 4, space group $P 2_1$, $\rho = 1.145$ g cm⁻³, intensities measured: 21983 ($2\theta_{\text{max}} = 49.68^{\circ}$), independent: 8604 ($R_{\text{int}} =$ 0.0549), 597 parameters refined, $R_1 = 0.0457$ for 8604 reflections with $I > 2\sigma(I)$, wR_2 (all data) = 0.1011, Goof = 0.994, maximum and minimum residual electron density 0.247 and -0.225 e·Å⁻³. Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited as supplementary publication no. CCDC-240722 (for **5b**), -240723 (for **9a**) and -240724 [for (2S, 2'R)-**17**] with the Cambridge Crystallographic Data Centre. Copies of the data can be obtained free of charge on application to The Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (Fax: +44-1223-336-033; E-mail: deposit@ccdc.cam.ac.uk).
- ^[13] For another, yet more complicated approach to such compounds, see also: A. de Meijere, S. Teichmann, D. Yu, J. Kopf, M. Oly, N. von Thienen, *Tetrahedron* **1989**, *45*, 2957–2968.
- [¹⁴] ^[14a] T. B. Durham, M. J. Miller, *J. Org. Chem.* 2003, 68, 35–42.
 [^{14b]} C. Palomo, J. M. Aizpurva, R. Galarza, A. Benito, U. K. Khamrai, A. Linden, *Tetrahedron* 2000, 56, 5563–5570. ^[14c] S.-K. Khim, J. M. Nuss, *Tetrahedron Lett.* 1999, 40, 1827–1830.
- ^[15] Ring opening of β -lactams with aryl substituents at C-4 under hydrogenolytic conditions has previously been observed: ^[15a] I. Ojima, S. Suga, R. Abe, *Tetrahedron Lett.* **1980**, *21*, 3907-3910. ^[15b] I. Ojima, N. Hatanaka, N. Yodo, R. Abe, M. Yatabe, M. Yamashita, in *Peptide Chemistry 1982* (Ed.: S. Sakakibara), Protein Research Foundation, Osaka, **1983**, pp. 29-34.
- ^[16] [^{16a]} G. D. Annis, E. M. Hebblethwaite, S. T. Hodgson, D. M. Hollinshead, S. V. Ley, *J. Chem. Soc., Perkin Trans. 1* **1983**, 2851–2856. ^[16b] J. Aszodi, A. Bonnet, G. Teutsch, *Tetrahedron* **1990**, *46*, 1579–1586.
- [17] [17a] V. Moreaux, H. Warren, J. M. Williams, *Tetrahedron Lett.* 1997, 38, 4655–4658. ^[17b] J. M. Blanco, O. Caamanno, F. Fer-

nandez, X. Garsia Mera, C. Lopez, G. Rodriguez, J. E. Rodriguez-Borges, A. Rodriguez-Hergueta, *Tetrahedron Lett.* **1998**, *39*, 5663–5666. ^[17c] B. Alcaide, A. Gomez, J. Plumet, J. Rodrigez-Lopez, *Tetrahedron* **1989**, *45*, 2751–2762. ^[17d] C. Palomo, M. Qiarbide, S. Bindi, *J. Org. Chem.* **1998**, *63*, 2469–2474.

M. Oiarbide, S. Bindi, *J. Org. Chem.* **1998**, *63*, 2469–2474. ^[18] ^[18a] A. de Meijere, S. I. Kozhushkov, T. Spaeth, N. S. Zefirov, J. Org. Chem. **1993**, 58, 502–505. ^[18b] A. de Meijere, S. I. Kozhushkov, T. Späth, Org. Synth. **2000**, 78, 142–151.

- ^[19] A. Vollmar, M. S. Dunn, J. Org. Chem. 1960, 25, 387-390.
- ^[20] N. N. Schwarz, J. N. Blumbergs, J. Org. Chem. 1964, 29, 1976–1979.

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