Synthesis of some tricyclic heteroaromatic systems and their A_1 and A_{2a} adenosine binding activity

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Summary — The syntheses, A_1 and A_{2a} adenosine receptor affinities and structure-activity relationships of some 2-aryl-1,2,4-triazolo[1,5-*a*]quinoxalines, 2-arylimidazo[1,2-*a*]quinoxalines, 1-arylimidazo[1,5-*a*]quinoxalines are reported and compared with that of a previously reported 2-phenylpyrazolo[1,5-*a*]quinoxaline. The results show that some triazoloquinoxalines are potent and specific A_1 adenosine receptor ligands and that the replacement of either nitrogen at position 1 or 3 of the triazoloquinoxaline moiety with a CH brought about a decrease in affinity at both adenosine receptors.

adenosine receptor ligand / 1,2,4-triazolo[1,5-a]quinoxaline / imidazo[1,2-a]quinoxaline / imidazo[1,5-a]quinoxaline / tricyclic heteroaromatic system

Introduction

Adenosine produces a variety of physiological responses by interacting with two major extracellular receptor subtypes, A1AR and A2AR, which were originally defined on the basis of whether their activation inhibited or stimulated adenyl cyclase, respectively [1]. The A_2AR has been further divided into high and low affinity subtypes, $A_{2a}AR$ and $A_{2b}AR$, respectively, after extensive structure-activity relationship studies [2, 3], biochemistry [4], binding studies [5-8] and the cloning of separate entities [9-12]. Another adenosine receptor, A_3AR , has recently been identified by cloning [13–15], while binding studies with [³H]CV 1808 [16] have led to the pharmacological identification of a new adenosine receptor, tentatively named A4AR. A3AR displays an unusual structural diversity for species homologues, and has a broad tissue distribution in human and sheep where it binds with high affinity some xanthines with acidic side chains or NH₂ groups[14].

Receptor subtypes are generally defined by their different affinities for specific synthetic agonists and antagonists. The synthesis of new adenosine receptor ligands would thus provide a valuable tool for defining the structural requirements of each receptor subtype. With this in mind, and as a further development of our previous work on the preparation of non-xanthine antagonists at A_1AR and A_2AR [17–19], we extended our investigations to the study of the tricyclic compounds 1–18 shown in figure 1. These

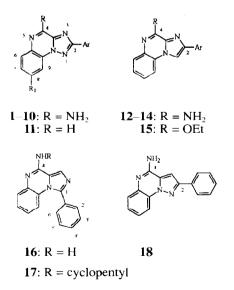


Fig 1. Chemical structures of the tricyclic systems 1-18.

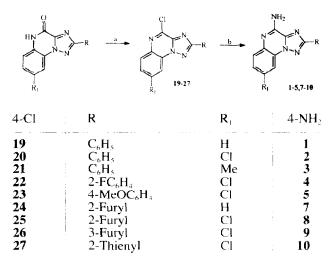
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investigations resulted in the synthesis of some 2aryl-1,2,4-triazolo[1,5-*a*]quinoxalin-4-amines 1–10, 2arylimidazo[1,2-*a*]quinoxalines 12–15 and 1-arylimidazo[1,5-*a*]quinoxalines 16–17. All these newly reported compounds, together with the previously reported 2-phenyl-1,2,4-triazolo[1,5-*a*]quinoxaline [20] 11 and 2-phenylpyrazolo[1,5-*a*]quinoxalin-4-amine [21] 18, were studied for their interactions with both A_1AR and $A_{2a}AR$.

Chemistry

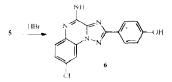
The final newly reported tricyclic heteroaromatic derivatives 1-5, 7-10, and 12-17 were prepared from the corresponding 4-chloro key intermediates 19-30and 34.

The syntheses of the 2-aryl-1,2,4-triazolo[1,5-*a*]quinoxalines 1–10 are illustrated in scheme 1. By reacting 2-aryl-1,2,4-triazolo[1,5-*a*]quinoxalin-4-ones [20, 22] with a mixture of POCl₃/PCl₅, the corresponding 4-chloro derivatives **19–27** were obtained. Nucleophilic replacement of the 4-chloro group with ammonia yielded compounds 1–5 and 7–10.

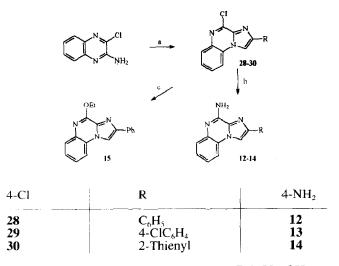


Scheme 1. a: POCl₃/PCl₅/pyridine: b: ammonia.

The 2-(4-hydroxyphenyl)-1,2,4-triazolo[1,5-a]quinoxalin-4-amine **6** ensued from dealkylation of the corresponding 2-(4-methoxyphenyl) derivative **5**.



The syntheses of the 2-arylimidazo[1,2-a]quinoxalines 12–15 are illustrated in scheme 2. By allowing 2-chloroquinoxalinamine [23] to react with bromomethyl-4-chlorophenylketone, following the procedure reported in the literature [22] for the preparation of compounds 28 and 30, compound 29 was obtained. Treatment with ammonia of 28–30 afforded compounds 12–14, while by reacting 28 with an ethanolic solution of sodium hydroxide the 4-ethoxy derivative 15 was prepared.

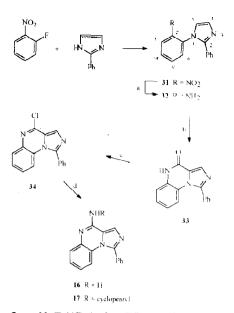


Scheme 2. a: RCOCH₂ Br; b: ammonia; c: EtOH/NaOH.

Finally, the syntheses of the 1-phenylimidazo[1,5-a]quinoxalines 16–17 are shown in scheme 3. Reaction of 1-fluoro-2-nitrobenzene with 2-phenylimidazole yielded the 1-(2-nitrophenyl)-2-phenylimidazole 31, which was catalytically reduced to the amino derivative 32. Treatment of compound 32 with 1,1'-carbonyldiimidazole afforded the tricyclic 1-phenylimidazo[1,5-a]quinoxalin-4-one 33 which, with POCl₃, gave the 4-chloro derivative 34. The latter with ammonia or cyclopentylamine gave the final compounds 16 and 17, respectively.

Biochemistry

Compounds 1–18 were tested for their ability to displace [3 H]- N^{6} -cyclohexyladenosine (CHA) on A₁AR in rat cerebral cortical membranes and [3 H]-2-[4-((2-carboxyethyl)phenethyl)amino]-5'-N-ethylcarboxamidoadenosine (CGS 21680) on A_{2a}AR in rat striatal membranes. The A₁AR and A_{2a}AR affinities of the tested compounds, expressed as their K_i values, are listed in table I.



Scheme 3. a: H₂/Pd/C; b: Im₂CO; c: POCl₃; d: ammonia or cyclopentylamine.

Results and conclusions

The tricyclic heteroaromatic systems are arranged in table I according to the modifications of the pentatomic ring, which range from 1,2,4-triazole, to differently fused imidazole and pyrazole.

Table I shows that the highest A AR affinities are displayed by the 2-aryl-1,2,4-triazolo[1,5-a]quinoxalin-4-amines 1-10. The adenosine receptor affinity within this subgroup of compounds is clearly dependent on the presence of the NH₂ group at position 4, since the 4-unsubstituted triazologuinoxaline 11 shows a substantial reduction in A₁AR affinity. For compounds 1–10, variation of the R_1 substituent at position 8 from hydrogen to chlorine or methyl produces little or no increase in potency. Variation in compounds 1-10 of the 2-substituent yielded different A_{2a}AR affinities. In fact, compounds 7-8 bearing a 2-(2-furyl) ring are the only ones that showed high $A_{2a}AR$ affinity. The substituent at position 2 was also important for A_1/A_{2a} selectivity. The 2-unsubstituted phenyl ring gave the non-selective adenosine ligands 1-3, while ortho or para substitution of the phenyl ring afforded the A_1 -selective ligands 4-6. The highly potent A_{2a} ligands 7-8 were however non-selective, displaying a 30 nM A₁ affinity. The 2-(3-furyl) and 2-(2-thienyl) substituents gave rise to the A1-selective ligands 9 and 10, respectively.

All the imidazo[1,2-*a*]quinoxalines 12–15 are less active at both A₁AR and A_{2a}AR than their 1-aza analogues triazolo-quinoxalines. In this subgroup of compounds the presence of the NH₂ group at position 4 is not necessary. In fact, 4-ethoxyimidazo[1,2-*a*] quinoxaline 15 displayed A_1 and A_{2a} potency comparable to that of the other 4-amino derivatives 12–14.

Variation of the kind of condensation between the imidazole and quinoxaline moiety occurring in compounds **16–17** does not enhance the inhibitory potency at either A_1AR or $A_{2a}AR$.

Finally, the 2-phenylpyrazolo[1,5-*a*]quinoxalin-4amine [21] **18**, displayed a 53-fold and 11-fold lower A_1 and A_{2a} binding activity, respectively, as compared to the 3-aza analogue 2-phenyl-1,2,4-triazolo[1,5-*a*] quinoxalin-4-amine **1**, thus discouraging further syntheses of this kind of compounds.

In conclusion, the syntheses of these tricyclic heteroaromatic systems have provided three potent and selective A_1 adenosine receptor ligands, compounds 4, 9 and 10, while the structure-activity relationships have shown that replacement of either nitrogen at position-1 or position-3 of the triazoloquinoxalines 1–10 with a CH resulted in a decrease in binding potency at A_1AR and/or $A_{2a}AR$. These findings are consistent with the chemical features of the natural ligand, the adenosine. The nitrogen atoms at position 1 and 3 of the triazole ring mimic those of position 7 and 9 of the adenosine and are thus necessary for the anchoring of a ligand to both A_1 and A_{2a} receptors.

Experimental protocols

Chemistry

Silica-gel plates (Merck F₂₅₄) and silica gel 60 (Merck, 70-230 mesh) were used for analytical and column chromatography, respectively. All melting points were determined on a Gallenkamp capillary melting point apparatus. Microanalyses were performed with a Perkin-Elmer 260 elemental analyzer for C, H, N and the results were within $\pm 0.4\%$ of the theoretical values. The IR spectra were recorded with a Perkin-Elmer 1420 spectrometer in nujol mull and are reported in cm-1. The ¹H-NMR spectra were run on a Varian Gemini 200 instrument in the Fourier transform mode at 200 MHz using an acquisition time of 1.5 s, a flip angle of 37° and a spectral width of 3000 Hz. The deuterium signal of the solvent provided the field frequency lock. Chemical shifts are reported in δ (ppm) and refer to the central peak of the solvent (CDCl₃ = 1385 Hz, DMSO $d_6 = 792$ Hz). The following abbreviations are used: s = singlet, d = doublet, dd = double doublet, t = triplet, q = quartet, m = multiplet, br = broad, ar = aromatic proton.

The physical data of the newly synthesized compounds are listed in table II.

2-Aryl-4-chloro-8- R_1 -1,2,4-triazolo[1,5-a]quinoxalines 19–27 A mixture of the suitable 2-aryl-8- R_1 -1,2,4-triazolo[1,5-a]quinoxalin-4-one [20, 22] (1 mmol), POCl₃ (5 mL), PCl₅ (0.3 mmol) and a catalytic amount of pyridine was heated under stirring and nitrogen flow at 120°C for 3 h. Evaporation of the excess POCl₃ at reduced pressure afforded a crude product which was used without further purification. The yields are calculated on the crude product. Only a small amount of crude product was recrystallized to determine the melting point and the ¹H-NMR. **Table I.** A_1 and A_{2a} binding constants of the reported tricyclic heteroaromatic systems^a.



Compound	R	X	Ŷ	Ζ	R_{I}	$K_i \pm SEM (nM)^{b}$	
						A_{I}^{c}	A_{2a}^{d}
1	NH ₂	N	C-C ₆ H ₅		Н	57 ± 6	300 ± 28
2	\mathbf{NH}_{2}	Ν	C-C ₆ H ₅	Ν	Cl	50 ± 4	161 ± 15
3	NH2	Ν	$C-C_{6}H_{5}$	Ν	Me	39 ± 3.7	107 ± 9.2
4	$\overline{NH_2}$	Ν	$C - C_6 H_4 F(2)$	Ν	Cl	13 ± 1.5	>10 000
5	NH_2	Ν	$C-C_6H_4OMe(4)$	Ν	Cl	150 ± 12	>10 000
6	NH2	Ν	$C-C_6H_4OH(4)$	Ν	Cl	36 ± 2	3170 ± 298
7	NH_2	Ν	C-furyl(2)	Ν	H	30 ± 2.3	11 ± 0.9
8	NH ₂	Ν	C-furyl(2)	Ν	Cl	30 ± 2	12 ± 1.1
9	NH_2	Ν	C-furyl(3)	Ν	Cl	24 ± 2.1	>10 000
10	NH ₂	Ν	C-thienyl(2)	Ν	Cl	12 ± 1	>10 000
11	н	Ν	C-C ₆ H ₅	Ν	Н	1300 ± 107	>10 000e
12	NH ₂	СН	C-C ₆ H ₅	Ν	Н	230 ± 20	720 ± 81^{e}
13	NH_2	СН	$C-C_6H_4Cl(4)$	Ν	Н	530 ± 51	4430 ± 519^{e}
14	NH,	CH	C-thienyl(2)	Ν	Н	730 ± 61	>10 000e
15	OEt	CH	C ₆ H ₅	Ν	H	680 ± 59	450 ± 39^{e}
16	NH ₂	$C-C_6H_5$	N	СН	н	130 ± 11	310 ± 30^{e}
17	NH-cyclopentyl	C-C ₆ H,	Ν	CH	Н	810 ± 73	>10 000
18	NH ₂	N	$C-C_6H_5$	CH	Н	3050 ± 298	$3380 \pm 328^{\circ}$

^aThe tests were carried out dissolving the tested compound in DMSO (DMSO/buffer 2%) unless otherwise stated. ^bThe K_i values are means \pm SEM of four separate assays, each performed in triplicate. ^cA₁ binding was measured as inhibition of [³H]CHA binding as described in the *Experimental protocols*. ^dA_{2a} binding was measured as inhibition of [³H]CGS 21680 binding as described in the *Experimental protocols*. ^eThe tests were carried out dissolving the tested compound in ethanol (ethanol/buffer 2%).

The title compounds displayed the following ¹H-NMR data: 19: (CDCl₃): 7.4-7.6 (m, 3Ĥ, H-3'-5'), 7.7-7.9 (m, 2H, H-7, 8), 8.14 (d, 1H, H-6, J = 8.2 Hz), 8.3-8.4 (m, 2H, H-2',6'), 8.52 (d, 1H, H-9, J = 8.2 Hz). 20: (CDCl₃): 7.5-7.6 (m, 3H, H-3'-5'), 7.68 (dd, 1H, H-7, J = 8.7, 2.2 Hz), 8.08 (d, 1H, H-6, J =8.7 Hz), 8.3–8.5 (m, 2H, H-2',6'), 8.53 (d, 1H, H-9, J = 2.2 Hz). 21: (DMSO-d₆): 2.66 (s, 3H, CH₃), 7.6–7.7 (m, 4H, H-3'-5', 7), 8.07 (d, 1H, H-6, J = 8.7 Hz), 8.3–8.4 (m, 3H, H-2',6',9). 22: (CDCl₃): 7.2-7.4 (m, 2H, H-3',5'), 7.5-7.6 (m, 1H, H-4'), 7.69 (dd, 1H, H-7, J = 8.7, 2.2 Hz), 8.09 (d, 1H, H-6, J = 8.7 Hz),8.3-8.4 (m, 1H, H-6'), 8.55 (d, 1H, H-9, J = 2.2 Hz). 23: $(CDCl_3)$: 3.91 (s, 3H, OCH_3), 7.05 (d, 2H, H-3',5', J = 8.8 Hz), 7.66 (dd, 1H, H-7, J = 8.7, 2.2. Hz), 8.06 (d, 1H, H-6, J = 8.7 Hz), 8.33 (d, 2H, H-2', 6', J = 8.8 Hz), 8.50 (d, 1H, H-9, J = 2.2Hz). 24: (CDCl₃): 6.60 (dd, 1H, H-4', J = 3.4, 1.7 Hz), 7.3–7.7 (m, 4H, H-7, 8, 3', 5'), 7.85 (d, 1H, H-6, J = 8.2 Hz), 8.30 (d, 1H, H-6, J = 8.2 Hz), 8.30H-9, J = 8.2 Hz). **25**: (DMSO- d_6): 6.79 (dd, 1H, H-4', J = 3.4, 1.7 Hz), 7.2–7.6 (m, 3H, H-6,7,3'), 7.98 (d, 1H, H-5', J = 1.7Hz), 8.07 (d, 1H, H-9, J = 2.2 Hz). 26: (CDCl₃): 7.18 (d, 1H, H-4', J = 2.1 Hz), 7.27 (dd, 1H, H-5', J = 2.1, 1.5 Hz), 7.46 (d, 1H, H-2', J = 1.5 Hz), 7.69 (dd, 1H, H-7, J = 8.7, 2.2 Hz), 8.08 (d, 1H, H-6, J = 8.7 Hz), 8.49 (d, 1H, H-9, J = 2.2 Hz). 27: (CDCl₃): 7.21 (dd, 1H, H-4', J = 4.8, 3.8 Hz), 7.4–7.5 (m, 2H, H-7,3'), 7.72 (d, 1H, H-6, J = 8.7 Hz), 8.02 (d, 1H, H-5', J = 4.8 Hz), 8.49 (d, 1H, H-9, J = 2.2 Hz).

2-Aryl-8- R_1 -1,2,4-triazolo[1,5-a]quinoxalin-4-amines 1–5, 7–10 A mixture of 4-chloro-derivative **19–27** (1 mmol) in absolute ethanol (30 ml) saturated with ammonia was heated at 130°C for 10 h in a sealed tube. Evaporation of the solvent at reduced pressure yielded a residue which was purified by column chromatography (eluting system conc NH₄OH/absolute ethanol/ CHCl₃/40–60° petroleum ether 4:32.5:170:30), and then recrystallized.

The title compounds displayed the following spectral data: 1: ¹H-NMR (CDCl₃): 5.7 (br s, 2H, NH₂), 7.4–7.6 (m, 5H, H-3'-5',7,8), 7.77 (d, 1H, H-6, J = 8.2 Hz), 8.3–8.4 (m, 3H, H-2',6',9). IR: 3320, 3460. 2: ¹H-NMR (CDCl₃): 5.7 (br s, 2H,

 NH_2), 7.5–7.6 (m, 4H, H-3'-5',7), 7.68 (d, 1H, H-6, J = 8.7 Hz), 8.3-8.4 (m, 3H, H-2',6',9). IR: 3310, 3480. 3: 1H-NMR $(DMSO-d_6)$: 2.51 (s, 3H, CH₃), 7.38 (dd, 1H, H-7, J = 8.6, 1.5 Hz), 7.5–7.6 (m, 6H, H-6,3',5'+ NH₂), 8.04 (d, 1H, H-9, J =1.5 Hz), 8.2–8.3 (m, 2H, H-2',6'). IR: 3310, 3470. 4: ¹H-NMR (CDCl₃): 5.8 (br s, 2H, NH₂), 7.2–7.4 (m, 2H, H-3',5'), 7.5–7.6 (m, 2H, H-7,4), 7.69 (d, 1H, H-6, J = 8.7 Hz), 8.2–8.3 (m, 1H, H-6'), 8.37 (d, 1H, H-9, J = 2.1 Hz). IR: 3140, 3310. 5: ¹H-NMR (CDCl₃): 3.90 (s, 3H, OCH₃), 5.7 (br s, 2H, NH₂), 7.04 (d, 2H, H-3',5', J = 8.9 Hz), 7.48 (dd, 1H, H-7, J = 8.7, 2.2 Hz), 7.67 (d, 1H, H-6, J = 8.7 Hz), 8.2–8.3 (m, 3H, H-2',6',9). IR 3310, 3470. 7: ¹H-NMR (CDCl₃): 5.7 (br s, 2H, NH_2), 6.62 (dd, 1H, H-4', J = 3.4, 1.8 Hz), 7.24 (d, 1H, H-3', J = 3.4 Hz), 7.4–7.6 (m, 2H, H-7.8), 7.66 (d, 1H, H-5', J =1.8 Hz), 7.76 (d, 1H, H-6, J = 8.2 Hz), 8.35 (d, 1H, H-9, J =8.2 Hz). IR: 3320, 3480. 8: ¹H-NMR (CDCl₃): 5.9 (br s, 2H, NH₂), 6.41 (dd, 1H, H-4', J = 3.4, 1.8 Hz), 7.23 (d, 1H, H-3', J = 3.4 Hz), 7.50 (dd, 1H, H-7, J = 8.7, 2.1 Hz), 7.6–7.7 (m, 2H, H-6,5'); 8.34 (d, 1H, H-9, J = 2.1 Hz). IR: 3150, 3320, **3480. 9**: ¹H NMR (CDCl₃): 5.8 (br s, 2H, NH₂), 7.10 (d, 1H, H-4', J = 2.1 Hz), 7.27 (dd, 1H, H-5', J = 2.1, 1.5 Hz), 7.4–7.5 (m, 2H, H-7,2'), 7.68 (d, 1H, H-6, J = 8.7 Hz), 8.30 (d, 1H, H-9, J = 2.1 Hz). IR: 3320, 3480. 10: ¹H-NMR (CDCl₃): 5.7 (br s, 2H, NH₂), 7.20 (dd, 1H, H-4', J = 4.8, 3.8 Hz), 7.5–7.6 (m, 2H, H-7,3'), 7.68 (d, 1H, H-6, J = 8.7 Hz), 7.93 (d, 1H, H-5', J =4.8 Hz), 8.33 (d, 1H, H-9, J = 2.1 Hz). IR: 3160, 3310.

2-(4-Hydroxyphenyl)-1,2,4-triazolo[1,5-a]quinoxalin-4-amine 6 A solution of 5 (1 mmol) in glacial acetic acid (8 mL) and HBr (48%, 9 mL) was heated at reflux for 3 h. The cooled solution was neutralized with a saturated solution of Na₂CO₃. The solvents were evaporated at reduced pressure and the residue was extracted with acetone in a Soxhelet apparatus. Evaporation of the acetone at reduced pressure afforded the crude title compound which after recrystallization displayed the following spectral data: ¹H-NMR (DMSO-d₆): 6.96 (d, 2H, H-3',5', J = 8.7 Hz), 7.5–7.8 (m, 4H, H-6,7 + NH₂), 8.0–8.2 (m, 3H, H-2',6',9), 10.01 (s, 1H, OH). IR: 2600–3300, 3380, 3480.

2-(4'-Chlorophenyl)-4-chloroimidazo[1,2-a]quinoxaline 29

The title compound was obtained following the procedure described in reference [22] to prepare its 2-phenyl- (**28**) and 2-(2-thienyl)- (**30**) analogues. Compound **29** displayed the following ¹H-NMR (DMSO- d_6): 7.6–7.9 (m, 4H, H-7,8,3',5'), 8.0–8.1 (m, 3H, H-6,2',6'), 8.38 (d, 1H, H-9, J = 7.9 Hz), 9.54 (s, 1H, H-1).

2-Arylimidazo[1,2-a]quinoxalin-4-amines 12-14

A mixture of 4-chloro derivative 28-30 (0.7 mmol) in absolute ethanol (10 mL) saturated with ammonia was heated at 110°C for 20 h in a sealed tube. Upon cooling a solid precipitated; this was collected, washed with water and recrystallized.

The title compounds displayed the following spectral data: **12**: ¹H-NMR (DMSO- d_6): 7.2 (br s, 2H, NH₂), 7.3–7.6 (m, 6H, H-6-8,3'-5'), 8.0–8.2 (m, 3H, H-9,2',6'), 9.15 (s, 1H, H-1). IR: 3320, 3480. **13**: ¹H-NMR (DMSO- d_6): 7.2–7.6 (m, 7H, H-6-8,3',5' + NH₂), 8.0–8.1 (m, 3H, H-9,2',6'), 9.19 (s, 1H, H-1). IR: 3310, 3490. **14**: ¹H-NMR (DMSO- d_6): 7.2–7.6 (m, 8H, H-6-8,3'-5' + NH₂), 8.11 (d, 1H, H-9, J = 7.9 Hz), 9.02 (s, 1H, H-1). IR: 3160, 3320.

2-Phenyl-4-ethoxyimidazo[1,2-a]quinoxaline 15

A mixture of 28 (0.43 mmol) in an ethanolic solution of NaOH (4.3 mmol in 10 mL absolute ethanol) was heated at reflux for 30 min. Upon cooling a solid was obtained which was collected, washed with water and recrystallized. ¹H-NMR

 $(DMSO-d_6)$: 1.51 (t, 3H, CH₃, J = 7.1 Hz), 4.67 (q, 2H, CH₂, J = 7.1 Hz), 7.3–7.7 (m, 5H, H-7,8,3',5'), 7.81 (d, 1H, H-6, J = 7.8 Hz), 8.0–8.1 (m, 2H, H-2',6'), 8.27 (d, 1H, H-9, J = 7.9 Hz), 9.29 (s, 1H, H-1).

1-(2'-Nitrophenyl)-2-phenylimidazole 31

Anhydrous K_2CO_3 (4.9 g) was added to a solution of 2-phenylimidazole (13.9 mmol) and 1-fluoro-2-nitrobenzene (34.6 mmol) in acetonitrile (20 mL). The mixture was heated at reflux for 80 h. Evaporation of the solvent at reduced pressure afforded a residue which was dissolved in CHCl₃ (250 mL). The organic solution was washed twice with water (200 mL each time), dried (anhydrous Na₂SO₄) and filtered through Norite. Evaporation of the CHCl₃ at reduced pressure afforded a resi-

Table II. Physical data of the newly synthesized compounds.

Compound	$Mp(^{\circ}C)(solvent)^{a}$	Yield (%) 20	
1	> 300 (A)		
2	> 300 (A)	47	
3	255-257 (B)	73	
4	> 300 (A)	65	
5	> 300 (A)	80	
6	> 300 (B)	60	
7	> 300 (A)	36	
8	> 300 (A)	30	
9	> 300 (A)	40	
10	> 300 (A)	36	
12	216-218 (A)	67	
13	> 300 (A)	85	
4	250–251 (A)	54	
.5	139–141 (A)	75	
6	292–293 (A)	50	
7	204–205 (C)	40	
.9	275-276 (A)	50	
0	> 300 (A)	30	
21	247–248 (A)	55	
22	> 300 (A)	54	
23	> 300 (A)	64	
24	> 300 (A)	65	
25	> 300 (A)	54	
26	> 300 (A)	50	
:7	> 300 (A)	40	
9	257-258 (D)	40	
31	168-169 (E)	75	
32	127-128 (F)	75	
33	289–290 (A)	80	
34	171–173 (G)	40	

^aRecrystallization solvents: A = absolute ethanol; B = dimethylformamide; C = cyclohexane/ethyl acetate; D = column chromatography, eluting system: chloroform/ethyl acetate, 9:1; E = ethyl acetate; F = ethanol/water; G = ethanol/diethyl ether.

due which was triturated with diethyl ether containing a few drops of diethyl acetate, collected and then recrystallized. ¹H-NMR (DMSO- d_6): 7.2–7.3 (m, 6H, 5 ar + H-4), 7.53 (s, 1H, H-5), 7.7–7.9 (m, 3H, ar), 8.08 (d, 1H, H-3', J = 9.4 Hz).

1-(2'-Aminophenyl)-2-phenylimidazole 32

Pd/C (0.34 mg) was added to a solution of **31** (4.3 mmol) in ethanol (200 mL). The mixture was hydrogenated in a Parr apparatus at 25 psi for 16 h. Elimination of the catalyst and evaporation of the solvent at reduced pressure afforded a waxy oil. When treated with diethyl ether, this gave rise to a white solid which was recrystallized. ¹H-NMR (DMSO- d_6): 4.9 (br s, 2H, NH₂), 6.5–6.6 (m, 1H, ar), 6.8–6.9 (m, 2H, ar), 7.1–7.3 (m, 6H, 4 ar + H-4,5), 7.4–7.5 (m, 2H, ar).

4.5-Dihydro-1-phenylimidazo[1,5-a]quinoxalin-4-one 33

1,1'-Carbonyldiimidazole (Im_2CO) (4.5 mmol) was added to a solution of **32** (3 mmol) in 1,2-dichlorobenzene (20 mL). The mixture was heated at reflux under nitrogen flow for 3 h. Upon cooling a solid was obtained which was collected, washed with diethyl ether and recrystallized. ¹H-NMR (DMSO- d_0): 6.9–7.0 (m, 1H, ar), 7.0–7.1 (m, 1H, ar), 7.2–7.4 (m. 2H, ar), 7.5–7.7 (m, 5H, ar), 7.98 (s, 1H, H-3), 11.5 (br s, 1H, NH). IR: 1690.

2-Phenyl-4-chloroimidazo[1,5-a]quinoxaline 34

N,*N*-Diethylaniline (0.3 mL) was added to a suspension of **33** (1.26 mmol) in POCl₃ (4 mL). The mixture was heated at reflux under nitrogen flow for 6 h. The excess of POCl₃ was evaporated at reduced pressure. The residue was treated twice with cyclohexane (20 mL) which was in its turn distilled off. The resulting oily residue became a crystalline white solid upon treatment with diethyl ether. ¹H-NMR (DMSO-d₆): 7.3–7.8 (m. 8H, ar), 7.91 (d, 1H, ar, J = 8.1 Hz), 8.18 (s, 1H, H-3).

2-Phenylimidazol[1,5-a]quinoxalin-4-amine 16

A suspension of 34 (0.47 mmol) in isopropanol saturated with ammonia (10 mL) was heated at 110°C for 40 h in a sealed tube. By quenching the mixture with water (20 mL) a solid was obtained which was collected, washed with water and recrystallized. ¹H-NMR (DMSO-*d*₆): 6.8–6.9 (m, 1H, ar), 7.1–7.3 (m, 4H, 2 ar + NH₂), 7.4–7.5 (m, 1H, ar), 7.5–7.7 (m, 5H, ar), 8.03 (s, 1H, H-3). IR: 3160, 3340.

2-Phenylimidazo[15-a]quinoxalin-4-cyclopentylamine 17

A mixture of **34** (0.6 mmol) and cyclopentylamine (0.9 mL) was heated at 120°C for 14 h in a sealed tube. Evaporation of the excess of cyclopentylamine at reduced pressure afforded a residue which was dissolved in CHCl₃ (20 mL). The solution was washed twice with water (20 mL each time) and dried (Na₂SO₄). Evaporation of the organic solvent at reduced pressure yielded a residue which was washed with a little diethyl ether and recrystallized. ¹H-NMR (DMSO-*d*₆): 1.5–1.8 (m, 6H, cyclopentyl protons), 2.0–2.2 (m, 2H, cyclopentyl protons), 4.5–4.6 (m, 1H, cyclopentyl proton), 6.8–6.9 (m, 1H, ar), 7.1–7.3 (m, 2H, ar), 7.5–7.7 (m, 7H, 6 ar + NH), 8.10 (s, 1H, H-3). IR: 3280.

Biochemistry

A₁ Receptor binding

Rat cerebral cortex was homogenized in ice-cold 0.32 M sucrose containing protease inhibitors ($20 \mu g/mL$ soybean trypsin inhibitor, $200 \mu g/mL$ bacitracine, and $160 \mu g/mL$ benz-amidine) in an ultra-turrax homogenizer. The homogenate was

centrifuged at 1000 g for 10 min at 4°C and the supernatant again centrifuged at 48 000 g for 15 min at 4°C. The resulting pellet was suspended in 10 volumes of ice-cold 40 mM Tris-HCl buffer at pH 7.7 containing 2 mM MgCl₂ and protease inhibitors (buffer T_i). It was then homogenized and centrifuged at 48 000 g for 15 min at 4°C.

The pellet was dispersed in 40 volumes of fresh T_1 buffer and incubated with adenosine deaminase (1 UI/mL) at 37°C for 60 min, then recentrifuged at 48 000 g for 15 min at 4°C.

The resulting pellet was frozen at -80° C until the time of assay.

The pellet was suspended in ice-cold T_1 buffer and A_1 binding assay was performed in triplicate by incubating at 25°C for 45 min in 0.5 mL T_1 buffer containing 1.3 nM [³H]CHA in the absence or presence of unlabelled 10 μ M *R*-phenylisopropyladenosine. The binding reaction was terminated by filtering through Whatman GF/B glass fiber filters under suction and washing twice with 5 mL ice-cold Tris-buffer. The filters were placed in scintillation vials and 4 mL Beckman Ready-Protein solvent scintillation fluid was added. The radioactivity was counted with an LS 1800 scintillation counter. Specific binding was obtained by subtracting non-specific binding from total binding and was approximated to 85–90% of the total binding.

A_{2a} receptor binding

Corpora striata were dissected from rat brain and the tissue was homogenized in 20 volumes of ice-cold 50 mM Tris-HCl buffer at pH 7.5 containing protease inhibitors as reported above and 10 mM MgCl₂ (buffer T_2). The homogenate was centrifuged at 48 000 g for 10 min at 4°C. The pellet was then suspended in 20 volumes of Tris-buffer (T_2) containing adenosine deaminase (1 Ul/mL) and incubated for 30 min at 37°C. The resulting pellet was diluted in 20 volumes of 50 mM Tris-HCl buffer at pH 7.5 containing 10 mM MgCl₂ and used in the binding assay.

 A_{2a} binding assay was performed in triplicate, by incubating aliquots of the membrane fraction (0.2–0.3 mg protein) in Tris-HCl buffer at pH 7.5, with approximately 4 nM [³H]CGS 21680 in a final volume of 0.5 mL. Incubation was carried out at 25°C for 90 min. Non-specific binding was defined in the presence of 10 µM CGS 21680. The binding reaction was concluded by filtration through Whatman GF/C glass fiber filters under reduced pressure. Filters were washed four times with 5 mL aliquots of ice-cold buffer and placed in scintillation vials. Specific binding was obtained by subtracting non-specific binding from total binding and approximated to 85–90% of the total binding. The receptor-bound radioactivity was measured as described above.

Compounds were dissolved in ethanol or DMSO (buffer/ concentration of 2%) and added to the assay mixture. Blank experiments were carried out to determine the effect of the solvent on binding.

Protein estimation was based on a reported method [24], after solubilization with 0.75 N sodium hydroxide, using bovine serum albumin as standard.

The concentration of tested compound that produce 50% inhibition of specific [³H]CHA or [³H]CGS 21680 binding (IC₅₀) was determined by log-probit analysis with seven concentrations of the displacer, each performed in triplicate. Inhibition constants (K_i) were calculated according the equation [25]: $K_i = IC_{50}/(1 + [L]/K_d)$ where [L] is the ligand concentration and K_d its dissociation constant. K_d of [³H]CHA binding to cortex membranes was 1.6 nM and K_d of [³H]CGS 21680 binding to striatal membranes was 15 nM [18].

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