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Structure-based hybridization of the bioactive natural products rhizonin A and ternatin leading to a selective fat-accumulation inhibitor against 3T3-L1 adipocytes

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Rhizonin A (1) and ternatin (2) are novel bioactive cyclic heptapeptides that were isolated from the fungus Rhizopus microsporus van Tieghem¹ and the mushroom Coliorus versicolor,^{2,3} respectively. (Fig. 1). They share common structural features in that they contain p-. N-methylated amino acids as well as unusual amino acids, for example, NMe-FurAla [NMe-3-(3-furvl)alanine] or B-OH-D-Leu [(2R,3R)-3-hydroxy-Leu]. Interestingly, they show extremely high structural homology: (i) identical sequence of DDDLLLL amino-acid configurations, (ii) positions of N-methylation, (iii) contiguous L-Leu⁴-NMe-L-Ala⁵ moieties. An analysis of the X-ray crystal structures of 1 and 2 demonstrated that the solid-state conformation of **1** resembles that of **2**, in particular with regard to a β turn structure in the region of positions $4 \rightarrow 7$, which might be mainly due to (i).⁴ Therefore, these two compounds could be classified as closely related cyclic peptides. However, their reported biological activities are quite different. Recently, we discovered that **2** significantly inhibited fat-accumulation both in vitro^{2,3} and in vivo.⁵ Meanwhile, **1** has only been investigated in vivo and is known to exhibit a potent hepatotoxic effect in rat which leads to death in rat and ducklings.^{6,7} To further examine the biological activity of 1, we recently achieved the chemical synthesis of 1 and evaluated its fat-accumulation inhibitory activity against 3T3-L1 adipocytes.⁸ While **1** exhibited a weak inhibitory effect at high concentration (IC₅₀ 55 μ M), it was also quite cytotoxic at

ABSTRACT

Based on the structural similarity between the naturally occurring cyclic heptapeptides rhizonin A and ternatin, two novel analogues were designed. The synthetic analogues were assessed with regard to their fat-accumulation inhibitory effect against 3T3-L1 adipocytes, and this led to the discovery of a potent and selective fat-accumulation inhibitor compared to the parent compound rhizonin A.

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the same concentration. To obtain a better understanding of structure–activity relationships (SAR) of $\mathbf{1}$ as well as of the relation between $\mathbf{1}$ and $\mathbf{2}$, we developed a synthesis of novel rhizonin A analogues, which led to the discovery of a selective fat-accumulation inhibitor against 3T3-L1 adipocytes.

First, we focused on the two NMe-FurAla moieties in **1**, which do not exist in **2**. Uniquely, rhizonin A (**1**) (and its sister compound rhizonin B¹) is the only natural product that contains an unusual FurAla moiety (including N-methylated variant) as a structural component. Thus, we designed the *di*-phenylalanine-substituted



Figure 1. Structures of rhizonin A (1) and ternatin (2).

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Figure 2. Design of rhizonin A analogues 3 and 4.

analogue [NMe-L-Phe³, NMe-D-Phe⁶]rhizonin A (**3**) to examine the influence of unusual NMe-FurAla moieties on bioactivity (Fig. 2). Moreover, with the concept of hybridization in mind, we sought to synthesize other analogue [NMe-L-Leu³, NMe-D-Phe⁶]rhizonin A (**4**), which has three amino-acid sequences as in **2** on the right side of the chemical structure.

The syntheses of analogues **3** and **4** were slightly modified from the synthetic route toward **1**. First, we prepared the right fragments **8a** and **8b**, which are shown in Scheme 1. The starting material *N*Me-L-Ala-OMe hydrochloride (**5**) was coupled with Boc-L-Leu-OH in the presence of PyBroP to give dipeptide **6**.

Removal of the Boc group in 50% TFA/CH₂Cl₂ followed by amide-bond formation with the corresponding Boc-amino acids provided tripeptides **7a** and **7b**, respectively. Finally, sequential Boc deprotection, coupling, and methyl ester hydrolysis afforded the desired **8a** and **8b**.

Next, the left fragment **11** was prepared from the known dipeptide 9^9 in the same manner (Scheme 2). Removal of the Boc group in **9** followed by coupling with Boc-*N*Me-D-Phe-OH afforded tripeptide **10**, which was then subjected to Boc deprotection to give



Scheme 1. Synthesis of the right fragments 8a and 8b. Reagents and conditions: (a) Boc-L-Leu-OH, PyBroP, DIPEA, CH₂Cl₂; (b) 50% TFA/CH₂Cl₂; (c) Boc-NMe-L-Phe-OH for 7a, Boc-NMe-L-Leu-OH for 7b, HATU, DIPEA, CH₂Cl₂; (d) Boc-L-Val-OH, HATU, DIPEA, CH₂Cl₂; (e) LiOH, *t*-BuOH, THF, H₂O.



3: R = Phe (13% from 12a) 4: R = *i*-Pr (8% from 12b)

Scheme 2. Synthesis of rhizonin A analogues 3 and 4. Reagents and conditions: (a) 50% TFA/CH₂Cl₂; (b) Boc-NMe-D-Phe-OH, HATU, DIPEA, CH₂Cl₂; (c) 8a (1.0 equiv) for 3, 8b (1.0 equiv) for 4, HATU, DIPEA, CH₂Cl₂ (d) LiOH, *t*-BuOH, THF, H₂O; (e) HATU, HOAt, DIPEA, DMF, CH₂Cl₂ (1.5 mM).

11. Fragment couplings of **11** with **8a** and **8b** were conducted between the carboxyl group in the L-Leu⁴ moiety and the amino group in the *N*Me-L-Ala⁵ moiety to give heptapeptides **12a** and **12b** in moderate yield. Sequential methyl ester hydrolysis and Boc deprotection of **12a** and **12b** provided the corresponding cyclic precursors. Finally, the key macrolactamizations between the carboxyl group in the D-Val¹ moiety and the amino group in the L-Val² moiety were performed in the presence of HATU (2.0 equiv) and HOAt (2.0 equiv) at low concentration (1.5 mM). After HPLC purification of the crude materials, rhizonin A analogues **3**¹⁰ and **4**¹¹ were obtained in 13% yield from **12a** and 8% yield from **12b** (in three steps), respectively.

The results of the in vitro fat-accumulation-inhibition assay for synthetic rhizonin A analogues **3** and **4**, as well as rhizonin A (**1**) and ternatin (**2**), are shown in Table 1. Cell viability was calculated independently to exclude undesired fat-accumulation inhibition arising from the toxicity of the tested compounds.

Based on the results, both **3** and **4** more strongly inhibited fataccumulation than the parent compound **1**, though **2** was the most bioactive among the compounds tested. Hybrid compound **4** was

Table 1

Inhibitory effects of synthetic analogues on fat-accumulation against 3T3-L1 adipocytes and their cell viability $^{\rm a}$

Compound	Fat-accumulation inhibitory effect: IC_{50} (μM)	Cell viability: IC ₅₀ (µM)
Rhizonin A (1)	55 ± 3.7	70% at 62 ^b (μM)
3	42 ± 1.1	>240
4	5.6 ± 2.1	>240
Ternatin (2)	0.027 ± 0.003	0.28 ± 0.03

^a Values are means of quadruplicate determinations.

^b Not tested at higher concentrations.

10-fold more potent than **1**. The overall potency of inhibitory effect is in the order (2>) 4 > 3 > 1. Thus, modifications on the right half of **1** that imitates the sequence of ternatin (2) should greatly strengthen the bioactivity.

On the other hand, the toxic profiles of the synthetic analogues showed an opposite tendency. Interestingly, **3** and **4** were found to be less toxic ($IC_{50} > 240 \mu$ M), while **1** showed considerable cytotoxicity (the expected IC_{50} value for cell viability is about 62–176 μ M) at IC_{50} value for fat-accumulation inhibition. The selectivity indices [SI; IC_{50} value for cytotoxicity/that for fat-accumulation] of **3** and **4** were >12.3 and >43, and therefore higher than that of **1** (the expected SI is about 1–3). This result strongly suggests that the two FurAla moieties are responsible for the potent cytotoxicity of **1** against 3T3-L1 adipocytes. Moreover, hybrid analogue **4** had a greater SI than **2** (SI = 10). Therefore, **4** may be a plausible candidate that possesses a selective and/or effective fat-accumulation inhibitory effect against 3T3-L1 murine adipocytes.

In summary, two novel analogues of rhizonin A (1) were designed on the basis of structural hybridization with the potent fat-accumulation inhibitor ternatin (2). The modification of the two NMe-FurAla moieties in the structure of 1 led to analogues with more potent and selective fat-accumulation inhibitory activities. Further chemical and biological studies on 1, including its SAR, are now underway.

Acknowledgments

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- 10. Spectroscopic data for 3: $[\alpha]^{23}_{D} 15.4^{\circ}$ (c = 0.15, CHCl₃); IR (CHCl₃) 3326, 2933, 2337, 1650, 1506 cm⁻¹; ¹H NMR (400 MHz, C₆D₆) δ 7.85 (d, *J* = 9.6 Hz, 1H), 7.77 (d, *J* = 9.2 Hz, 1H), 7.41 (d, *J* = 7.2 Hz, 1H), 7.35 (s, 1H), 7.22–6.93 (m, 10H), 5.93 (dd, *J* = 12.4, 5.2 Hz, 1H), 5.89 (d, *J* = 10.0 Hz, 1H), 5.70–5.62 (m, 1H), 4.90 (t, *J* = 9.2 Hz, 1H), 4.71 (t, *J* = 9.2 Hz, 1H), 4.38 (q, *J* = 6.8 Hz, 1H), 4.00–3.96 (m, 1H), 3.89 (dd, *J* = 15.2, 5.2 Hz, 1H), 3.66–3.58 (m, 1H), 3.40 (dd, *J* = 8.4, 4.8 Hz, 1H), 2.80 (dd, *J* = 15.2, 12.4 Hz, 1H), 2.71 (s, 3H), 2.69 (s, 3H), 2.63 (s, 3H), 2.33–2.25 (m, 1H), 2.24–2.12 (m, 1H), 1.90–1.75 (m, 2H), 1.55–1.47 (m, 1H), 1.37–1.27 (m, 2H), 1.25 (d, *J* = 6.8 Hz, 6H), 1.18–1.13 (m, 1H), 1.05 (d, *J* = 6.8 Hz, 3H), 1.02 (d, *J* = 6.8 Hz, 3H), 0.96 (t, *J* = 7.2 Hz, 3H), 0.94 (d, *J* = 6.8 Hz, 3H), 0.91 (d, *J* = 6.4 Hz, 3H), 0.79 (d, *J* = 6.8 Hz, 3H), 0.47 (d, *J* = 7.2 Hz, 3H); RRMS (FAB) calcd for C₄₆H₆₉Ny₇₀Na (M+Na)* 854.5196, found 854.5175.
- To C46H69N7070K4 (MFTNA) 634-5150, round 634-5175. 11. Spectroscopic data for 4: $[\alpha]^{23}_{D} - 2.5^{\circ}$ (c = 0.15, CHCl₃); IR (CHCl₃) 3300, 2966, 2881, 2363, 1644, 1499 cm⁻¹; ¹H NMR (400 MHz, C₆D₆) δ 7.87 (d, *J* = 9.2 Hz, 1H), 7.62 (d, *J* = 9.2 Hz, 1H), 7.36 (s, 1H), 7.18–6.90 (m, 5H), 6.85 (d, *J* = 6.4 Hz, 1H), 6.09–5.90 (m, 2H), 5.71–5.62 (m, 1H), 5.43–5.26 (m, 1H), 4.98 (t, *J* = 9.2 Hz, 1H), 4.79 (t, *J* = 9.2 Hz, 1H), 4.39 (q, *J* = 7.2 Hz, 1H), 3.48–3.35 (m, 2H), 3.06 (s, 3H), 2.65 (s, 3H), 2.58 (s, 3H), 1.23 (d, *J* = 6.8 Hz, 3H), 1.06–0.80 (m, 4H), 1.05 (d, *J* = 6.8 Hz, 3H), 1.00 (d, *J* = 6.8 Hz, 3H), 0.99 (d, *J* = 6.8 Hz, 3H), 0.96 (t, *J* = 4.8 Hz, 3H), 0.92 (d, *J* = 6.4 Hz, 3H), 0.75 (d, *J* = 6.4 Hz, 3H), 0.84 (d, *J* = 6.4 Hz, 3H), 0.82 (d, *J* = 6.4 Hz, 3H), 0.75 (d, *J* = 6.4 Hz, 3H); HRMS (FAB) calcd for C4₃H₇₁N₇₀7Na (M+Na)^{*} 820.5313, found 820.5291.