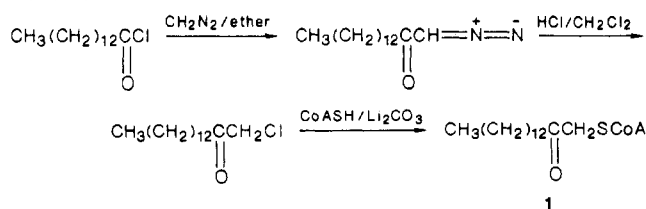


Scheme I



Department of Chemistry, The Ohio State University.

Registry No. 1 (free acid), 121124-66-1; 1-4Li, 121124-67-2; CoA, 85-61-0; NMT, 110071-61-9; $\text{CH}_3(\text{CH}_2)_{12}\text{COCl}$, 112-64-1; $\text{N}_2=\text{CHCO}(\text{CH}_2)_{12}\text{CH}_3$, 90670-23-8; $\text{ClCH}_2\text{CO}(\text{CH}_2)_{12}\text{CH}_3$, 121097-11-8.

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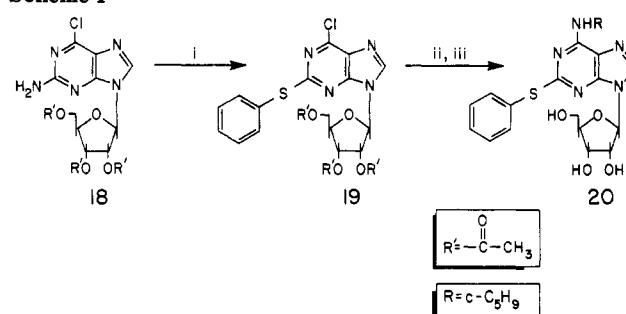
C2,N⁶-Disubstituted Adenosines: Synthesis and Structure-Activity Relationships

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Extracellular adenosine receptors have been divided into two major subtypes, called A₁ and A₂. Substitution of the adenosine molecule with appropriate groups at C2 or N⁶ is known to impart selectivity for the A₂ receptor over the A₁ receptor. In the present study, we investigated whether substitution at both C2 and N⁶ would have additive effects on the A₂/A₁ affinity ratio, thereby providing compounds with greater A₂ selectivity than presently available agents. Disappointingly, additivity appeared to hold only when an A₁-selective group was present at N⁶. For instance, 2-(phenylamino) substitution of the A₁-selective agonist N⁶-cyclopentyladenosine resulted in a 70-fold shift in selectivity in favor of the A₂ receptor, but the same substitution applied to the A₂-selective agonist N⁶-[2-(3,5-dimethoxyphenyl)-2-(2-methylphenyl)ethyl]adenosine resulted in a 100-fold loss of affinity with no change in A₂ selectivity.

Adenosine causes a variety of physiological responses, which are mediated by two subtypes of extracellular receptors, called A₁ and A₂. These two receptor subtypes can be distinguished on the basis of structure-activity relationships,¹⁻³ and specific receptor binding assays exist for both subtypes.^{4,5} Considerable effort has been devoted to the search for adenosine agonists with improved selectivity for A₁ or A₂ receptors. Although agonists with 1000-fold or greater selectivity for the A₁ receptor are known,⁶ until recently the most A₂-selective agonist was 2-(phenylamino)adenosine (CV-1808, compound 7 in Table II),⁷ which shows only 5-fold A₂ selectivity.⁵ Very recently, N⁶-[2-(3,5-dimethoxyphenyl)-2-(2-methylphenyl)ethyl]adenosine (compound 50 in Table IV) was shown to possess about a 30-fold selectivity for the A₂ receptor.⁸ Because 7 is substituted at C2, whereas 50 is substituted at N⁶ we became interested in the possibility that the functional groups responsible for conferring selectivity on these two compounds might interact with independent sites on the adenosine receptor, thereby allowing additive enhancement of selectivity by combining structural modifications at both positions. Because many other C2 and N⁶ groups with widely differing effects on A₁ and A₂ affinity have been reported (see Tables II and IV),^{7,9-11} we also tested representative combinations of these groups for

Scheme I^a

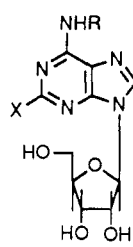
^a (i) PhSSPh, isoamyl nitrite, CH₃CN, Δ; (ii) RNH₂, DME, Et₃N, room temperature; (iii) MeOH-NH₃, room temperature.

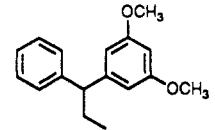
their effects on adenosine-receptor selectivity. Our results indicate that the effects of C2 and N⁶ substitution are only

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Table I. Physical Properties of Novel Adenosine Agonists


| example | R | X | mp, °C | formula analysis |
|-----------------|--|----------------------|---------|---|
| 13 | cyclopentyl | NH ₂ | 210–211 | C ₁₅ H ₂₂ N ₆ O ₄ |
| 14 | cyclohexyl | NH ₂ | 214–215 | C ₁₆ H ₂₄ N ₆ O ₄ |
| 15 | (R)-1-methyl-2-phenylethyl | NH ₂ | 108–110 | C ₁₉ H ₂₄ N ₆ O ₄ |
| 16 | 2,2-diphenylethyl | NH ₂ | 134–137 | C ₂₄ H ₂₆ N ₆ O ₄ ·0.5H ₂ O |
| 17 | 9-fluorenylmethyl | NH ₂ | 154–158 | C ₂₄ H ₂₄ N ₆ O ₄ ·0.5CH ₃ OH |
| 20 | cyclopentyl | SPh | 105–110 | C ₂₁ H ₂₅ N ₆ O ₄ S |
| 21 | cyclohexyl | SPh | 103–108 | C ₂₂ H ₂₇ N ₆ O ₄ S |
| 23 | 2,2-diphenylethyl | SPh | 116–119 | C ₃₀ H ₂₉ N ₆ O ₄ S |
| 24 | 1-naphthylmethyl | SPh | 115–120 | C ₂₇ H ₂₅ N ₆ O ₄ S |
| 27 | cyclopentyl | SO ₂ Ph | 72–75 | C ₂₁ H ₂₅ N ₆ O ₆ S |
| 28 | cyclohexyl | SO ₂ Ph | 87–93 | C ₂₂ H ₂₇ N ₆ O ₆ S·0.5C ₂ H ₅ OH |
| 29 | 1-naphthylmethyl | SO ₂ Ph | 117–122 | C ₂₇ H ₂₅ N ₆ O ₆ S |
| 36 | 1-naphthylmethyl | F | 131–135 | C ₂₁ H ₂₀ N ₆ O ₄ F |
| 37 | 1-naphthylmethyl | cyclohexylamine | 137–140 | C ₂₄ H ₃₂ N ₆ O ₄ ·0.25H ₂ O |
| 38 | 1-naphthylmethyl | NHCH ₂ Ph | 205–208 | C ₂₈ H ₂₈ N ₆ O ₄ |
| 44 | 1-naphthylmethyl | NHPh | 205–210 | C ₂₇ H ₂₆ N ₆ O ₄ |
| 45 | 2,2-diphenylethyl | NHPh | 116–121 | C ₃₀ H ₃₀ N ₆ O ₄ ·0.4H ₂ O |
| 47 | 9-fluorenylmethyl | NHPh | 220–222 | C ₃₀ H ₂₈ N ₆ O ₄ |
| 49 | | NHPh | 118–121 | C ₃₂ H ₃₄ N ₆ O ₆ |
| 51 |  | NHPh | 126–130 | C ₃₃ H ₃₆ N ₆ O ₆ |
| 53 | CH ₂ Ph | NHPh | 222–224 | C ₂₃ H ₂₄ N ₆ O ₄ |
| 55 | cyclopropyl | NHPh | 200–202 | C ₁₉ H ₂₂ N ₆ O ₄ |
| 30 | cyclopentyl | NHPh | 172–175 | C ₂₁ H ₂₆ N ₆ O ₄ ·0.4H ₂ O |
| 56 | cyclohexyl | NHPh | 229–231 | C ₂₂ H ₂₈ N ₆ O ₄ |
| 58 | 2-endo-norbornyl | NHPh | 210–212 | C ₂₃ H ₂₈ N ₆ O ₄ |
| 60 | (S)-2-hydroxypropyl | NHPh | 120–123 | C ₁₉ H ₂₄ N ₆ O ₅ ·0.25H ₂ O |
| 62 ^a | 1-pyrrolidinyl | NHPh | 222–224 | C ₂₀ H ₂₄ N ₆ O ₄ |

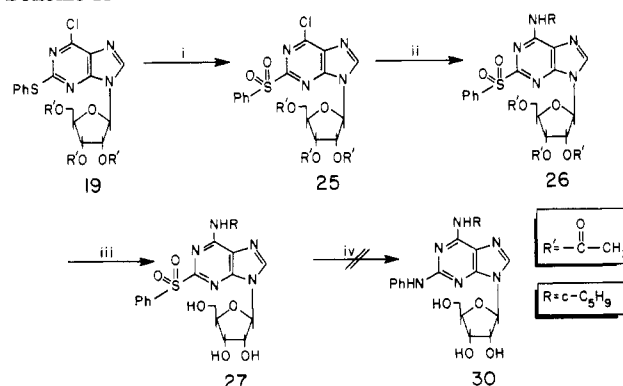
^a Compound 62 is 2-(phenylamino)-6-(1-pyrrolidinyl)-9-β-D-ribofuranosyl-9H-purine

partially additive, with the least additivity unfortunately being seen with the most A₂-selective parent groups.

Chemistry

The 2-amino analogues (examples 13–17; Table I) were synthesized in a standard fashion by reacting 2-amino-6-chloropurine ribonucleoside with an appropriate amine in the presence of a base in refluxing ethanol. Synthesis of 2-phenylthio derivatives (Tables II and III) was achieved

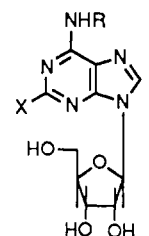
Scheme II^a



^a (i) KMnO₄, AcOH, 0–10 °C, 3 h; (ii) RNH₂, Et₃N, DME, room temperature, 2.5 h; (iii) MeOH–NH₃, 4 h; (iv) PhNH₂, DMF, Δ.

by utilizing a recently reported method¹² shown in Scheme I. The corresponding phenylsulfone derivatives (examples 27–29) were synthesized as follows (Scheme II): the in-

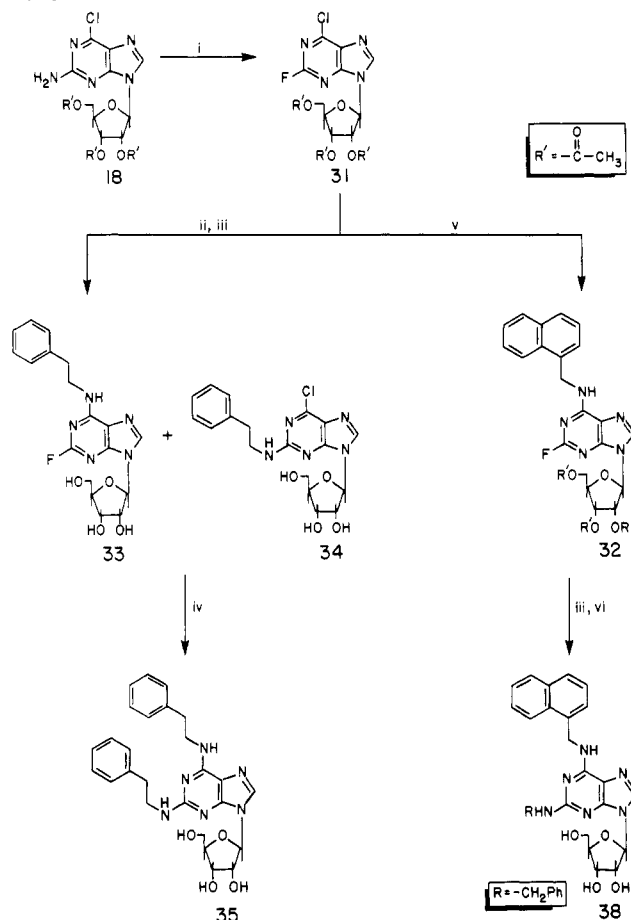
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Table II. Affinities of C2-Modified Adenosines and Reference Agents in A₁ and A₂ Receptor Binding Assays


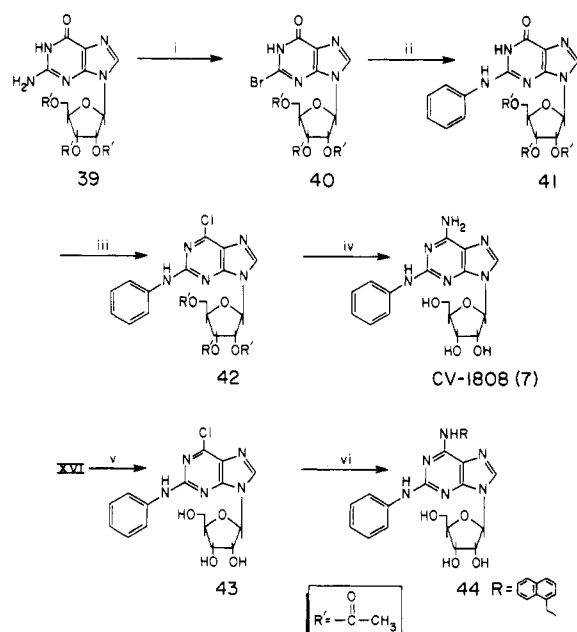
| example | X | R | K _i , nM ^a | | |
|---------|--------------------|----------------------------|----------------------------------|----------------|--------------------------------|
| | | | A ₁ | A ₂ | A ₂ /A ₁ |
| 1 | H | H | 12.8 ^b | 37.0 | 2.9 |
| 2 | Cl | H | 9.3 | 63 | 6.8 |
| 3 | OH | H | 94 | 330 | 3.5 |
| 4 | SPh | H | 2200 | 2000 | 0.90 |
| 5 | SO ₂ Ph | H | 2600 | 1230 | 0.46 |
| 6 | 4-OMePh | H | 1320 | 600 | 0.45 |
| 7 | NHPh | H | 600 | 116 | 0.190 |
| 8 | H | cyclopentyl | 0.59 | 460 | 780 |
| 9 | H | cyclohexyl | 1.42 | 610 | 430 |
| 10 | H | (R)-1-methyl-2-phenylethyl | 1.17 | 124 | 106 |
| 11 | H | 2,2-diphenylethyl | 6.8 | 25 | 3.6 |
| 12 | H | 1-naphthyl-methyl | 24 | 9.4 | 0.38 |

^a A₁ affinities were determined in [³H]-N⁶-cyclohexyladenosine binding to rat whole-brain membranes, and A₂ affinities were determined in [³H]NECA binding to rat striatal membranes.⁵ Values for compounds 2, 3, 6, and 7 are from ref 5. All values are means of three or more independent determinations. ^b The affinity of adenosine cannot be determined directly because of the necessity for adenosine deaminase in the binding assays.⁵ The values given are derived from Free-Wilson analysis of mono- and disubstituted adenosine analogues.⁵

intermediate 6-chloro-2-(phenylthio)purine ribonucleoside triacetate (19) was first oxidized with KMnO₄ in acetic acid to the sulfone derivative 25, followed by treatment with an appropriate amine to afford the adenosine derivative 26. The deprotection was carried out in methanolic ammonia at room temperature to yield the target phenyl sulfone adenosines 27–29. Attempts to convert these sulfone derivatives to the corresponding CV-1808 analogues by direct displacement with aniline under various conditions failed, mainly due to the poor nucleophilicity of aniline. Alternatively (Scheme III), the 2-amino-6-chloropurine ribonucleoside triacetate (18) upon treatment with HF-pyridine complex at low temperature¹³ afforded the 2-fluoro-6-chloro compound 31 in a good yield. Reaction of 31 with (1-naphthylmethyl)amine yielded the 2-fluoro-adenosine derivative 32. However, the specificity seen in the above reaction did not generalize to all nucleophiles. Attempts to displace chlorine selectively in a similar fashion with 2-phenethylamine failed, yielding a 1:1 mixture of regioisomeric products 33 and 34 following deprotection. This was confirmed by proton NMR spectra in which the sugar protons as well as the C8 proton showed separate chemical shifts. This mixture upon further treatment with 2-phenethylamine in refluxing ethanol gave *N*-(2-phenylethyl)-2-[(2-phenylethyl)amino]adenosine (35). Similarly, reaction of 31 with (2,2-diphenylethyl)amine gave a 2:1 ratio of corresponding regioisomeric products. Thus we surmise that, to some extent, the regioselective displacement seen with (1-naphthylmethyl)amine is due to a steric factor. Nevertheless, compound 32, following deprotection, can be reacted with a variety of primary

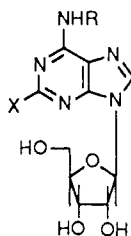
Scheme III^a

^a (i) HF-pyridine, *tert*-butyl nitrite, -55 to -30 °C; (ii) Ph(CH₂)₂NH₂, DME, Et₃N, room temperature; (iii) MeOH-NH₃, room temperature; (iv) Ph(CH₂)₂NH₂ (xs) EtOH, Δ; (v) (1-naphthylmethyl)amine Et₃N, DME, room temperature; (vi) RNH₂, EtOH, Et₃N, Δ.

Scheme IV^a

^a (i) CHBr₃, *n*-amyl nitrite, Δ; (ii) PhNH₂, MeOH, Δ; (iii) POCl₃, *N,N*-dimethylaniline, Et₄NCl, CH₃CN, Δ; (iv) NH₃-MeOH, Δ; (v) MeOH-NH₃, room temperature; (vi) RNH₂, Et₃N, EtOH, Δ.

amines to afford analogues 37–38. Once again, attempts to incorporate a phenylamino group at the C2 position

Table III. Affinities of C2,N⁶-Disubstituted Adenosines in A₁ and A₂ Receptor Binding Assays

| example | R | X | K _i , nM ^a | | |
|---------|------------------------------------|--------------------------------------|----------------------------------|----------------|--------------------------------|
| | | | A ₁ | A ₂ | A ₂ /A ₁ |
| 13 | cyclopentyl | NH ₂ | 8.3 | 6100 | 730 |
| 14 | cyclohexyl | NH ₂ | 19.3 | 3500 | 181 |
| 15 | (R)-1-methyl-2-phenylethyl | NH ₂ | 19.2 | 1530 | 80 |
| 16 | 2,2-diphenylethyl | NH ₂ | 61 | 135 | 2.2 |
| 17 | 9-fluorenylmethyl | NH ₂ | 18.5 | 22 | 1.20 |
| 20 | cyclopentyl | SPh | 37 | 4000 | 107 |
| 21 | cyclohexyl | SPh | 160 | 6700 | 42 |
| 22 | (R)-1-methyl-2-phenylethyl | SPh | 210 | 1000 | 4.7 |
| 23 | 2,2-diphenylethyl | SPh | 840 | 800 | 0.95 |
| 24 | 1-naphthylmethyl | SPh | 1470 | 610 | 0.42 |
| 27 | cyclopentyl | SO ₂ Ph | 96 | 2300 | 24 |
| 28 | cyclohexyl | SO ₂ Ph | 420 | 5100 | 12.3 |
| 29 | 1-naphthylmethyl | SO ₂ Ph | 2000 | 270 | 0.133 |
| 35 | (CH ₂) ₂ Ph | NH(CH ₂) ₂ Ph | 1620 | 2000 | 1.26 |
| 36 | 1-naphthylmethyl | F | 18.1 | 14.9 | 0.82 |
| 37 | 1-naphthylmethyl | cyclohexylamine | 45000 | 10300 | 0.23 |
| 38 | 1-naphthylmethyl | NHCH ₂ Ph | 530 | 350 | 0.66 |

^a A₁ affinities were determined in [³H]-N⁶-cyclohexyladenosine binding to rat whole-brain membranes, and A₂ affinities were determined in [³H]NECA binding to rat striatal membranes.⁵ K_i values for compounds 16 and 29 are means of three and two independent experiments, respectively. Other results are from single determinations.

under various conditions failed.

Finally, we resorted to an alternate route in which the aniline function is incorporated in the beginning of the synthesis. Such a synthetic methodology¹² is shown in Scheme IV. Using this efficient scheme, we prepared several 2-(phenylamino)adenosine analogues (Table IV) and evaluated these compounds in the A₁ and A₂ receptor binding assays.

Receptor Binding and Structure-Activity Relationships. Incorporation of NH₂ at the C2 position had a deleterious effect on both A₁ and A₂-receptor affinities (examples 13–17; Table III). Interestingly, although the phenylthio group is similar to a phenylamino group in size and shape, the phenylthio analogues (4, 20–24, Tables II and III) also showed weak binding at both receptors; in contrast, 2-(phenylamino)adenosine (7) had an A₂ affinity of 116 nM and a 5-fold A₂ selectivity. The 2-(phenylthio) derivatives retained similar A₂/A₁ affinity ratios as compared to the parent N⁶ derivatives (Tables II and III). The corresponding sulfone derivatives (27–29) began to show good A₂ selectivity as represented by 2-(phenylsulfonyl)-N⁶-(1-naphthylmethyl)adenosine (29). This doubly modified adenosine analogue had an A₂-binding affinity similar to that of CV-1808 (7), but was slightly more selective. The improvement in selectivity compared to that of the parent N⁶ derivative 25 is primarily due to greater loss of affinity at the A₁ receptor than at the A₂ receptor. This indicates that there is a more significant tolerance at the 2-position domain of the A₂ receptor than at the A₁ receptor. The loss of A₁ affinity may be due to either steric bulk or unfavorable charge interactions from the sulfone moiety.

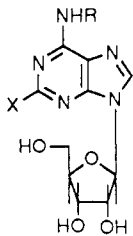
The 2-fluoro derivative (36) served as a precursor to various C2-substituted adenosines (37 and 38). Interestingly, the 2-(cyclohexylamino) derivative 37, although A₂ selective, had a very weak affinity at both receptors, especially compared to the corresponding 2-(phenylamino) derivative 44. The major difference between the two

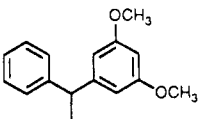
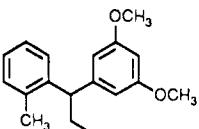
analogues is that the saturated six-membered ring in the former occupies a larger space than the planar aromatic ring present in the latter compound. This suggests that a limited pocket favorable for hydrophobic interactions may exist near the C2 position of adenosine at the A₂ receptor.

The compounds discussed so far were synthesized for SAR studies preliminary to the synthesis of the corresponding 2-(phenylamino) analogues. Our primary interest was to generate a series of CV-1808 analogues having a wide variety of N⁶ substituents. We first synthesized analogues in which we incorporated the N⁶ side chains of highly A₂-selective agonists⁸ as represented by compounds 47, 49, and 51. The receptor-binding results were discouraging (Table IV), since the anticipated additivity was not observed in these molecules. However, lack of additivity has also been reported with N⁶, 5' doubly modified agonists.^{8,14} Compound 47 loses significant affinity at both receptors, yet gains some selectivity for the A₂ receptor. Similarly, compounds 49 and 51, although highly selective, lose potency significantly at both the receptors. Compound 47 shows the highest loss of affinity at the A₂ receptor (400-fold) when compared to its parent, 46.

Several 2-(phenylamino) derivatives of A₁-selective N⁶-modified adenosines were synthesized in order to determine whether the lack of additivity seen with A₂-selective N⁶ derivatives would also pertain to A₁-selective compounds. Most of the resultant compounds (30, 55, 56, 58, and 60) showed increased A₂ affinity and, although still A₁-selective, were much less so than the parent N⁶ derivatives. These results indicate that 2-(phenylamino) sub-

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Table IV. Effects of 2-(Phenylamino) Substitution on A₁ and A₂ Affinities of N⁶-Modified Adenosines


| example | R | X | K _i , nM ^a | | A ₂ /A ₁ |
|-----------------|---|------|----------------------------------|----------------|--------------------------------|
| | | | A ₁ | A ₂ | |
| 7 | H | NHPh | 600 | 116 | 0.190 |
| 12 | 1-naphthylmethyl | H | 24 | 9.4 | 0.38 |
| 44 | 1-naphthylmethyl | NHPh | 560 | 230 | 0.41 |
| 11 | 2,2-diphenylethyl | H | 6.8 | 25 | 3.6 |
| 45 | 2,2-diphenylethyl | NHPh | 2700 | 650 | 0.24 |
| 46 | 9-fluorenylmethyl | H | 5.2 | 4.9 | 0.94 |
| 47 | 9-fluorenylmethyl | NHPh | 8900 | 2100 | 0.24 |
| 48 |  | H | 30 | 6.1 | 0.20 |
| 49 | same as above | NHPh | 9000 | 470 | 0.052 |
| 50 |  | H | 142 | 4.4 | 0.031 |
| 51 | same as above | NHPh | 10300 | 340 | 0.034 |
| 52 | CH ₂ Ph | H | 120 | 280 | 2.4 |
| 53 | CH ₂ Ph | NHPh | 1630 | 7100 | 4.4 |
| 54 | cyclopropyl | H | 3.2 | 1240 | 390 |
| 55 | cyclopropyl | NHPh | 68 | 960 | 14.1 |
| 8 | cyclopentyl | H | 0.59 | 460 | 780 |
| 30 | cyclopentyl | NHPh | 12.4 | 144 | 11.6 |
| 9 | cyclohexyl | H | 1.42 | 610 | 430 |
| 56 | cyclohexyl | NHPh | 54 | 450 | 8.4 |
| 57 | 2-endo-norbornyl | H | 0.42 | 770 | 1850 |
| 58 | 2-endo-norbornyl | NHPh | 7.7 | 472 | 61 |
| 59 | (S)-2-hydroxypropyl | H | 5.0 | 9300 | 1870 |
| 60 | (S)-2-hydroxypropyl | NHPh | 50 | 1700 | 34 |
| 10 | (R)-1-methyl-2-phenylethyl | H | 1.17 | 124 | 106 |
| 61 | (R)-1-methyl-2-phenylethyl | NHPh | 152 | 240 | 1.56 |
| 62 ^b | 1-pyrrolidinyl | NHPh | 18400 | 58000 | 3.2 |

^a A₁ affinities were determined in [³H]-N⁶-cyclohexyladenosine binding to rat whole-brain membranes, and A₂ affinities were determined in [³H]-NECA binding to rat striatal membranes.⁵ Results for compounds 7 and 52 are from ref 5; those for 48 and 50 are from ref 8; that of 11 is from ref 9; those for 12 and 46 are from ref 10; that of 58 is from ref 11; those for 8 and 9 are from ref 15; that of 59 is from ref 16. A₁ and A₂ affinities for the above compounds and A₂ affinities for compounds 44, 47, 51, 58, and 61 are means of three separate experiments. Other values are single determinations. ^b Compound 62 is 2-(phenylamino)-6-(1-pyrrolidinyl)-9-β-D-ribofuranosyl-9H-purine.

stitution can improve A₂ affinity as well as reduce A₁ affinity, implying that the 2-position domain of the A₂ receptor may contain a hydrophobic binding region that is absent in the A₁ receptor. An interesting exception to the rule that A₁-selective N⁶ derivatives show additive effects on 2-(phenylamino) substitution is 61, a combination between R-PIA (10) and CV-1808 (7). This compound shows a 2-fold loss of A₂ affinity compared to R-PIA (10), suggesting that the lack of additivity seen with A₂-selective agonists is not due to A₂ selectivity per se, but rather to structural features that the A₂-selective compounds share with R-PIA but not with the other A₁-selective agonists in Table IV. The pertinent structural feature is probably the presence of one or more aromatic rings distal to N⁶. R-PIA and the A₂-selective compounds possess aralkyl side chains, whereas all of the remaining A₁-selective compounds have compact alkyl or cycloalkyl groups at N⁶.

We can envision several possible explanations for the lack of additivity between C2 and N⁶ modifications. The first would involve direct steric interference between the two side chains due to a partial overlap between the C2 and N⁶ aryl-binding pockets. Thus, distal aryl groups at

C2 and N⁶ would occupy their respective pockets when present alone, but could not both occupy the overlapping portion of the two pockets when present in the same molecule. In this case, one of the two aryl groups would be forced into an unfavorable position, resulting in loss of affinity. This interpretation is consistent with present knowledge of adenosine-receptor structure-activity relationships. In particular, the N⁶ side chain most likely points in the direction of N¹ rather than N⁷,¹¹ so the more distal parts of the N⁶ chain could easily wrap around into the C2 pocket. Whether the N⁶ chain actually does bend in that direction is unclear, since the distal portion of the N⁶ domain has not yet been mapped. However, this hypothesis does explain why additivity was seen with alkyl and cycloalkyl groups at N⁶, since these groups would not extend far enough to encroach into the C2 pocket.

Alternatively, the two side chains could interfere with each other in an indirect manner. For instance, binding of the N⁶ side chain could induce an allosteric change in the receptor, resulting in closure of the C2 pocket. Conversely, the presence of a side chain at C2 or N⁶ might cause a shift in the position of adenosine on the receptor,

thereby displacing the other side chain to an unfavorable location. Although the present data do not distinguish between direct and indirect steric interference, the previously reported lack of additivity between N⁶ and 5' substitution^{5,14} must be due to indirect interactions, since the N⁶ and 5' positions are too far apart for any direct steric interactions.

2-(Phenylamino) substitution lowered A₁ affinity in all cases, with the magnitude of the effect ranging from 10-fold for compound 60 to 1700-fold for compound 47. These results indicate that the C2 domain of the A₁ receptor does not readily accept the phenylamino group. Interestingly, compound 60 also showed the greatest enhancement of A₂ affinity with 2-(phenylamino) substitution, while compound 47 showed the greatest deterioration. These parallelisms suggest that the A₁ receptor may also demonstrate disruptive interactions between N⁶-aralkyl groups and the 2-(phenylamino) substituent.

Finally, compound 62 confirms the importance of an N⁶-hydrogen for binding affinity at both the receptors, implying that the SAR of the 6-amino group itself is unaltered.

In summary, we have demonstrated that there is a hydrophobic binding site near the C2 position of adenosine that is specific for the aromatic function. Occupation of this site increases A₂ affinity and selectivity when N⁶ is occupied by an alkyl or cycloalkyl group, but not when N⁶ is occupied by an aralkyl side chain.

Experimental Section

Melting points are uncorrected. Analytical thin-layer chromatography (TLC) was done with precoated glass plates (EM Science silica gel 60F-254). Flash column chromatography was performed on silica gel 60 (230–400 mesh). ¹H NMR spectra were obtained on Varian EM-390 or Varian XL-200 spectrometer. Mass spectra were recorded on a Finnegan 4500 mass spectrometer with an INCOS data system or a VG 7070 E/HR mass spectrometer with an 11/250 data system. Solvents and reagents were commercially available unless otherwise noted and were used directly. Elemental analyses were determined at Parke-Davis.

General Method for the Preparation of Compounds 13–17. **N-Cyclopentyl-2-aminoadenosine (13).** A mixture of 6-chloro-2-amino-9-β-D-ribofuranosyl-9H-purine (2.0 g, 6.6 mmol), cyclopentylamine (0.85 g, 9.9 mmol), and Et₃N (1.17 g, 11.0 mmol) in ethanol (50 mL) was refluxed under nitrogen for 18 h. The volatiles were evaporated, and the residue was purified by flash column chromatography on silica gel with 5% methanol–chloroform as an eluent. Evaporation of solvent from pure fractions followed by crystallization from ethanol–hexane afforded 1.9 g (82.6%) of *N*-cyclopentyl-2-aminoadenosine (13): mp 210–211 °C; ¹H NMR (DMSO-*d*₆) δ 1.4–2.0 (m, 8 H, cyclopentane), 3.46–3.6 (m, 2 H, 2 H₅), 3.9 (m, 1 H, CH-cyclopentane), 4.1 (m, 1 H, H₄), 4.4–4.7 (m, 2 H, H₃ and H₂), 5.1 (d, 1 H, 3'-OH), 5.3–5.6 (m, 2 H, 5'-OH and 2'-OH), 5.75 (d, 1 H, H₁), 5.8 (s, 2 H, NH₂), 7.15 (d, 1 H, NH), 7.95 (s, 1 H, H₈); mass spectrum, *m/z* 350. Anal. (C₁₅H₂₂N₆O₄) C, H, N.

General Method for the Preparation of the Compounds of Scheme II. **6-Chloro-2-(phenylsulfonyl)-9-(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)-9H-purine (25).** A solution of 6-chloro-2-(phenylthio)-9-(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)-9H-purine (19¹²) (1.7 g, 3.25 mmol) in acetic acid (30 mL) was added to a solution of potassium permanganate (1.50 g, 9.8 mmol) in water (15 mL) at 0 °C in an ice bath. The reaction mixture was stirred at 0 °C for 3 h, and then water was added until a clear yellow solution resulted. The solution was extracted with chloroform (3 × 150 mL) and washed with 5% NaHCO₃ (2 × 25 mL), followed by brine (1 × 50 mL). The organic phase was dried over Na₂SO₄ and filtered, and the solvent was evaporated to yield 1.5 g (82%) of a solid (25): ¹H NMR (CDCl₃) δ 2.1 (s, 3 H, COCH₃), 2.13 (s, 3 H, COCH₃), 2.21 (s, 3 H, COCH₃), 4.43–4.51 (m, 3 H, 1 H₄, 2 H₅), 5.6 (t, 1 H, H₃), 5.76 (t, 1 H, H₂), 6.28 (d, 1 H, H₁), 7.56–7.68 (m, 3 H, phenyl), 8.20 (d, 2 H, phenyl), and 8.48 (s, 1 H, H₈); mass spectrum (FAB), *m/z* 552.9 (M⁺).

N-Cyclopentyl-2-(phenylsulfonyl)adenosine 2',3',5'-Triacetate (26). A solution of 6-chloro-2-(phenylsulfonyl)-9-(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)-9H-purine (25) (1.09 g, 1.97 mmol) and triethylamine (0.29 g, 2.8 mmol) in DME (35 mL) was added to a solution of cyclopentylamine (0.2 g, 2.8 mmol) in DME (5 mL) and the mixture was stirred at room temperature for 3 h. The precipitated solid was filtered (Et₃N⁺HCl⁻) and washed with DME, and the volatiles were evaporated. The residue was purified by flash column chromatography on silica gel with CHCl₃ as an eluent. Evaporation of the solvent from the pure fractions (*R*_f = 0.65 in 5% MeOH–CHCl₃) afforded 0.96 g (81%) of *N*-cyclopentyl-2-(phenylsulfonyl)adenosine 2',3',5'-triacetate (26): mp 77–81 °C; ¹H NMR (CDCl₃) δ 1.67–2.08 (brm, 8 H, cyclopentane), 2.09 (s, 3 H, COCH₃), 2.13 (s, 3 H, COCH₃), 2.18 (s, 3 H, COCH₃), 4.38–4.45 (m, 4 H, 2 H₅, H₄, and CH-cyclopentane), 5.57 (t, 1 H, H₃), 5.72 (t, 1 H, H₂), 5.9 (d, 1 H, H₄), 6.2 (brd, 1 H, NH), 7.55–7.64 (m, 3 H, phenyl), 7.99 (s, 1 H, H₈), and 8.17 (d, 2 H, phenyl); mass spectrum (FAB), *m/z* 602.2. Anal. (C₂₇H₃₁N₅O₉S) C, H, N, S.

N-Cyclopentyl-2-(phenylsulfonyl)adenosine (27). A mixture of *N*-cyclopentyl-2-(phenylsulfonyl)adenosine 2',3',5'-triacetate (26) (0.81 g, 1.3 mmol) in saturated methanolic ammonia (25 mL) was stirred at room temperature for 3 h. The volatiles were evaporated, and the residue was purified by flash column chromatography on silica gel with 4% MeOH–CHCl₃ as an eluent. Evaporation of solvent from the pure fraction gave 0.36 g (56%) of *N*-cyclopentyl-2-(phenylsulfonyl)adenosine (27): mp 72–75 °C; ¹H NMR (DMSO-*d*₆) δ 1.45–2.0 (m, 8 H, cyclopentane), 3.52–3.65 (m, 2 H, 2 H₅), 3.95 (m, 1 H, CH-cyclopentane), 4.12 (m, 2 H, H₃ and H₄), 4.58 (q, 1 H, H₂), 4.97 (t, 1 H, 5'-OH), 5.25 (d, 1 H, 3'-OH), 5.49 (d, 1 H, 2'-OH), 5.91 (d, 1 H, H₁), 7.66 (m, 3 H, phenyl), 7.99 (d, 2 H, phenyl), 8.54 (d, 1 H, NH), and 8.58 (s, 1 H, H₈); mass spectrum (FAB), *m/z* 476. Anal. (C₂₁H₂₅N₅O₆S) C, H, N, S.

General Methods for the Preparation of the Compounds of Scheme III. **N-(1-Naphthylmethyl)-2-fluoroadenosine 2',3',5'-Triacetate (32).** A mixture of 6-chloro-2-fluoro-9-(2,3,5-tri-*O*-acetyl-β-D-ribofuranosyl)-9H-purine (31)¹³ (2.0 g, 4.6 mmol), (1-naphthylmethyl)amine (0.8 g, 5.0 mmol), and Et₃N (0.52 g 5.0 mmol) in dimethoxyethane (25 mL) was stirred at room temperature for 20 h. The precipitated solid (Et₃NH⁺Cl⁻) was filtered and washed with DME, and the volatiles were evaporated. The residue was purified by flash column chromatography on silica gel with CHCl₃ as an eluent. Evaporation of solvent from pure fractions afforded 1.5 g (58.6%) of *N*-(1-naphthylmethyl)-2-fluoroadenosine 2',3',5'-triacetate (32): mp 76–80 °C; ¹H NMR (CDCl₃) δ 2.08 (s, 3 H, COCH₃), 2.13 (s, 3 H, COCH₃), 2.14 (s, 3 H, COCH₃), 4.4 (m, 3 H, 2 H₅), 5.8 (t, 1 H, H₂), 6.1 (d, 1 H, H₁), 6.4 (brs, 1 H, NH), and 7.27–8.08 (m, 8 H, phenyl and H₈); mass spectrum, *m/z* (relative intensity) 551 (50), 293 (60), 256 (60), 139 (100). Anal. (C₂₇H₂₆N₅O₇F) C, H, N, F.

N-(1-Naphthylmethyl)-2-fluoroadenosine (36). A solution of *N*-(1-naphthylmethyl)-2-fluoroadenosine 2',3',5'-triacetate (32) (1.2 g, 2.18 mmol) in saturated methanolic ammonia (20 mL) was stirred at room temperature for 2.5 h. The volatiles were evaporated, and the residue upon treatment with CHCl₃–hexane afforded a precipitate, which was filtered and dried, resulting in 0.89 g (96%) of *N*-(1-naphthylmethyl)-2-fluoroadenosine (36): mp 131–135 °C; ¹H NMR (DMSO-*d*₆) δ 3.5–3.67 (m, 2 H, 2 H₅) 3.95 (d, 1 H, H₄), 4.15 (q, 1 H, H₃), 4.55 (q, 1 H, H₂), 5.06 (t, 1 H, 5'-OH), 5.13 (d, 2 H, CH₂ naphthyl), 5.22 (d, 1 H, 3'-OH), 5.49 (d, 1 H, 2'-OH), 5.82 (d, 1 H, H₁), 7.5–8.2 (m, 7 H, phenyl), 8.42 (s, 1 H, 8 H), and 9.09 (brs, 1 H, NH); mass spectrum (FAB), *m/z* 426.1 (M⁺). Anal. (C₂₁H₂₀N₅O₄F) C, H, N, F.

N-(1-Naphthylmethyl)-2-[(phenylmethyl)amino]adenosine (38). A mixture of *N*-(1-naphthylmethyl)-2-fluoroadenosine (36) (0.5 g, 1.1 mmol), benzylamine (0.25 g, 2.3 mmol), and triethylamine (0.23 g, 2.3 mmol) in ethanol (20 mL) was refluxed for 24 h. Upon cooling, crystalline material was obtained, which was filtered and dried, affording 0.35 g (58%) of *N*-(1-naphthylmethyl)-2-[(phenylmethyl)amino]adenosine (38): mp 205–208 °C; ¹H NMR (DMSO-*d*₆) δ 3.42–3.64 (m, 2 H, 2 H₅), 3.85 (d, 1 H, H₄), 4.08 (q, 1 H, H₃), 4.38 (d, 2 H, CH₂Ph), 4.52 (q, 1 H, H₂), 5.09 (brd, 4 H, CH₂-naphthyl, 5'-OH, and 3'-OH), 5.32 (d, 1 H, 2'-OH), 5.71 (d, 1 H, H₁), 6.92 (brt, 1 H, NH), 7.13–8.24 (m, 12 H, phenyl). Anal. (C₁₈H₂₈N₆O₄) C, H, N.

***N*-(2-Phenylethyl)-2-[(2-phenylethyl)amino]adenosine (35).** A reaction mixture of 2-fluoro-6-chloro-9-(2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl)-9H-purine (31) (1.2 g, 2.7 mmol), 2-phenylethylamine (0.37 g, 3.0 mmol), and triethylamine (0.3 g, 3.0 mmol) in DME (20 mL) was stirred at room temperature for 20 h. TLC (5% MeOH-CHCl₃) showed the absence of starting material. The precipitated solid (Et₃NH⁺Cl⁻) was filtered and washed with DME, and the volatiles were removed from the filtrate. The residue was then dissolved in saturated methanolic ammonia (15 mL) and stirred at room temperature for 3 h. The volatiles were removed, and the residue was treated with CHCl₃-MeOH-ether. The solid (1.1 g) was filtered and dried. ¹H NMR CDCl₃ showed a 1:1 mixture of compounds 33 and 34. All the sugar protons were duplicate and the C8 proton from both the compounds showed two separate singlets at δ 8.37 and 8.39 in a 1:1 ratio.

This 1:1 mixture of 33 and 34 was further reacted with an excess of 2-phenylethylamine in refluxing ethanol for 20 h. The volatiles were evaporated, and the residue upon crystallization from chloroform-ether (1:4) gave a solid, which was filtered and dried, affording 0.9 g (62.5% from 31 of *N*-(2-phenylethyl)-2-[(2-phenylethyl)amino]adenosine (35): mp 137-142 °C; ¹H NMR (DMSO-*d*₆) δ 2.7-3.0 (m, 4 H, CH₂Ph), 3.42-3.65 (m, 6 H, 2 CH₂NH and 2 H₅), 3.88 (brd, 1 H, H₄), 4.10 (brt, 1 H, H₃), 4.57 (t, 1 H, H₂), 4.6-5.5 (br, 3 H, 3 OH), 5.73 (d, 1 H, H₁), 6.34 (s, 1 H, NH), 7.22 (brs, 10 H, aromatic), 7.29 (s, 1 H, NH), and 7.90 (s, 1 H, H₈); mass spectrum (FAB), *m/z* 491.1 (M⁺). Anal. (C₂₆H₃₀N₆O₄·1.5H₂O) C, H, N.

Receptor Binding. Affinities of compounds for inhibition of binding of [³H]-*N*⁶-cyclohexyladenosine to A₁ receptors in rat brain membranes and for inhibition of [³H]NECA binding to A₂ re-

ceptors in rat striatal membranes in the presence of 50 nM *N*⁶-cyclopentyladenosine were determined as previously described.⁵

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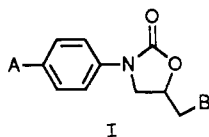
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Antibacterials. Synthesis and Structure-Activity Studies of 3-Aryl-2-oxooxazolidines. 1. The "B" Group

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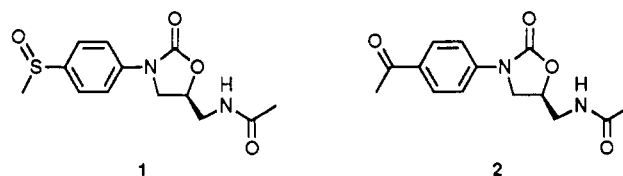
The synthesis and structure/activity studies of the effect of varying the "B" group in a series of oxazolidinone antibacterials (I) are described. Two synthetic routes were used: (1) alkylation of aniline with glycidol followed



by dialkyl carbonate heterocyclization to afford I (A = H, B = OH), whose arene ring was further elaborated by using electrophilic aromatic substitution methodology; (2) cycloaddition of substituted aryl isocyanates with epoxides to give A and B with a variety of values. I with B = OH or Br were converted to other "B" functionalities by using S_N2 methodology. Antibacterial evaluation of compounds I with A = acetyl, isopropyl, methylthio, methylsulfinyl, methylsulfonyl, and sulfonamido and a variety of different "B" groups against *Staphylococcus aureus* and *Enterococcus faecalis* concluded that the compounds with B = aminoacyl, and particularly acetamido, were the most active of those examined in each A series, possessing MICs in the range of 0.5-4 μ g/mL for the most active compounds described.

The oxazolidinones,¹ exemplified by DuP 105 (1) and DuP 721 (2), are a new class of orally active, synthetic antibacterial agents, derived from a random screening lead, whose in vitro spectrum includes activity against staphylococci, streptococci, enterococci, anaerobic bacteria, and mycobacteria.²

As a class, the oxazolidinones have equal activity against methicillin-sensitive and -resistant staphylococcal strains and β -lactamase positive and negative strains.^{3,4} Pharmacokinetic studies on DuP 721 indicate that peak serum



levels exceeding the MIC₉₀'s can readily be achieved following single doses per os.⁵ Mechanism of action studies

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(1) Fugitt, R. B.; Luckenbaugh, R. W. U.S. Patent 4,340,606, July 20, 1982. Gregory, W. A. U.S. Patent 4,461,773, July 24, 1984. Gregory, W. A. U.S. Patent, 4,705,799, November 10, 1987.