

Tetrahedron Letters 39 (1998) 3953-3956

TETRAHEDRON LETTERS

Synthesis of D-erythro-Sphingosine and D-erythro-Sphinganine Via 3-Ketosphinganine

Robert V. Hoffman* and Junhua Tao

Department of Chemistry and Biochemistry

New Mexico State University

Las Cruces, NM 88003-001

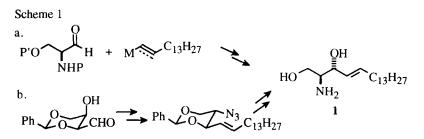
Received 19 February 1998; accepted 24 March 1998

Abstract: D-erythro- sphingosine and D-erythro-sphinganine can be produced in protected form from serine by a synthetic approach in which the normal biological intermediate 3-ketosphinganine in protectyed form, is a key synthetic intermediate. The sequence is short and convergent, proceeds in good overall yields ($\approx 30\%$ for 6 steps) and with excellent stereocontrol (>91% de, >95% ee). © 1998 Elsevier Science Ltd. All rights reserved.

The finding that sphingolipids are involved in "essentially all aspects of cell regulation"¹ has led to an explosion of interest in sphingolipid chemistry.¹⁻⁴ Sphingosine **1** is the core structure of most sphingolipids **2**, which can vary in the nature of the head group R_1 , the structure of the N-acyl group R_2 (if one is present), and the structure of the sphingosine tail R_3 .⁵ For example, the saturated analog sphinganine **3** lacks the biological activity of sphingosine highlighting the importance of the double bond.



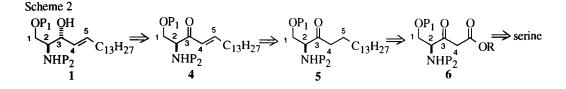
Since most sphingolipids are prepared from sphingosine, quite a number of syntheses of sphingosine and its derivatives have been reported. In general these syntheses fall into three main categories.⁶ The first uses stereoselective addition of an organometallic reagent (often a lithium acetylide) to a protected serinal. This approach forms the 3,4 carbon-carbon bond and sets the stereochemistry of the C-3 hydroxyl group in one step (Scheme 1a).⁷ The second major strategy uses carbohydrate precursors as the source of stereochemistry at C-2 and C-3. The tail is then attached by an anionic addition of some type (Scheme 1b).^{6,8} The third major category



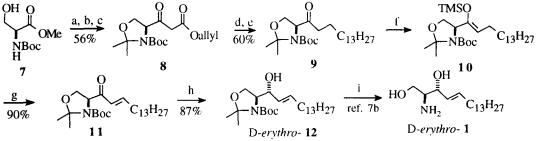
0040-4039/98/\$19.00 © 1998 Elsevier Science Ltd. All rights reserved. *PII:* S0040-4039(98)00733-3 uses a variety of chiral precursors to build up the structure by nucleophilic addition processes.⁹ All these approaches set the stereochemistry of the head groups early and attach the tail as a nucleophile.

The D-erythro stereochemistry of 1 is most common, but all four possible diastereomers of the 2,3-amino alcohol unit are known and are all bioactive to different degrees.¹⁰ Thus stereochemical control at C-2 and C-3 is crucial to any synthesis. Moreover, the *trans* geometry of the sphingosine $C_{13}H_{27}$ alkene tail is crucial for both activity and ease of purification since the separation of *cis* and *trans*-sphingosines is very tedious.

Our interest in the synthesis of densely functionalized molecules¹¹ led to the retrosynthetic scheme for protected D-*erythro*-sphingosine 1 shown in Scheme 2. This strategy is fundamentally different from previous syntheses in both sequence and polarity. The tail is attached as an electrophile to 6. After introduction of the double bond in ketosphinganine 5, stereoselective reduction of the ketone group of 3-ketosphingosine 4 sets the stereochemistry of the C-3 hydroxyl group in the last stage of the synthesis. This approach mimics to some degree the biological route to sphingosines which also passes through 3-ketosphingosine 1 requires that the reduction of 5 proceeds by a chelated transition state. Chelation control using an N-Boc oxazolidine for P₁ and P₂ provides a means to cleanly control the stereochemical outcome of the reduction.^{7a}



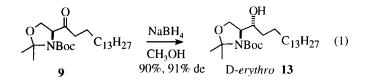
This strategy was successfully reduced to practice as seen in Scheme 3. Commercially available L-(*N*-Boc) serine methyl ester **7** was cyclized to the corresponding 2,2-oxazolidine with 2,2-dimethoxypropane.¹³ Conversion to β-ketoester **8** in 56% yield with CDI and lithio allyl acetate followed a standard procedure.¹⁴ Ketoester **8** was alkylated by treatment with NaH followed by 1-tetradecyl triflate at room temperature for 6 h. 1-Bromotetradecane could also be used to alkylate the enolate of **8** by refluxing in THF-HMPA (5:1) with 10% NaI for 6 h.The milder triflate alkylation procedure is preferred in order to minimize the chances of epimerization. Treatment of the crude alkylation product with Pd(PPh₃)₄ and morpholine gave deallylation and decarboxylation Scheme 3



a. $(CH_3)_2CH(OCH_3)_2$, TsOH; b. LiOH; c. (i) CDI, (ii) LiCH₂CO₂allyl; d. (i) NaH, (ii) TfOCH₂C₁₃H₂₇; e. Pd(PPh₃)₃, morpholine; f.(i) NaHMDS, -78°C, (ii)TMSCl; g. Pd(OAc)₂, CH₃CN; h. NaBH₄, CeCl₃, -20°C; i. 1N HCl to 3-ketosphinganine derivative 9 in 60% yield. The use of allyl esters in β -ketoester 8 is a significant improvement over *t*-butyl esters used earlier.^{11b} Palladium [0] not only removes the allyl group under mild, neutral conditions, but it also catalyzes the decarboxylation of the resulting β -keto acid which occurs smoothly at room temperature.

Treatment of 9 with NaHMDS followed by TMSCl gave TMS-enol ether of 10^{15} which was oxidized with Pd(OAc)₂ to the α , β -unsaturated ketone 11 (90%).¹⁶ The nmr spectrum of 11 had only one set of vinyl signals with J= 15.7 Hz indicating that the *trans* isomer was produced exclusively. In practice, 11 was not purified but carried on as the crude product. Reduction with NaBH₄/ CeCl₃ (87%) gave the known D-*erythro*-sphingosine derivative 12.^{7b} The CeCl₃ was needed to suppress conjugate reduction of 11 which occurred to the extent of 25% in its absence. The diastereoselectivity of the reduction was excellent (92% de) and the *anti*-stereochemistry results from the expected chelation controlled reduction. An LIS study (Eu(hfc)₃) of the major diastereomer 12 showed the sequence was highly enantioselective (>95% ee). Deprotection of 12 to D-*erythro*-sphingosine 1 using 1 N HCl is straightforward.^{7b}

As expected, reduction of 3-ketosphinganine 9 with sodium borohydride gave D-*erythro*-sphinganine derivative 13 (90% yield, 91% de) without the need for CeCl₃ because conjugate reduction is not an issue (eq 1).



In summary, D-erythro- sphingosine and D-erythro-sphinganine can be produced in protected form from serine by a synthetic approach in which 3-ketosphinganine is a key synthetic intermediate and thus mimics to some degree the biological route. The relatively short sequence proceeds in good overall yields ($\approx 30\%$ for 6 steps) and with excellent stereocontrol (>91% de, >95% ee). The extension of this methodology to the synthesis of other sphingosine derivatives and analogs is currently underway.

Acknowledgement This work was supported by a grant from the National Science Foundation (CHE 9520431).

References and Notes

- For an excellent overview of sphingolipid functions see Merrill, A. H., J.r; Sweeley, C. C. in "Biochemistry of Lipids, Lipoproteins and Membranes", Vance, D. E.; Vance, J. E. Eds.; Elsevier Science B. V.: Amsterdam, 1996; Cpt. 12, pp 309-339.
- 2. Hannun, Y. A.; Loomis, C. R.; Merrill, A. H., Jr.; Bell, R. M. J. Biol. Chem. 1986, 261, 12604.
- 3. Hannun, Y. A.; Bell, R. M. Science 1989, 243, 500.
- 4. a. Merrill, A. H., Jr.; Hannun, Y. A.; Bell, R. M. in "Advances in Lipid Research, Vol. 25 Sphingolipids Part A: Functions and Breakdown Products" Bell, R. M.; Hannun, Y. A.; Merrill, A. H., Jr., Eds.; Academic Press: San Diego, CA; 1993, pp 1-23. b. Bell, R. M.; Hannun, Y. A.; Merrill, A. H., Jr., Eds."Advances in

Lipid Research, Vol. 25 Sphingolipids Part A: Functions and Breakdown Products"; Academic Press: San Diego, CA; 1993. c. Cevc, G, Ed. "Phospholipids Handbook"; Marcel Dekker: New York, 1993.

- 5. Karlsson, K.-A. Lipids 1970, 5, 878.
- For an excellent discussion of synthetic approaches see a. Schmidt, R. R.; Bar, T.; Wild, R. Synthesis 1995, 868. b. Schmidt, R. R. in "Synthesis in Lipid Chemistry", Tyman, J. H. P., Ed; Royal Society of Chemistry: Cambridge, UK; 1996, pp 93-118 and references therein.
- For example: a. Nimkar, S.; Menaldino, D.; Merrill, A. H.; Liotta, D. Tetrahedron Lett. 1988, 29, 3037. b. Garner, P.; Park, J. M.; Malecki, E. J. Org. Chem. 1988, 53, 4395. c. Herold, P. Helv. Chim. Acta. 1988, 71, 354. d. Polt, R.; Peterson, M. A.; DeYoung, L. J. Org. Chem. 1992, 57, 5469 and references therein. e. Soai, K.; Takahashi, K. J. Chem. Soc., Perkins Trans. I 1994, 1258. f. Dondoni, A.; Perrone, D.; Turturici, E. J. Chem. Soc., Perkins Trans. I 1997, 2389.
- For example: a. Hirata, N.; Yamagiwa, Y.; Kamikawa, T. J. Chem. Soc., Perkins Trans. 1 1991, 2279. b. Li, Y.-L.;
 Wu, Y.-L. Liebigs Ann. 1996, 2079. c. Murakami, T.; Hato, M. J. Chem. Soc. Perkins Trans. 1 1996, 823.
- See for example: a. Katsumura, S.; Yamamoto, N.; Fukuda, E.; Iwama, S. Chem. Lett. 1995, 393 and references therein. b. Davis, F. A.; Reddy, G. V. Tetrahedron Lett. 1996, 37, 4349. c. Hudlicky, T.; Nugent, T.; Griffith, W. J. Org. Chem. 1994, 59, 7944. d. Garigipati, R. S.; Weinreb, S. M. J. Am. Chem. Soc. 1983, 105, 4499. e. Spanu, P.; Rassu, G.; Pinna, L.; Battistini, L.; Casiraghi, G. Tetrahedron: Assym. 1997 8, 3237. f. Solladié-Cavallo, A.; Koessler, J. L. J. Org. Chem. 1994, 59, 3240.
- Sachs, C. W.; Ballas, L. M.; Mascarella, S. W.; Safa, A. R.; Lewin, A. H.; Loomis, C.; Carroll, F. I.; Bell, R. M.; Fine, R. L. *Biochem. Pharmacol.* 1996, *52*, 603.
- a. Hoffman, R. V.; Tao, J.-H. J. Org. Chem. 1997, 62, 2292. b. Tao, J.-H, Hoffman, R. V.; J. Org. Chem. 1997, 62, 6240.
- 12. Geeraert, L.; Mannaerts, G. P.; Van Veldhoven, P. P. Biochem. J. 1997, 327, 125.
- 13. Garner, P.; Park, J. M. J. Org. Chem. 1987, 52, 2361.
- 14. Harris, B. D.; Joullie, M. M.; . Tetrahedron 1988, 44, 3489.
- 15. It is known that bulky substituents on the nitrogen of α-aminoketones promote enolization at the α'-position. Lubell, W. D.; Rapoport, H. J. Am. Chem. Soc. 1988, 110, 7447. b. Alexander, C. W.; Liotta, D. C. Tetrahedron Lett. 1996, 37, 1961 and references therein.
- 16. Ito, Y.; Hirao, T.; Saegusa, T. J. Org. Chem. 1978, 43, 1011.