

Supporting Information

Experimental and Computational Probes of a Self-Assembled Capsule

Tetsuo Iwasawa, Dariush Ajami, and Julius Rebek, Jr.*

The Skaggs Institute for Chemical Biology and the Department of Chemistry, The Scripps Research Institute

MB-26, 10550 North Torrey Pines Road, La Jolla, California 92037

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(1) General NMR spectra were recorded on a Bruker DRX-600 spectrometer. Proton (^1H) chemical shifts, reported in parts per million (ppm), were indirectly referenced to external tetramethylsilane employing resonances due to trace monoprotio-solvent as an internal reference. Deuterated NMR solvents were obtained from Cambridge Isotope Laboratories, Inc., and used without further purification.

(2) Materials Compounds **3a**, **6a – 6c**, and **5** were purchased from Aldrich Chemical Company, and were used as received. The compound **6e** was purchased from Acros Organics, and were used as received. The starting materials for other biaryls, 3-bromopyridine, 2-bromopyridine, 2-bromo-5-methylpyridine, 4-bromotoluene, 5-bromopyrimidine, *p*-tolylboronic acid, 4-ethylphenylboronic acid,

4-*n*-propylphenylboronic acid, and 4-*n*-butylphenylboronic acid, were purchased from Aldrich Chemical Company., and were used without further purification. The 5-bromo-2-methylpyridine was purchased from Apollo Scientific Ltd., and was used without further purification. The reagents for cross-coupling reactions, Pd(OAc)₂, K₃PO₄, K₂CO₃, and tetra-*n*-butylammonium bromide were purchased from Aldrich Chemical Company. The catalyst Pd(OAc)₂(Cy₂NH)₂ was prepared according to the literature.¹ The cylindrical cavitand **1** was prepared according to the literature.²

(3) Complete list of author for Guassian, Inc., Pittsburgh, PA, 2002: Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, Jr., J. A.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.; Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; and Pople, J. A.; Gaussian, Inc., Wallingford CT, 2004.

(4) Preparation of biaryls in Figure 1b. The biaryls **2a – 2g**, **3b – 3g**, **6d**, and **7a – 7c** were prepared according to the Suzuki-Miyaura cross-coupling under aerobic conditions.¹ The synthesis of biaryl **4** was referred to the literature.³ Typical procedures about **2f** and **4** were described below, as well as data of other biaryls. The ¹H NMR and ¹³C NMR spectra of **2a – 2g**, **3b – 3g**, **4**, **6d**, and **7a – 7c** are also attached in from page 8S – 44S.

2f: In the air, to a 50 mL one neck flask K₃PO₄ (850 mg, 4 mmol), Pd(OAc)₂(Cy₂NH)₂ (23 mg, 0.04 mmol), 5-bromo-2-methylpyridine (344 mg, 2 mmol), 4-*n*-propylboronic acid (394 mg, 2.4 mmol), and EtOH (6 mL) were added. After stirring at ambient temperature

for overnight, EtOAc (50 mL) was added, and the mixture was washed with water and brine, and dried over Na₂SO₄. Evaporation of the solvent provided the crude product, which was purified by column chromatography to afford the desired products in 50% yield (210 mg) as pale yellow crystals. ¹H NMR (600 MHz, C₆D₆) δ 8.91 (d, *J* = 2.4 Hz, 1H), 7.40 (dd, *J* = 2.4 Hz, 7.8 Hz, 1H), 7.30 (d, *J* = 8.4 Hz, 2H), 7.04 (d, *J* = 8.4 Hz, 2H), 6.74 (d, *J* = 7.8 Hz, 1H), 2.47 (s, 3H), 2.43 (t, *J* = 7.2 Hz, 2H), 1.52 (dt, *J* = 7.2 Hz, 7.2 Hz, 2H), 0.849 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 157.3, 148.0, 142.2, 136.0, 134.1, 133.8, 129.4, 127.1, 122.9, 37.9, 24.8, 24.2, 13.9. HRMS (ESI, MH⁺): Calcd. For C₁₅H₁₈N: 212.1434. Found: 212.1440.

4³: Under a nitrogen atmosphere, a mixture of K₂CO₃ (1.10 g, 8 mmol), Pd(OAc)₂ (90 mg, 0.4 mmol), tetra-*n*-butylammonium bromide (1.29 g, 4 mmol) and 5-bromo-2-methylpyridine (1.37 g, 8 mmol) in a mix of DMF (0.9 mL) / H₂O (0.35 mL) was stirred for 2 minutes at 115 °C. Isopropanol (480 mg, 8 mmol) was added. After stirring for 48 h, the reaction mixture was allowed to cool to ambient temperature. To the mixture was water and ether, and the organic phase was washed with water and brine, and dried over Na₂SO₄. Evaporation of the solvent provided the crude product, which was purified by column chromatography to afford the desired products in 51% yield (377 mg) as white crystals. ¹H NMR (600 MHz, C₆D₆) δ 8.70 (d, *J* = 2.4 Hz, 2H), 7.18 (dd, *J* = 2.4 Hz, 7.8 Hz, 2H), 6.66 (d, *J* = 7.8 Hz, 2H), 2.44 (s, 6H). ¹³C NMR (150 MHz, C₆D₆) δ 157.9, 147.9, 134.0, 130.8, 123.0, 24.2. HRMS (ESI, MH⁺): Calcd. For C₁₂H₁₃N₂: 185.1073. Found: 185.1067.

2a⁵: ¹H NMR (600 MHz, C₆D₆) δ 8.97 (d, *J* = 1.2 Hz, 1H), 8.52 (d, *J* = 3.6 Hz, 1H), 7.39 – 7.37 (m, 1H), 7.21 (d, *J* = 8.4 Hz, 2H), 6.96 (d, *J* = 8.4 Hz, 2H), 6.78 – 6.77 (m, 1H), 2.10 (s, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 148.8, 148.7, 137.7, 136.6, 135.5, 133.6, 129.9, 127.2, 123.4, 21.0. HRMS (ESI, MH⁺): Calcd. For C₁₂H₁₁N: 170.0964. Found: 170.0970.

2b: ¹H NMR (600 MHz, C₆D₆) δ 8.89 (d, *J* = 2.4 Hz, 1H), 7.39 (dd, *J* = 2.4 Hz, 7.8 Hz, 1H), 7.25 (d, *J* = 8.4 Hz, 2H), 6.99 (d, *J* = 8.4 Hz, 2H), 6.73 (d, *J* = 7.8 Hz, 1H), 2.48 (s, 3H), 2.11 (s, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 157.3, 148.0, 137.4, 135.7, 134.1, 133.8, 129.9, 127.1, 122.9, 24.2, 21.0. HRMS (ESI, MH⁺): Calcd. For C₁₃H₁₃N: 184.1121. Found: 184.1123.

2c: ^1H NMR (600 MHz, C_6D_6) δ 8.98 (d, $J = 1.8$ Hz, 1H), 8.52 (dd, $J = 1.8$ Hz, 4.8 Hz, 1H), 7.41 – 7.39 (m, 1H), 7.25 (d, $J = 7.8$ Hz, 2H), 7.02 (d, $J = 7.8$ Hz, 2H), 6.78 (dd, $J = 4.8$ Hz, 8.4 Hz, 1H), 2.44 (q, $J = 7.8$ Hz, 2H), 1.08 (t, $J = 7.8$ Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 148.8, 148.7, 144.1, 136.6, 135.8, 133.7, 128.8, 127.4, 123.4, 28.8, 15.7. HRMS (ESI, MH^+): Calcd. For $\text{C}_{13}\text{H}_{14}\text{N}$: 184.1121. Found: 184.1128.

2d: ^1H NMR (600 MHz, C_6D_6) δ 8.91 (s, 1H), 7.40 (dd, $J = 2.4$ Hz, 7.8 Hz, 1H), 7.29 (d, $J = 8.4$ Hz, 2H), 7.03 (d, $J = 8.4$ Hz, 2H), 6.74 (d, $J = 7.8$ Hz, 1H), 2.47 (s, 3H), 2.45 (q, $J = 7.8$ Hz, 2H), 1.09 (t, $J = 7.8$ Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 157.3, 148.0, 143.8, 135.9, 134.1, 133.8, 128.7, 127.2, 122.9, 28.8, 24.2, 15.8. HRMS (ESI, MH^+): Calcd. For $\text{C}_{14}\text{H}_{16}\text{N}$: 198.1277. Found: 198.1280.

2e: ^1H NMR (600 MHz, C_6D_6) δ 8.99 (d, $J = 2.4$ Hz, 1H), 8.52 (dd, $J = 2.4$ Hz, 4.8 Hz, 1H), 7.41 – 7.39 (m, 1H), 7.25 (d, $J = 8.4$ Hz, 2H), 7.01 (d, $J = 8.4$ Hz, 2H), 2.42 (t, $J = 7.8$ Hz, 2H), 1.52 (dt, $J = 7.8$ Hz, 7.8 Hz, 2H), 0.84 (t, $J = 7.8$ Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 148.8, 148.7, 142.5, 136.6, 135.8, 133.7, 129.7, 127.3, 123.4, 37.9, 24.8, 13.9. HRMS (ESI, MH^+): Calcd. For $\text{C}_{14}\text{H}_{16}\text{N}$: 198.1277. Found: 198.1284.

2g⁶: ^1H NMR (600 MHz, C_6D_6) δ 9.00 (d, $J = 2.4$ Hz, 1H), 8.53 – 8.52 (m, 1H), 7.42 – 7.40 (m, 1H), 7.26 (d, $J = 8.4$ Hz, 2H), 7.03 (d, $J = 8.4$ Hz, 2H), 6.79 – 6.77 (m, 1H), 2.46 (t, $J = 7.8$ Hz, 2H), 1.49 (dd, $J = 7.8$ Hz, 7.8 Hz, 2H), 1.24 (dt, $J = 7.2$ Hz, 7.8 Hz, 2H), 0.86 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 148.8, 148.7, 142.8, 136.6, 135.8, 133.7, 129.4, 127.3, 123.4, 35.6, 33.9, 22.6, 14.1. HRMS (ESI, MH^+): Calcd. For $\text{C}_{15}\text{H}_{18}\text{N}$: 212.1434. Found: 212.1434.

3b⁷: ^1H NMR (600 MHz, C_6D_6) δ 8.49 (d, $J = 2.4$ Hz, 1H), 8.15 (d, $J = 7.8$ Hz, 2H), 7.35 (d, $J = 8.4$ Hz, 1H), 7.11 (d, $J = 7.8$ Hz, 2H), 6.99 (dd, $J = 2.4$ Hz, 8.4 Hz, 1H), 2.13 (s, 3H), 1.85 (s, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 155.0, 150.5, 138.5, 137.3, 136.8, 131.0, 129.7, 127.0, 119.2, 21.2, 17.8. MS (EI, m/z): 183 (M^+).

3c: ^1H NMR (600 MHz, C_6D_6) δ 8.55 – 8.53 (m, 1H), 8.02 (d, $J = 7.8$ Hz, 2H), 7.36 (d, $J = 7.8$ Hz, 1H), 7.24 – 7.21 (m, 1H), 7.12 (d, $J = 7.8$ Hz, 2H), 6.75 – 6.73 (m, 1H), 2.55 (q, $J =$

7.2 Hz, 1H), 1.20 (t, J = 7.2 Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 157.6, 149.9, 145.3, 137.5, 136.2, 128.5, 127.3, 121.7, 119.8, 28.9, 15.6. HRMS (ESI, MH^+): Calcd. For $\text{C}_{13}\text{H}_{14}\text{N}$: 184.1121. Found: 184.1128.

3d: ^1H NMR (600 MHz, C_6D_6) δ 8.49 – 8.48 (m, 1H), 8.17 (d, J = 7.8 Hz, 2H), 7.36 (d, J = 7.8 Hz, 2H), 7.16 – 7.14 (m, 1H), 6.99 (dd, J = 2.4 Hz, 7.8 Hz, 1H), 2.47 (q, J = 7.8 Hz, 2H), 1.85 (s, 3H), 1.09 (t, J = 7.8 Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 155.0, 150.4, 144.9, 137.5, 136.9, 131.1, 128.5, 127.1, 119.4, 28.9, 17.8, 15.7. HRMS (ESI, MH^+): Calcd. For $\text{C}_{14}\text{H}_{16}\text{N}$: 198.1277. Found: 198.1283.

3e: ^1H NMR (600 MHz, C_6D_6) δ 8.60 – 8.58 (m, 1H), 8.14 (d, J = 8.4 Hz, 2H), 7.38 (d, J = 7.8 Hz, 1H), 7.14 – 7.11 (m, 3H), 6.65 (m, 1H), 2.44 (t, J = 7.2 Hz, 2H), 1.51 (dt, J = 7.2 Hz, 7.8 Hz, 2H), 0.83 (t, J = 7.8 Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 157.6, 150.0, 143.7, 137.5, 136.3, 129.1, 127.2, 121.7, 119.8, 38.0, 24.7, 13.9. HRMS (ESI, MH^+): Calcd. For $\text{C}_{14}\text{H}_{16}\text{N}$: 198.1277. Found: 198.1279.

3f: ^1H NMR (600 MHz, C_6D_6) δ 8.51 – 8.49 (m, 1H), 8.19 (d, J = 8.4 Hz, 2H), 7.37 (d, J = 8.4 Hz, 1H), 7.16 – 7.14 (m, 3H), 7.00 – 6.97 (m, 1H), 2.45 (t, J = 7.8 Hz, 2H), 1.85 (s, 3H), 1.53 (dt, J = 7.8 Hz, 7.2 Hz, 2H), 0.85 (t, J = 7.2 Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 155.1, 150.5, 143.3, 137.6, 136.9, 131.1, 129.1, 127.0, 119.3, 38.0, 24.8, 17.8, 13.9. HRMS (ESI, MH^+): Calcd. For $\text{C}_{15}\text{H}_{18}\text{N}$: 212.1434. Found: 212.1436.

3g⁸: ^1H NMR (600 MHz, C_6D_6) δ 8.60 – 8.59 (m, 1H), 8.15 (d, J = 7.8 Hz, 2H), 7.38 – 7.37 (m, 1H), 7.13 (d, J = 7.8 Hz, 2H), 7.12 – 7.10 (m, 1H), 6.64 (dd, J = 4.8 Hz, 7.2 Hz, 1H), 2.48 (t, J = 7.8 Hz, 2H), 1.49 (dd, J = 7.8 Hz, 7.2 Hz, 2H), 1.23 (dt, J = 7.8 Hz, 7.2 Hz, 2H), 0.85 (t, J = 7.8 Hz, 3H). HRMS (ESI, MH^+): Calcd. For $\text{C}_{15}\text{H}_{18}\text{N}$: 212.1434. Found: 212.1439.

6d⁹: ^1H NMR (600 MHz, CDCl_3) δ 7.49 (d, J = 8.4 Hz, 2H), 7.47 (d, J = 7.8 Hz, 2H), 7.10 (d, J = 7.8 Hz, 2H), 7.07 (d, J = 8.4 Hz, 2H), 2.49 (q, J = 7.8 Hz, 2H), 2.16 (s, 3H), 1.12 (t, J = 7.8 Hz, 3H). ^{13}C NMR (150 MHz, C_6D_6) δ 143.1, 139.1, 138.9, 136.6, 129.8, 128.6, 127.3, 127.2, 28.8, 21.1, 15.9. MS (EI, m/z): 196 (M^+).

7a¹⁰: ¹H NMR (600 MHz, CDCl₃) δ 9.18 (s, 1H), 8.93 (s, 2H), 7.48 (d, *J* = 8.4 Hz, 2H), 7.31 (d, *J* = 8.4 Hz, 2H), 2.42 (s, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 157.1, 154.7, 139.1, 134.2, 131.3, 130.1, 126.8, 21.2. MS (EI, *m/z*): 170 (M⁺).

7b: ¹H NMR (600 MHz, CDCl₃) δ 9.27 (s, 1H), 8.69 (s, 2H), 7.02 (d, *J* = 8.4 Hz, 2H), 6.94 (d, *J* = 8.4 Hz, 2H), 2.40 (q, *J* = 7.2 Hz, 2H), 1.06 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 157.9, 154.7, 144.9, 134.2, 132.2, 128.9, 127.1, 28.8, 15.6. HRMS (ESI, MH⁺): Calcd. For C₁₂H₁₃N₂: 185.1073. Found: 185.1079.

7c: ¹H NMR (600 MHz, C₆D₆) δ 9.27 (s, 1H), 8.69 (s, 2H), 7.03 (d, *J* = 7.8 Hz, 2H), 6.95 (d, *J* = 7.8 Hz, 2H), 2.38 (t, *J* = 7.2 Hz, 2H), 1.48 (dd, *J* = 7.8 Hz, 7.2 Hz, 2H), 0.83 (t, *J* = 7.8 Hz, 3H). ¹³C NMR (150 MHz, C₆D₆) δ 158.0, 154.7, 143.4, 134.1, 132.2, 129.5, 127.0, 37.8, 24.7, 13.8. HRMS (ESI, MH⁺): Calcd. For C₁₃H₁₅N₂: 199.1230. Found: 199.1226.

(5) Competitive encapsulation studies

In the competitive encapsulations in Table 1 and Table 2, the concentration of **1** was 4 mM, and that of each guest was performed in 50 mM. The mixture of the host and guests was warmed and sonicated to dissolve the host-guest complexes, and placed at ambient temperature for at least 30 min. The ratios were based on the peaks of alkyl side-chains located at the upfield of ¹H NMR spectra, and determined on the integral values and/or weighing papers of the magnified peaks. All the ¹H NMR spectra of encapsulated guests and the pairwise competition of Table 1 and Table 2 are shown in from page 45S to 57S.

(6) Reference

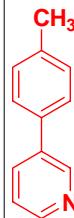
- 1) Tao, B.; Boykin, D. W. *J. Org. Chem.* **2004**, *69*, 4330 – 4335.
- 2) Hayashida, O.; Sebo, L.; Rebek, J., Jr. *J. Org. Chem.* **2002**, *67*, 8291 – 8298.
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- 4) Cherioux, F.; Attias, A.-J.; Maillette, H. *Advanced Functional Materials* **2002**, *12*, 203 – 208.
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- 9) Cho, C.-H.; Yun, H.-S.; Park, K. *J. Org. Chem.* **2003**, 68, 3017 -3025.
- 10) Kano, S.; Yuasa, Y.; Shibuya, S.; Hibino, S. *Heterocycles* **1982**, 19, 1079 – 1082.

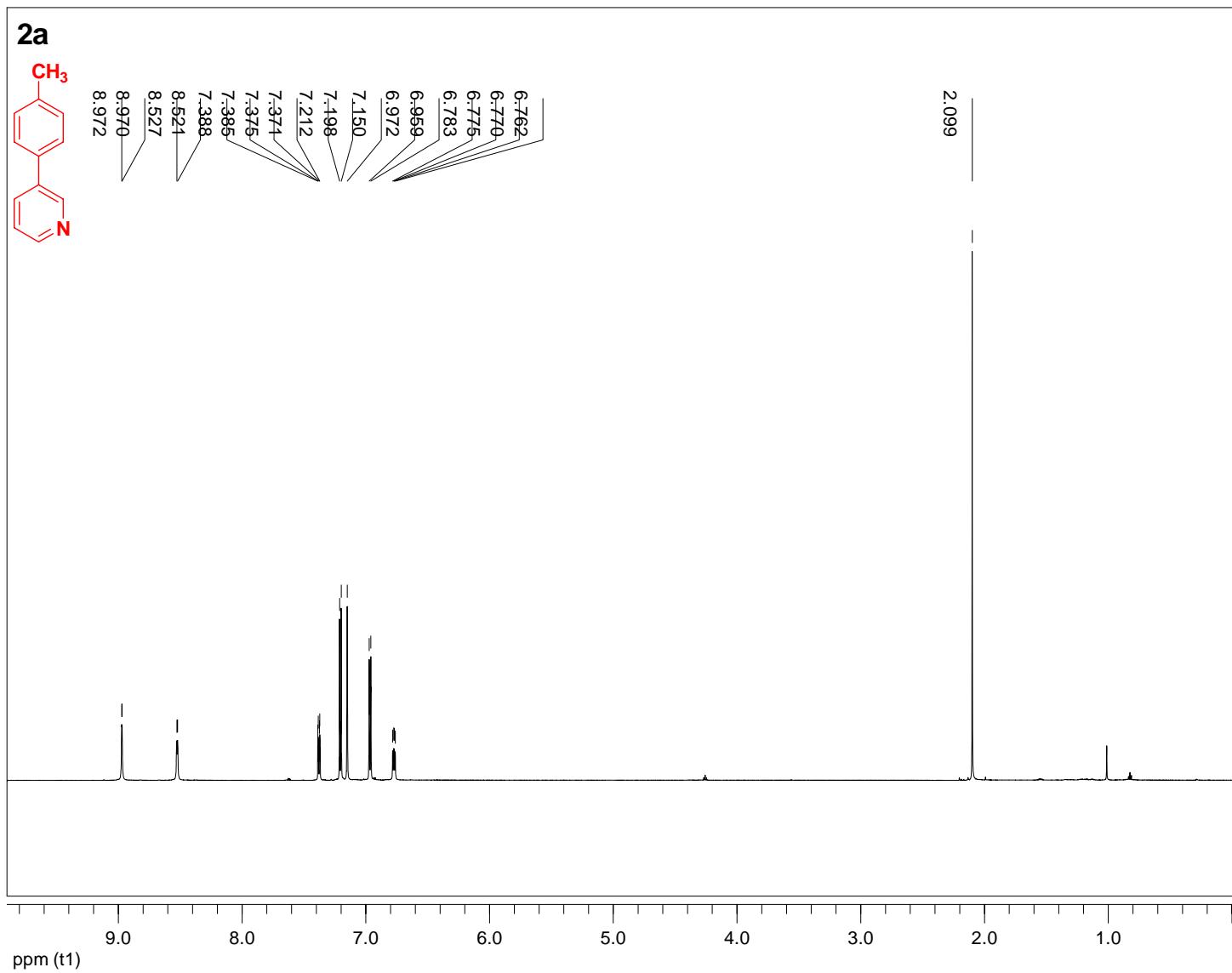
(7) ^1H and ^{13}C NMR spectra of biaryls 2a-g (page 8S – 21S), 3b-g (page 22S – 34S) 4 (page 35S – 36S), 6d (page 37S – 38S), 7a-c (page 39S – 44S), and all ^1H NMR spectra of encapsulated biaryls (page 45S – 52S) and competitive encapsulation studies (page 53S – 57S).

2a



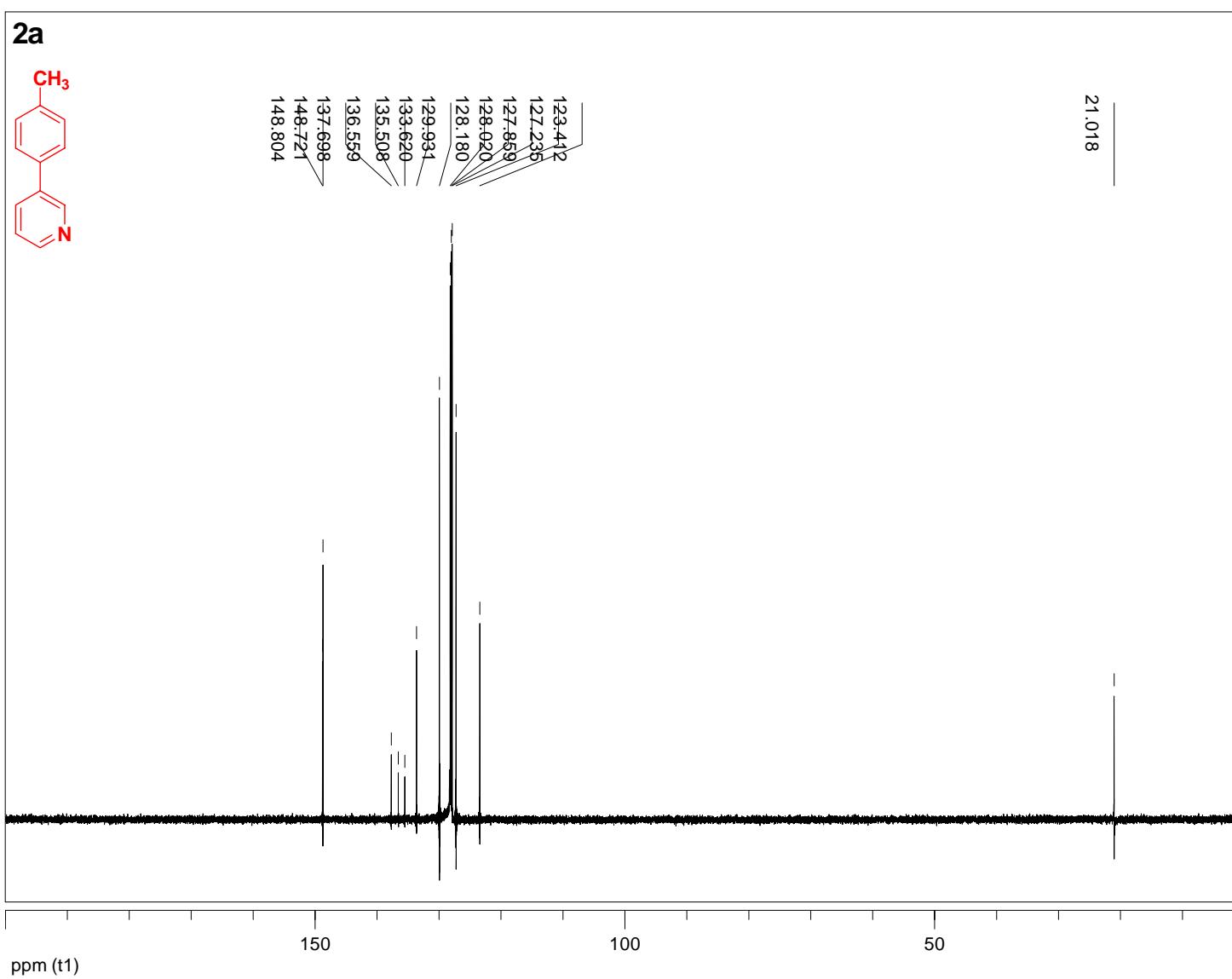
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6.776
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6.959
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7.150
7.198
7.212
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7.385
7.388
8.521
8.527
8.970
8.972

2.099



8S

2a



2b



8.898

7.390

7.376

7.372

7.263

7.249

7.150

6.995

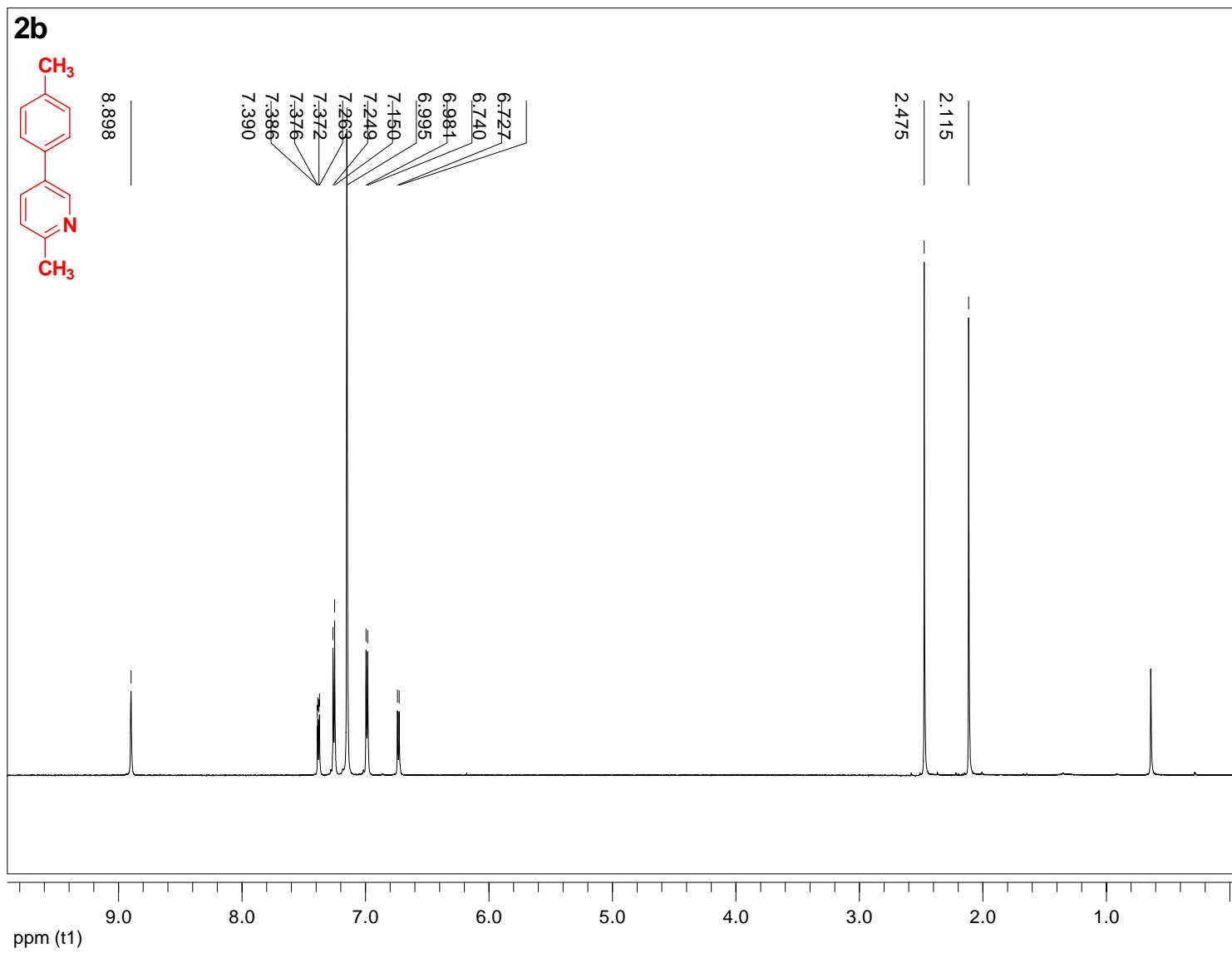
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6.740

6.727

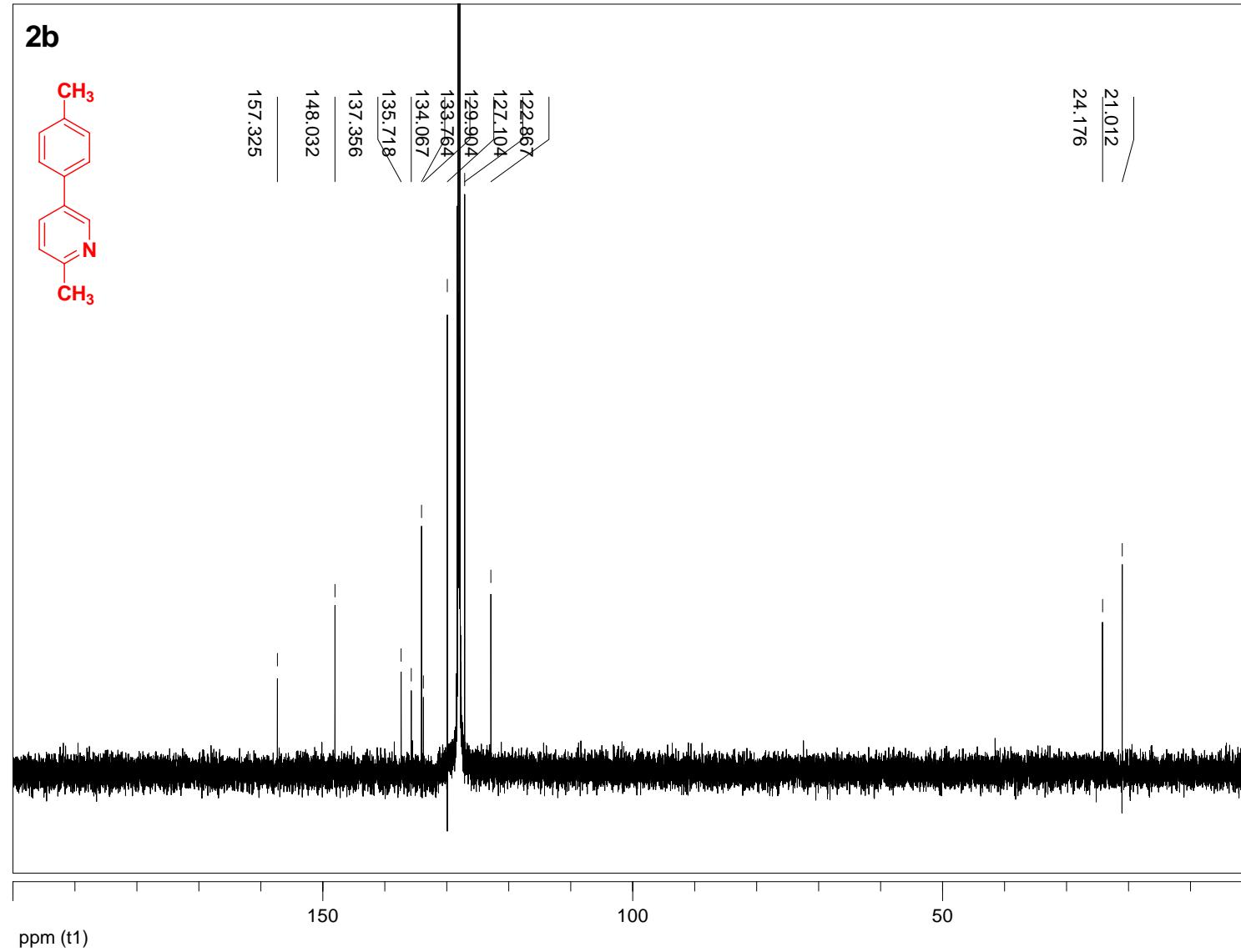
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2.115



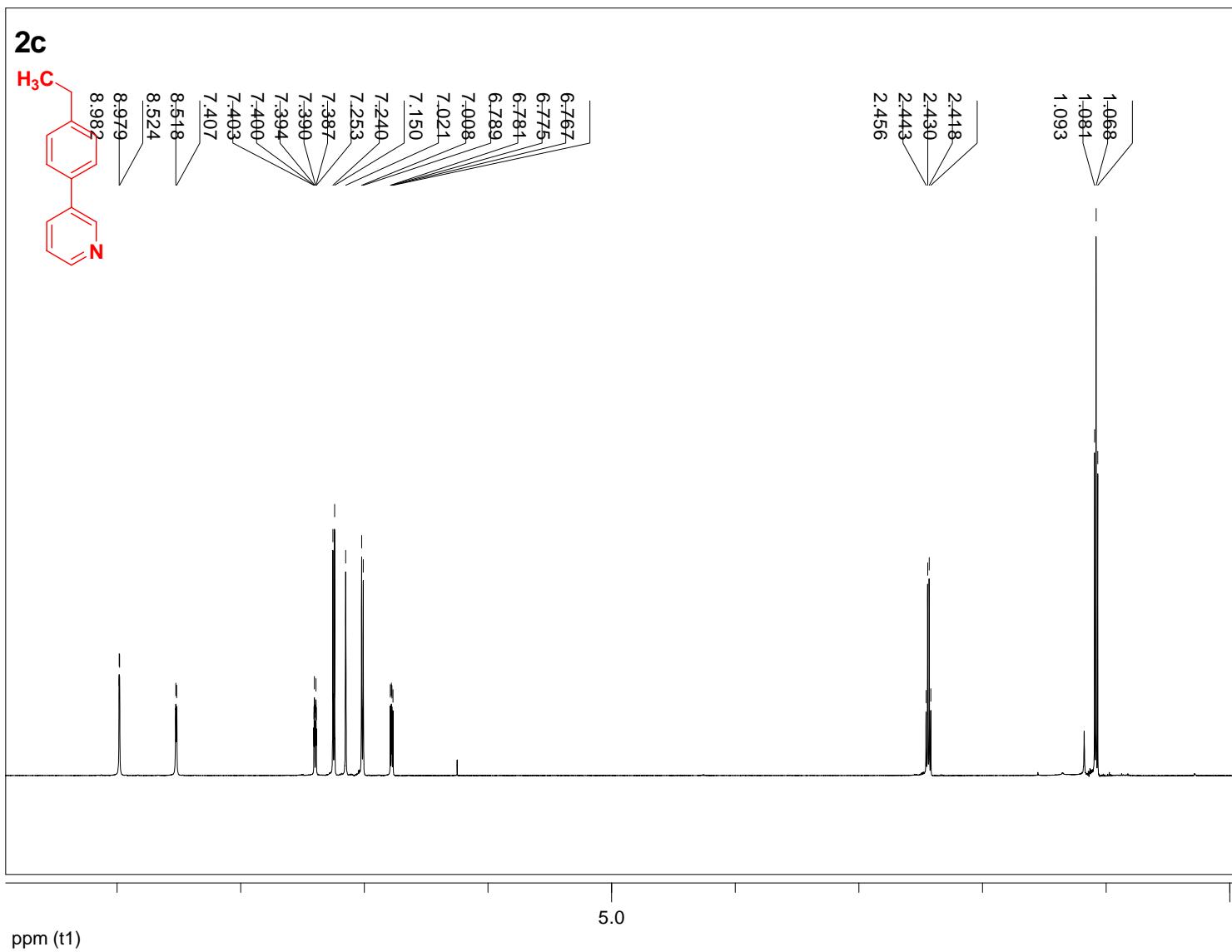
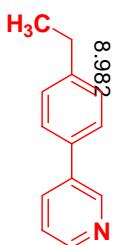
10S

2b



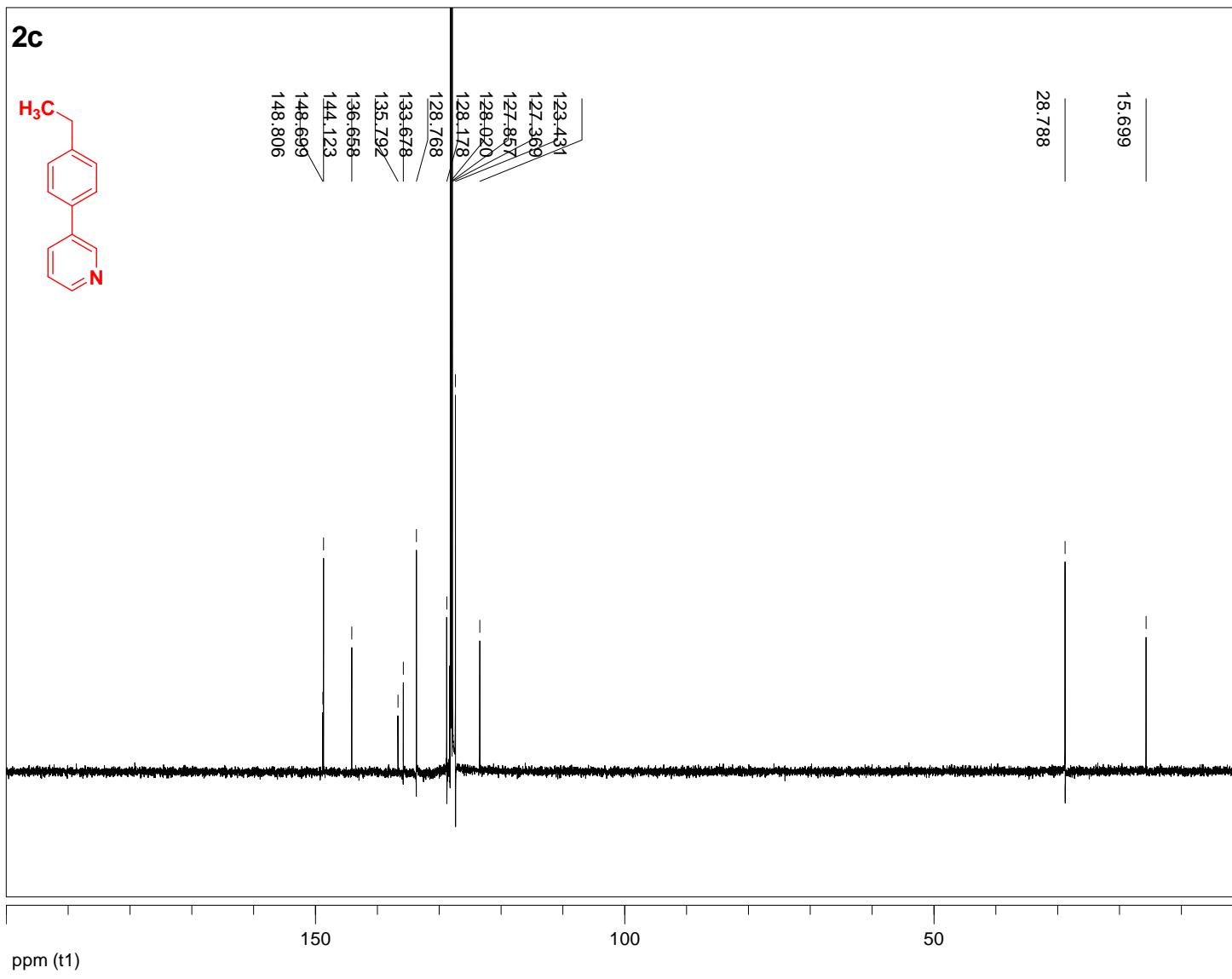
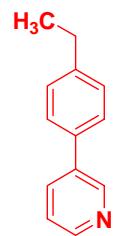
11S

2c



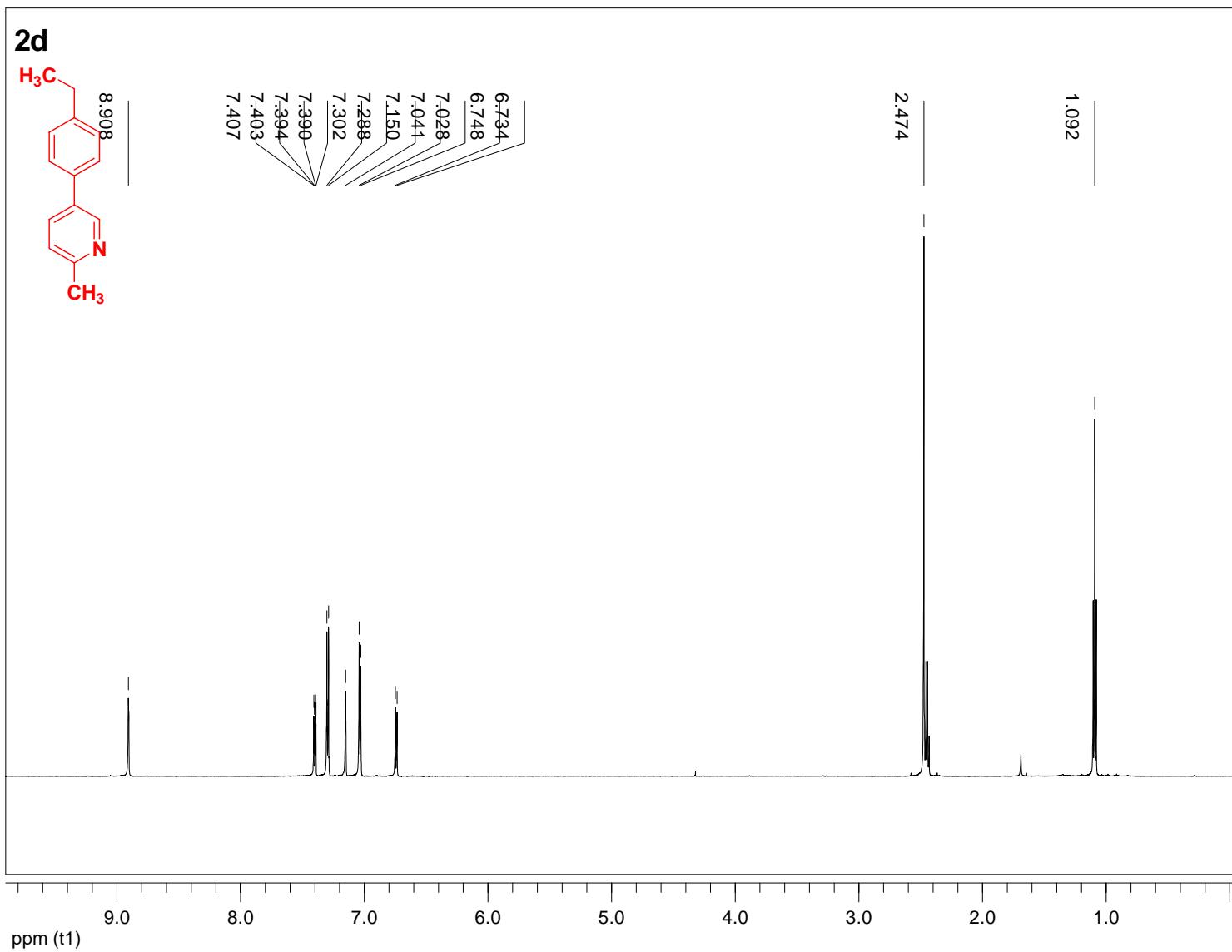
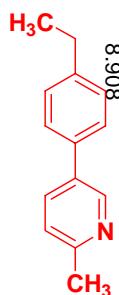
12S

2c



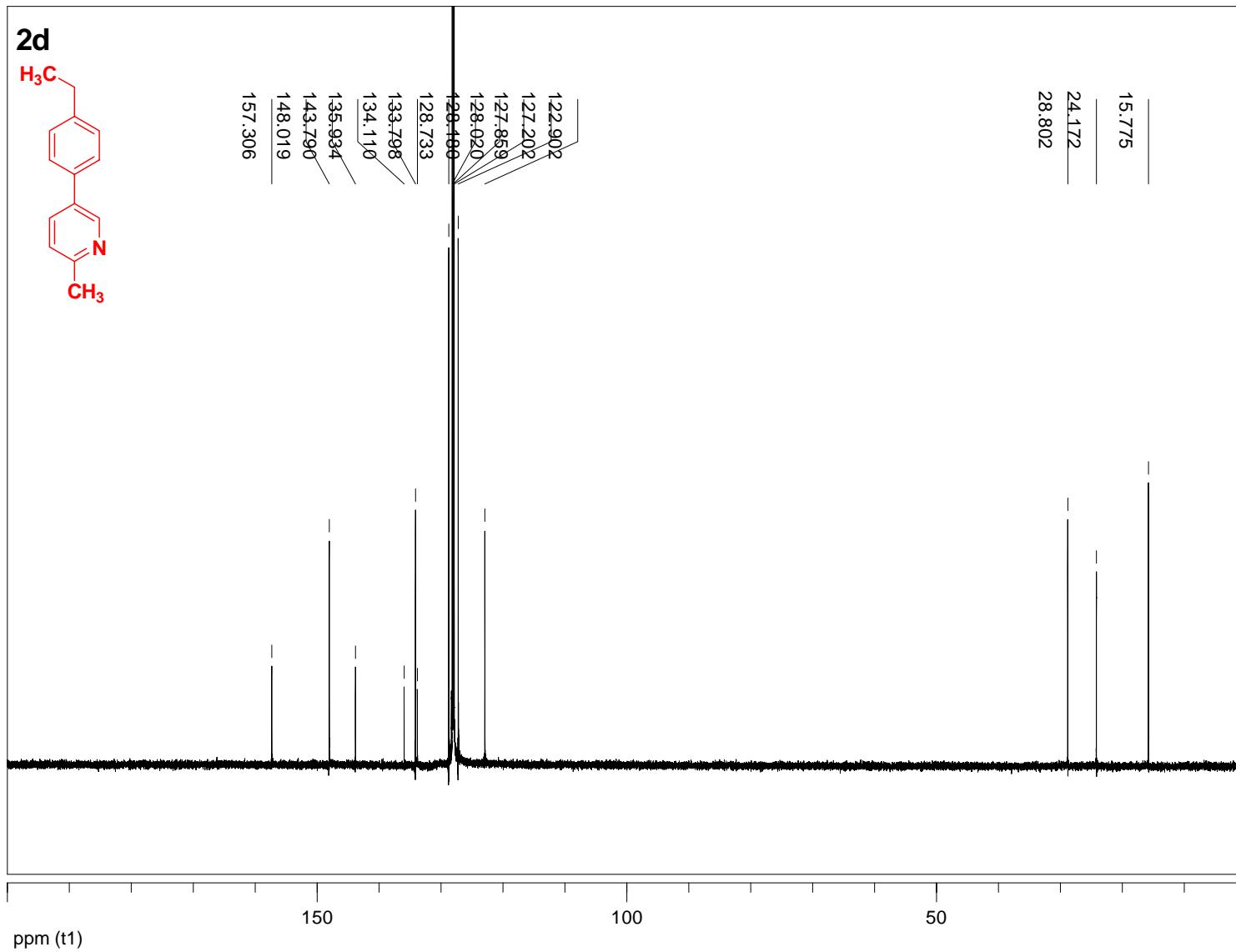
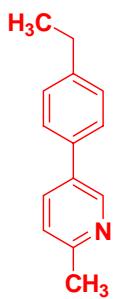
^{13}S

2d



14S

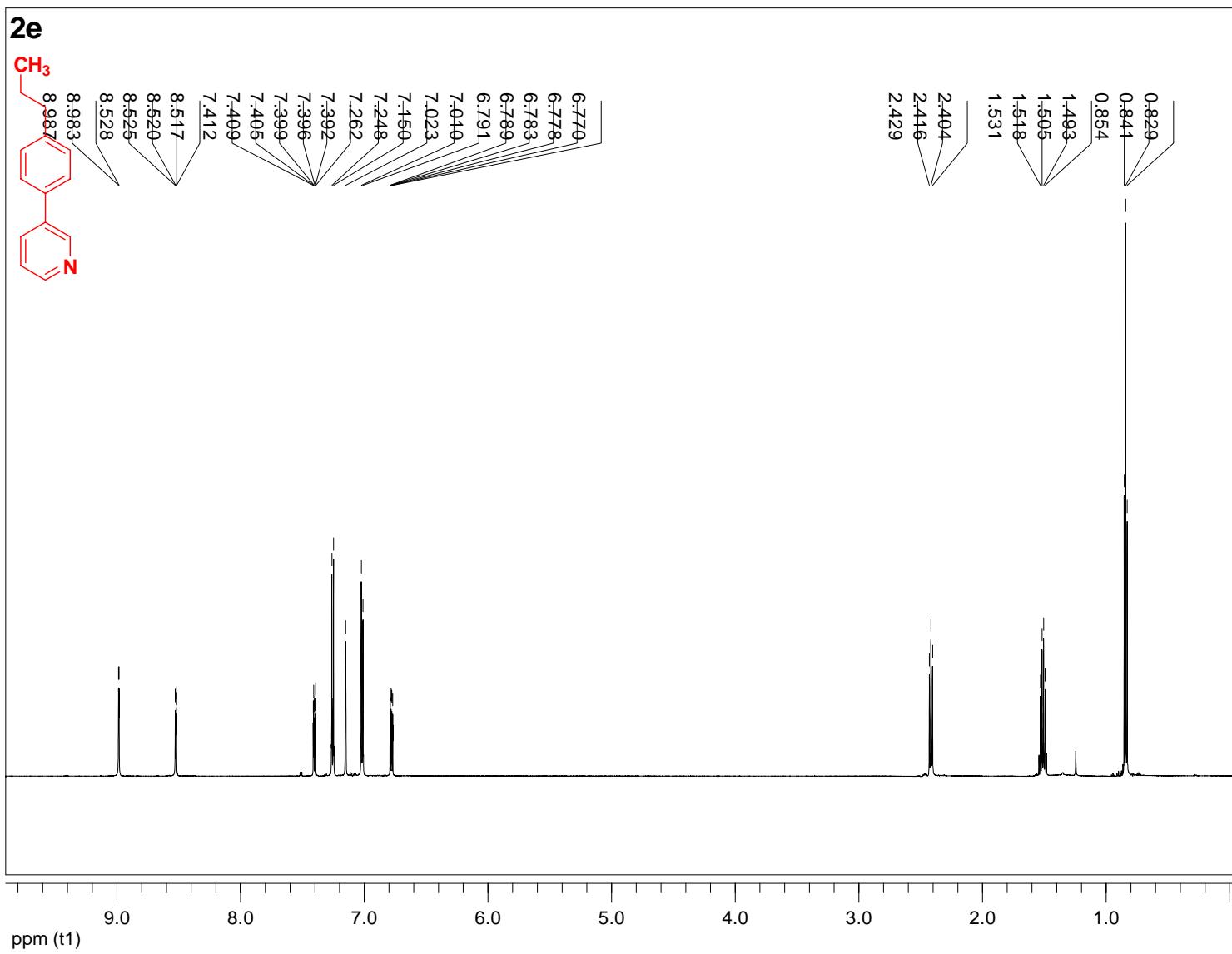
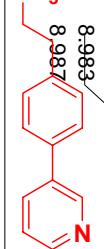
2d



15S

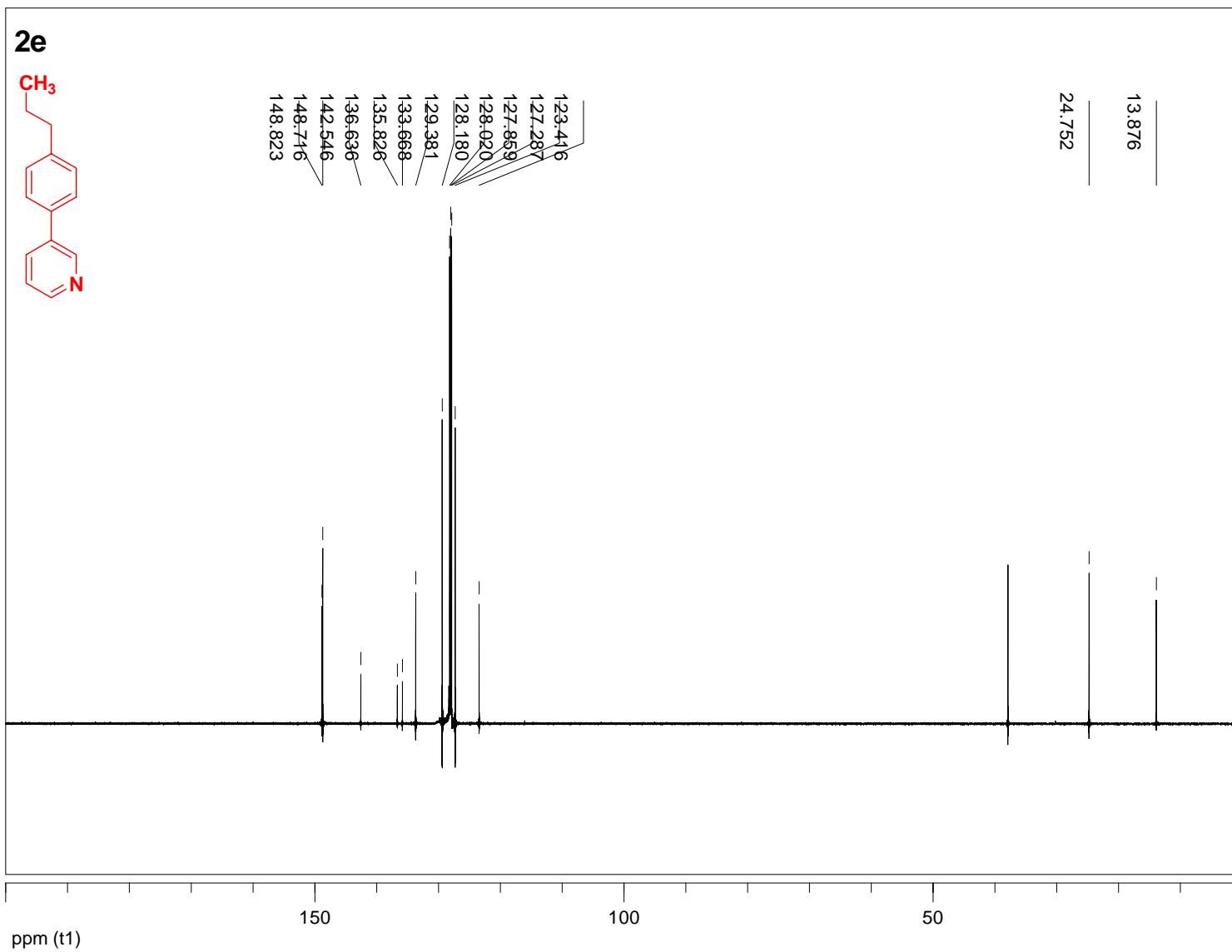
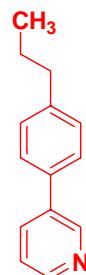
2e

CH₃



16S

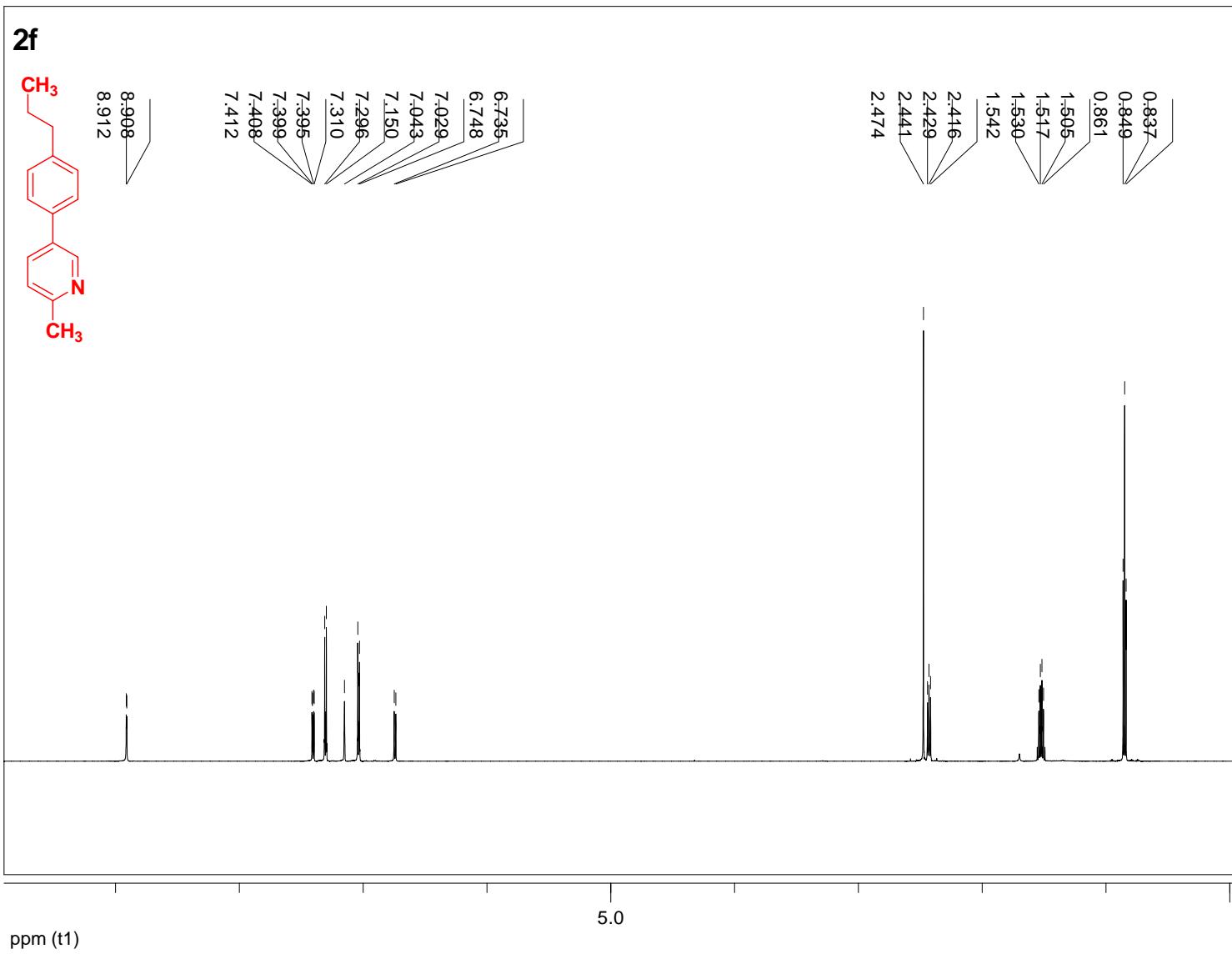
2e



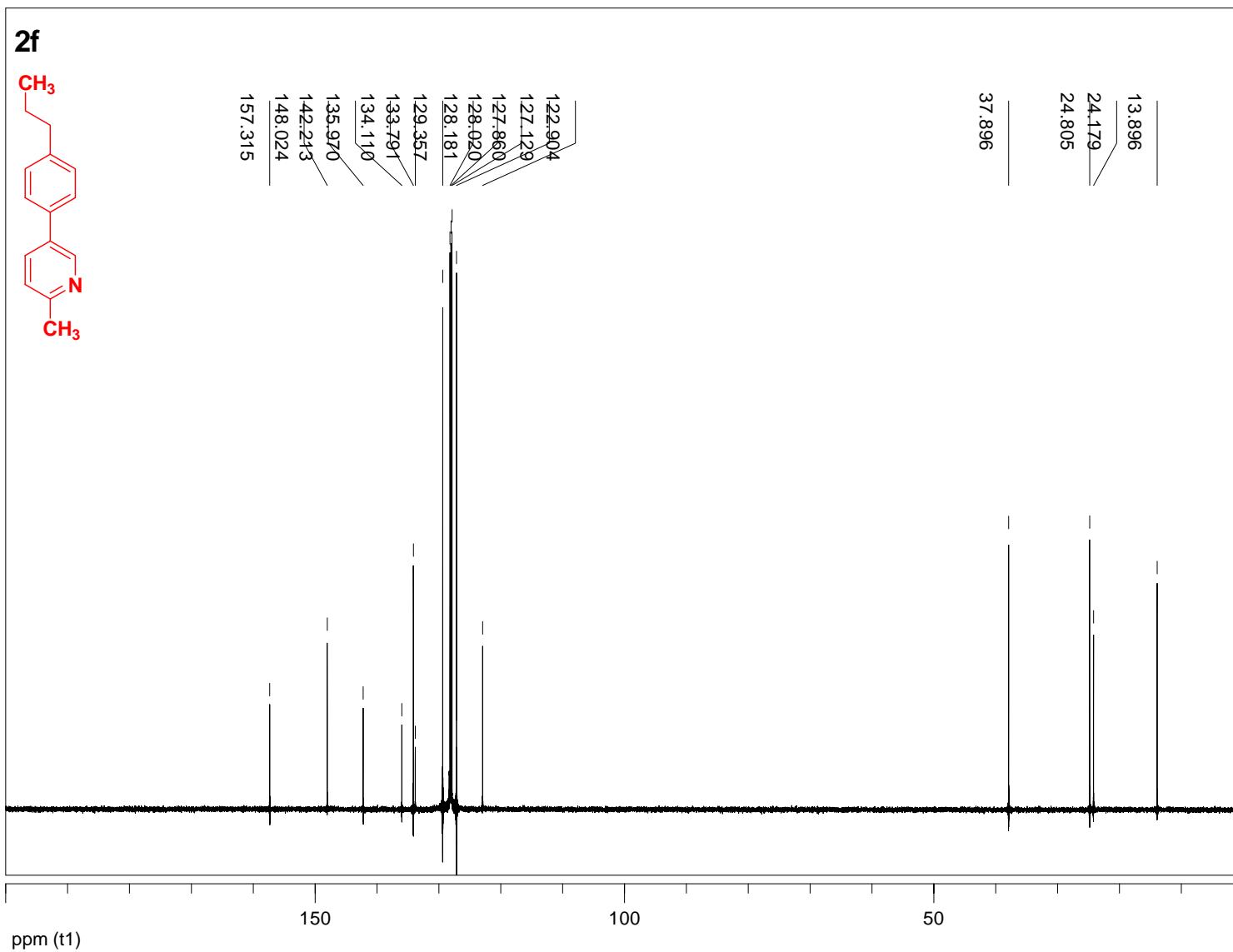
ppm (t1)

17S

2f

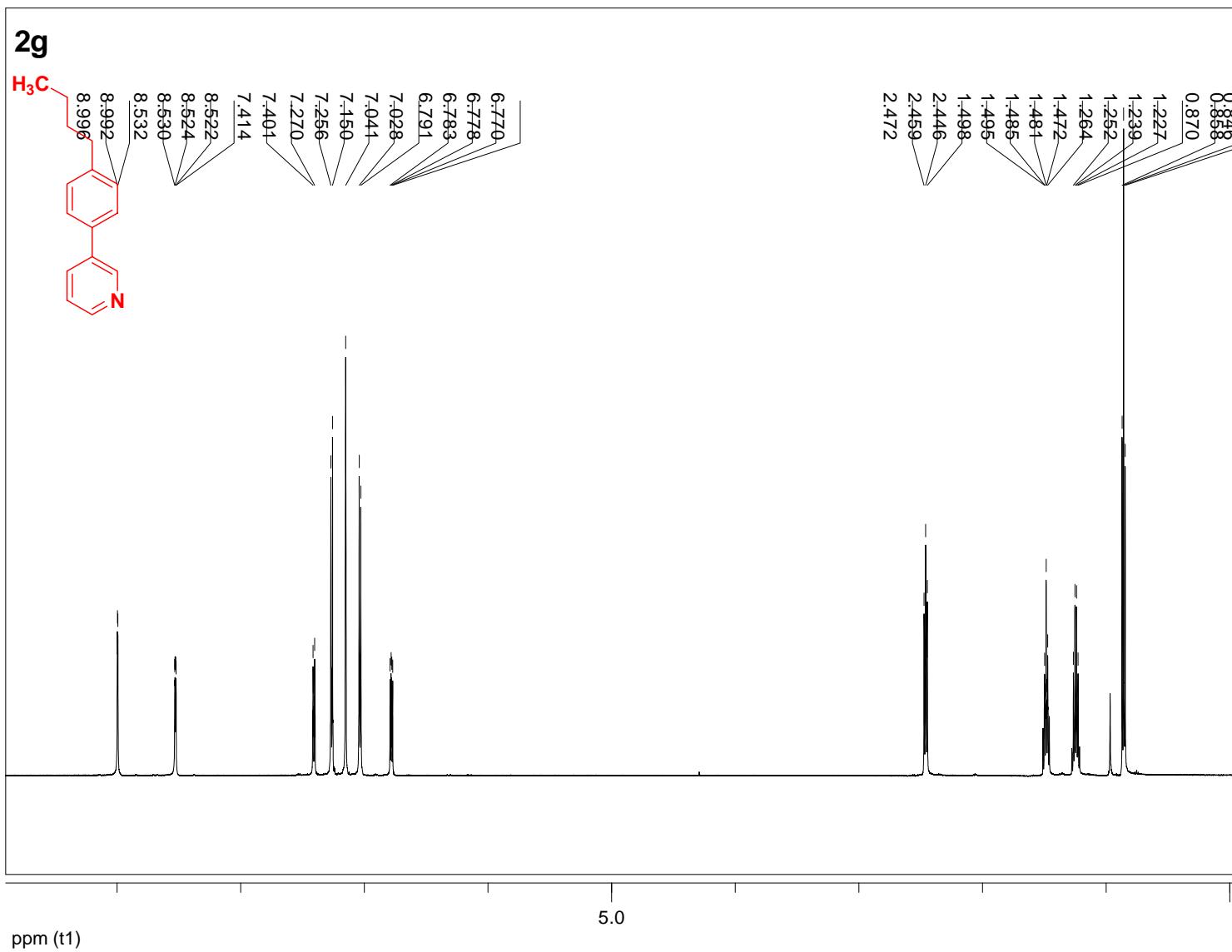
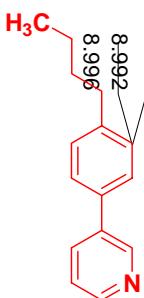


2f



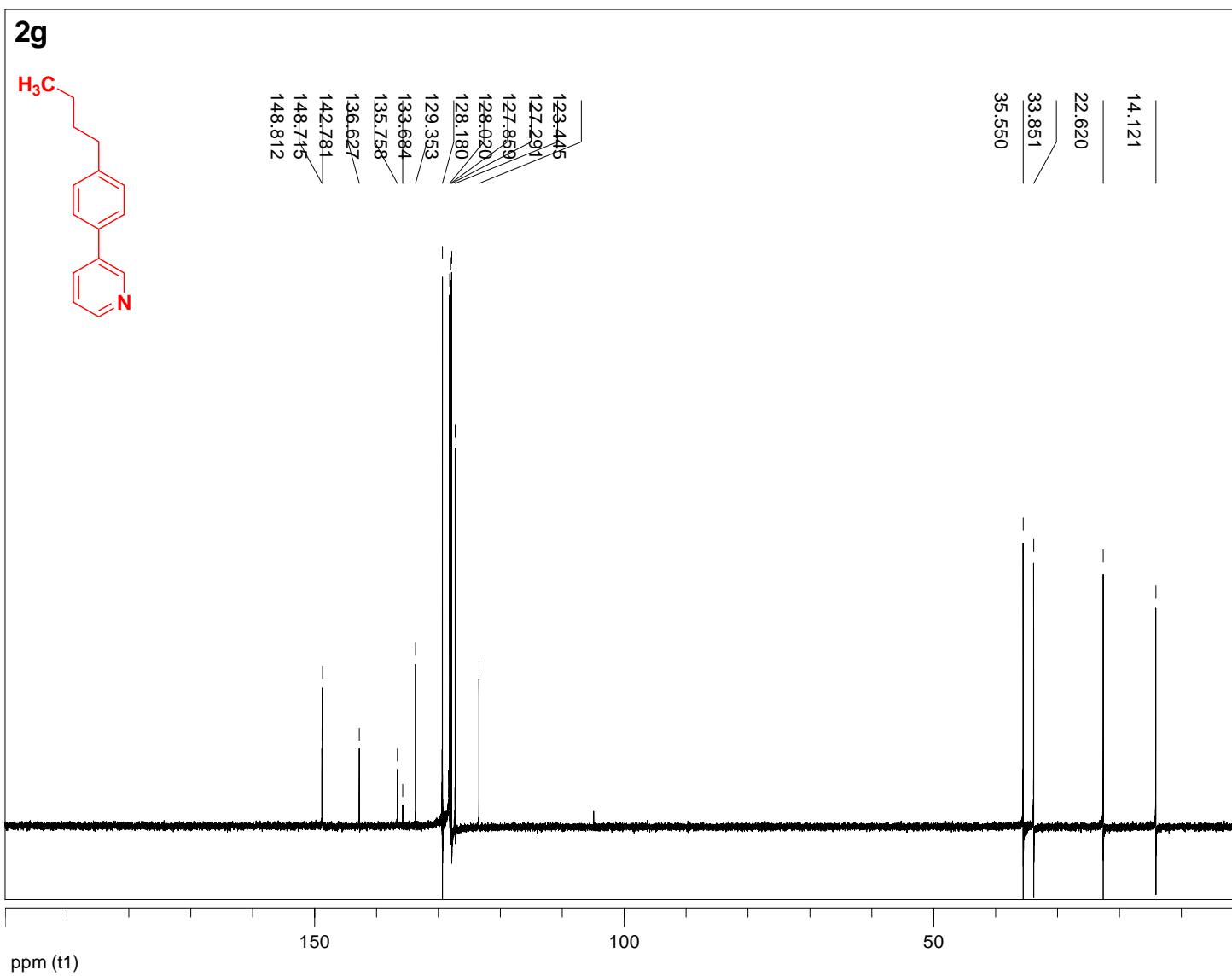
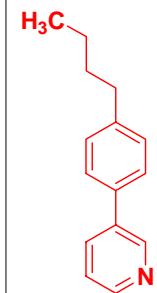
^{19}S

2g



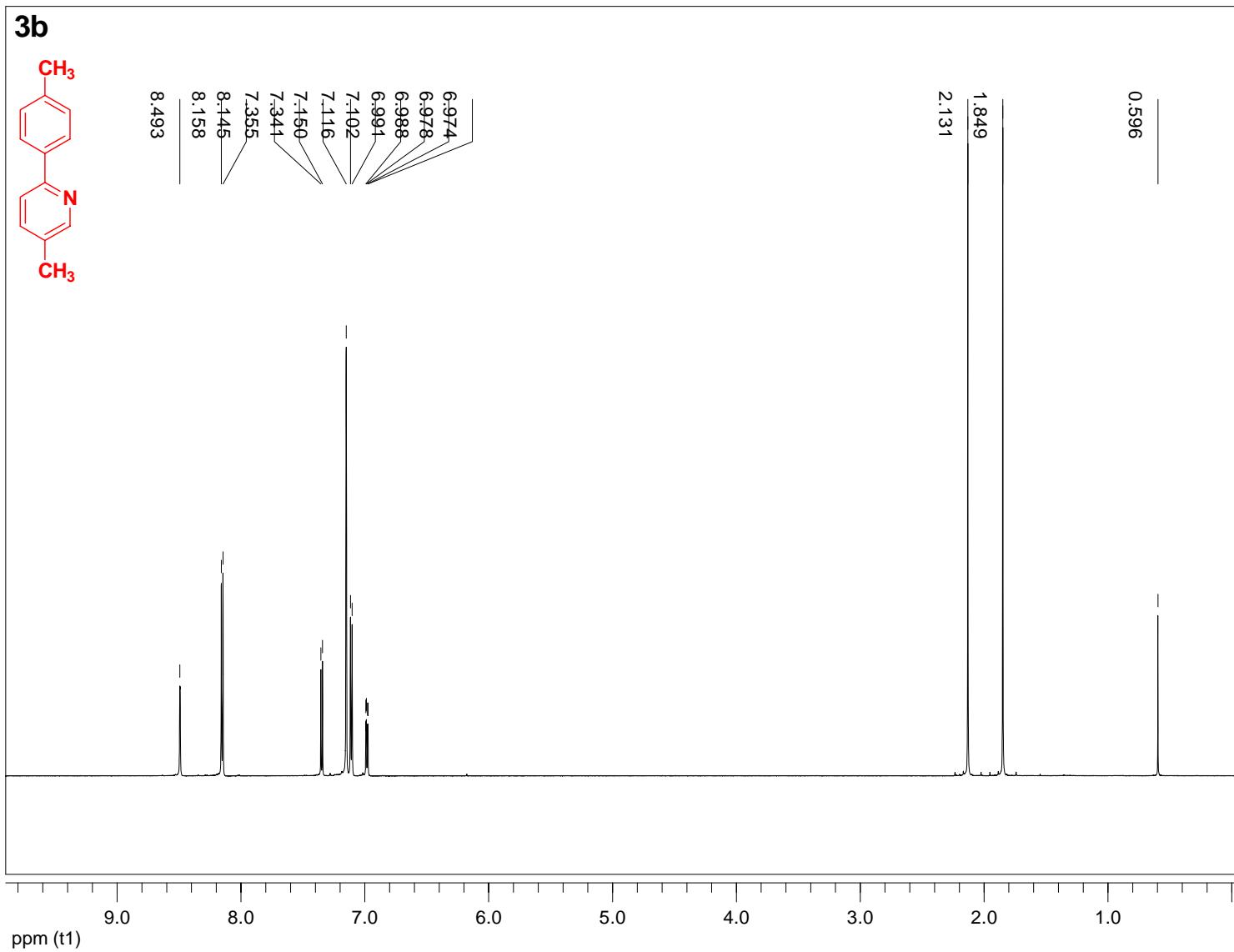
20S

2g



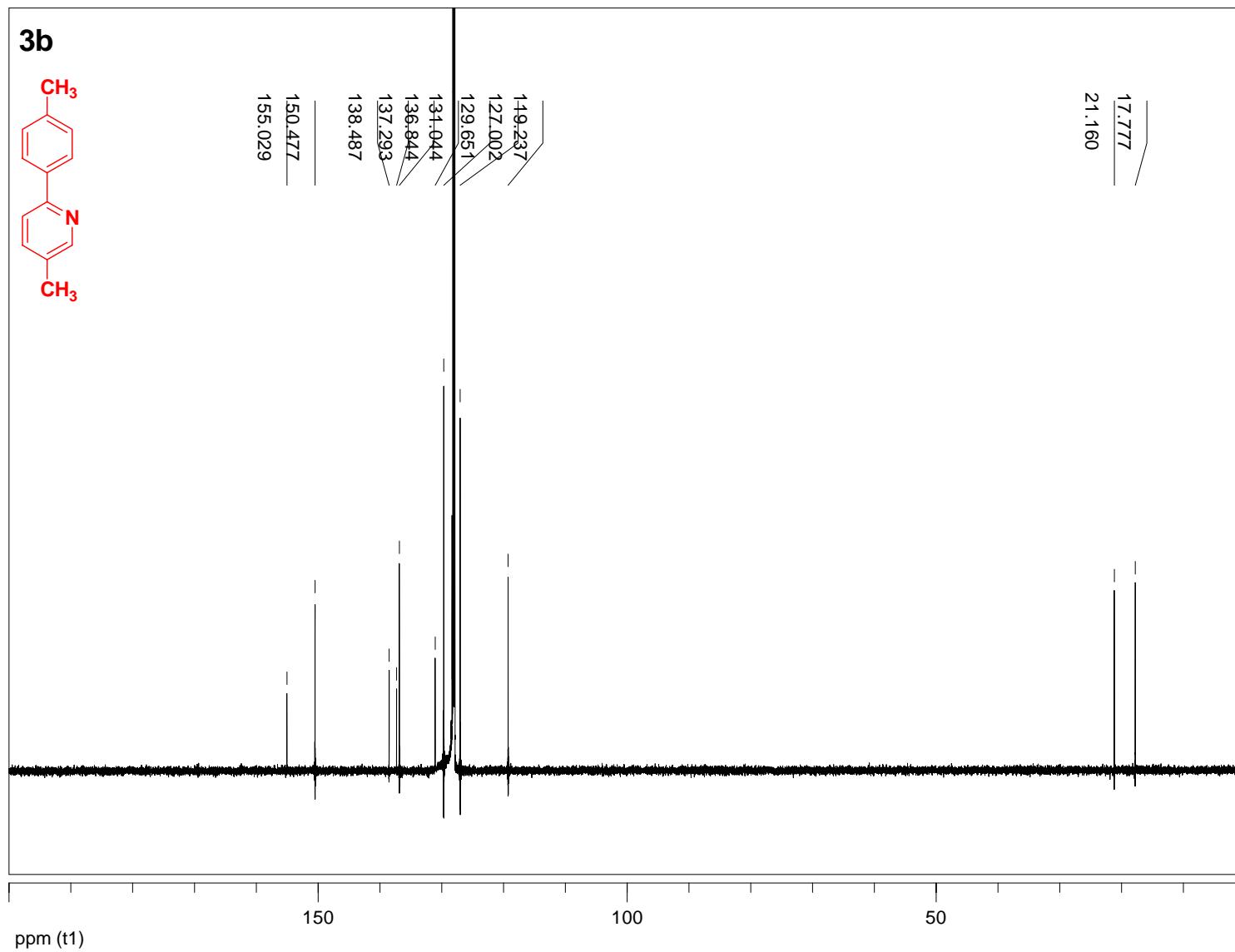
21S

3b



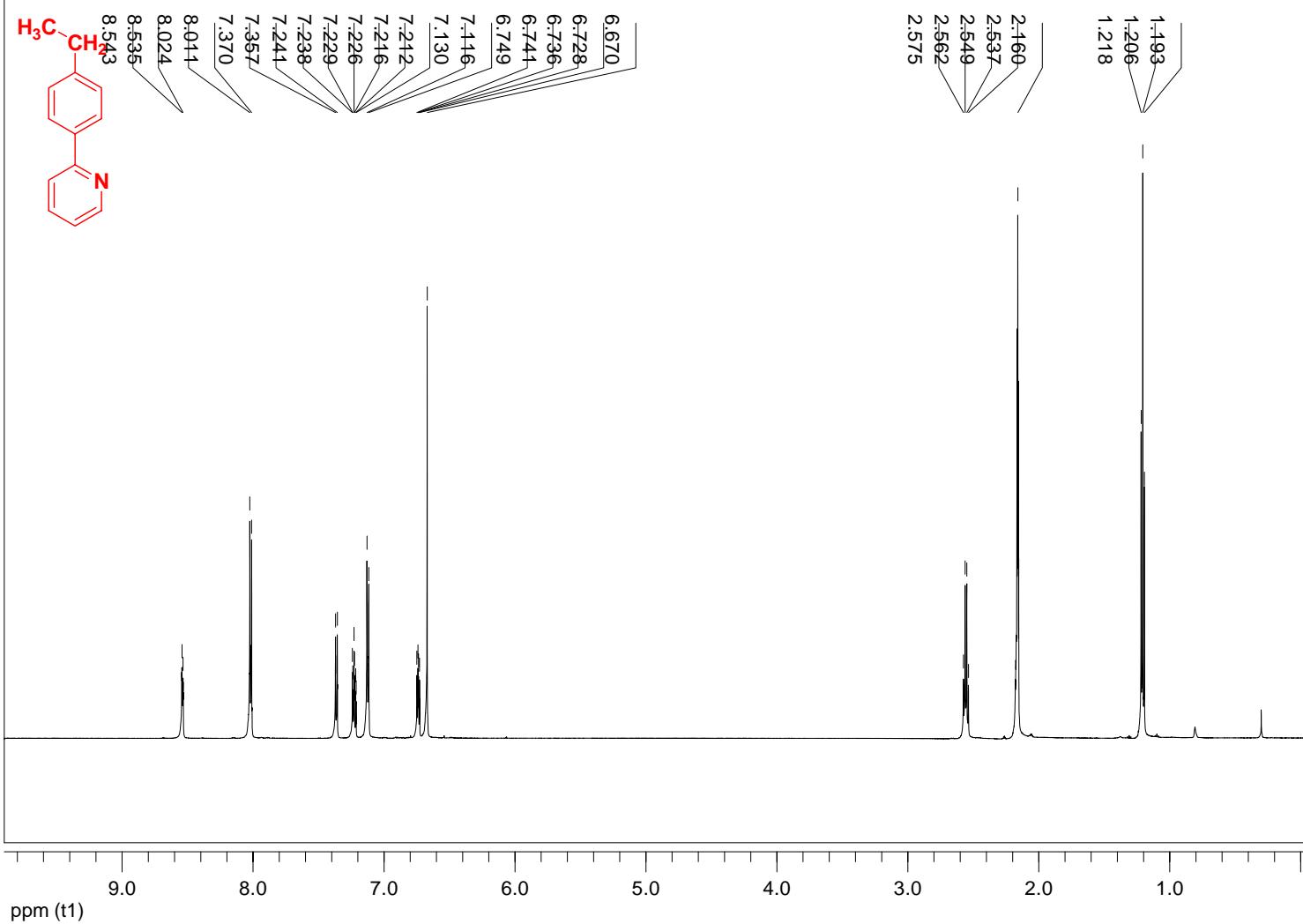
22S

3b



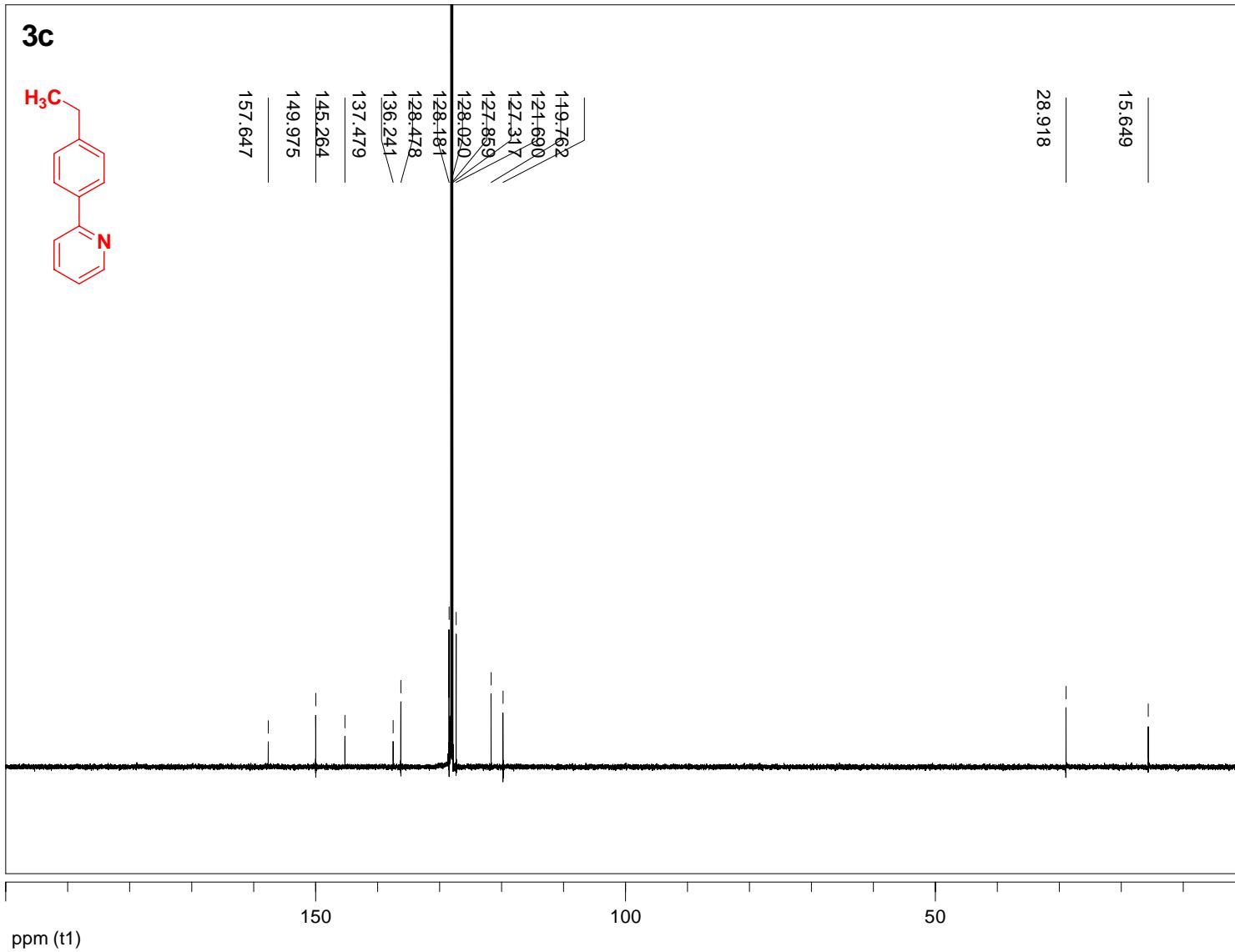
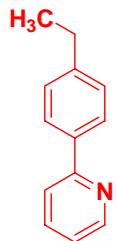
23S

3c



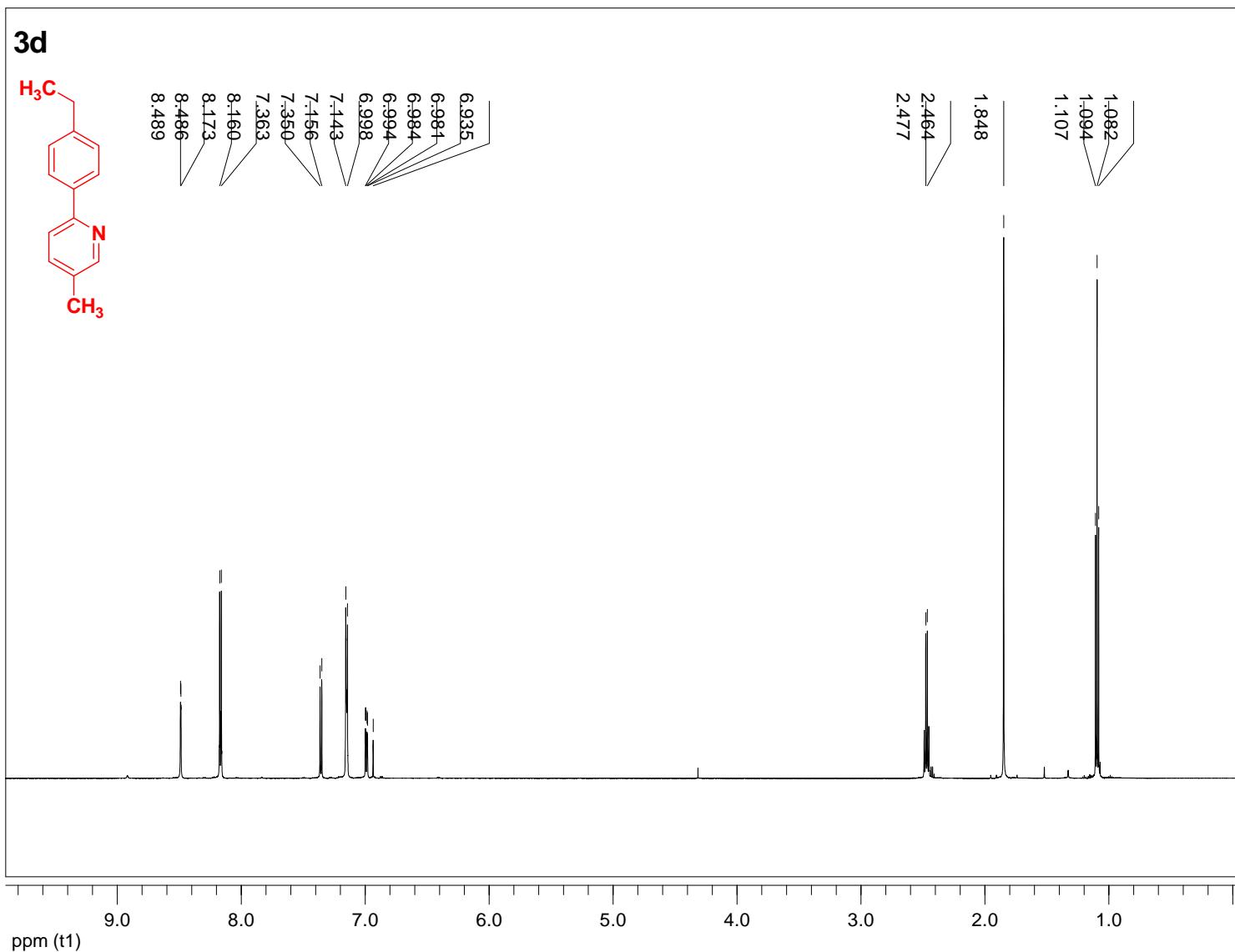
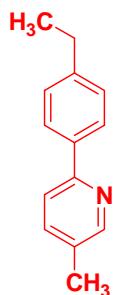
24S

3c



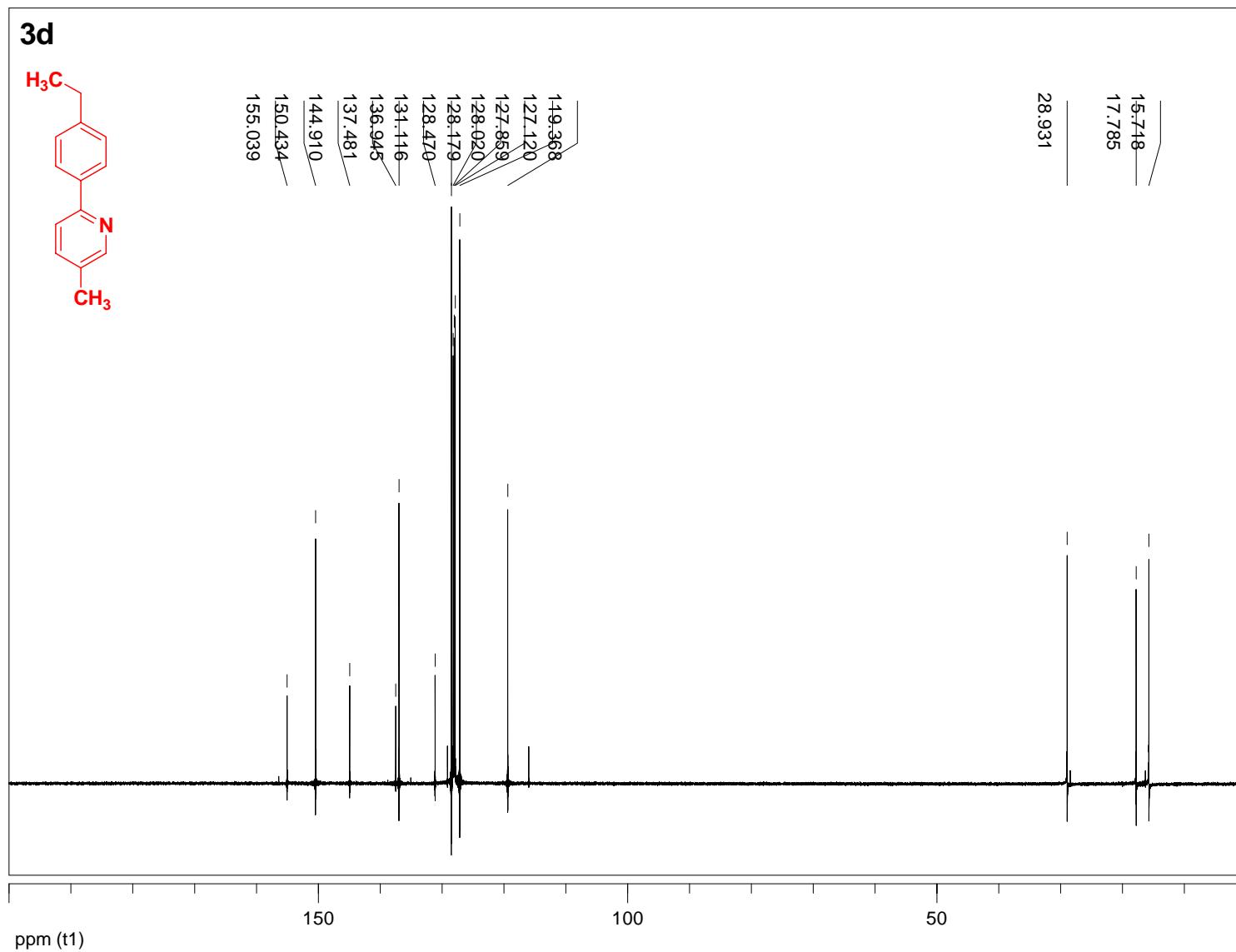
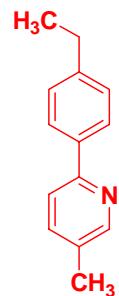
25S

3d



26S

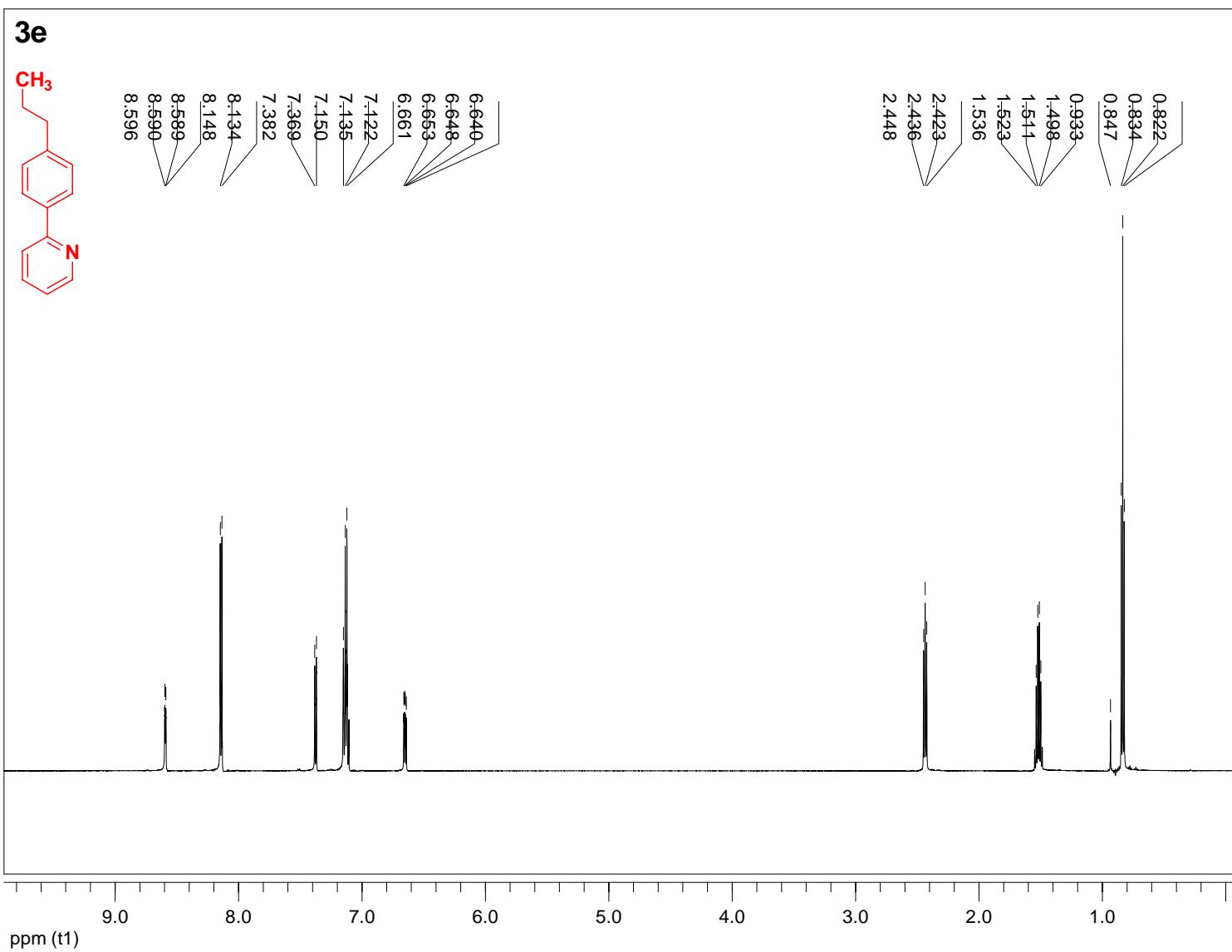
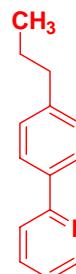
3d



ppm (t1)

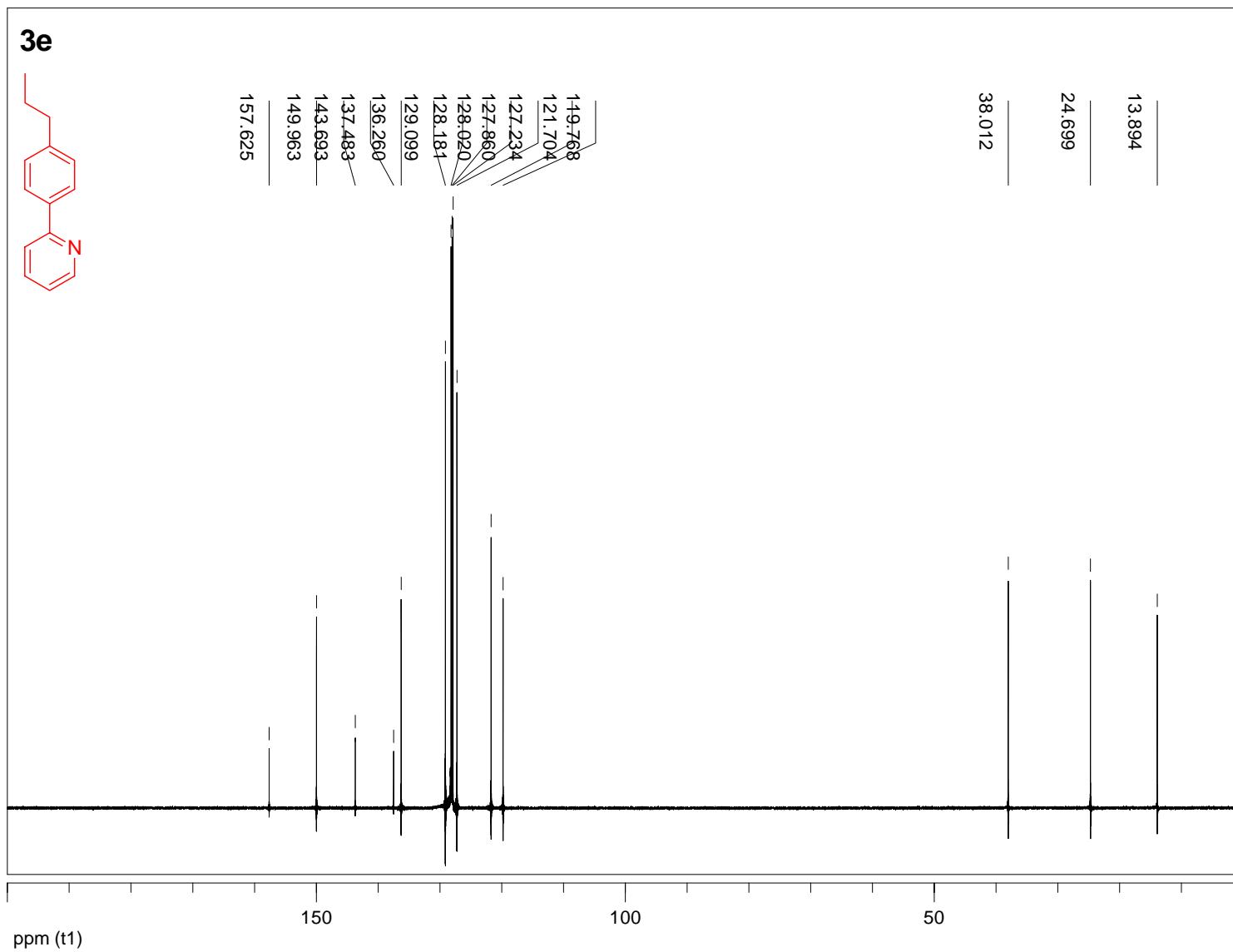
27S

3e



28S

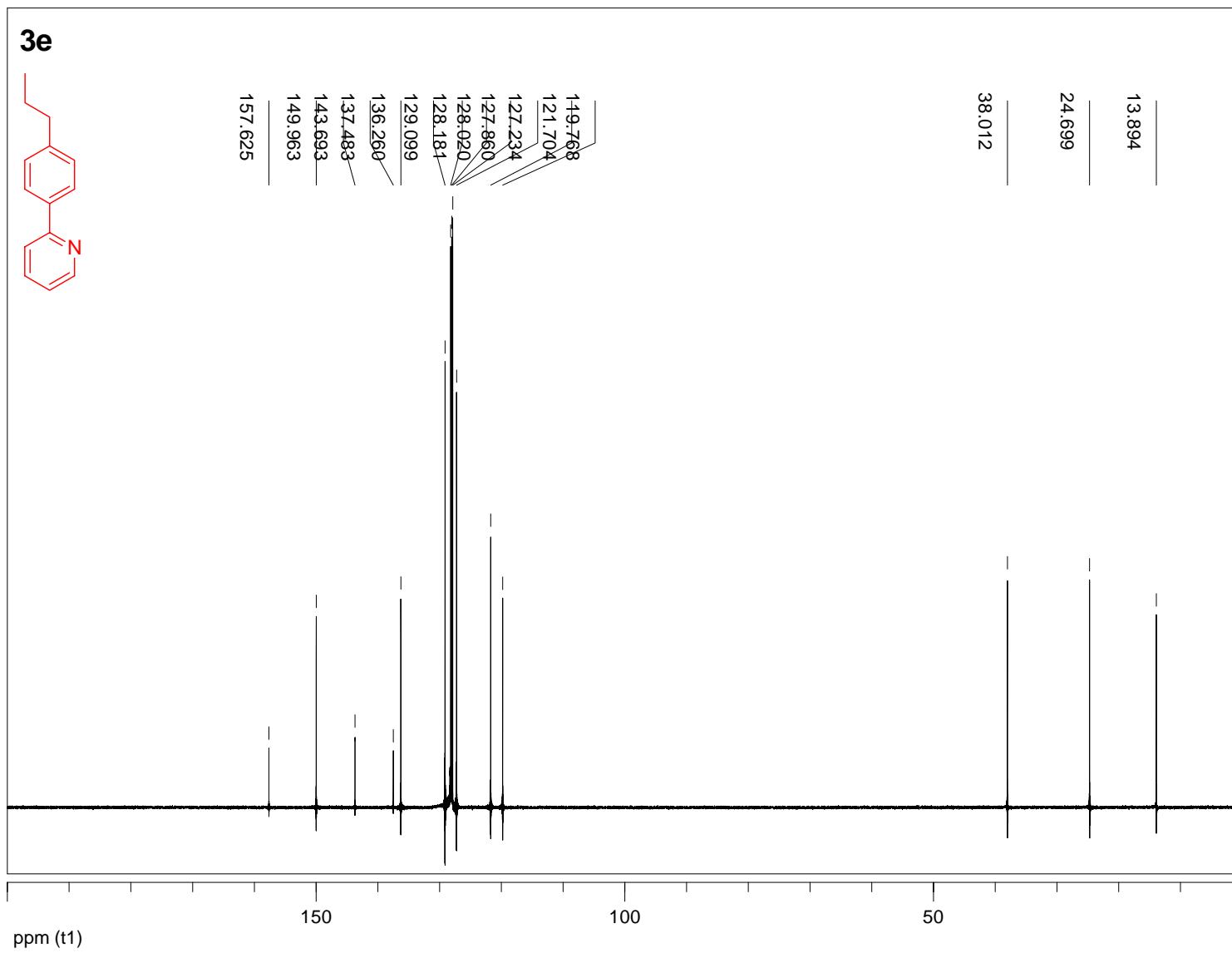
3e



ppm (t1)

^{29}S

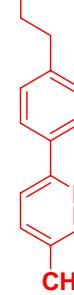
3e



30S

3f

CH₃



8.502

8.498

8.481

8.195

7.978

7.979

6.970

6.982

6.992

6.991

6.995

6.996

7.450

7.360

7.374

7.460

2.464

2.439

2.451

1.849

1.551

1.538

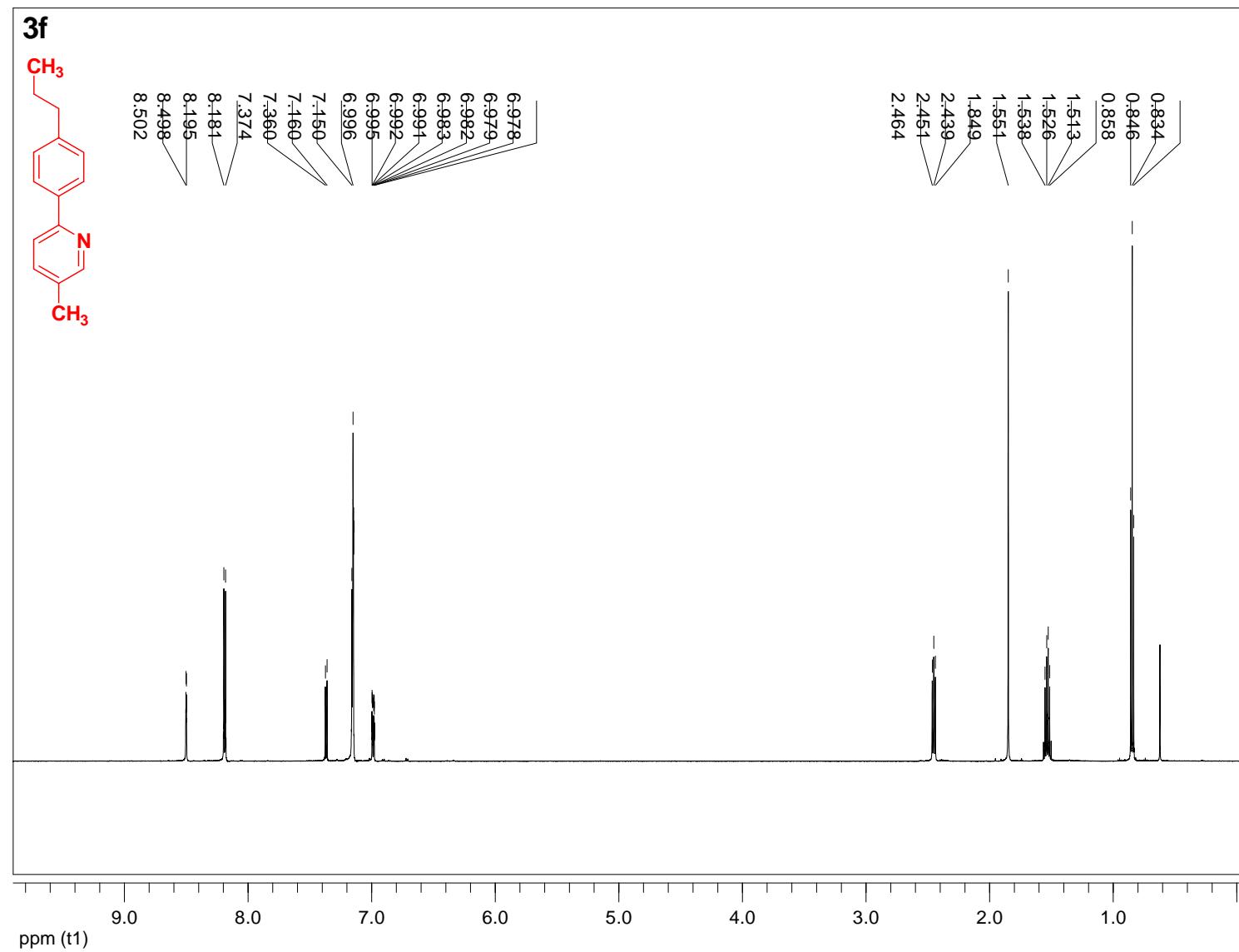
1.526

1.513

0.846

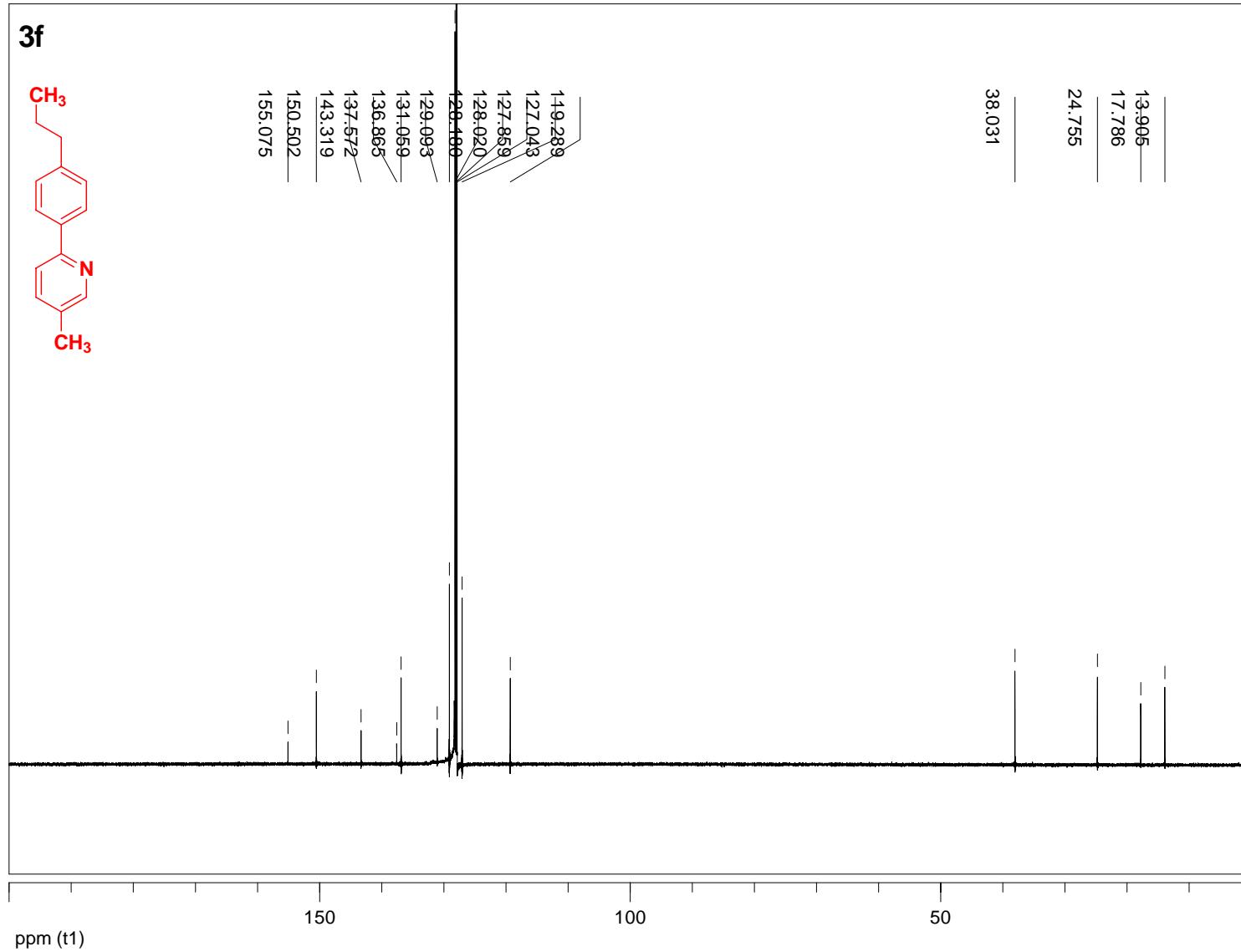
0.834

0.858



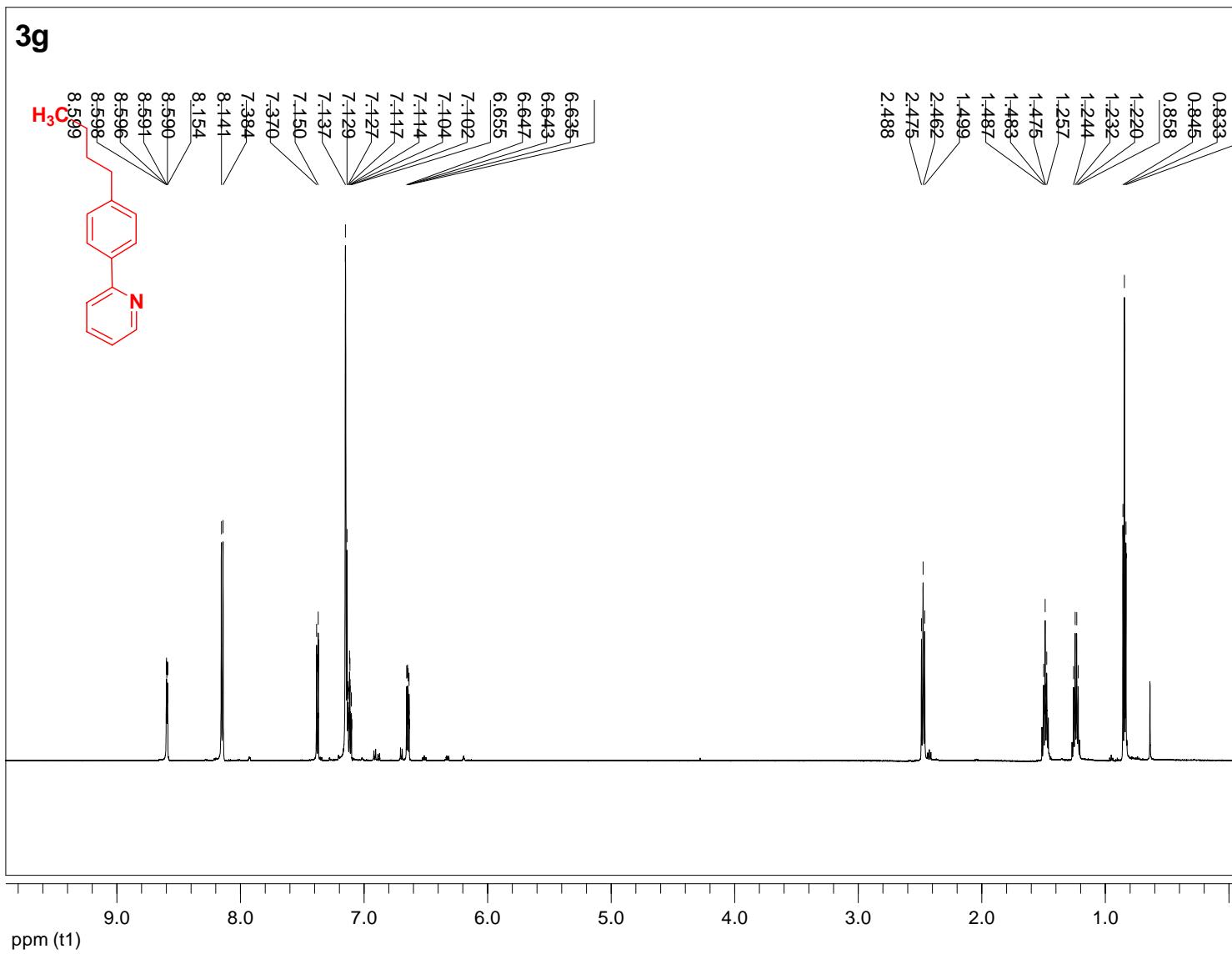
31S

3f

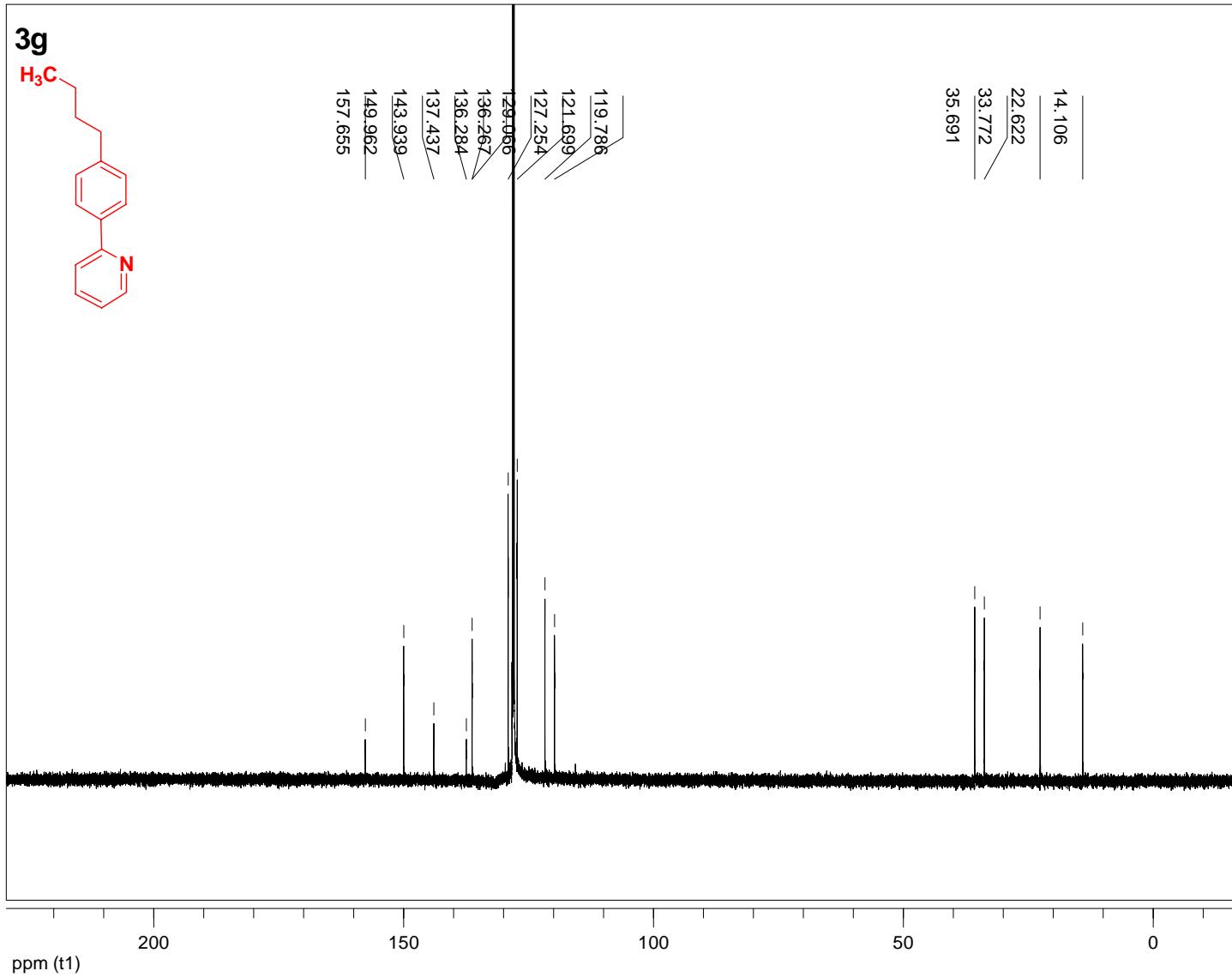


32S

3g



³³S



³⁴S

4

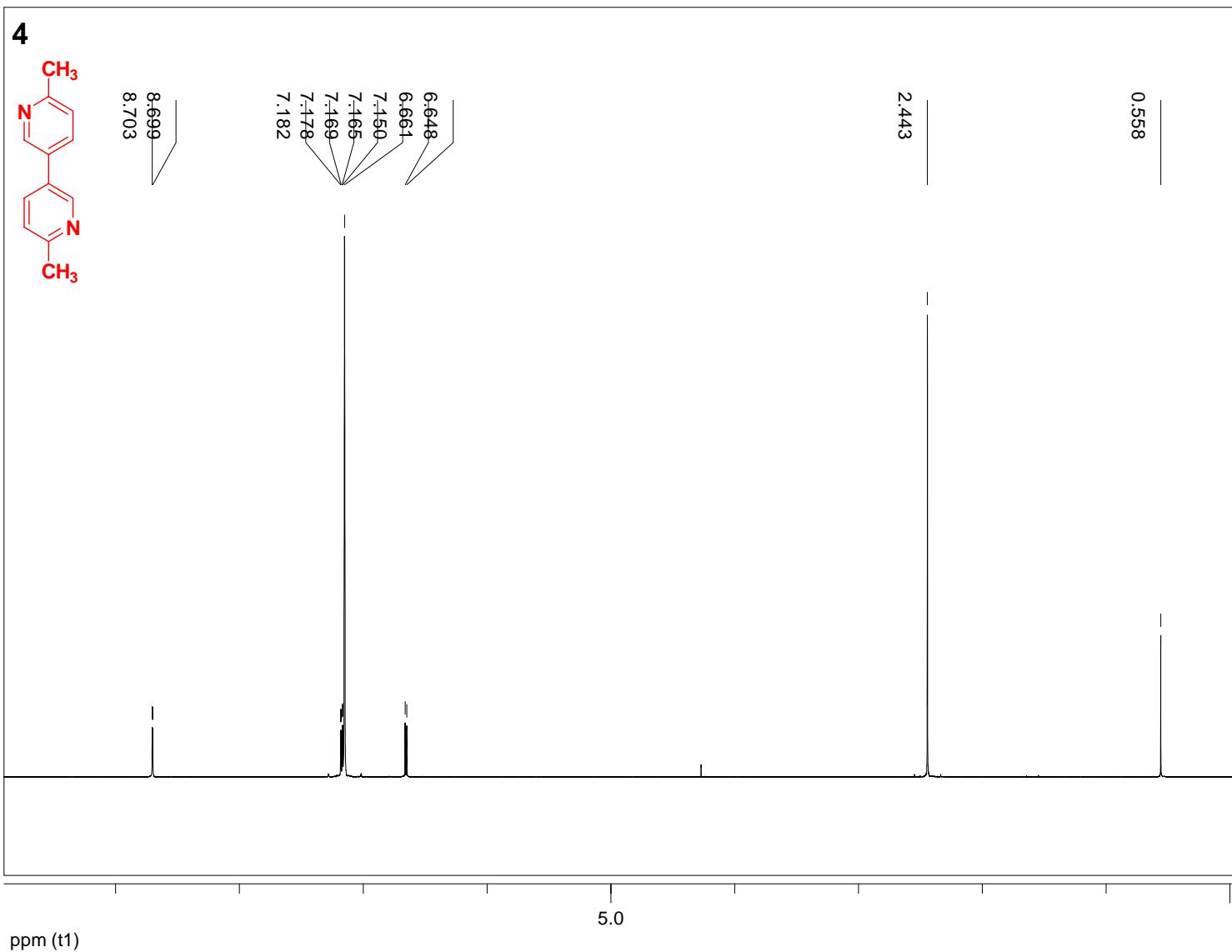


8.699
8.703

7.182
7.478
7.469
7.465
7.460
6.661
6.648

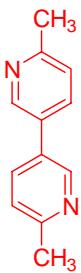
2.443

0.558



35S

4

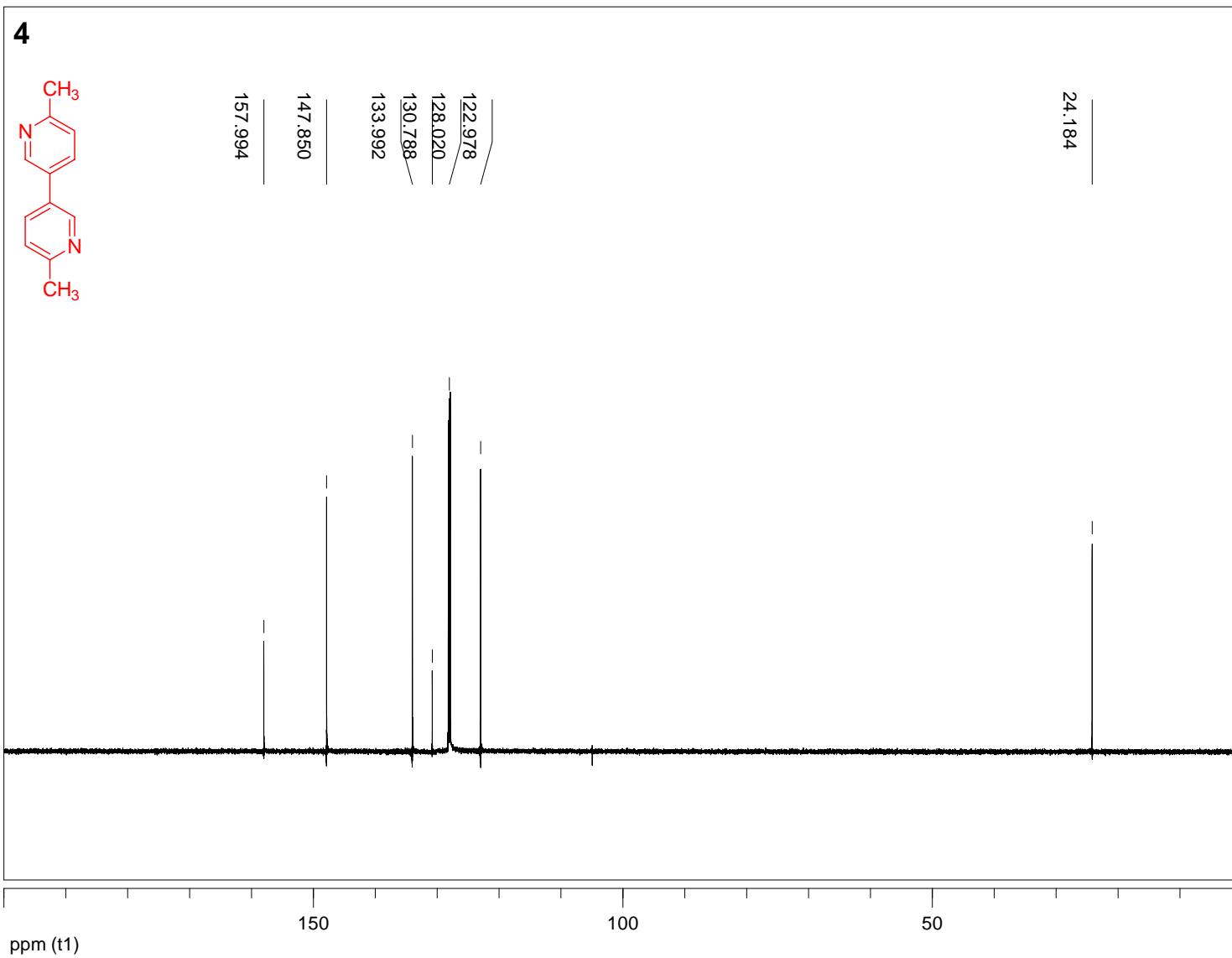


122.978
128.020
130.788
133.992

147.850

157.994

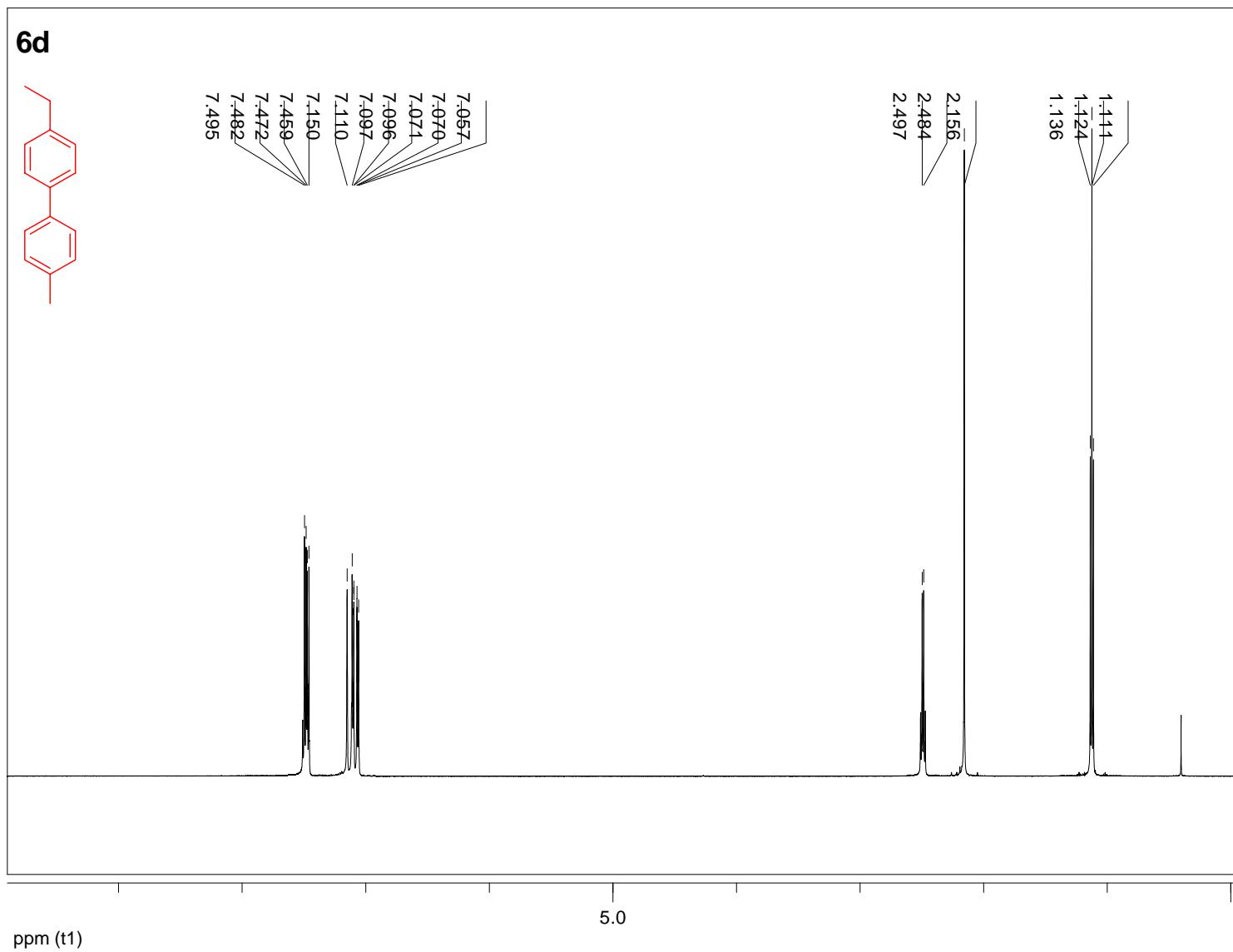
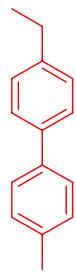
24.184



ppm (t1)

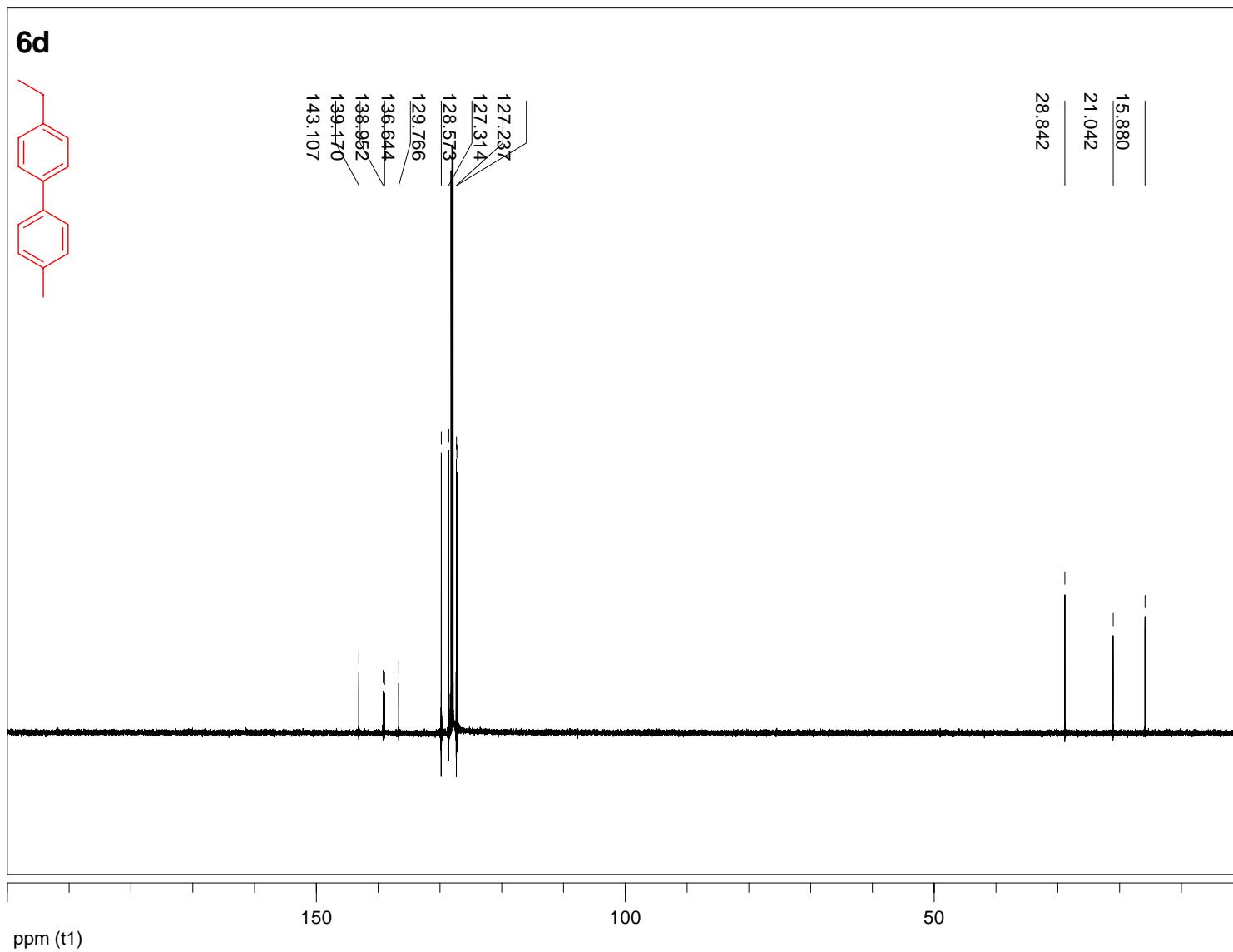
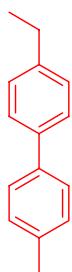
36S

6d



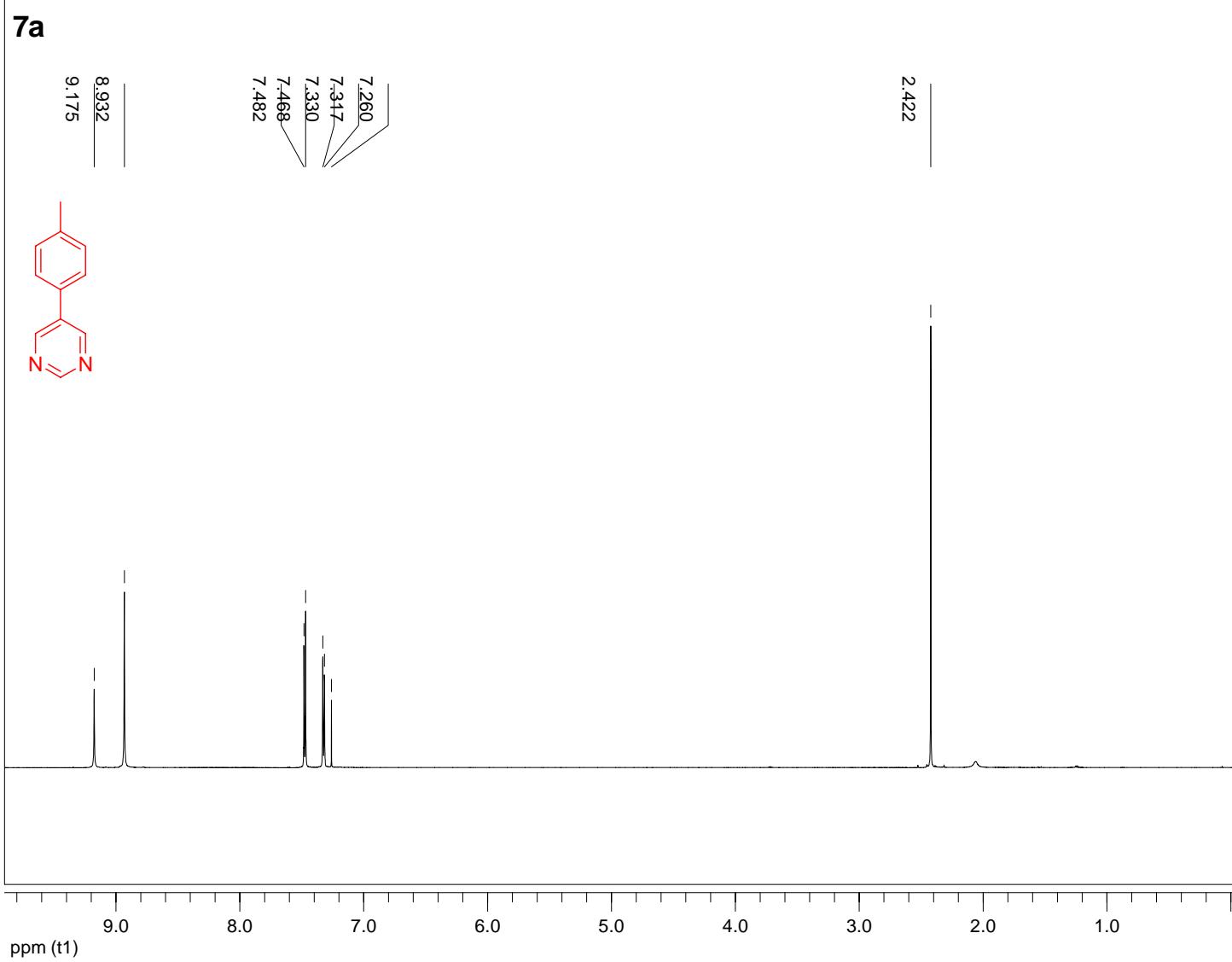
³⁷S

6d



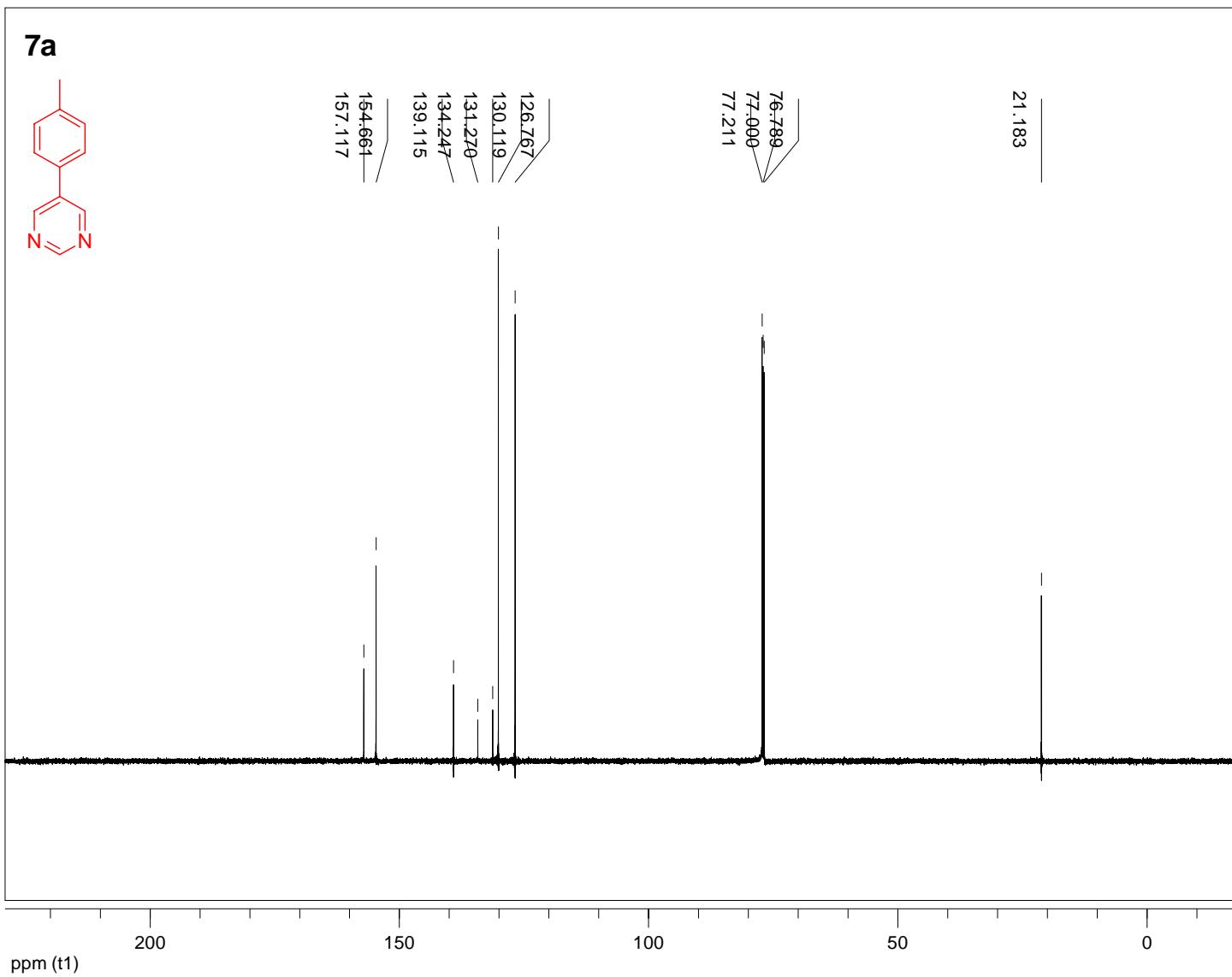
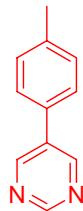
38S

7a



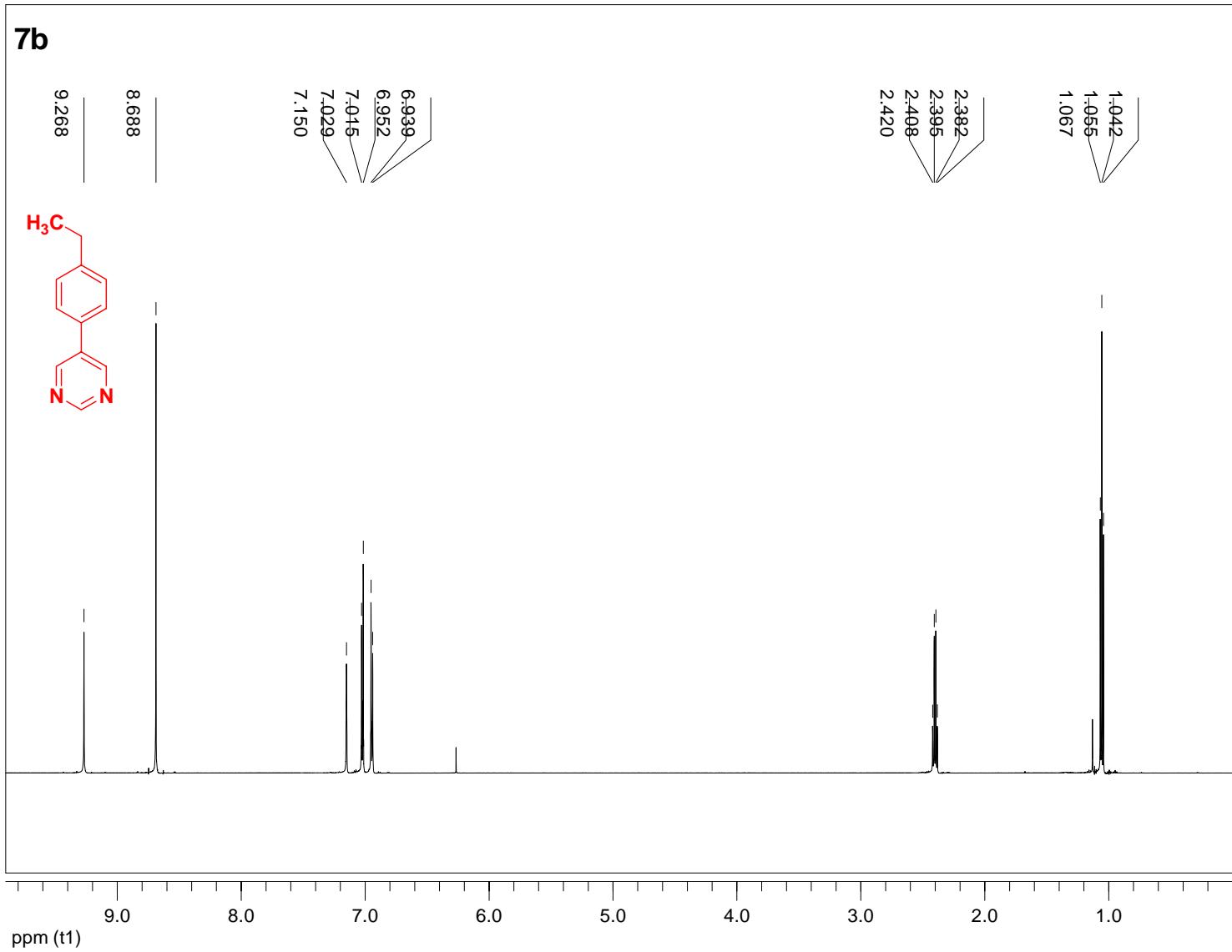
^{39}S

7a



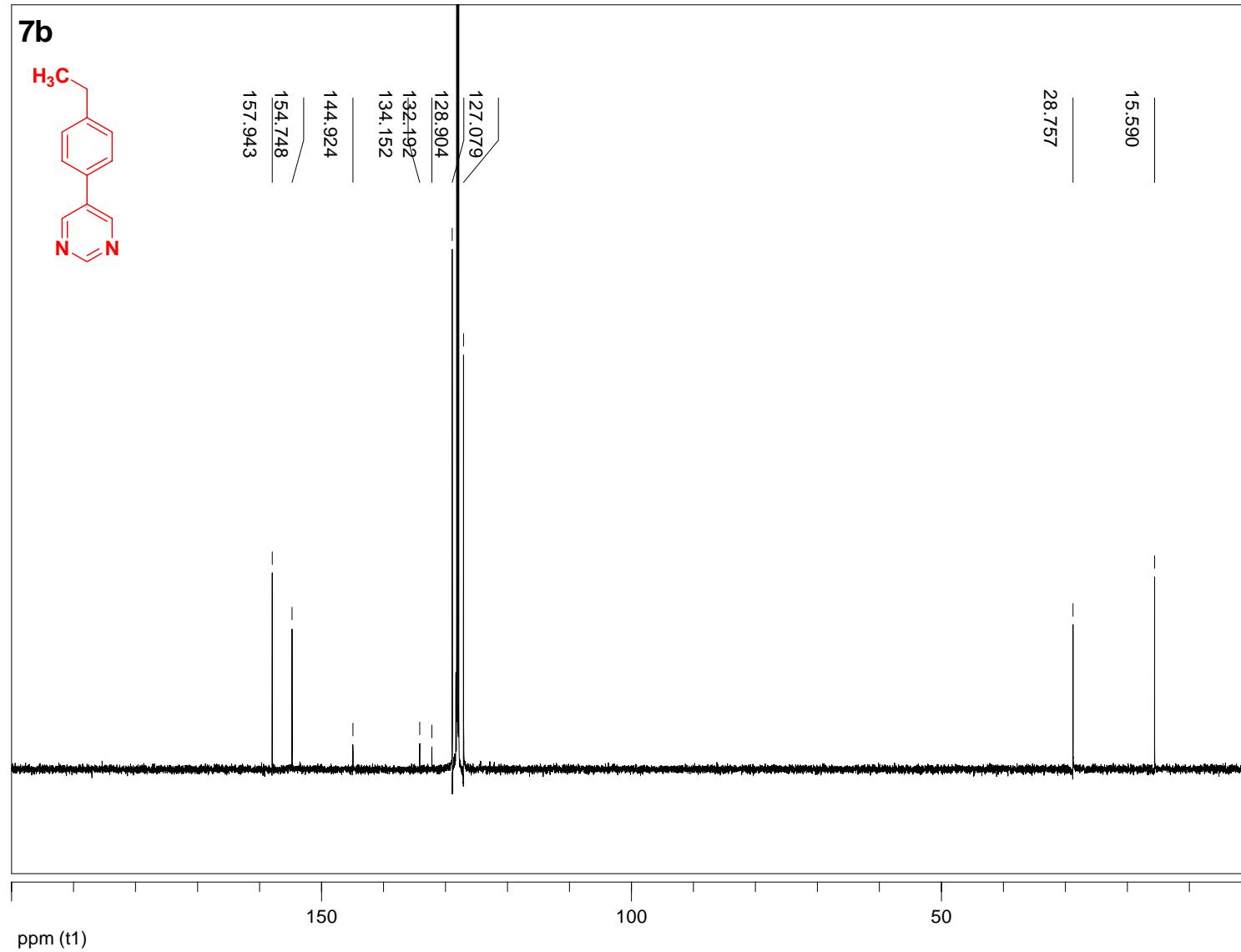
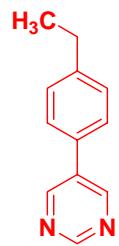
40S

7b

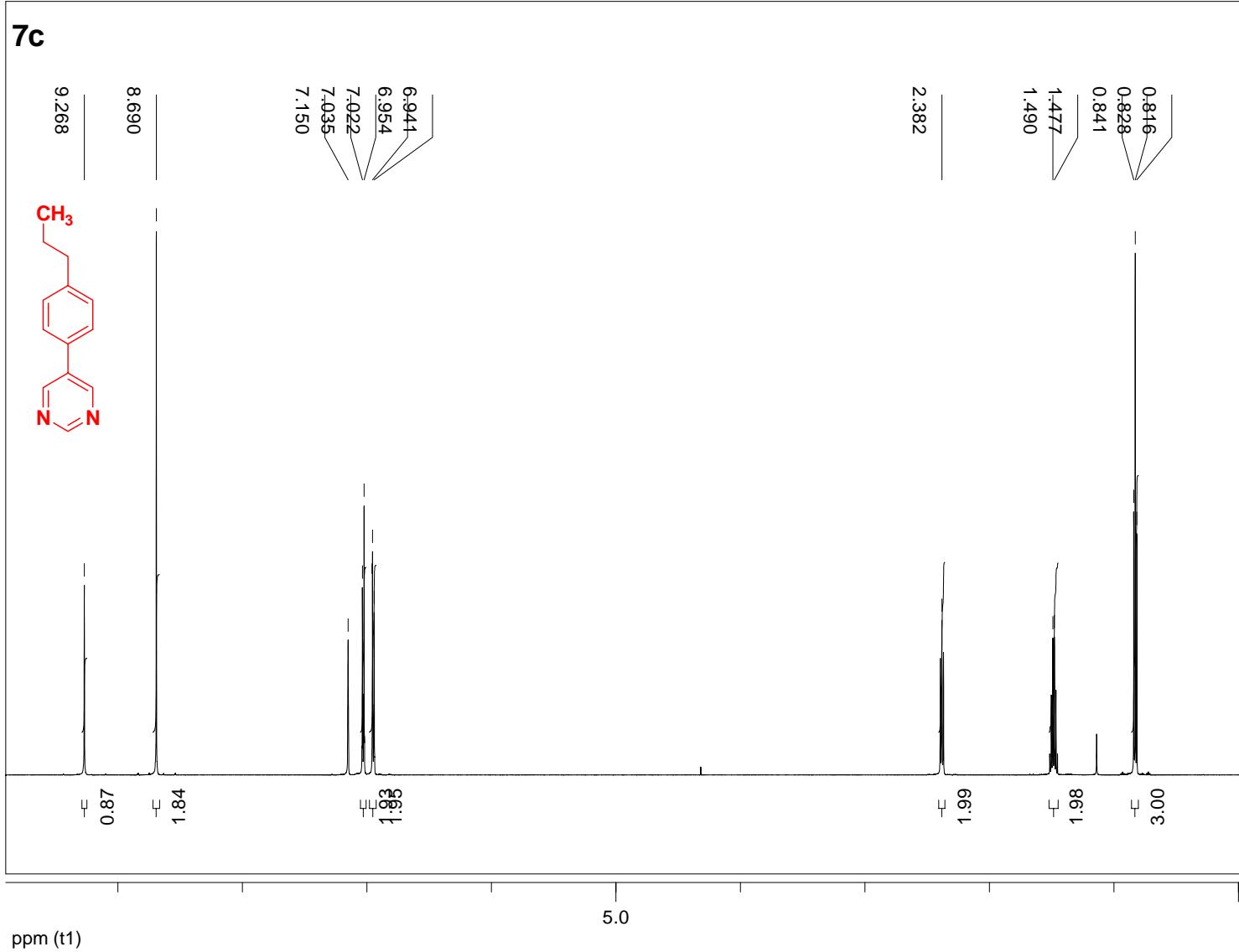


41S

7b

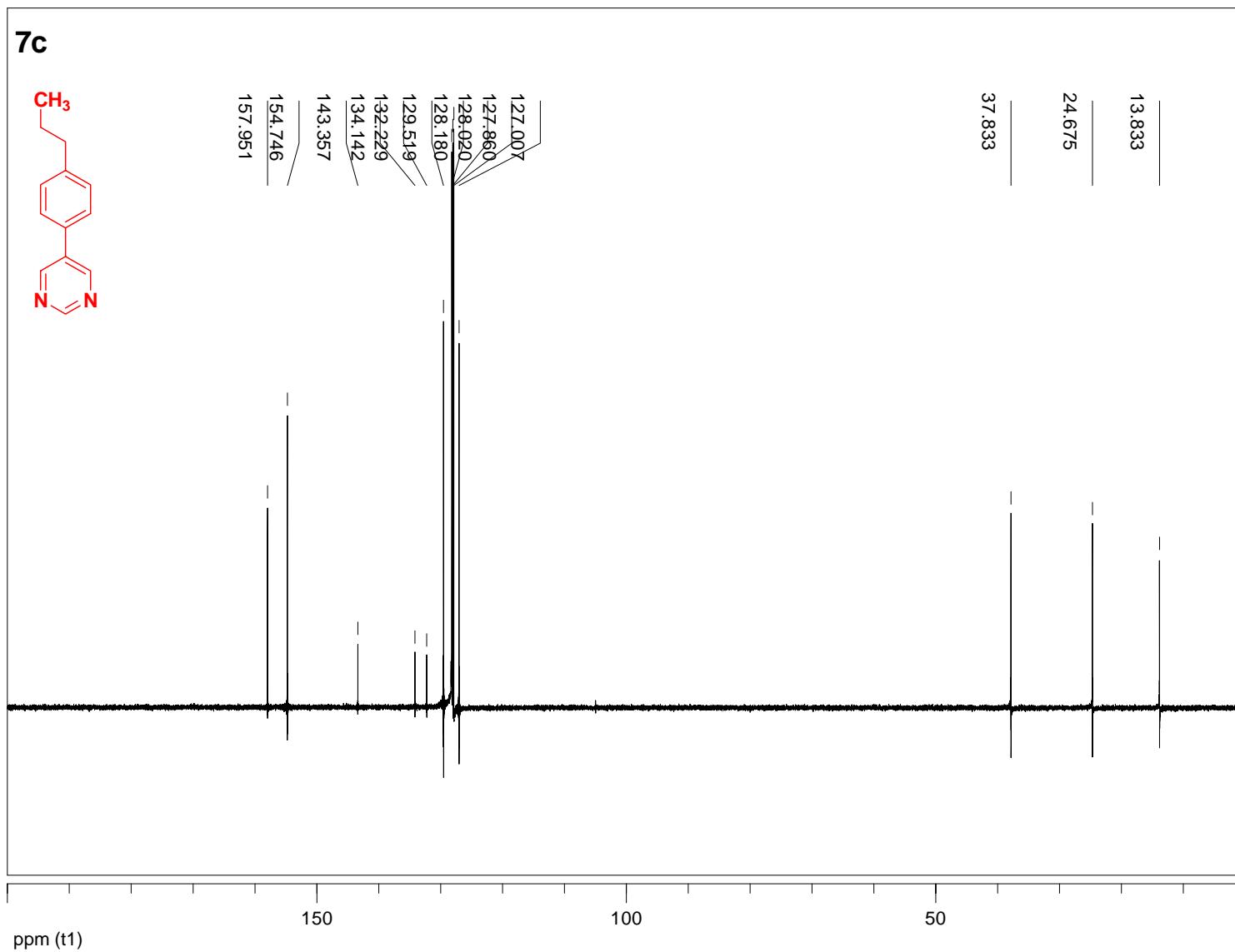


7c



43S

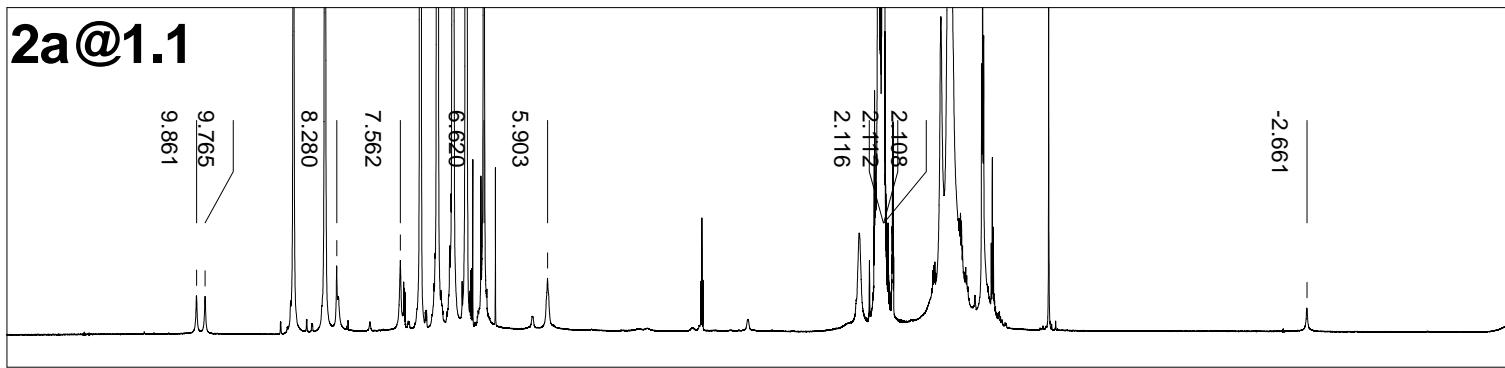
7c



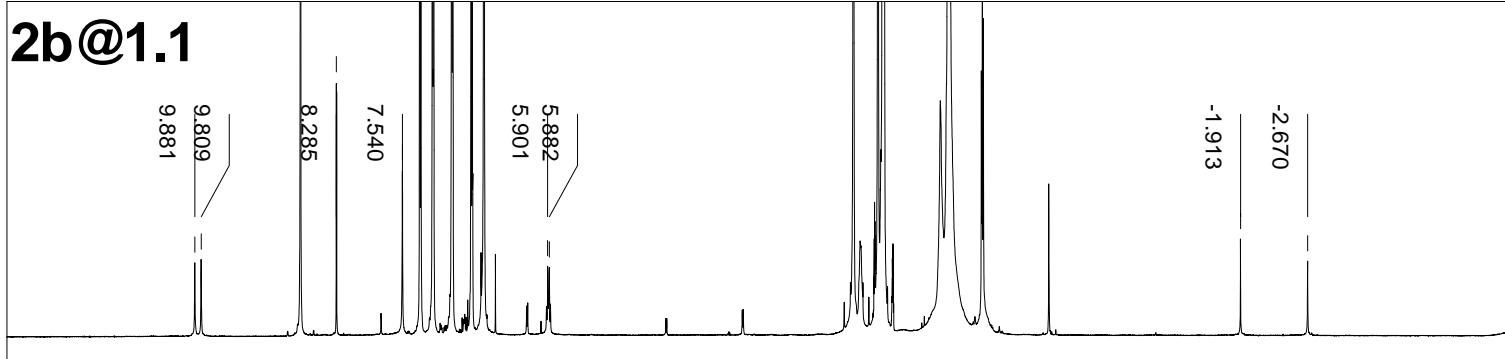
ppm (t1)

44S

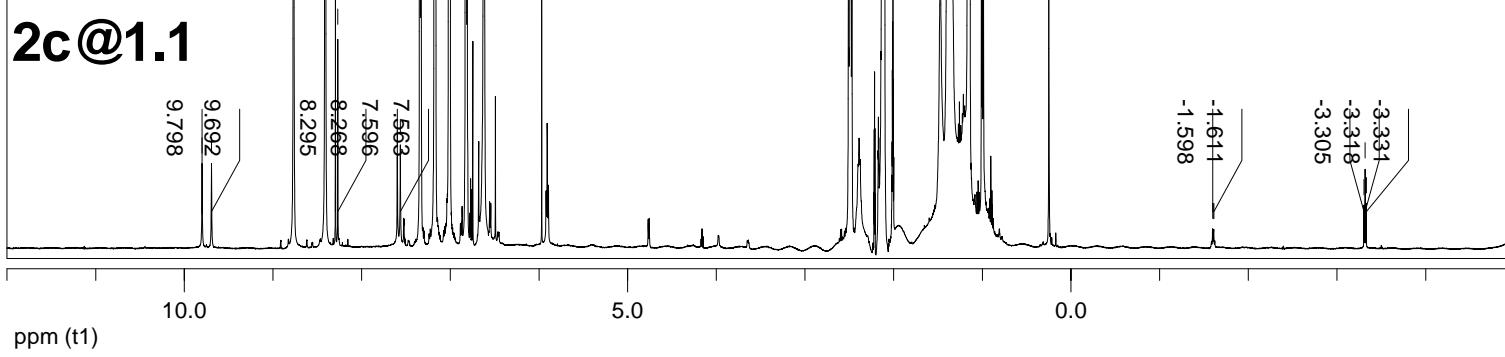
2a@1.1



2b@1.1

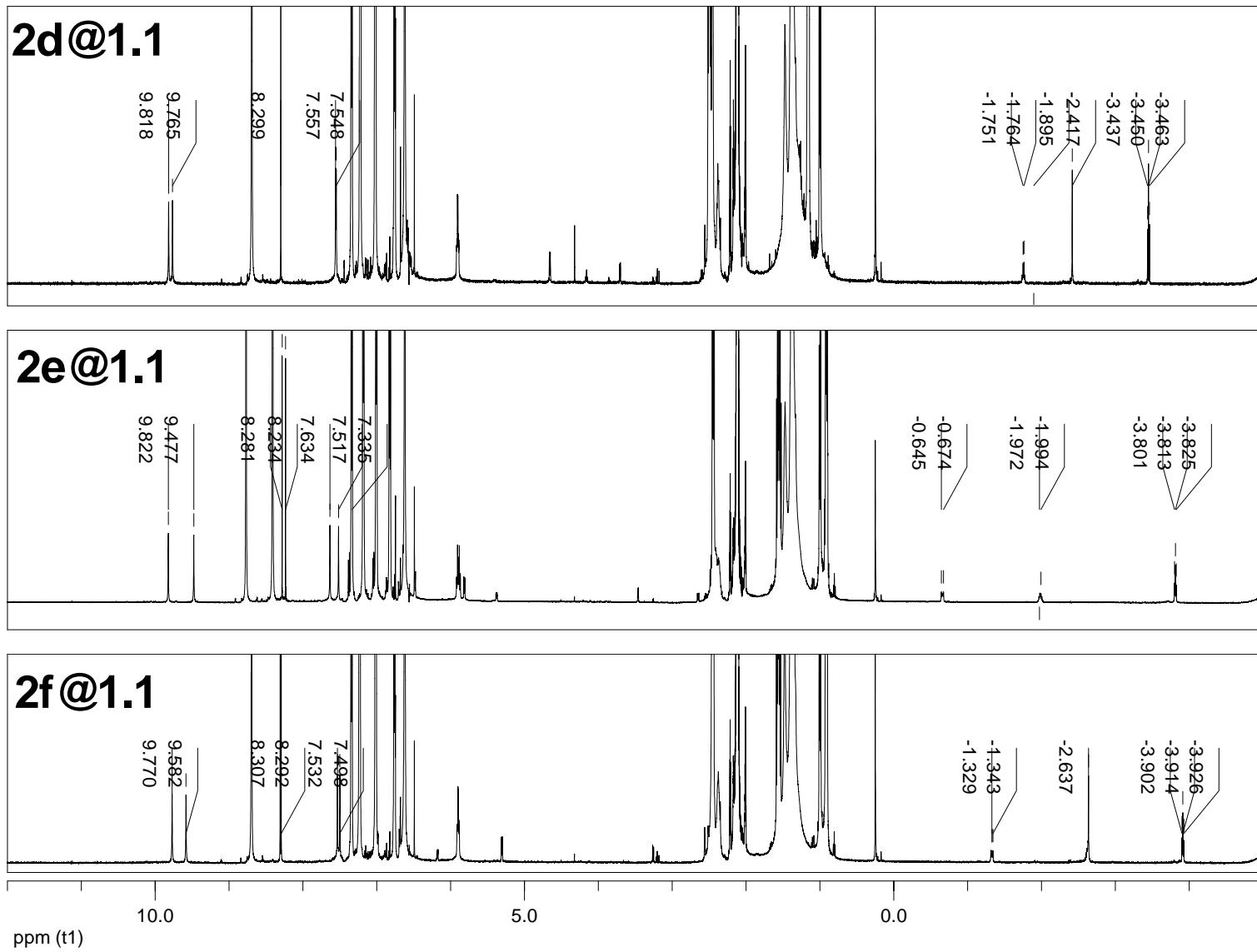


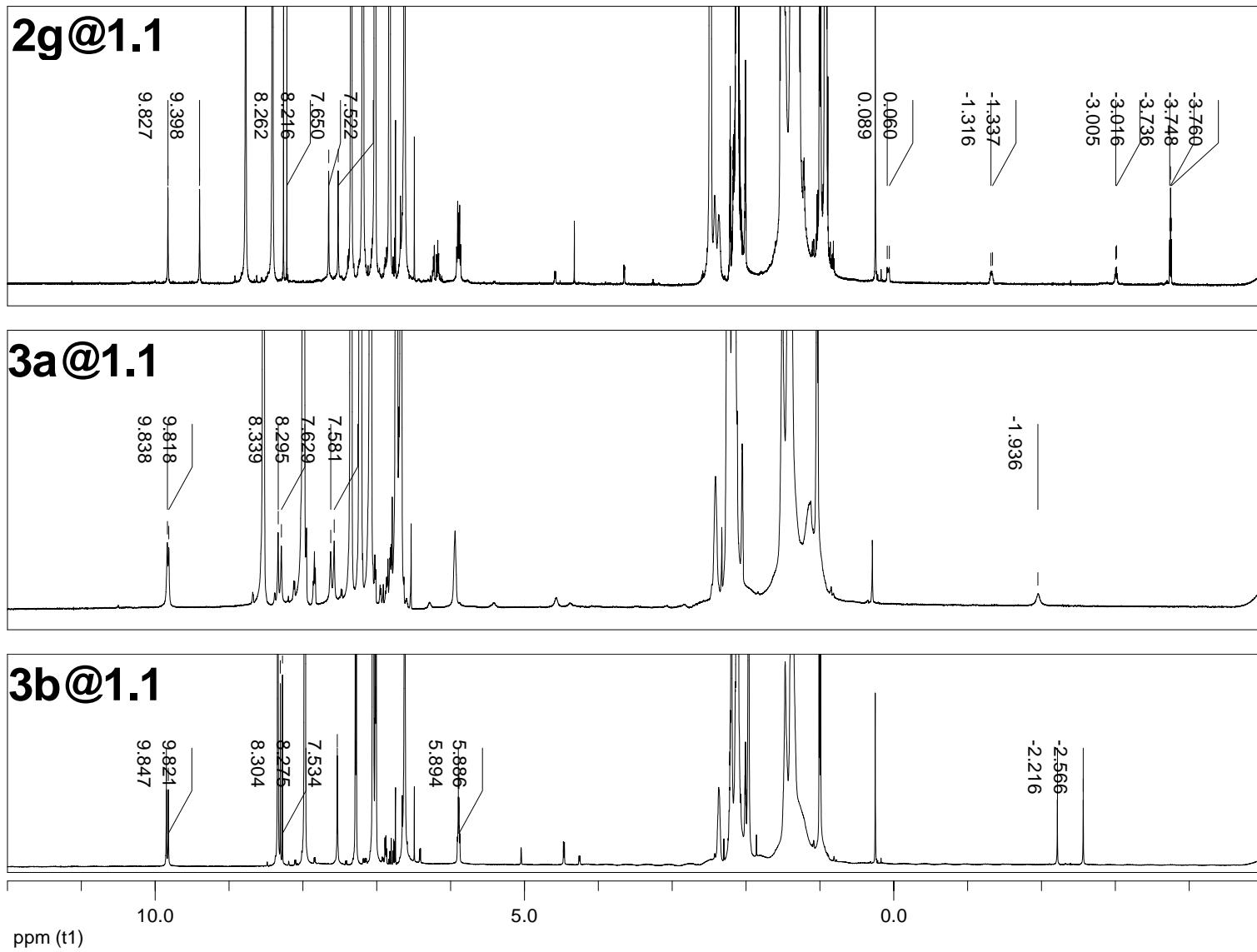
2c@1.1

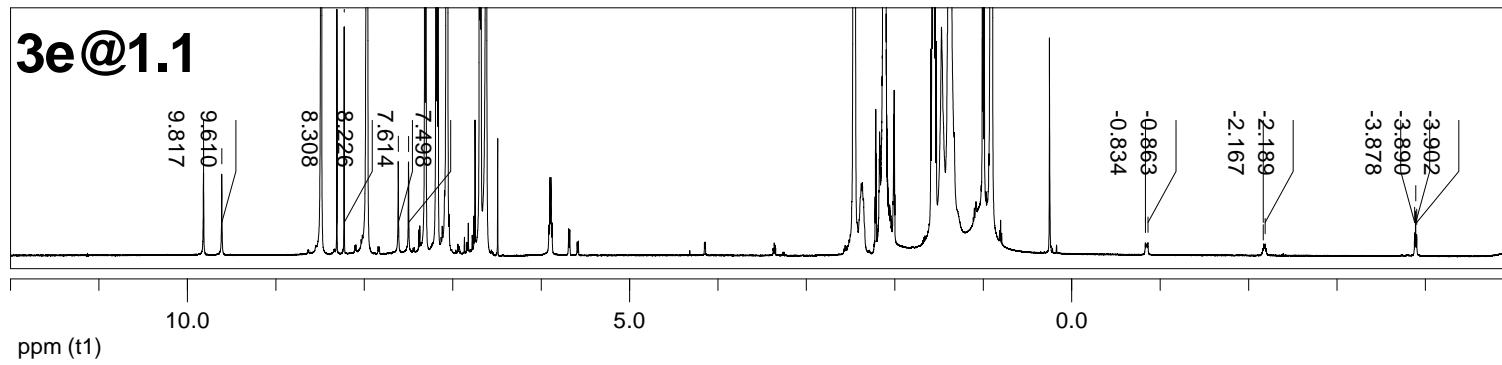
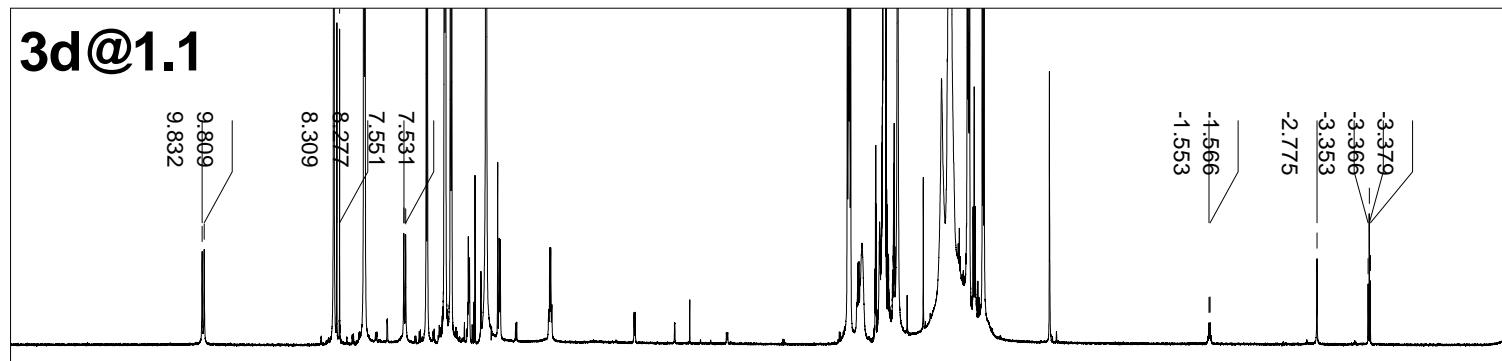
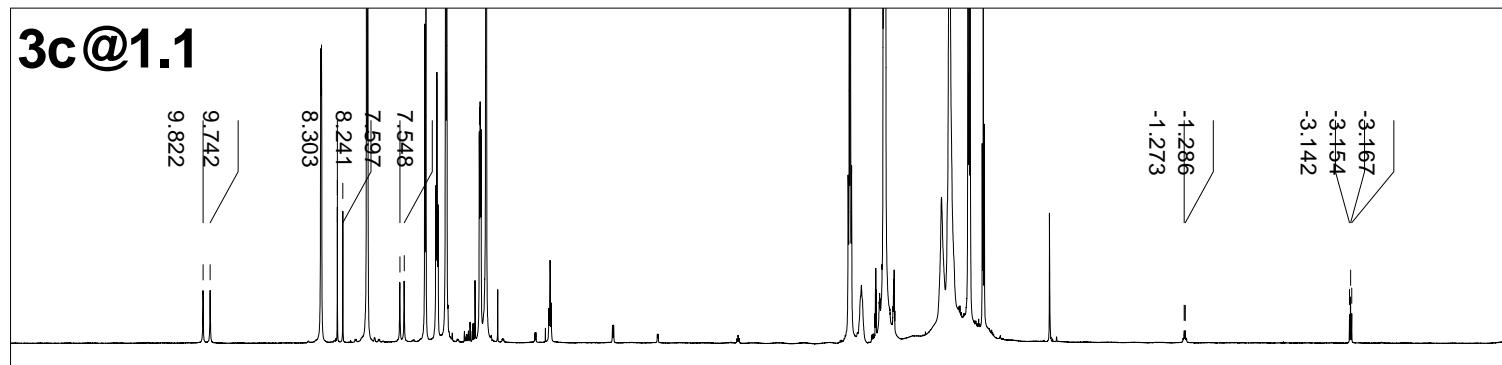


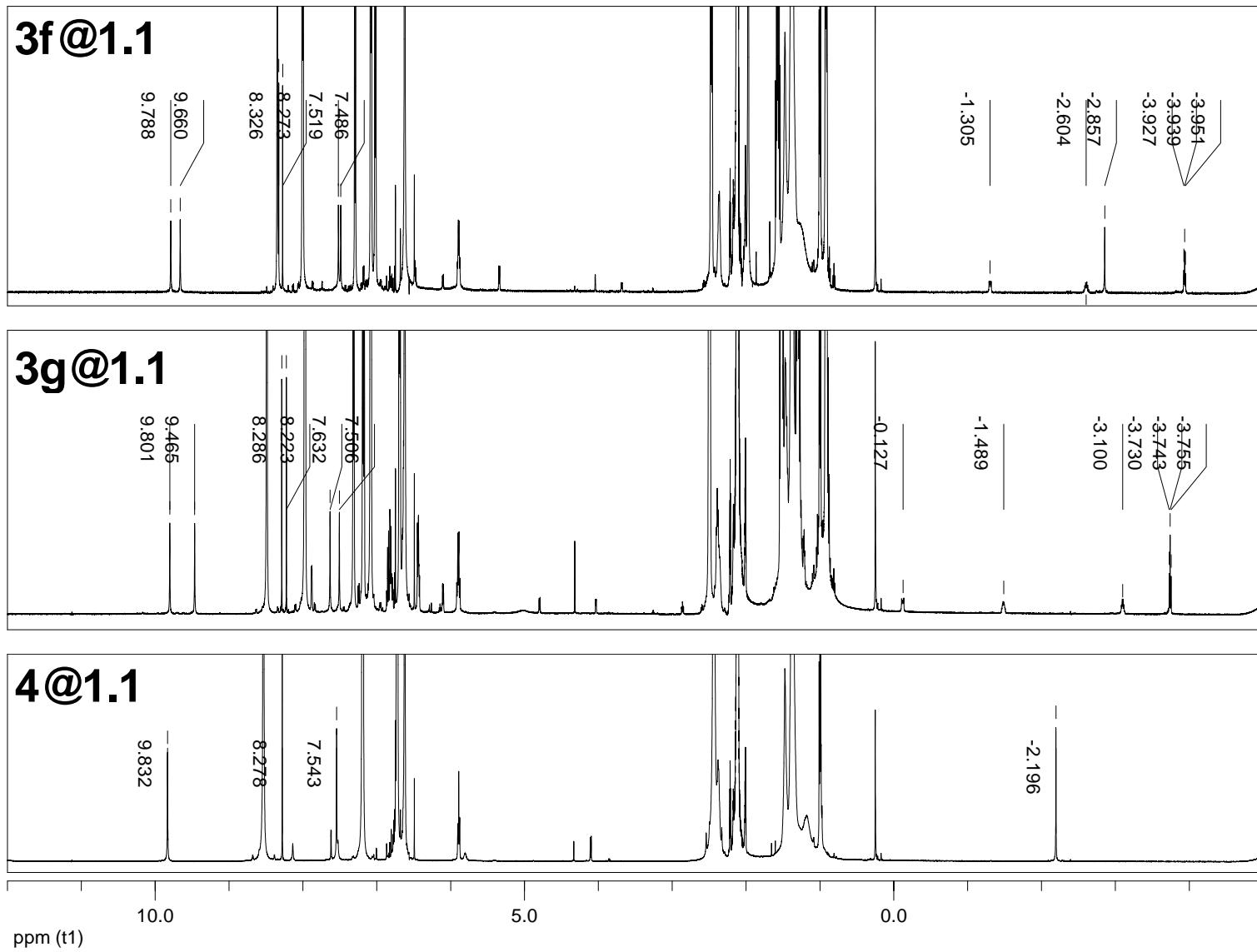
ppm (t1)

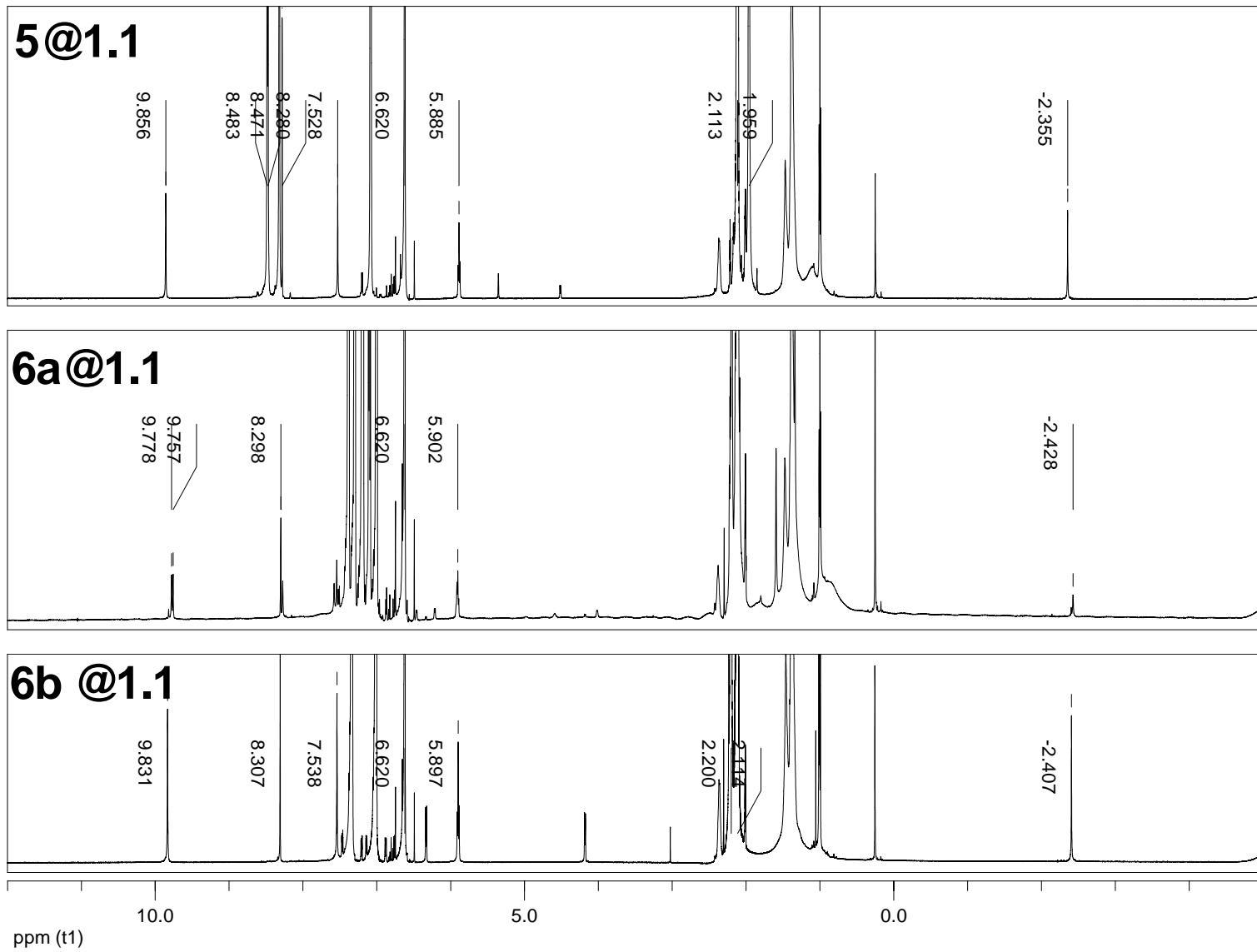
45S



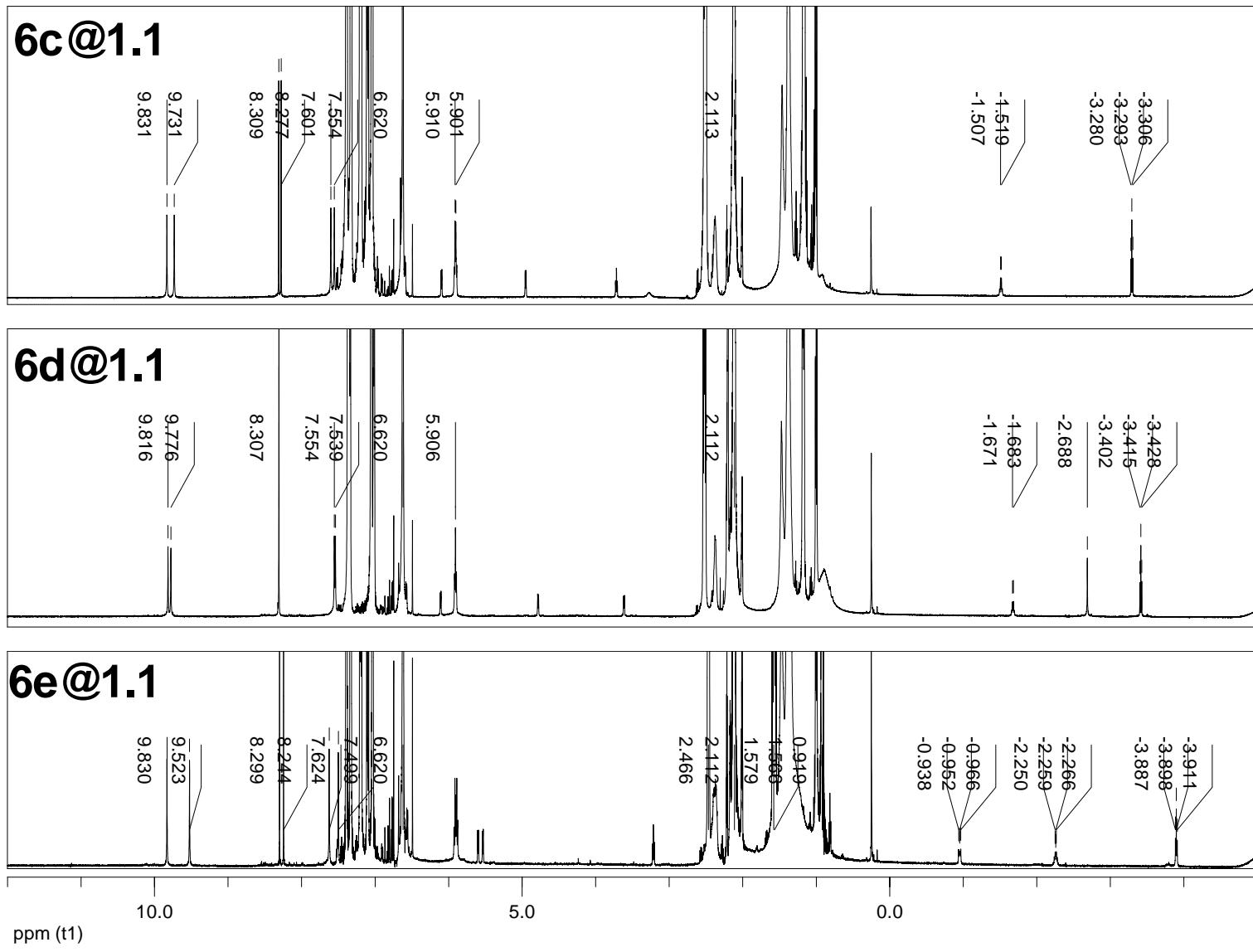




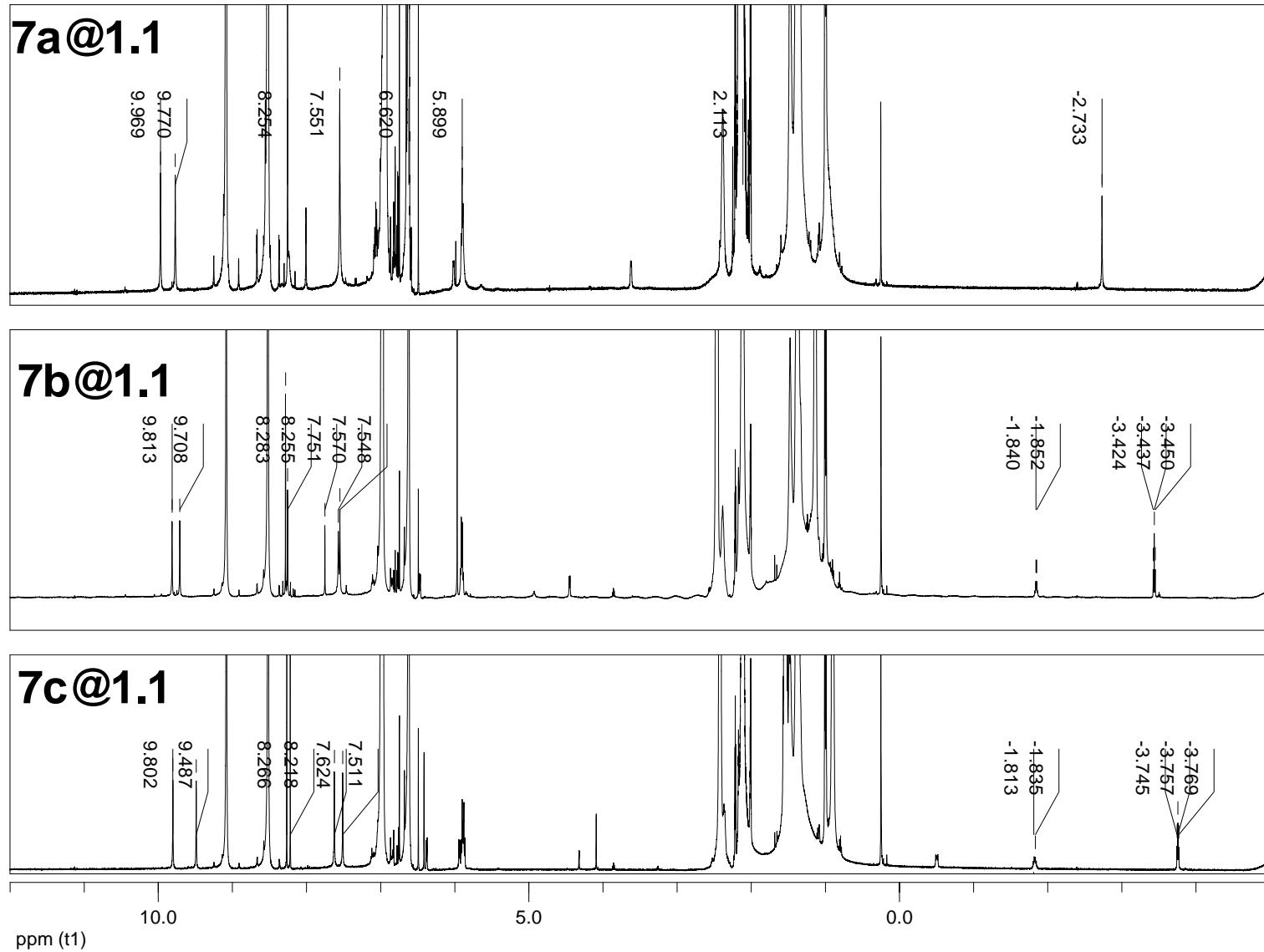




50S



51S



52S

Table1, entry1

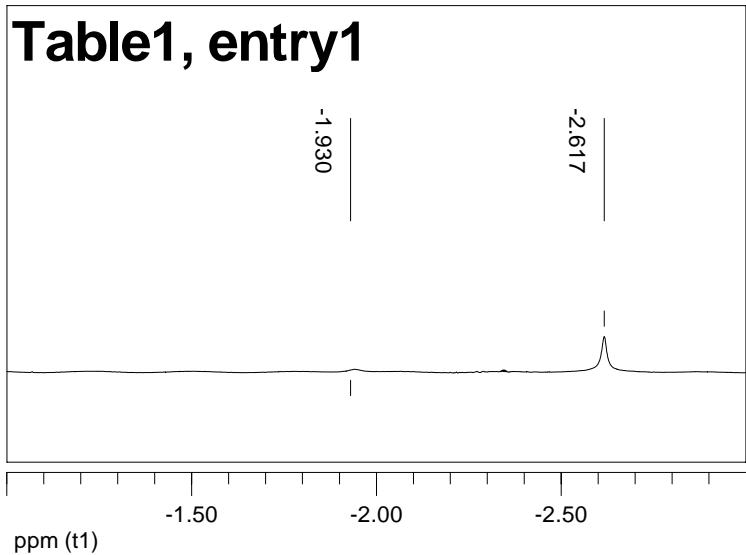


Table1, entry2

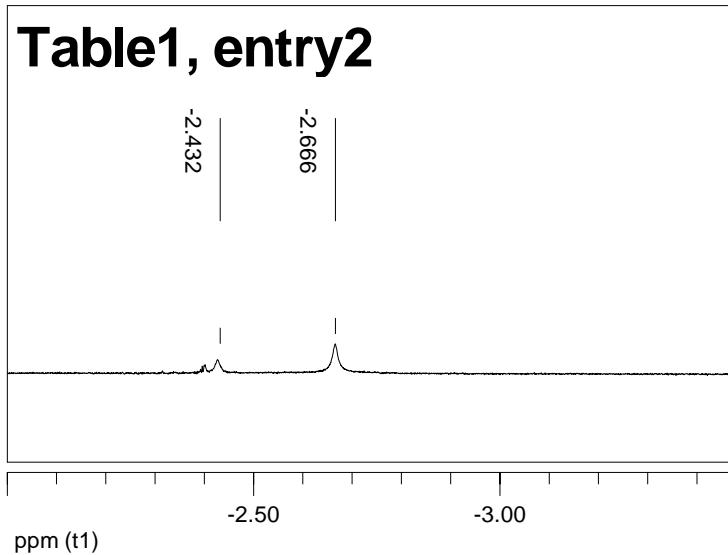


Table1, entry3

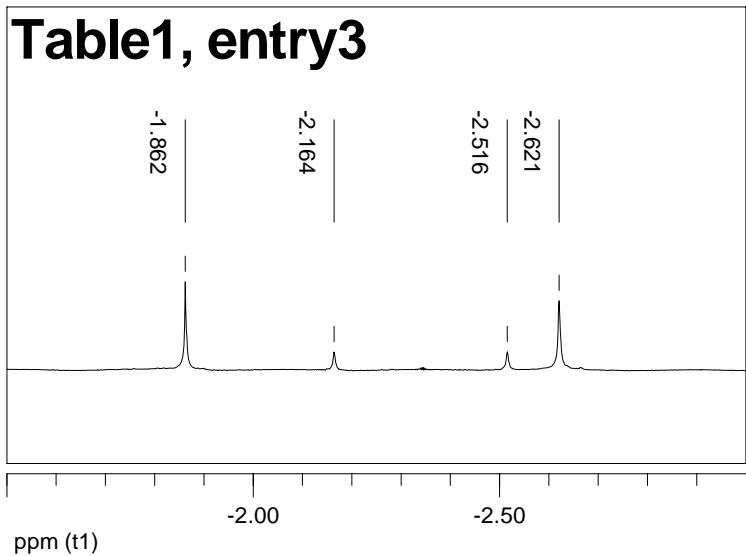


Table1, entry4

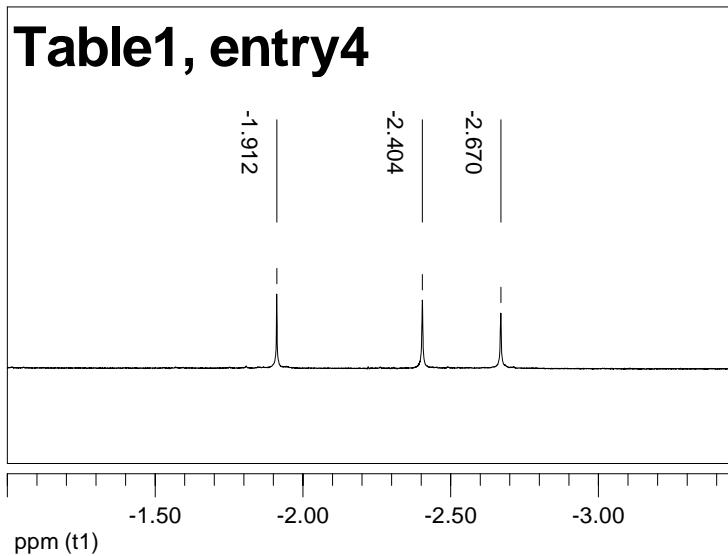


Table1, entry5

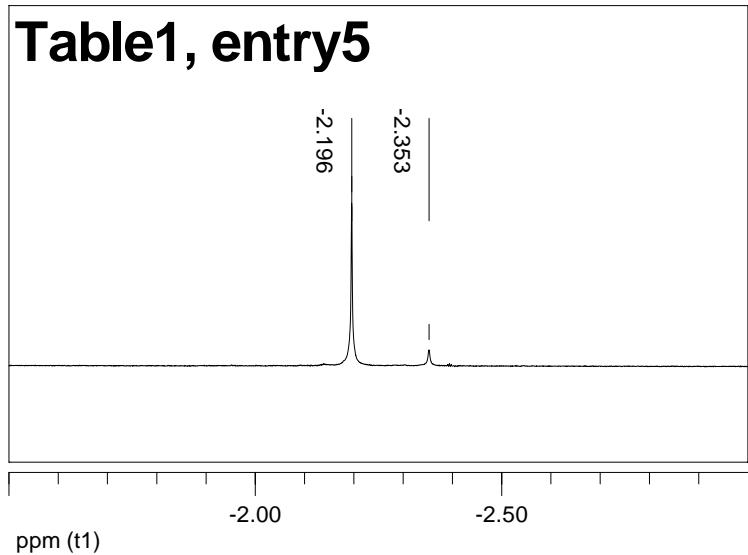


Table1, entry6

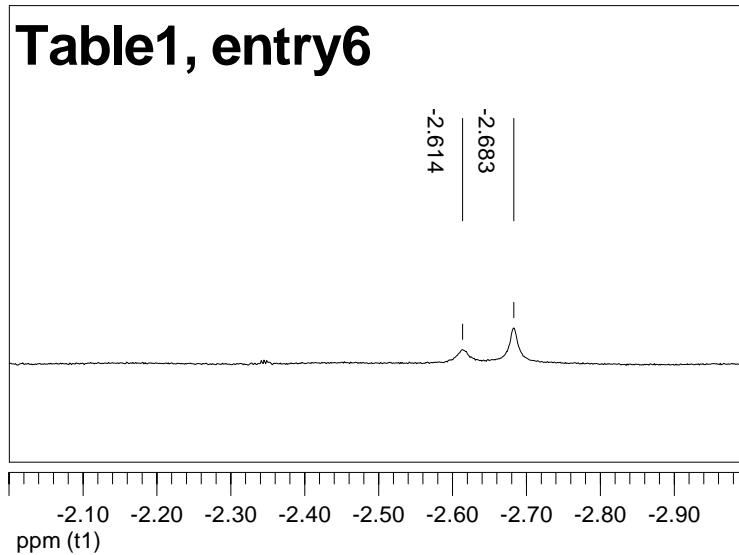


Table2, entry1

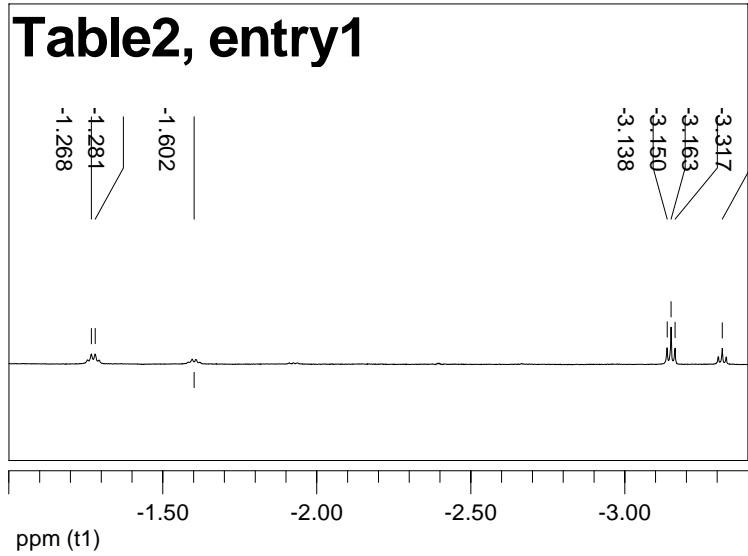


Table2, entry2

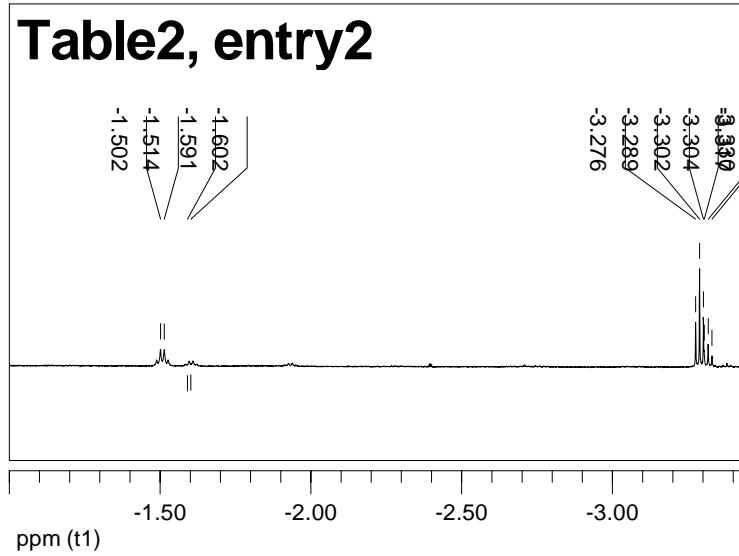


Table2, entry3

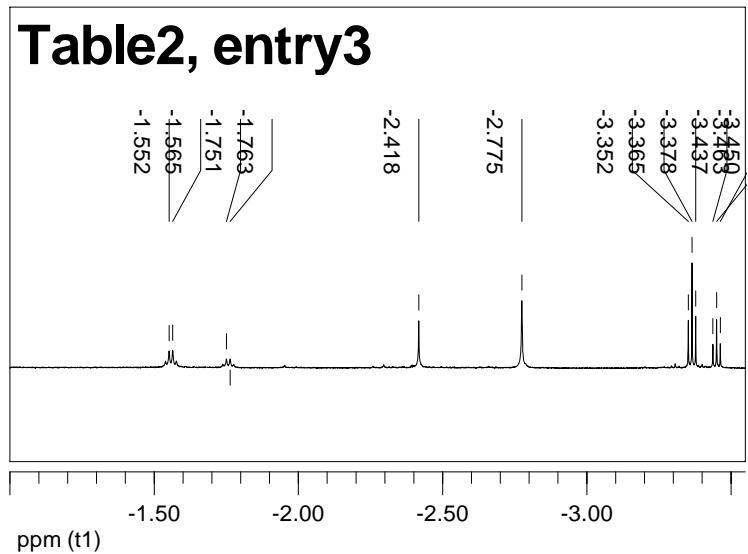


Table2, entry4

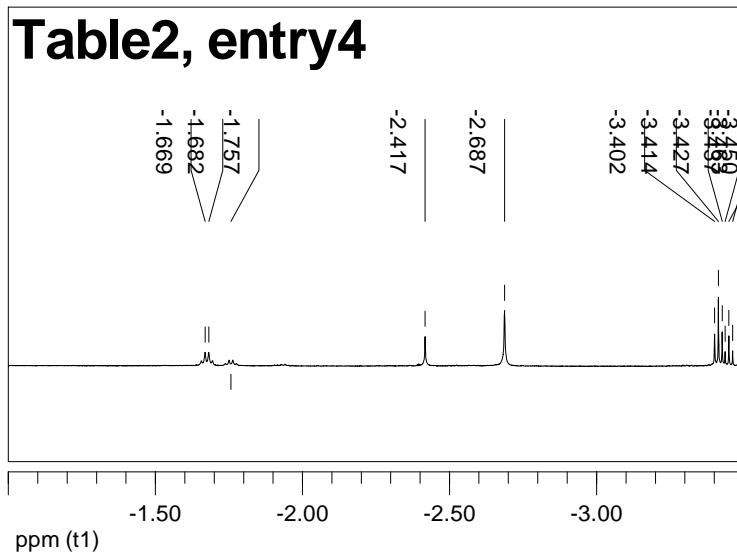


Table2, entry5

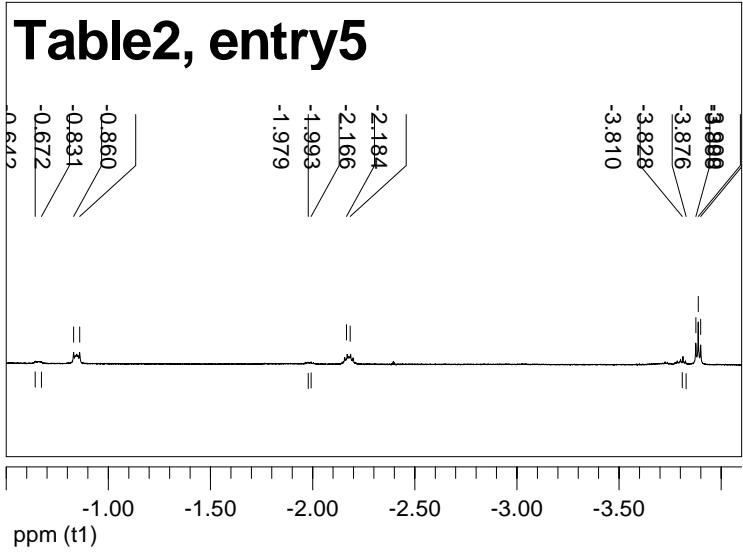


Table2, entry6

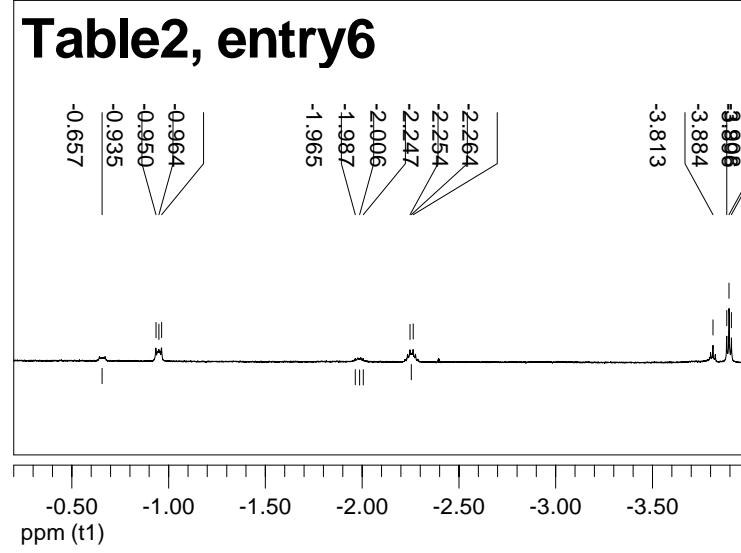


Table2, entry7

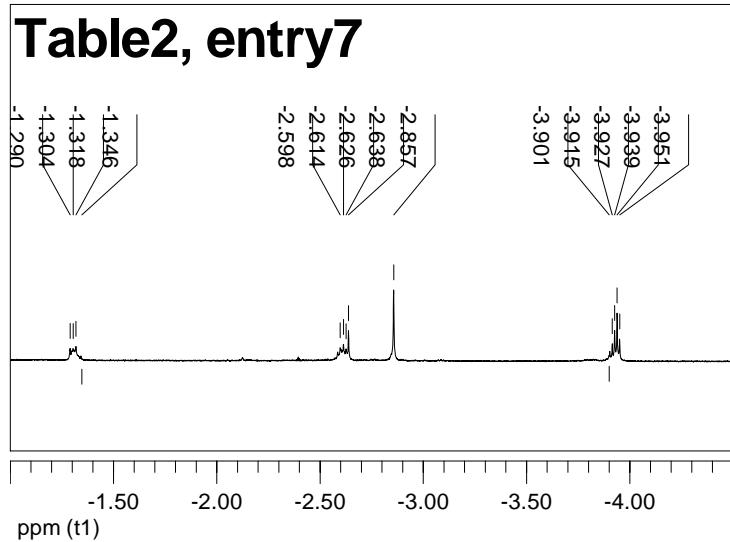


Table2, entry7

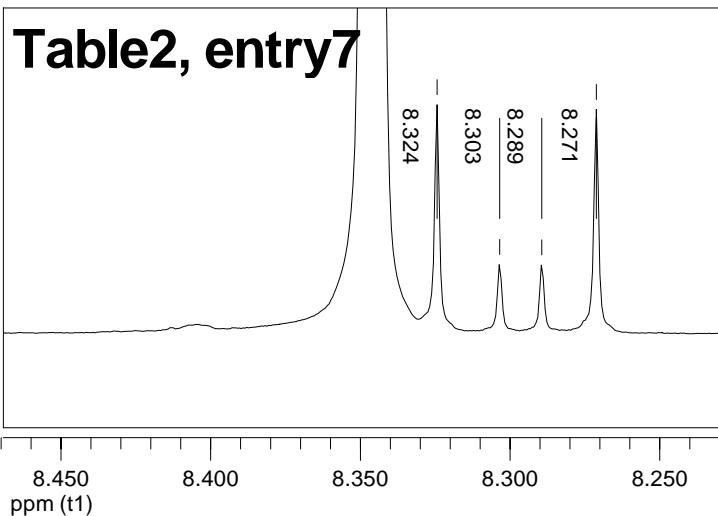


Table2, entry8

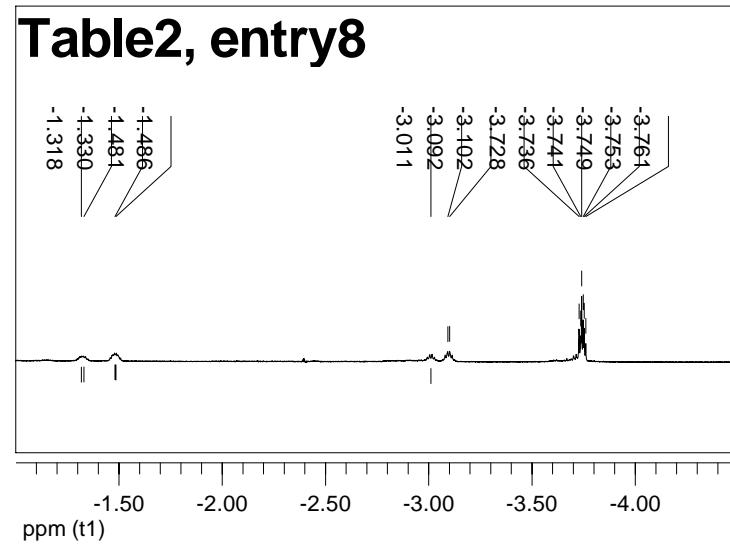


Table2, entry9

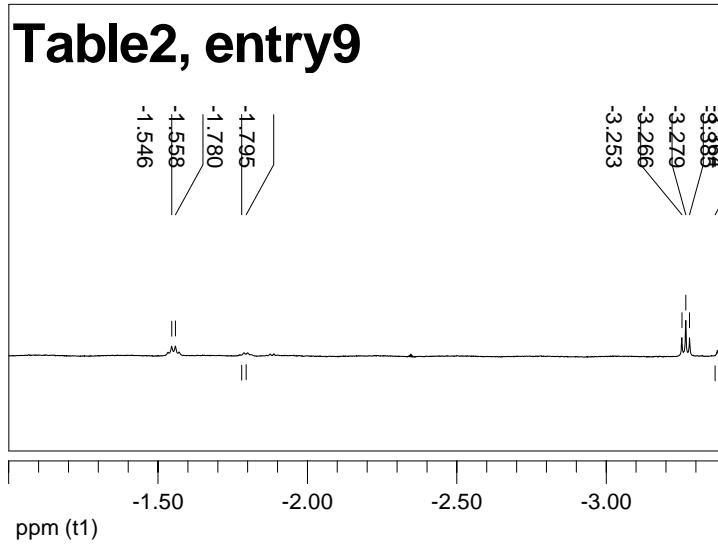


Table2, entry10

