ORIGINAL RESEARCH





Synthesis and cytotoxic evaluation of malachite green derived oleanolic and ursolic acid piperazineamides

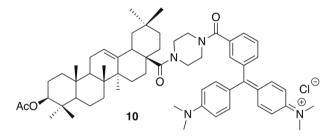
Sander Friedrich¹ · Immo Serbian¹ · Sophie Hoenke¹ · Ratna Kancana Wolfram¹ · René Csuk¹

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Abstract

The coupling of acetylated piperazinylamide spacered triterpenoic oleanolic acid and ursolic acid with *meta* or *para* substituted carboxylated malachite green analogs gave conjugates **10**, **11**, **15**, and **16** that were cytotoxic for several human tumor cell lines. Especially, an oleanolic acid-derived compound **10** was cytotoxic for MCF-7 human breast carcinoma cells $(EC_{50} = 0.7 \,\mu\text{M})$. These derivatives represent first examples of triterpenoic acid derivatives holding a cationic scaffold derived from malachite green.

Graphical Abstract



 $EC_{50} = 0.7 \,\mu M$ (MCF7 breast adenocarcinoma cells)

Keywords Oleanolic acid · Ursolic acid · Cytotoxicity · Terpenoids · Malachite green

Introduction

Chemotherapeutic treatment of cancer represents still a scientific challenge. Although nowadays many patients suffering from cancer can be cured or—at least—their life span can be increased. Several types of cancer, however, are

difficult to be handled, and the cure rate (Holzel et al. 2017; Laffman-Johnson 2012; Vliek et al. 2018) remained low over all the years despite intensive research.

Recently the potential of scaffolds holding a cationic functional group came into the focus of renewed interest inasmuch as their transport into cells might be facilitated by cation-transporters (Cai et al. 2016; Everett et al. 2013; Qian et al. 2016). Some of these transporters were reported to be overexpressed in malignant cells (Cai et al. 2014; Cai et al. 2016; Everett et al. 2013). Several molecules holding a cationic residue are cytotoxic and seem to target the mitochondria, such as ammonium (Biedermann et al. 2010; Kataev et al. 2014) or phosphonium salts (Spivak et al. 2013, 2017) of more complex molecules. Thus, these compounds are mitocans ("mitochondrially targeted anticancer drugs").

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René Csuk rene.csuk@chemie.uni-halle.de

¹ Organic Chemistry, Martin-Luther-University Halle-Wittenberg, Kurt-Mothes-Str. 2, D-06120 Halle (Saale), Germany

Some conjugates of pentacyclic triterpenes are mitocans, too. Thus, hybrids holding an extra cationic functional group have shown promising to excellent cytotoxic results. While "simple" quaternary ammonium salts (Biedermann et al. 2010; Kataev, Strobykina, and Zakharova 2014) were only of moderate cytotoxicity with their EC₅₀ values being in the same potency range as phosphonium salts (Spivak et al. 2017, 2013), hybrids consisting of a suitable pentacyclic triterpene, an amine spacer and a BODIPY-FL group (Brandes et al. 2020; Krajcovicova et al. 2018) held lower EC₅₀ values against a variety of different human cancer cell lines. Superior cytotoxicity, however, was found for those triterpenoids holding one or two O-acetyl groups on ring A, an amide spacer at C-28 (preferentially a piperazinyl residue), and a rhodamine B moiety attached to this spacer (Sommerwerk et al. 2017). EC_{50} values in the low micromolar (Kahnt et al. 2018; Wolfram et al. 2018a, 2018b) and even nano-molar range (Sommerwerk et al. 2017) were reported for these conjugates.

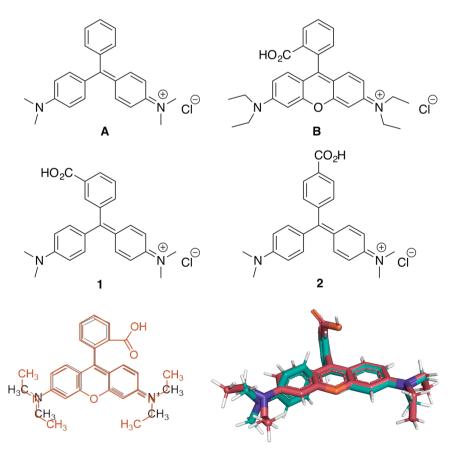
Results and discussion

In extension of these findings and due to the close structural similarity between malachite green (**A**) and rhodamine B (**B**, Fig. 1) we became interested in the synthesis and

Fig. 1 Structure of malachite green (a) and rhodamine B (b) as well as of scaffolds 1 and 2. The lower section displays the close structural similarity between malachite green and rhodamine B biological evaluation of conjugates holding a cationic triphenylmethane moiety especially of scaffolds of the "malachite green type", i.e., 1 and 2; these scaffolds differ from malachite green by the presence of an additional carboxyl group; the latter is necessary for the attachment to the triterpene-spacer adduct.

Recently conjugates holding this type of a malachite green moiety (Müller et al. 1997; Rassow and Gruber 1915; Yang et al. 2007) have been used for the production of antibodies to be used in environmental analyses (Yang et al. 2007), as part of a fluorophore to label antimicrobial peptides (Zhao et al. 2016) and for the detection of nerve gas simulants by chromogenic chemodosimeters (Costero et al. 2012).

The synthesis of 1 (Scheme 1) started with the reaction of *m*-formyl-benzoic acid (3) with *N*, *N*-dimethylaniline in the presence of zinc chloride to yield 4 (Rassow and Gruber 1915; Sinev et al. 1978). Similarly, from *p*-formylbenzoic acid (5) compound 6 was obtained in 89% yield as a blue-greenish compound. Reaction of 4 with tetrachloro-*p*-benzoquinone afforded 41% of 1 while from 6 (Harle et al. 2018; Mueller et al. 1981; Rassow and Gruber 1915) under the same conditions 2 was obtained; both compounds are dark green solids. Compounds of this type are also known as "Mordant Green" or "Mordant Blue".



Scheme 1 Synthesis of malachite green analogs 1 and 2: reactions and conditions:
a ZnCl₂, EtOH, *N*, *N*-dimethylaniline, reflux, 1 day, 89% (of 4), 69% (of 6);
b tetrachloro-*p*-benzoquinone, CHCl₃, 35 °C, 2 h, 41% (of 1), 65% (of 2)

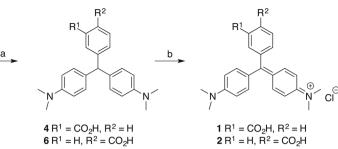
 $\begin{array}{c} CHO \\ H \\ R^2 \end{array} \xrightarrow{a}$

3 $R^1 = CO_2H, R^2 = H$ **5** $R^1 = H, R^2 = CO_2H$

Oleanolic acid (7) was acetylated (Sommerwerk et al. 2017) (Scheme 2) to yield acetate 8; the latter compound was coupled with piperazine as previously reported to yield 9 (Sommerwerk et al. 2017). Compound 1 was activated in situ with oxalyl chloride and coupled with 9 to afford target compound 10. Reaction of 9 with 2 gave 11.

Acetylation of ursolic acid (12) gave acetate 13 followed by its reaction with piperazine to yield amide 14 (Sommerwerk et al. 2017). Coupling of the latter compound with 1 or 2 as described above afforded the hybrids 15 and 16, respectively.

The compounds were screened for their cytotoxic activity using sulforhodamine B assays; the results from these assays are summarized in Table 1. Betulinic acid and doxorubicin were used as standards. While ursolic acid (12) is of similar cytotoxicity as standard betulinic acid, no significant cytotoxicity was found for oleanolic acid (7). Cytotoxicity increased about tenfold for the actylated piperazinylamides 9 and 14. Except for the HT29 human adenocarcinoma cells, the EC₅₀ values determined for the carboxylated malachite green derivatives 1 and 2 were found in the low one-digit µM range. As far as the coupling products 10, 11, 15, and 16 are concerned, oleanolic acid-derived compounds 10 and 11 were of significantly higher cytotoxicity than ursolic acidderived 15 and 16. The selectivity between tumor cells and the nonmalignant fibroblasts is-by and large-the same as in betulinic acid but significantly better than that of doxorubicin. Furthermore, meta substituted malachite green carboxylates 10 and 15 were about twice as cytotoxic than para substituted analogs 11 and 16, respectively. Thus, this makes oleanolic acid-derived 10 to the most cytotoxic compound of this series holding EC_{50} values between 0.7 and 0.9 μ M. Furthermore, 10 is ~17 times more cytotoxic than standard triterpenoic acid betulinic acid and circa 50 times more cytotoxic than starting material oleanolic acid. Interestingly, while the lowest cytotoxicity was observed for HT29 cells, for an analogous rhodamine B derivative especially for this cell line the highest cytotoxicity has been noted. This seems to prove that both the type of triterpenoic acid, the type of amide linkage, the type of cationic residue, and its substitution pattern are of great importance both with respect to a tumor cell line-specific cytotoxicity and to cytotoxicity in general. Ongoing studies in our laboratory try to gain a deeper insight into these observations.



Conclusion

Coupling of acetylated triterpenoic oleanolic acid and ursolic acid holding a piperazinylamide spacer with *meta* or *para* substituted carboxylated malachite green analogs gave conjugates **10**, **11**, **15**, and **16**, respectively. These compounds were cytotoxic for several human tumor cell lines. Thereby, oleanolic acid-derived compound **10** was especially cytotoxic for MCF-7 human breast carcinoma cells $(EC_{50} = 0.7 \,\mu\text{M})$.

Experimental

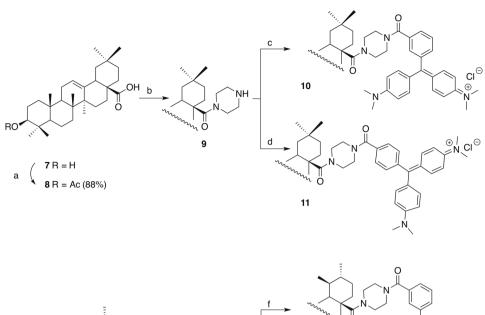
NMR spectra were recorded using the Varian spectrometers Gemini 2000 or Unity 500 (δ given in ppm, J in Hz; typical experiments: H-H-COSY, HMBC, HSQC, NOESY), MS spectra were taken on a Finnigan MAT LCQ 7000 (electrospray, voltage 4.1 kV, sheath gas nitrogen) instrument. The optical rotations were measured on a Perkin-Elmer polarimeter at 20 °C; TLC was performed on silica gel (Merck 5554, detection with cerium molybdate reagent); melting points are uncorrected (Leica hot stage microscope), and elemental analyses were performed on a Foss-Heraeus Vario EL (C-HNS) unit. IR spectra were recorded on a Perkin Elmer FT-IR spectrometer Spectrum 1000. The solvents were dried according to usual procedures. The purity of the compounds was determined by HPLC and found to be >96%. Oleanolic acid (7) and ursolic acid (12) were obtained from Betulinines (Stříbrná Skalice, Czech Republic) in bulk quantities.

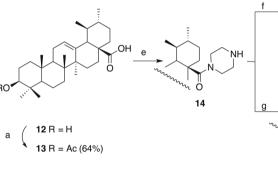
Synthesis

N-(4-((3-carboxyphenyl)(4-(dimethylamino)phenyl) methylene)cyclohexa-2,5-dien-1-yliden)-*N*methylmethanaminium chloride (1)

Following the procedure given for the synthesis of **2** from **4** (1.34 mmol, 500 mg), tetrachloro-*p*-benzoquinone (393 mg, 1.6 mmol) and glacial acetic acid (1 mL; 0.1 mmol) followed by crystallization of the solid from water (100 mL) **1** (224 mg, 41%) was obtained as a purple

Scheme 2 Synthesis of compounds 7–16: reactions and conditions: a Ac₂O, NEt₃, DMAP, DCM, 88% (of 8), 64% (of 13); b oxalyl chloride, THF; DCM, piperazine 49%; c oxalyl chloride, THF; NEt₃, DMAP, 25 °C, 2 h, 50%; d oxalyl chloride, THF; NEt₃, DMAP, 25 °C, 2 h, 50%; e oxalyl chloride, THF; DCM, piperazine 57%; f oxalyl chloride, THF; NEt₃, DMAP, 25 °C, 2 h, 50%; g oxalyl chloride, THF; NEt₃, DMAP, 25 °C, 2 h, 41%





 $NH = \begin{bmatrix} f & & & & & \\ & & & & & & \\ & & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\ & & & & & & \\$

Table 1 Cytotoxicity of selected
compounds; SRB assay EC_{50}
values [μ M] after 72 h of
treatment; averaged from three
independent experiments
performed each in triplicate;
confidence interval CI = 95%

#	FaDu	A2780	HT29	MCF-7	SW1736	NIH 3T3
1	6.1 ± 1.0	9.7 ± 1.1	18.2 ± 1.4	6.4 ± 0.8	8.1 ± 1.5	6.3 ± 1.7
2	8.2 ± 0.9	4.6 ± 0.7	12.4 ± 1.2	6.5 ± 1.1	9.7 ± 0.8	5.2 ± 1.1
7	>30	>30	>30	>30	>30	>30
9	n.d.	1.7 ± 0.1	1.3 ± 0.1	1.7 ± 0.1	1.8 ± 0.9	2.2 ± 1.0
10	0.8 ± 0.2	0.9 ± 0.2	4.3 ± 0.5	0.7 ± 0.1	0.9 ± 0.1	0.6 ± 0.2
11	1.5 ± 0.1	2.1 ± 0.3	4.6 ± 0.2	1.4 ± 0.3	1.7 ± 0.1	1.5 ± 0.3
12	n.d.	11.7 ± 0.6	10.6 ± 0.7	12.7 ± 0.1	14.3 ± 0.4	5.2 ± 1.4
14	n.d.	2.1 ± 0.1	1.9 ± 0.3	2.0 ± 0.1	3.1 ± 0.8	9.7 ± 0.8
15	1.5 ± 0.1	1.4 ± 0.3	2.4 ± 0.2	1.2 ± 0.1	1.7 ± 0.4	1.2 ± 0.3
16	2.3 ± 0.2	4.9 ± 0.6	7.9 ± 0.5	2.9 ± 0.7	4.0 ± 0.5	2.6 ± 0.7
Betulinic acid	n.d.	12.7 ± 1.8	18.4 ± 2.0	12.0 ± 1.7	16.4 ± 1.9	16.1 ± 1.4
Doxorubicin	n.d.	0.01 ± 0.01	0.9 ± 0.2	1.1 ± 0.3	1.7 ± 0.3	0.008 ± 0.001

Human cancer cell lines: FaDu (hypopharyngeal carcinoma), A2780 (ovarian carcinoma), HT29 (colorectal carcinoma), MCF-7 (breast adenocarcinoma), SW1736 (thyroid carcinoma) NIH 3T3 (nonmalignant fibroblasts); cutoff $30\,\mu M$

n.d. not determined

solid; $R_F = (ACN/H_2O, 85:15) = 0.44$; m.p. 207–210 °C; ¹H NMR (400 MHz, DMSO-d₆): $\delta = 10.23$ (s, 1H, COOH), 8.25 (d, J = 7.8 Hz, 1H, 4'-H), 7.80 (t, J =1.8 Hz, 1H, 2'-H), 7.73 (t, J = 7.7 Hz, 1H, 5'-H), 7.56 (dt, 7.7, 1.5 Hz, 1H, 6'-H), 7.34-7.29 (m, 4H, 2-H, 7-H), 7.08 $(d, J = 9.3 \text{ Hz}, 4\text{H}, 3\text{-H}, 8\text{-H}) \text{ ppm}; {}^{13}\text{C NMR} (400 \text{ MHz},$ DMSO-d₆): $\delta = 174.0$ (C-5), 172.4 (C-OOH), 167.0 (C-9), 156.9 (C-1), 140.5 (C-7), 138.5 (C-6'), 134.9 (C-2'), 133.5 (C-4'), 131.6 (C-1'), 129.6 (C-3'), 126.9 (C-6, C4), 121.1 (C-5'), 114.7 (C-3, C8), 41.0 (1-NMe₂), 21.5 (9-NMe₂) ppm; IR (KBr): $\nu = 3447 sbr$, 1617w, 1584m, 1369*m*, 1168*m* cm⁻¹; UV–Vis (CHCl3): λ_{max} (log ε) = 220 (3.4), 345.17 (2.82), 475.22 (2.72), 682.72 (3.06), 714.92 (3.27) nm; MS (ESI, MeOH): m/z (%) = 373.33 $(100\%, [M-Cl]^{-})$; analysis calcd for $C_{24}H_{25}N_2O_2Cl$ (408.93): C 70.49, H 6.16, N 6.85; found: C 70.35, H 6.38, N 6.73.

N-(4-((4-carboxyphenyl)(4-(dimethylamino)phenyl) methylene)cyclohexa-2,5-dien-1-yliden)-*N*methylmethanaminium chloride (2)

To a solution of 6 (500 mg 1.34 mmol) and tetrachloro-pbenzoquinone (393 mg, 1.6 mmol) in CHCl₃ (50 mL) glacial acetic acid (1 mL, 0.1 mmol) was added, and stirring at 35 °C was continued for 2 h. The volatiles were removed under reduced pressure, and the residue was crystallized from water (100 mL) to yield 2 (970 mg, 65%) as a dark green solid; $R_F = 0.3$ (ACN/H₂O, 85:15); m.p. 230–232 °C; ¹H NMR (400 MHz, DMSO-d₆): $\delta = 10.21$ (s, 1H, COOH), 8.12 (d, J = 8.7 Hz, 2H, 3'-H), 7.42 (d, J = 8.5 Hz, 2H, 2'-H), 7.32 (d, J = 9.2 Hz, 2H, 3-H), 7.08 (d, J = 9.2 Hz, 2H, 2-H), 6.97 (d, J = 9.2 Hz, 2H, 7-H), 6.61 (d, J = 9.2 Hz, 2H, 8-H), 3.31 (s, 12H, 1-NMe₂), 2.85 (s, 6H, 9-NMe₂) ppm; ¹³C NMR (100 MHz, DMSO-d₆): $\delta = 173.6$ (C-5), 166.7 (C-OOH), 156.7 (C-1), 154.3 (C-9), 140.3 (C-3), 136.0 (C-1'), 134.9 (C-2'), 134.3 (C-4'), 129.7 (C-3'), 128.9 (C-7), 121.0 (C-6), 111.9 (C-8), 41.1 (1-NMe₂), 40.6 (9-NMe₂) ppm; IR (KBr): $\nu = 3447s$, 1589*m*, 1363*m*, 1169*m* cm⁻¹; UV-vis (CHCl3): λ_{max} (log ε) = 345 (4.38), 480 (4.14), 675 (4.43), 725 (4.65) nm; MS (ESI, MeOH): *m/z* (%) = 373.33 $(100, [M-Cl]^{-})$; analysis calcd for $C_{24}H_{25}N_2O_2Cl$ (408.93): C 70.49, H 6.16, N 6.85; found: C 70.30, H 6.32, N 6.69.

3-[Bis(4-dimethylamino)phenyl]methyl-benzoic acid (4)

To a solution of **3** (1 g, 6.67 mmol) and ZnCl₂ (2.7 g, 20.01 mmol) in ethanol (50 mL) *N*, *N*-dimethylaniline (2.5 g, 20.01 mmol) was added, and stirring under reflux was continued for 1 day. Work-up as described for **6** gave **4** (2.23 g, 89%) as a greenish/blue solid; $R_F = 0.89$ (*n*-hexane/EtOAc, 8:1); m.p. 149–151 °C; ¹H NMR (400 MHz,

DMSO-d₆): δ = 7.92 (d, J = 13.8 Hz, 2H, 2'-H, 4'-H), 7.29 (d, J = 19.0 Hz, 2H, 6'-H, 3'-H), 6.94 (d, J = 13.8 Hz, 4H, 3-H), 6.67 (d, J = 13.8 Hz, 4H, 2-H), 5.4 (s, 1H, 5-H), 2.88 (s, 12H, 1–NMe₂) ppm; ¹³C-NMR (400 MHz, DMSO-d₆): δ = 148.8 (C-1), 145.5 (C-1'), 134.1 (C-2'), 132.8 (C-3'), 131.1 (C-5'), 129.9 (C-3, C-4), 129.1 (C-6'), 128.1 (C-4'), 113.0 (C-2), 54.9 (C-5), 40.9 (NMe₂) ppm; IR (KBr): ν = 3448*s*, 1614*m*, 1519*m*, 1351*w* cm⁻¹; UV–vis (CHCl₃): λ_{max} (log ε) = 289.65 (4.3 nm); MS (ESI, MeOH): *m*/*z* (%) = 375.27 (100%, [M + H]⁺); analysis calcd for C₂₄H₂₄N₂O₂ (374.48): C 76.98, H 7.00, N 7.48; found: C 76.83, H 7.15, N 7.31.

4-[Bis(4-dimethylamino)phenyl]methyl-benzoic acid (6)

To a solution of 5 (1.44 g, 9.6 mmol) and $ZnCl_2$ (3.9 g, 28.8 mmol) in ethanol (100 mL) N, N-dimethylaniline (3.6 mL, 28.8 mmol) was slowly added, and the mixture was stirred under reflux for 1 day. The mixture was diluted with methanol (40 mL), the pH value was adjusted to 5 by adding aq. HCl, and the precipitate was filtered off. Compound 6 (2.45 g, 69%) was obtained as light green/blue solid; $R_F = 0.95$ (*n*-hexane/EtOAc, 3:1); m.p. 252 °C (decomp.) (lit: 250 °C, decomp. (Yang et al. 2007)); ¹H NMR (400 MHz, DMSO-d₆): $\delta = 12.76$ (s, 1H, 5'-H), 7.88–7.82 (m, 2H, 3'-H), 7.21–7.16 (m, 2H, 2'-H), 6.93-6.85 (m, 4H, 2-H), 5.38 (s, 1H, 5-H), 2.83 (s, 12H, NMe₂) ppm; ¹³C NMR (400 MHz, DMSO-d₆): $\delta = 167.8$ (C-5'), 151.1 (C-1'), 149.3 (C-1), 131.9 (C-4), 129.8 (C-2), 129.7 (C-3'), 129.4 (C-2'), 112.8 (C-3'), 54.59 (C-5), 40.7 (NMe₂) ppm; IR (KBr): $\nu = 3447s_{br}$, 2883*m*, 1683*s*, 1610*s*, 1571w, 1519s, 1479w, 1443w, 1428w, 1349w, 1314w, 1293w, 1227w, 1202w, 1177w, 1164w, 1061w, 1017w cm⁻¹; UV-vis (CHCl₃): λ_{max} (log ε) = 270.37 (4.16) nm; MS (ESI, MeOH): m/z (%) = 375.33 (100%, [M + H]⁺); anlysis calcd for C₂₄H₂₄N₂O₂ (374.48): C 76.98, H 7.00, N 7.48; found: C 76.77, H 7.19, N 7.29.

3β-Acetyloxy-olean-12-en-28-oic acid (8)

Acetylation of **7** as previously described gave **8** (88%) as a colorless solid; $R_F = 0.7$ (silica gel, toluene/ethyl acetate/formic acid/heptane, 80:26:5:1); m.p.: 263–265 °C (lit.: 260–261 °C) (Sommerwerk et al. 2017); $[\alpha]_D =$ +117.7° (*c* 0.37, CHCl₃), (lit.: +119° (*c* 0.1, CHCl₃) (Sommerwerk et al. 2017); MS (ESI, MeOH): *m*/*z* = 497.5 (75%, $[M - H]^-$), 995.2 (100%, $[2M - H]^-$), 1017.7 (29%, $[2M - 2H + Na]^-$).

3β-Acetyloxy-olean-12-en-28-oyl piperazine (9)

Reaction of **8** with oxalyl chloride and piperazine as previously described gave **9** (49%); $R_F = 0.46$ (silica gel, CHCl₃/MeOH, 9:1); m.p. 172–176 °C (lit.: 170–176 °C) (Sommerwerk et al. 2017); MS (ESI, MeOH): m/z = 567.4 (45%, $[M + H]^+$).

N-(4-((4-dimethylamino)phenyl)-3-((4-(3β-acetyloxyolean-12-en-28-carboxy)piperazin)-1-yl)oxy)carbonylphenylmethylencyclohexa-2,5-dien-1-yliden)-*N*methylmethanaminium chloride (10)

Following the procedure given for the synthesis of 11, from 1 (200 mg, 0.49 mmol) compound 10 (240 mg, 49.8%) was obtained as a dark green solid; R_F (CHCl₃/MeOH, 9:1) = 0.35; m.p. 212–215 °C; ¹H NMR (400 MHz, CDCl₃): $\delta =$ 8.02 (s, 1H, 37-H), 7.78-7.69 (m, 1H, 39-H), 7.49-7.3 (m, 4H, 2'-H, 7'-H), 7.05–6.91 (d, J = 8.9 Hz, 1H, 40-H), 6.99 (d, J = 8.8 Hz, 4H, 3'-H, 8'-H), 6.72–6.58 (m, 1H, 38-H), 5.21 (t, J = 3.6 Hz, 1H, 12-H), 4.49 (m, 1H, 3-H), 3.72 (q, J = 7.0 Hz, 4H, 33-H), 3.39 (s, 6H, N-Me₂), 3.23 (d, J =15.5 Hz, 4H, 34-H), 2.96 (s, 1H, 18-H), 2.17 (m, 1H, 16-H_a), 2.04 (s, 3H, 32-H), 1.94–1.86 (m, 2H, 11-H), 1.63–1.54 (m, 8H, 6-H, 16-H_b, 2-H, 15-H_a, 7-H), 1.53–1.20 (m, 11H, 22-H, 15-H_b, 21-H, 1-H, 19-H, 9-H), 0.79 (m, 1H, 5-H), 0.89–0.80 (m, 12H, 23-H, 24-H, 25-H, 27-H), 0.72 (s, 3H, 30-H), 0.70 (s, 6H, 26-H, 29-H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C-28), 171.0 (C-31), 169.3 (C-35), 157.0 (C-1', C-9'), 143.6 (C-7'), 140.9 (C-8'), 140.8 (C-2'), 138.5 (C-41), 134.9 (C-39), 133.4 (C-37), 131.5 (C-40), 129.5 (C-38), 125.0 (C-12), 121.9 (C-36), 120.4 (C-6'), 114.3 (C-3'), 81.3 (C-3), 56.0 (C-30), 47.4 (C-9), 46.6 (C-34), 43.4 (C-33), 41.2 (1-NCH₃), 39.4 (C-8), 39.0 (C-5), 38.8 (C-17), 38.6 (C-1), 37.5 (C-4), 36.5 (C-10), 36.0 (C-18), 34.9 (C-21), 33.3 (C-22), 31.9 (C-14), 30.4 (C-20), 30.3 (C-27), 28.3 (C-24), 23.7 (C-15), 23.6 (C-2), 23.5 (C-11), 23.4 (C-19), 23.0 (C-16), 21.6 (C-32), 21.2 (C-29), 18.4 (C-6), 17.0 (C-23), 16.1 (C-26), 15.9 (C-25), 8.4 (C-7), 0.89 (C-5') ppm; IR (KBr): $\nu = 3447s_{br}$, 2925*m*, 1618*m*, 1584*m*, 1370*m*, 1247*m*, 1170*m* cm⁻¹; UV–vis (CHCl3): λ_{max} (log ε) = 229.01 (3.56), 244.02 (4.26), 347.2 (3.87), 485.3 (3.71), 731.09 (4.35) nm; MS (ESI, MeOH): m/z (%): 921.73 (100%, $[M - Cl]^{-}$); $C_{26}H_{81}N_4O_4Cl$ (957.78): C 75.24, H 8.52, N 5.85; found: C 75.13, H 8.81, N 5.43.

N-(4-((4-dimethylamino)phenyl)-4-((4-(3β-acetyloxyolean-12-en-28-carboxy)piperazin)-1-yl)oxy)-

carbonylphenylmethylencyclohexa-2,5-dien-1-yliden)-*N*-methylmethanaminium chloride (11)

To a solution of **2** (500 mg, 1.23 mmol) in dry dichloromethane (DCM, 50 mL), oxalyl chloride (0.62 mg, 4.92 mmol) and three drops of dry dimethylformamide (DMF) were added. After stirring for 30 min, the volatiles were removed under reduced pressure, the residue was dissolved in dry tetrahydrofuran (THF, 3×20 mL), and the solvents were evaporated DMF. The solid was dissolved in dry DCM (50 mL), triethylamine (0.2 g, 0.3 mL, 10 mmol), DMAP (4-dimethylaminopyridine, 15 mg, 0.1 mmol), and a solution of 9 (849 mg, 1.5 mmol) in dry DCM (20 mL) were added. The mixture was stirred at 25 °C for 2 h; usual workup followed by chromatography (silica gel, CHCl₃/MeOH, 9:1) gave 11 (594 mg, 50%) as a dark green solid; R_F $(CHCl_3/MeOH, 9:1) = 0.35; m.p. 192-195 ^{\circ}C; ^{1}H NMR$ (400 MHz, CDCl₃): $\delta = 7.99$ (s, 2H, 37-H), 7.56 (d, J =8.2 Hz, 2H, 38-H), 7.37 (dd, J = 8.73, 3.5 Hz, 4H, 2'-H, 8'-H), 6.97 (d, J = 9.2 Hz, 4H, 3'-H, 7'-H), 5.21 (m, 1H, 12-H), 4.47 (t, J = 8.3 Hz, 1H, 3-H), 3.69 (m, 4H, 33-H), 3.36 (s, 6H, N-Me₂), 3.1 (q, J = 14.0, 7.3 Hz, 4H, 34-H), 2.94 (s, 1H, 18-H), 2.12 (m, 1H, 16-H_a), 2.02 (s, 3H, 32-H), 1.89 (m, 2H, 11-H), 1.79–1.54 (m, 8H, 6-H, 16-H_b, 2-H, 15-H_a, 7-H), 1.53–1.25 (m, 8H, 22-H, 21-H, 1-H_a, 19-H, 9-H), 1.22 (s, 3H, 27-H), 1.06 (m, 2H, 1-H_b, 15-H_b), 0.92 (m, 6H, 25-H, 29-H), 0.79 (m, 1H, 5-H), 0.88–0.77 (dd, J = 10.9, 6.2 Hz, 9H, 23-H, 24-H, 30-H), 0.72 (s, 3H, 26-H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 171.0$ (C-28), 169.3 (C-35), 162.5 (C-31), 157.0 (C-1', C9'), 143.6 (C-7'), 140.9 (C-8'), 140.8 (C-2'), 140.8 (C-13), 139.3 (C-39), 134.7 (C-36), 127.3 (C-38), 127.2 (C-37), 125.2 (C-12), 119.4 (C-6'), 114.1 (C-3'), 80.9 (C-3), 55.3 (C-30), 47.5 (C-9), 45.9 (C-34), 42.1 (C-33), 41.2 (1-NMe₂), 39.4 (C-8), 38.8 (C-5), 38.7 (C-17), 38.2 (C-1), 37.7 (C-4), 36.9 (C-10), 36.5 (C-18), 34.4 (C-21), 33.0 (C-22), 31.4 (C-14), 30.4 (C-20), 29.7 (C-27), 28.1 (C-24), 24.1 (C-2), 23.5 (C-15), 23.3 (C-19), 23.0 (C-16), 21.3 (C-32), 21.2 (C-29), 18.2 (C-6), 16.9 (C-23), 16.7 (C-26), 15.5 (C-25), 8.6 (C-7), 0.98 (C-5') ppm; IR (KBr): $\nu = 3445s_{br}$, 2956w, 1636m, 1584m, 1371*m*, 1170*w* cm⁻¹; UV–vis (CHCl3): λ_{max} (log ε) = 242 (3.16), 345 (3.83), 475 (3.61), 617.15 (1.79), 732.12 nm (3.22) nm; MS (ESI, MeOH): m/z (%) = 921.80 (100%, [M $-Cl]^{-}$; analysis calcd for $C_{26}H_{81}N_4O_4Cl$ (957.78): C 75.24, H 8.52, N 5.85; found: C 75.02, H 8.76, N 5.39.

3β-Acetyloxy-urs-12-en-28-oic acid (13)

Acetylation of **12** as previously described (Sommerwerk et al. 2017) gave **13** (64%) as a colorless solid; $R_F = 0.7$ (silica gel, toluene/ethyl acetate/formic acid/heptane, 80:26:5:1); m.p. 242–244 °C (lit.: 242.7–244.1 °C); $[\alpha]_D = +69.89^{\circ}$ (*c* 0.86, CHCl₃), (lit.: +71.2° (*c* 1.0, CHCl₃); MS (ESI, MeOH): m/z = 497.5 (64%, $[M - H]^{-}$), 542.9 (30%, $[M + HCO_2]^{-}$), 995.1 (68%, $[2M - H]^{-}$), 1017.5 (100%, $[2M - 2H + Na]^{-}$).

3β-Acetyloxy-olean-12-en-28-oyl piperazine (14)

Reaction of **13** with oxalyl chloride and piperazine as previously (Sommerwerk et al. 2017) described gave **14** (57%); $R_F = 0.44$ (silica gel, CHCl₃/MeOH, 9:1); m.p. 158–161 °C (lit.: 158–161 °C); MS (ESI, MeOH): m/z = 567.4 (61%, $[M + H]^+$).

N-(4-((4-dimethylamino)phenyl)-3-((4-(3β-acetyloxyurs-12en-28-carboxy)piperazin1-1-yl)oxy)carbonylphenylmethylencyclohexa-2,5-dien-1-yliden)-*N*methylmethanaminium chloride (15)

Following the procedure given for the synthesis of 11 from 1 (200 mg, 0.49 mmol) and 14 (333 mg, 0.59 mmol) compound 15 (200 mg, 50%) was obtained as a dark green solid; R_F (CHCl₃/MeOH, 9:1) = 0.35; m.p. 214–218 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.00$ (s, 1H, 37-H), 7.70 (d, J = 3.8 Hz, 1H, 39-H), 7.62 (t, J = 3.7 Hz, 1H, 38-H), 7.44-7.28 (m, 5H, 2'-H, 7'-H, 40-H), 7.01 (d, J = 5.4 Hz, 4H, 3'-H, 8'-H), 5.23 (t, J = 3.7 Hz, 1H, 12-H), 4.47 (t, J = 7.9 Hz, 1H, 3-H), 3.70 (m, 4H, 33-H), 3.38 (s, 12H, NMe₂), 3.14 (m, 4H, 34-H), 2.94 (m, 1H, 18-H), 2.12 (m, 1H, 16-H_a), 2.02 (s, 3H, 32-H), 1.97–1.79 (m, 2H, 11-H), 1.78–1.45 (m, 16H, 19-H, 6-H, 16-H_b, 2-H, 15-H, 7-H, 22-H, 21-H, 1-H), 1.44-1.28 (m, 1H, 20-H); 1.19 (m, 2H, 5-H, 9-H), 1.11 (s, 3H, 23-H), 0.89 (s, 12H, 27-H, 25-H, 24-H), 0.84 (s, 6H, 26-H, 30-H), 0.69 (s, 3H, 29-H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.1$ (C-28), 171.0 (C-31), 169.1 (C-35), 157.0 (C-1', C-9'), 144.6 (C-13), 144.5 (C-36), 140.7 (C-2', C7'), 140.0 (C-5'), 135.8 (C-41),132.8 (C-40), 131.2 (C-39), 129.0 (C-38), 127.3 (C-4', C6'), 121.6 (C-12), 114.2 (C-3', C8'), 80.9 (C-3), 55.3 (C-5), 47.7 (C-9), 47.6 (C-17), 47.6 (C-34), 46.3 (C-33), 43.7 (C-18), 43.6 (C-14), 41.9 (C-19), 41.3 (C-10), 39.3 (C-8), 39.1 (C-22), 38.2 (C-1), 38.1 (C-7), 37.8 (C-4), 37.7 (C-21), 36.9 (C-15), 33.0 (C-20), 30.3 (C-16), 28.0 (C-11), 27.9 (C-27), 24.0 (C-2), 23.5 (C-30), 23.4 (C-32), 21.3 (C-6), 18.2 (C-29), 16.9 (26C), 16.7 (C-23), 15.4 (C-25) ppm; IR (KBr): $\nu = 3447 s_{hr}$, 2990w, 1636w, 1584w, 1371w, 1169w cm⁻¹; UV-vis (CHCl₃): λ_{max} $(\log \epsilon) = 220.98$ (3.54), 346.19 (3.87), 484.15 (3.76), 682.69 (4.11), 746.26 (4.32) nm; MS (ESI, MeOH): m/z $(\%) = 921.73 (100\%, [M - Cl]^{-}); C_{26}H_{81}N_4O_4Cl (957.78):$ C 75.24, H 8.52, N 5.85; found: C 75.11, H 8.69, N 5.49.

N-(4-((4-dimethylamino)phenyl)-4-((4-(3β-acetyloxyurs-12en-28-carboxy)piperazin)-1-yl)oxy)carbonylphenylmethylencyclohexa-2,5-dien-1-yliden)-Nmethylmethanaminium chloride (16)

Following the procedure given for the synthesis of **11**, from **2** (200 mg, 0.49 mmol) and **14** (333 mg, 0.59 mmol) **16** (164 mg, 41%) was obtained as a green solid; R_F (CHCl₃/MeOH, 9:1) = 0.35; m.p. 212–215 °C; ¹H NMR (400 MHz, CDCl₃): $\delta = 8.01$ (d, J = 7.3 Hz, 2H, 37-H), 7.56 (d, J = 7.7 Hz, 2H, 38-H), 7.38 (d, J = 7.7 Hz, 2H, 3'-H), 7.08 (d, J = 9.2 Hz, 2H, 2'-H), 6.99 (d, J = 8.2 Hz, 2H, 7'-H), 6.61 (d, J = 9.2 Hz, 2H, 2'H, 5.25 (t, J = 3.5 Hz, 3H, 12-H), 4.46

(t, J = 8.0 Hz, 1H, 3-H), 3.74 (m, 4H, 33-H), 3.38 (s, 6H, 33-H), 3.38 (1'-NMe₂), 3.10 (q, 7.5 Hz, 5H, 34-H, 18-H), 2.15 (s, 1H, 16-H_a), 2.02 (s, 3H, 32-H), 1.96–1.46 (m, 4H, 11-H, 6-H_a, 16-H_b), 1.39 (m, 8H, 2-H, 15-H_a, 19-H, 7-H, 9-H, 21-H_a), 1.28–1.14 (m, 9H, 20-H, 6-H_b, 22-H 21-H_b, 1-H, 15-H_b, 5-H), 1.12 (s, 3H, 27-H), 0.90 (s, 3H, 30-H), 0.89 (s, 3H, 25-H), 0.85 (s, 3H, 29-H), 0.83 (s, 3H, 24-H), 0.71 (s, 3H, 23-H), 0.65 (s, 3H, 26-H) ppm; ¹³C NMR (100 MHz, CDCl₃): $\delta = 175.3$ (C-28), 171.0 (C-31), 169.2 (C-35), 157.0 (C-1', C-9'), 144.7 (C-13), 144.5 (C-7'), 140.9 (C-8'), 140.8 (C-2'), 139.4 (C-36), 134.7 (C-37), 127.2 (C-38), 121.6 (C-12), 114.1 (C-3'), 80.9 (C-3), 55.3 (C-30), 52.5 (C-18), 47.7 (C-17), 47.6 (C-9), 47.6 (C-34), 45.9 (NMe₂), 43.6 (C-33), 43.5 (C-14), 41.9 (C-8), 41.3 (9'-NMe₂), 39.1 (C-5), 38.1 (C-20), 37.6 (C-1), 36.9 (C-4), 33.9 (C-10), 33.0 (C-22), 30.4 (C-21), 29.9 (C-15), 28.0 (C-24), 27.8 (C-16), 25.9 (C-11), 24.1 (C-27), 23.5 (C-2), 23.4 (C-19), 21.3 (C-32), 18.2 (C-29), 16.9 (C-23), 16.6 (C-26), 15.4 (C-25), 8.7 (C-7), 0.9 (C-5') nm; IR (KBr): $\nu = 3442s_{hr}$, 2940m, 2678m, 1618m, 1584m, 1371m, 1248w, 1171m cm⁻¹; UV-vis (CHCl₃): λ_{max} (log ε) = 226.9 (3.9), 256.0 (4.1), 486.2 (3.98), 682 (4.21), 737.0 (4.6) nm; MS (ESI, MeOH): *m/z* (%) = 921.73 $(100\%, [M - Cl]^{-}); C_{26}H_{81}N_4O_4Cl (957.78): C 75.24, H$ 8.52, N 5.85; found: C 74.99, H 8.68, N 5.44.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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