

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 14 (2004) 1339–1342

## A series of spirocyclic analogues as potent inhibitors of bacterial phenylalanyl-*t*RNA synthetases

Xiang Y. Yu,<sup>a,\*</sup> John Finn,<sup>a</sup> Jason M. Hill,<sup>a</sup> Zhong G. Wang,<sup>a</sup> Dennis Keith,<sup>a</sup> Jared Silverman<sup>b</sup> and N. Oliver<sup>b</sup>

<sup>a</sup>Department of Medicinal Chemistry, Cubist Pharmaceuticals, Inc., 65 Hayden Ave, Lexington, MA 02421, USA <sup>b</sup>Department of Biology, Cubist Pharmaceuticals, Inc., 65 Hayden Ave, Lexington, MA 02421, USA

Received 27 March 2003; accepted 26 November 2003

**Abstract**—We have identified a series of spirocyclic furan and pyrrolidine inhibitors of *Enterococcus faecalis* and *Staphylococcus aureus* phenylalanyl-*t*RNA synthetases. The most potent analogue **1b** showed IC<sub>50</sub> = 5 nM (*E. faecalis* PheRS) and IC<sub>50</sub> = 2 nM (*S. aureus* PheRS) with high selectivity over the human enzyme. The crystal X-ray structure of analogue **1b** was determined.  $\bigcirc$  2004 Elsevier Ltd. All rights reserved.

Emergence of resistant bacteria has given new urgency to the search for new antibacterial agents that act via novel mechanisms of action.<sup>1-3</sup> Aminoacyl-tRNA synthetases are essential enzymes for biological cell growth.<sup>4–8</sup> We have previously described several series of aminoacyl-tRNA synthetase inhibitors.9-11 As part of our high throughput screening efforts identifying t-RNA synthetase inhibitors, we discovered that a series of spirocyclic compounds had good inhibition of Enterococcus faecalis and Staphylococcus aureus Phenylalanyl-tRNA synthetases (EfPheRS and SaPheRS). The lead compound **1a** inhibited EfPheRS (IC<sub>50</sub> = 0.82  $\mu$ M) and SaPheRS (IC<sub>50</sub>=0.38  $\mu$ M) with high selectivity over the human enzyme (IC<sub>50</sub> > 100  $\mu$ M). In this communication, we report on the structure-activity relationships of the series of spirocyclic analogues based on compound 1a.

The spirocyclic furan analogues were prepared by synthetic route outlined in Scheme 1. Reaction of 1,3-indandione **2** with aldehydes in the presence of piperidine produced 2-benzylidene-1,3-indandiones **3**. Treatment with 30% hydrogen peroxide in methanol formed 3-phenylspirocyclic [oxirane-2,2'-indan]-1',3'-dione **4**.<sup>12</sup> Heating the epoxide **4** gave an intermediate carbonyl ylide which underwent 1,3-dipolar cycloaddition with maleimides **5**<sup>13</sup> to give spirocyclic isomers **1** and **6–14**.<sup>14</sup>

0960-894X/\$ - see front matter  $\odot$  2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2003.11.081

In the case of the 3,4-dichlorophenyl analogue, the two isomers, cis (1a) <sup>15</sup> and trans (1b), were formed in a ratio of 2:1 and separated by silica gel chromatography (20% hexane in dichloromethane).





<sup>\*</sup> Corresponding author. Tel.: +1-781-372-4840; fax: +1-781-274-9129; e-mail: xyu@activbiotics.com

The structure of these compounds was determined by <sup>1</sup>H NMR experiments and mass spectra. X-ray crystallography of analogue **1b** (Fig. 1) further confirmed the stereochemistry.

Scheme 2 shows the preparation of spirocyclic pyrrolidine analogues. Reaction of ninhydrin 15a, phenyl glycine 16 and substituted maleimide 5a provided two isomers (17a and b),<sup>16</sup> which were separated by Florisil<sup>®</sup> chromatography. Analogue 22 was synthesized by coupling with amine 20, aldehyde 21 and substituted maleimide 5a. Because formation of carbomethoxy analogues in one step was unsuccessful, analogue 25 was prepared stepwise by generating ylide first followed by treatment with 5a.<sup>17,18</sup>

Spirocyclic analogues were evaluated for inhibition of the aminoacylation activity<sup>19</sup> of EfPheRS and SaPheRS and the results of spirocyclic furan analogues are shown in Table 1. The stereochemistry dramatically affects the activity. The *trans* spirocyclic analogues are significantly

OID

C(20

∠ cne



Ø

CIII

C(26)

C(23

0125

CI24

 $\cap$ 

CID

CI12

(122) (122) 003

more potent than the *cis* spirocyclic analogues. The *trans* 3,4-dichlorophenyl analogue **1b** is the most potent compound in this series. It inhibits EfPheRS (IC<sub>50</sub> = 5 nM) and SaPheRS (IC<sub>50</sub>=2 nM) with high selectivity over the human enzyme (IC<sub>50</sub> > 100  $\mu$ M). This trend is also evident in comparing the cis 3-chloro-4-methylphenyl analogue 6a and the trans chloro-4-methylphenyl analogue **6b**. The substitution pattern on the  $R_2$  position also affects the potency. 3,4-Methylenedioxy benzene analogue 7 has moderate activity whereas 3-(trifluoromethyl)-4-chlorophenyl analogue 9 is less active. Analogue 13 with an electron-donating methoxy group and analogue 14 with an electron-withdrawing nitro group show a significant loss in potency. Interestingly, only limited variations appear to be tolerated at the  $R_1$  position. The 4-methyl and 4-chlorophenyl analogues (8 and 10) showed a significant loss in potency. The 2-methyl-phenyl analogue 12 is ten times less active than the phenyl analogue **11**.

The results of spirocyclic pyrrolidine analogues are shown in Table 2. It is worthy to note that the *trans* spirocyclic pyrrolidine analogues were also more potent than the *cis* spirocyclic analogues. 3,4-Dichlorophenyl *cis* analogue **17a** is 100 times less active than *trans* analogue **17b**. Analogues without carbonyl groups on the spirocyclic ring (**19a** and **b**) were inactive. Benzoyl analogues **22a** and **22b** were only moderately active whereas the carbomethoxy analogue **25** was inactive.



Figure 2. WT S. aureus macromolecular labeling with PMA.



Scheme 2.

Table 1. Inhibition of EfPheRS and SaPheRS by furan analogues



Compd	$\mathbf{R}^1$	$\mathbf{R}^2$	IC <sub>50</sub> (μM) (EfPheRS)	IC <sub>50</sub> (μM) (SaPheRS)	IC <sub>50</sub> (μM) (Human PheRS)
<b>1a</b> ( <i>cis</i> )	Ph	3,4-Cl <sub>2</sub> -Ph	0.85	0.56	>100
<b>1b</b> (trans)	Ph	3,4-Cl <sub>2</sub> -Ph	0.005	0.002	>100
<b>6a</b> ( <i>cis</i> )	Ph	3-Cl-4-Me-Ph	8.3	13.7	>100
<b>6b</b> (trans)	Ph	3-Cl-4-Me-Ph	0.22	0.07	>100
7 (cis:trans = 25:1)	Ph	3,4-(OCH <sub>2</sub> O)-Ph	2.7	0.97	>100
<b>8</b> ( <i>cis</i> )	4-Me-Ph	3,4-(OCH <sub>2</sub> O)-Ph	>100	>100	>100
<b>9</b> ( <i>cis</i> )	Ph	3-CF <sub>3</sub> -4-Cl-Ph	32	45	>100
<b>10</b> ( <i>cis</i> )	4-Cl-Ph	3-CF <sub>3</sub> -4-Cl-Ph	>100	>100	>100
<b>11</b> ( <i>cis:trans</i> $=$ 18:1)	Ph	2-OMe-Ph	6.1	1.8	>100
<b>12</b> ( <i>cis</i> )	2-Me-Ph	2-OMe-Ph	57	13	>100
<b>13</b> ( <i>cis</i> )	Ph	4-OMe-Ph	>100	>100	>100
<b>14</b> ( <i>cis</i> )	Ph	4-NO <sub>2</sub> -Ph	>100	>100	>100

These spirocyclic furan and pyrrolidine compounds also exhibit *Escherichia coli* PheRS inhibitory activity. For example, the most potent analogue **1b** possesses good activity against *E. coli* PheRS ( $IC_{50} = 30$  nM).

The analogues were tested for antimicrobial activity against a panel of organisms. Analogue 1b has weak

whole cell activity (MIC=50  $\mu$ g/mL, *S. aureus* ATCC6538P). Inhibitors of tRNA synthetase activity produce a unique profile in assays monitoring macromolecular synthesis in *S. aureus*. As shown in Figure 2, pseudomonic acid (PMA),<sup>20–24</sup> a known inhibitor of isoleucine-*t*RNA synthetase, inhibits synthesis of both protein and RNA. RNA synthesis is inhibited due to

Table 2. Inhibition of EfPheRS and SaPheRS by pyrrolidine analogues



<sup>a</sup> The stereochemical center adjacent to the benzoyl or methoxy carbonyl centers was not assigned.



Figure 3. WT S. aureus macromolecular labeling with analogue 1b.

induction of the stringent response.<sup>25</sup> Induction of the stringent response is inhibited by addition of the protein synthesis inhibitor chloramphenicol (CAM), allowing RNA synthesis to continue uninhibited until the dose of PMA is increased by 10–100-fold. As shown in Figure 3, analogue **1b** also triggers the stringent response, as indicated by the altered profile of RNA synthesis inhibition in the presence of CAM. This result suggests a synthetase component to the mechanism of action.

In conclusion, a novel structural class of spirocyclic furan and pyrrolidine analogues with potent activity against *E. faecalis* and *S. aureus* PheRSs with high selectivity over the human enzyme has been discovered. These compounds also exhibit *E. coli* PheRS inhibitory activity. Further study shows that an analogue **1b** was unstable in Mueller Hinten Broth (MHB) bacterial media. The results of our efforts to improve the stability and the antibacterial activity of this series are described in the next paper.

## Acknowledgements

The authors would like to thank the Biological and Analytical staff at Cubist for enzyme assay data and spectra, Professor J. A. Golen at University of Masechusetts at Dartmouth for the crystal X-ray structure, and the staff at Bristol-Myers for <sup>1</sup>H NMR NOE experiments.

## **References and notes**

1. Fernandez-Lopez, S.; Kim, H.; Choi, E. C.; Delgado, M.; Granja, J. R.; Khasanov, A.; Kraehenbuehl, K.; Long, G.; Weinberger, D. A.; Wilcoxen, K. M.; Ghadiri, M. R. *Nature (London, U. K.)* **2001**, 452.

- 2. Frosco, M.; Barrett, J. F. Exp. Opin. Invest. Drugs 1998, 7, 175.
- 3. Sanders, C. C. Clin. Infect. Dis 1992, 14, 1089.
- 4. Davis, M. W.; Buechter, D. D.; Schimmel, P. Biochemistry 1994, 33, 9904.
- 5. Delarue, M.; Moras, D. BioEssays 1993, 15, 675.
- 6. Hou, Y. M.; Francklyn, C.; Schimmel, P. *Trends Biochem. Sci.* **1989**, *14*, 233.
- 7. Schulman, L. H.; Abelson, J. Science 1988, 240, 1591.
- 8. Warner, J. R.; Soeiro, R. N. Engl. J. Med. 1967, 12, 675.
- Yu, X. Y.; Hill, J. M.; Yu, G.; Yang, Y.; Kluge, A. F.; Keith, D.; Gallant, P.; Silverman, J.; Lim, A. *Bioorg. Med. Chem. Lett.* 2001, 11, 541.
- Yu, X. Y.; Hill, J. M.; Yu, G.; Wang, W.; Kluge, A. F.; Wendler, P.; Gallant, P. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 375.
- Hill, J. M.; Yu, G.; Shue, Y.; Zydowsky, T. M.; Rebek, J. Jr. U.S. patent, 5,726,195.
- Weigele, M.; Tengi, J. P.; De Bernardo, S.; Czajkowski, R.; Leimgruber, W. J. Org. Chem. 1976, 41, 388.
- 13. Cava, M. P.; Deana, A. A.; Muth, K.; Mitchell, M. J. *Organic Synthesis* **1961**, *41*, 93.
- 14. Krysin, M. Y.; Anokhina, I. K.; Zalukaev, L. P. Khim. Geterotsikl. Soedin 1987, 11, 1463.
- 15. Studies on the mechanism of action for cis (1a) shows that it is a competitive inhibitor with respect to phenylalanine  $(K_i=0.39 \ \mu\text{M})$  and ATP  $(K_i=0.39 \ \mu\text{M})$ . Attempts to separate the chiral racemic mixtures using various chiral columns were unsuccessful.
- Vaidyanathan, G.; Wilson, J. W. J. Org. Chem. 1989, 54, 1810.
- 17. Galley, G.; Liebscher, J.; Patzl, M. J. Org. Chem. 1995, 60, 5005.
- Hamper, B. C.; Duckesherer, D. R.; South, M. S. *Tetra*hedron Lett. **1996**, 37, 3671.
- The detailed aminoacylation assays were described in the patent: Finn, J.; Yu, X. Y.; Wang, Z.; Hill, J. M.; Keith, D.; Gallant, P.; Wendler, P. *PCT int. Appl. (2000), WO* 0018772
- Brown, P.; Davies, D. T.; O'Hanlon, P. J.; Wilson, J. M. J. Med. Chem. 1996, 39, 446.
- 21. Class, Y. J.; DeShong, P. Chem. Review 1995, 95, 1843.
- 22. Hughes, J.; Mellows, G. Biochem. J. 1980, 191, 209.
- 23. Hughes, J.; Mellows, G. J. Antibiotics 1978, 31, 330.
- 24. Hughes, J.; Mellows, G. Biochem. J. 1978, 176, 305.
- 25. Cashel, M. Encycl. Microbiol. (2nd Ed.) 2000, 4, 467.