2980, 2853, 1601, 1468, 1345, 1284, 1210, 1077, 982, 921, 778 cm⁻¹; ¹³C NMR (CDCl₃) δ 154.84, 129.87, 126.96, 118.99, 116.95, 113.21, 43.49, 40.43, 19.59; EIMS, m/z (relative intensity) 149 (75.8, M⁺), 148 (84.7), 132 (25.8), 120 (100), 91 (40.0). Anal. (C₉H₁₁NO·HCl) C, H, N.

Radiochemical Assay for PNMT Activity. The assay employed in this investigation has been described elsewhere.^{32,48} Briefly, a typical assay mixture consisted of 50 μ L of 0.5 M phosphate buffer (pH 8.0), 25 μ L of a 10 μ M solution of unlabeled AdoMet, 5 μ L of [methyl-³H]AdoMet, containing approximately 2×10^6 dpm (specific activity approximately 15 mCi/mmol), 25 μ L of substrate solution, 25 μ L of inhibitor solution (if added), 25 μ L of the enzyme preparation, and sufficient water to achieve a final volume of 250 μ L. After incubation for 30 min at 37 °C, the reaction was terminated by the addition of 250 μ L of 0.5 M borate buffer (pH 10) and extracted with 2 mL of toluene/isoamyl alcohol (7:3). The organic layer was removed and transferred to a scintillation vial and diluted with cocktail for counting. The mode of inhibition was ascertained by inspection of the 1/V vs 1/S plot of the data.

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Registry No. 12, 31404-61-2; 13, 91-21-4; 14, 102877-50-9; 14·HCl, 102879-34-5; 15, 14446-24-3; 15·HBr, 59839-23-5; 16, 30798-64-2; 16·HBr, 110192-19-3; 17, 32999-37-4; 17·HBr, 110192-20-6; 18, 103030-70-2; 18·HCl, 103030-69-9; 19, 42923-77-3; 19·HCl, 57196-62-0; 20, 43207-78-9; 21, 34146-68-4; 22, 103904-80-9; 23, 88081-58-7; 24, 103904-88-7; 25, 103904-81-0; 26, 103904-85-4; 27, 103904-89-8; 28, 2039-67-0; 29, 110192-21-7; 30, 22246-62-4; 31, 74904-29-3; 32, 55-81-2; 33, 91247-71-1; 34, 22246-04-4; 35, 2439-04-5; 36, 110192-22-8; 37, 110192-23-9; 38, 110192-24-0; PNMT, 9037-68-7.

Novel 1H-Benzimidazol-4-ols with Potent 5-Lipoxygenase Inhibitory Activity

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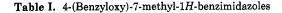
The synthesis and structure-activity profile of 2-substituted benzimidazol-4-ols as inhibitors of cell-free RBL-1 5-lipoxygenase are discussed, and their potency is compared with that of the standard inhibitors phenidone, AA 861, BW 755C, and nordihydroguaiaretic acid. In contrast to the standard compounds, most did not inhibit the release of slow-reacting substance of anaphylaxis (SRS-A) in vivo when administered at 200 μ M ip to rats subjected to peritoneal anaphylaxis, although five compounds containing a methoxylated benzyl group (compounds 36, 39, 42, and 43) or hydroxylated benzyl group (41) showed similar activity to that of phenidone, nordihydroguaiaretic acid, and AA 861. Of the many compounds tested, two, 5-tert-butyl-7-methyl-2-(trifluoromethyl)-1H-benzimidazol-4-ol (57) and 2-(4-methoxybenzyl)-7-methyl-1H-benzimidazol-4-ol (36), like dexamethasone, inhibited monocyte accumulation in a pleural exudate model of inflammation. Standard lipoxygenase inhibitors such as phenidone, BW 755C, and AA 861 were inactive in this system.

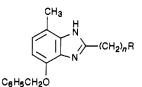
Oxidative metabolites of arachidonic acid and their subsequent derivatives have been implicated in the pathology of a variety of inflammatory and allergic diseases,¹ and modulation of their formation has attracted considerable attention.² While the early work of Vane and others³ showed that the nonsteroidal antiinflammatory drugs owed their activity to the inhibition of cyclooxygenase and the consequent reduction in the formation of thromboxanes and prostaglandins, little interest was shown in other oxidative pathways until the late 1970s. Indeed, it was the characterization of the slow-reacting substance of anaphylaxis (SRS-A), a potent bronchoconstrictor, as a mixture of the leukotrienes LTC_4 , LTD_4 , and LTE_4^1 and the identification of LTB_4 , a potent chemotaxin for mononuclear cells⁴ and polymorphonuclear cells,⁵ that focused attention on the 5-lipoxygenase pathway of arachidonic acid metabolism and the range of products available through it. It was this increased awareness of the breadth of the arachidonic acid cascade and the enzymes involved that led to the development of inhibitors of leukotriene synthesis. Early inhibitors such as phenidone⁶ and BW $755C^7$ were shown to exhibit activity on both the major oxidative pathways, although subsequent work has identified an ever-increasing number of more selective inhibitors of 5-lipoxygenase.⁸ Few of these, however, have shown useful in vivo activity in potentially relevant animal models of allergic or inflammatory disease.⁹

- Samuelson, B. Science (Washington, D.C.) 1983, 220, 568.
 See, for example: Musser, J. H.; Kreft, A. F.; Lewis, A. J.
- Annu. Rep. Med. Chem. 1985, 20, 71. (3) Vane, J. R. Nature (London) New Biol. 1971, 231, 232.
- (4) Higgs, G. A.; Mugridge, K. G. Adv. Prostaglandin, Thromb-
- oxane, Leukotriene Res. 1983, 12, 19. (5) Ford-Hutchinson, A. W.; Bray, M. A.; Doig, M. V.; Shipley, M.
- E.; Smith, M. J. M. Nature (London) 1980, 286, 2646.
 (6) (a) Blackwell, G. J.; Flower, R. J. Prostaglandins 1978, 16, 417.
 (b) Blackwell, G. J.; Flower, R. J. Br. J. Pharmacol. 1978, 63,
- (7) Higgs, G. A.; Flower, R. J.; Vane, J. R. Biochem. Pharmacol.
- (1) Higgs, G. A.; Flower, R. S.; Valle, S. R. Biochem. 1 narmatol.
 (1) 1979, 28, 1959.
 (2) (1) Niceleux K. C. Beterie, N. A. Seite, S. B. J. Cham. Sec.
- (8) (a) Nicolaou, K. C.; Petasis, N. A.; Seitz, S. P. J. Chem. Soc., Chem. Commun. 1981, 1195. (b) Corey, E. J.; Kang, J. Tetrahedron Lett. 1982, 23, 1651. (c) Yoshimoto, T.; Furukawa, M.; Yamamoto, S.; Horie, T.; Watanabe-Kohno, S. Biochem. Biophys. Res. Commun. 1983, 116, 612. (d) Sun, F. F.; McGuire, J. C.; Prostaglandins 1983, 26, 211. (e) Corey, E. J.; Cashman, J. R.; Kantner, S. S.; Wright, S. W. J. Am. Chem. Soc. 1984, 106, 1503. (f) Ashida, Y.; Saijo, T.; Kuriki, H.; Makino, H.; Terao, S.; Maki, Y. Prostaglandins 1983, 26, 955.

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[‡] Medicinal Research Centre.

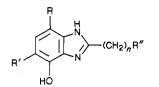




no.	R	n	method ^a	yield, %	crystn solvent	mp, °C	formula	anal.
10	C ₆ H ₅	0	B	50	xylene	221-223	C ₂₁ H ₁₈ N ₂ O	C, H, N
11	C_6H_5	1	Α	55	EtOAc-hexane	198 - 200	$C_{22}H_{20}N_{2}O$	C, H, N
12	C_6H_5	2	Α	65	EtOH	203 - 204	$C_{23}H_{22}N_2O$	C, H, N
13	4-CH ₃ C ₆ H ₄	1	Α	67	EtOAc	184-186	$C_{23}H_{22}N_{2}O$	C, H, N
14	4-CH ₃ OC ₆ H ₄	Ó	Α	64	EtOAc	223 - 225	$C_{22}H_{22}N_2O_2$	C, H, N
15	4-CH ₃ OC ₆ H ₄	1	Α	56	EtOAc	181 - 183	$C_{23}H_{22}N_2O_2$	C, H, N
16	3-CH ₃ OC ₆ H ₄	1	Α	80	EtOAc	144 - 145	$C_{23}H_{22}N_2O_2$	C, H, N
17	$2-CH_{3}OC_{6}H_{4}$	1	Α	73	EtOAc	174 - 176	$C_{23}H_{22}N_2O_2$	C, H, N
18	$4 - C_2 H_5 O C_6 H_4$	1	Α	72	$EtOAc-Et_2O$	161 - 162	$C_{24}H_{24}N_2O_2$	C, H, N
19	4-HOČ ₆ H ₄	1	Α	22	Me ₂ CO	121 - 123	$C_{22}H_{20}N_2O_2 \cdot 0.5C_2H_5OH$	C, H, N
20	$3,4-(CH_{3}O)_{2}C_{6}H_{3}$	1	Α	71	EtŐAc	140 - 143	$C_{24}H_{24}N_2O_3$	C, H, N
21	3,4,5-(CH ₃ O) ₃ C ₆ H ₂	1	Α	86	EtOAc	165 - 167	$C_{25}H_{26}N_{2}O_{4}$	C, H, N
22	3-CF ₃ C ₆ H ₄	0	В	31 ^b	-	166 - 167	$C_{22}H_{17}F_{3}N_{2}O$	C, H, N
23	$4-N(CH_3)_2C_6H_4$	0	В	52	-	260 - 262	$C_{23}H_{23}N_{3}O$	
24	$1 - C_{10}H_7$	1	Α	71	EtOAc	171 - 172	$C_{26}H_{22}N_2O$	C, H, N
25	$2 - C_{10} H_7$	1	Α	62	EtOAc	172 - 174	$C_{26}H_{22}N_{2}O$	C, H, N
26	2-furyl	0	В	91°	-		$C_{19}H_{16}N_2O_2$	
27	2-thienyl	1	Α	52	EtOAc	194-196	$C_{20}H_{18}N_{2}OS$	H, N, C^d
28	3-pyridyl	.1	A	е			$C_{21}H_{19}N_{3}O$	

^aSee text and Experimental Section. ^bPurified by column chromatography; CH_2Cl_2 on SiO₂. ^cCrude yield after partial chromatographic purification; CH_2Cl_2 on SiO₂. ^dC: calcd, 71.82; found, 71.37. ^eNot isolated.

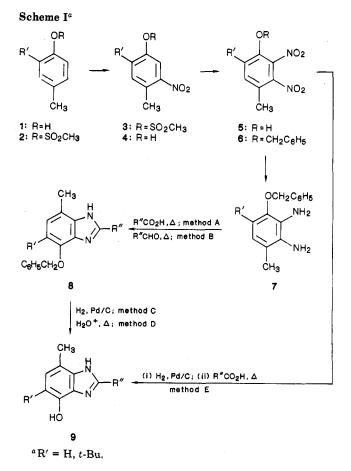
In this paper we present data relating to a series of 1H-benzimidazol-4-ols I with potent 5-lipoxygenase activity in vitro. Several compounds have been shown to inhibit



I: R, R'=H, alkyl; R"=H, CF3, aryl, heteroaryl; n=0-3

the release of SRS-A in vivo in a rat model of immediate hypersensitivity. Furthermore, representatives of the series have shown the ability to inhibit the accumulation of monocytes in a rat pleural exudate model of inflammation. The monocyte is thought to play a key role in chronic inflammation through its interactions with other cells and the release of soluble mediators such as interleukin 1.¹⁰ Compounds such as the 1*H*-benzimidazol-4-ols that inhibit SRS-A release and monocyte accumulation could prove useful therapy in allergic and inflammatory conditions in humans.

Chemistry. Benzimidazoles were prepared by the classic route¹¹ from the appropriate *o*-phenylenediamines as shown in Scheme I. Thus, prior mesylation of the readily available phenols 1 to give 2 allows nitration to proceed at the meta position to give good yields of the mononitro derivatives 3. Alkaline hydrolysis of the mesyl protecting group and further nitration result in the predominant formation of the ortho dinitro compounds 5. Further protection of the phenolic moiety as its benzyl ether 6 is generally essential at this stage in order to



prevent competitive cyclization to benzoxazoles in subsequent steps.¹² This is not a requirement, however, when the C-6 substituent (of 6) is a sterically demanding group such as *tert*-butyl, as shown by the synthesis of compounds

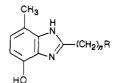
⁽⁹⁾ Spicer, B. A.; Laycock, S. M.; Smith, H. Agents Actions 1985, 17, 498.

⁽¹⁰⁾ Krane, S. M.; Dayer, J. M.; Simon, L. S.; Byrne, M. S. Collagen Relat. Res. 1985, 5, 99.

⁽¹¹⁾ Grimmett, M. R. In Comprehensive Heterocyclic Chemistry; Katritsky, A. R., Rees, C. W., Eds.; Pergamon: Oxford, 1984; Vol. 5, p 457.

 ⁽¹²⁾ Cornforth, J. W. In *Heterocyclic Compounds*; Elderfield, R. C., Ed.; Wiley: London, 1957, p 418.

Table II. 7-Methyl-1H-benzimidazol-4-ols



									inhibn of RBL-1
no.	R	n	methodª	yield, %	crystn solvent	mp, °C	formula	anal.	5-lipoxygenase: IC ₅₀ , ^b μM
29	C_6H_5	0	С	73	EtOH-Et ₂ O	252-256	$C_{14}H_{12}N_2O\cdot HCl$	C, H, N, Cl	1.2(1.0-1.5)
30	C_6H_5	1	С	85	$EtOH-Et_2O$	204 - 205	$C_{15}H_{14}N_2O$	C, H, N	1.2(0.4-2.2)
31°	C_6H_5	1	\mathbf{E}	67	CHCl ₃ -EtOAc~pentane	226 - 228	$C_{19}H_{22}N_2O$	C, H, N	1.3(1.1-1.6)
32	C_6H_5	2	С	86	EtOH-Et ₂ O	238 - 240	$C_{16}H_{16}N_2O\cdot HCl$	C, H, N	2.1(1.3-2.8)
33	$4 - CH_3C_6H_4$	1	С	19	EtOH	217 - 218	$C_{16}H_{16}N_2O$	C, H, N	2.54(2.46 - 2.63)
34	4-ClC ₆ H ₄	1	-	67	$EtOH-Et_2O$	275 - 277	C ₁₅ H ₁₃ ClN ₂ O·HBr	C, H, N	1.0(0.3-1.6)
35	$4 - CH_3OC_6H_4$	0	С	51	EtOH	283 - 285		C, H, N, Cl	2.2(1.4-3.2)
							C ₂ H ₅ OH		
36	$4-CH_3OC_6H_4$	1	С	87	EtOH	232 - 235	C ₁₆ H ₁₆ N ₂ O ₂ ·HCl	C, H, N, Cl	2.1(1.7-2.6)
37^{c}	4-CH ₃ OC ₆ H ₄	1	\mathbf{E}	38	EtOH-Et ₂ O	232 - 235	C ₂₀ H ₂₄ N ₂ O ₂ ·HCl	C, H, N, Cl	2.2(0.9-4.0)
38	3-CH ₃ OC ₆ H ₄	1	С	48	EtOH	239 - 241		C, H, N, Cl	1.7(1.0-2.4)
	-						C ₂ H ₅ OH		
39	$2-CH_3OC_6H_4$	1	С	54	EtOH	244 - 245	C ₁₆ H ₁₆ N ₂ O ₂ ·HCl	C, H, N	3.4(3.3-3.5)
40	$4-C_2H_5OC_6H_4$	1	С	88	EtOH	227 - 229	C ₁₇ H ₁₈ N ₂ O ₂ ·HCl	C, H, N, Cl	3.0(2.3 - 3.8)
41	4-HOC ₆ H ₄	1	С	32	EtOH	273 - 275	$C_{15}H_{14}N_2O_2HCl$	C, H, N	6.9(4.9-11.6)
42	$3,4-(CH_3O)_2C_6H_3$	1	С	52	EtOH	264 - 265	C ₁₇ H ₁₈ N ₂ O ₃ ·HCl	C, H, N, Cl	5.2(3.8-7.1)
43	3,4,5-(CH ₃ O) ₃ C ₆ H ₂	1	\mathbf{C}	87	EtOH	250 - 251	C ₁₈ H ₂₀ N ₂ O ₄ ·HCl	H, N, Cl, C^d	9.7 (7.5-11.8)
44	3-CF ₃ C ₆ H ₄	0	С	59	$EtOH-H_2O$	>280	$C_{15}H_{11}F_3N_2O\cdot HCl$	C, H, N, Cl	е
45	$4 - N(CH_3)_2 C_6 H_4$	0	С	82	EtOH	194	C ₁₆ H ₁₇ N ₃ O·HCl	C, H, Cl, N^{f}	1.3(1.0-1.7)
46	$1-C_{10}H_7$	1	С	69	EtOH	262 - 263	$C_{19}H_{16}N_2O \cdot HCl$	C, H, N, Cl^g	2.3(1.8-2.8)
47	$2 - C_{10} H_7$	1	С	92	EtOH	160 - 165	$C_{19}H_{16}N_2O \cdot HCl$	C, H, N, Cl	e
48	2-furyl	0	С	30	EtOH	256 - 258	$C_{12}H_{10}N_2O_2 \cdot 1.1$	C, H, N, Cl	3.3(1.8-5.7)
							HCl-0.5C ₂ H ₅ OH		
49	2-thienyl	1	D	98	-	206 - 208	$C_{13}H_{12}N_2OS \cdot HCI$	C, H, N, Cl, S	2.1 (1.9 - 2.4)
50	3-pyridyl	1	D	4	MeOH-EtOAc-Et ₂ O	264 - 268	C ₁₄ H ₁₃ N ₃ O	C, H, N	5.0 (4.0-6.4)

^aSee text and Experimental Section. ^bIC₅₀ values for standard compounds: NDGA, 0.14 (0.03–0.29) μ M; BW 755C, 0.89 (0.72–1.07) μ M; AA 861, 2.0 (1.9–2.1) μ M; phenidone, 2.1 (1.07–5.54) μ M; 95% confidence limits in brackets. ^c5-C(CH₃)₃. ^dC: calcd, 57.14; found, 57.65. ^eInhibition >90% at 20 μ M; IC₅₀ not determined. ^fN: calcd, 13.83; found, 13.40. ^gCl: calcd, 10.92; found, 11.49.

Table III. Ra	t Passive	Peritoneal	Anaphylaxis	Data
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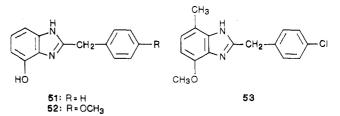
compound	% inhibn \pm SEM of SRS-A release at compd concn of 2×10^{-4} M ip $(n)^a$
36	$58 \pm 8^{**}$ (6)
39	$64 \pm 12^{**}$ (7)
41	$58 \pm 6^{***}$ (6)
42	$55 \pm 12^{***}$ (7)
43	$59 \pm 7^{***}$ (6)
phenidone	$70 \pm 8^{***}$ (7)
AA 861	$77 \pm 5^{***}$ (7)
BW 755C	$25 \pm 15^*$ (14)
NDGA	$64 \pm 9^{***}$ (21)

^a Asterisks denote differences from controls as assessed by Student's t test; (*) p < 0.05, (**) p < 0.01, (***) p < 0.001; n = number of animals used. Concentrations are those in the 6 mL injected. Compounds 29, 30, 32-35, 38, 40, 45, 46, 48, 49, 51, 52, 56, and 57 were tested and shown to have no effect on the release of SRS-A at this dose.

31 and 37 (method E). Raney nickel reduction of the protected dinitro derivatives 6 then proceeds well to give the requisite diamines 7, which were generally used without further purification. The transformation of 7 to the required benzimidazol-4-ols may be effected by either of two standard routes: by the reaction with a suitable carboxylic acid in the absence of solvent at elevated temperatures (method A) or, alternatively, with an aryl carboxaldehyde in xylene at reflux (method B). Moderate to good yields of the protected benzimidazol-4-ols 8 (Table I) are obtained via either route. Reductive or hydrolytic cleavage of the O-benzyl groups from 8 (methods C and D respectively) gives the unprotected benzimidazol-4-ols 9 (Table II).

Results and Discussion

Early studies with the first member of the series, 30, showed it to be a potent inhibitor of 5-lipoxygenase in vitro (IC₅₀ = 1.2 μ M). In contrast it was a poor inhibitor of bovine seminal vesicle cyclooxygenase, having little activity at 20 μ M.¹³ The interesting activity found with 30 prompted a program in which compounds bearing a variety of substituents in the benzimidazole ring were prepared (Table II). A free hydroxyl group appeared essential for good activity (cf. 30 with 15, 25, and 53, which had IC₅₀ values in excess of 20 μ M), as did the presence of an alkyl group at C-7 (cf. 30 with 51 and 52, which have IC₅₀ values of ca. 20 μ M).



Similar in vitro potency was observed when the C-2 substituent contained an aromatic ring either directly bonded to the imidazole nucleus or one or two carbon atoms removed from it (cf. 29, 30, and 32). In vivo results suggested that the C-2 benzyl moieties were preferred. With the possible exception of the 4-hydroxy and trimethoxy compounds, 41 and 43 respectively, there was a

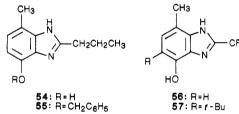
(13) Thody, V. E., unpublished results.

Table IV. Rat Pleural Exudate Model

			% inhibition ^c		
compound ^a	dose, mg/kg po	n ^b	DNA content	exudate volume	
57	24	10	38*	48*	
	12	10	57***	23	
	6	10	57***	49*	
	3	10	54**	52**	
dexamethasone	45 μg	10	71***	80***	
36	12	9	33*	35*	
dexamethasone	45 μg	10	59***	56***	

^a Compounds 29-32, 37, 42-45, and 47 were tested and shown to be inactive. ^b Number of animals used. ^cAsterisks denote differences from controls as assessed by Student's t test; (*) p < 0.05, (**) p < 0.01. (***) p < 0.001.

tendency for similar potency to be shown in vitro regardless of the nature of the aromatic substitution at C-2 (cf. **30**, **33**, **34**, **36**, and **46**). Potency was also retained by replacement of the benzenoid ring at C-2 by both thiophene, as in **49**, and pyridine, as in **50**. However, complete removal of the aromatic appendage and substitution by *n*-propyl or trifluoromethyl (compounds **54** and **56** respectively) resulted in a reduction in potency (IC₅₀ ~ 10 μ M). Interestingly, while introduction of a *tert*-butyl group at C-5 had little effect on potency in compounds **31** and **37**, the 5-*tert*-butyl homologue **57** of the trifluoromethyl compound **56** was the most potent inhibitor of this series, with IC₅₀ = 0.25 (0.18–0.31) μ M, comparable to that of nordihydroguaiaretic acid (NDGA), the most potent of the standard compounds in this screen (Table II).



Significant differences between the more potent members of the series were seen in vivo, with only a narrow band of compounds (Table III) showing activity against the release of SRS-A in a rat passive peritoneal model of anaphylaxis using homocytotropic antibody.¹⁴ Thus, the 2- and 4-methoxybenzyl derivatives, 39 and 36 respectively, significantly inhibited the release of SRS-A when administered at 200 μ M ip while two close analogues, the 3methoxy (38) and 4-ethoxy (40) compounds, did not show in vivo activity. It is of some interest that the 4-hydroxy compound 41 and the two multiply methoxylated analogues 42 and 43 were also active whereas the (methoxyphenyl)benzimidazol-4-ol 35 was inactive at the doses tested. Phenidone, NDGA and AA 861 show similar potency when administered ip in this model, but BW 755C was markedly less potent (Table III).

Compound 36 and the trifluoromethyl analogue 57 were also active in the 72-h pleural exudate model given orally, showing the same profile as dexamethasone (Table IV), although all other compounds tested were inactive. At this time point the monocyte is the predominant cell type.^{15,16} Cyclooxygenase inhibitors such as indomethacin and the mixed cyclooxygenase/lipoxygenase inhibitors such as phenidone¹⁷ and BW 755C¹⁸ have been shown to exacer-

(16) Mangan, F. R., et al., unpublished results.

bate the cellular reaction measured at 72 h post carrageenan injection.¹⁹ The selective 5-lipoxygenase inhibitor AA 861⁸⁶ has been shown to be inactive in this model.¹⁹ The low number of compounds active in the passive peritoneal anaphylaxis model that are also active in the pleural model may, of course, simply reflect the absence of oral absorption or different metabolism or pharmacokinetics, following oral administration in the latter system. (The compounds are given intraperitoneally in the anaphylaxis model and are thus immediately at the site of the inflammatory response.) Compound 57, however, proved active in the pleural model in spite of being inactive in the anaphylaxis system. This may suggest that activities of some of these compounds other than inhibition of 5-lipoxygenase may play a role in reducing monocyte accumulation.

While the importance of 5-lipoxygenase products in allergic disease and chronic inflammation must await the successful progression and evaluation of a selective inhibitor in humans, the design of relevant animal models remains a major priority. The compounds described herein might be of value as pharmacological tools in the development of such animal models.

Experimental Section

Melting points were taken on a Büchi melting point apparatus and are uncorrected. ¹H NMR spectra were recorded on a JEOL JMN-GX270 270-MHz spectrometer using Me₄Si as internal standard. All compounds had spectra consistent with their assigned structures and had elemental analyses within $\pm 0.4\%$ of the calculated values except where noted.

2-tert-Butyl-4-methyl-5-nitrophenyl Methanesulfonate (3) $[\mathbf{R}' = \mathbf{C}(\mathbf{CH}_3)_3]$. Mesyl chloride (147 g, 1.28 mol) was added dropwise over 30 min to a solution of 2-tert-butyl-4-methylphenol (1) $[\mathbf{R}' = \mathbf{C}(\mathbf{CH}_3)_3]$ (200 g, 1.22 mol) in pyridine (500 mL) at 0 °C. The solution was stirred at 10 °C for 1 h and room temperature for 3 h. It was poured into excess 5 M HCl and extracted with CH₂Cl₂. The organic layer was washed with 5 M HCl and H₂O and dried (Na₂SO₄). Evaporation to dryness in vacuo gave crude 2-tert-butyl-4-methylphenyl methanesulfonate (2) $[\mathbf{R}' = C(\mathbf{CH}_3)_3]$ (250 g, 1.03 mol), which was dissolved in concentrated H₂SO₄ (750 mL) and cooled to 0 °C. A mixture of concentrated HNO₃ (93 g, 1.03 mol) and concentrated H₂SO₄ (175 mL) was added dropwise over 1 h at 0 °C, and the solution was poured onto ice. The resulting solid was collected, washed with H₂O, and dried to afford 260 g (74%) of 3 $[\mathbf{R}' = C(\mathbf{CH}_3)_3]$: mp 91–92 °C (CHCl₃-pentane); ¹H NMR (CDCl₃) δ 1.4 (9 H, s, CH₃), 2.55 (3 H, s, Ar CH₃), 3.30 (3 H, s, SO₂CH₃), 7.40 (1 H, s), 8.15 (1 H, s). Anal. (C₁₂H₁₇NO₅S) C, H, N.

2-tert-Butyl-4-methyl-5-nitrophenol (4) $[\mathbf{R}' = \mathbf{C}(\mathbf{CH}_3)_3]$. A suspension of 3 $[\mathbf{R}' = \mathbf{C}(\mathbf{CH}_3)_3]$ (260 g, 0.906 mol) in 4.06 M aqueous NaOH (800 mL) was stirred at 60–70 °C for 1.5 h. The solution was filtered and poured onto excess iced 5 M HCl, and the mixture was extracted with $\mathbf{CH}_2\mathbf{Cl}_2$. The organic layer was washed (H₂O), dried (Na₂SO₄), and evaporated to dryness to give 140 g (74%) of 4 $[\mathbf{R}' = \mathbf{C}(\mathbf{CH}_3)_3]$: mp 96–98 °C ($\mathbf{CH}_2\mathbf{Cl}_2$ -pentane); ¹H NMR (\mathbf{CDCl}_3) δ 1.40 (9 H, s, \mathbf{CH}_3), 2.50 (3 H, s, $\mathbf{Ar} \mathbf{CH}_3$), 5.20 (1 H, br, exchangeable OH), 7.20 (1 H, s), 7.43 (1 H, s). Anal. ($\mathbf{C}_{11}\mathbf{H}_{15}\mathbf{NO}_3$) C, H, N.

6-tert-Butyl-2,3-dinitro-4-methylphenol (5) $[R' = C(CH_3)_3]$. A solution of fuming HNO₃ (10 mL, 0.22 mol) in glacial AcOH (15 mL) was added dropwise over 15 min to a solution of 4 $[R' = C(CH_3)_3]$ (35 g, 0.17 mol) in AcOH (60 mL), with a temperature of 18–20 °C maintained. After 10 min, the solution was poured onto ice and the solid was collected, washed with H₂O, dried (Na₂SO₄), and evaporated in vacuo. The residue was separated

⁽¹⁴⁾ Ross, J. W.; Smith, H.; Spicer, B. A. Int. Arch. Allergy Appl. Immunol 1976, 51, 226.

 ⁽¹⁵⁾ Ackerman, N.; Tomolomis, A.; Miram, L.; Kheifets, J.; Martinez, S.; Carter, A. J. Pharmacol. Exp. Ther. 1980, 215, 588.
 (16) Manzan E. D. et al. manufacture in the interval of the second s

⁽¹⁷⁾ Foster, S. J.; McCormick, M. E.; Howarth, A. Agents Actions 1985, 17, 358.

⁽¹⁸⁾ Salmon, J. A.; Simmons, P. M.; Moncada, S. J. Pharm. Pharmacol. 1983, 35, 808.

⁽¹⁹⁾ Thomson, M. J.; Bermudez, J.; Mangan, F. R., unpublished results.

by column chromatography on silica eluting with CH₂Cl₂ to afford 25 g (59%) of the title compound after recrystallization from CH₂Cl₂-pentane: mp 104-105 °C; ¹H NMR (CDCl₃) δ 1.45 (9 H, s, CH₃), 2.30 (3 H, s, Ar CH₃), 7.50 (1 H, s, H-6), 11.20 (1 H, s, exchangeable OH). Anal. (C₁₁H₁₄N₂O₅) C, H, N.

4-(Benzyloxy)-2,3-dinitrotoluene (6) (R' = H). A mixture of 4-methyl-2,3-dinitrophenol²⁰ (12 g, 0.04 mol), anhydrous K₂CO₃ (15 g, 0.11 mol), potassium iodide (0.1 g), and benzyl chloride (25 g, 0.2 mol) in acetone (150 mL) was stirred under reflux for 18 h. The cooled mixture was filtered and the filtrate evaporated to dryness in vacuo. The residue was taken up in CHCl₃ (400 mL), washed with H₂O, and dried (Na₂SO₄). The solution was evaporated to half-volume in vacuo, pentane (150 mL) added, and the title compound (16.2 g, 93%) recovered by filtration: mp 116–118 °C (CHCl₃-hexane); ¹H NMR (CDCl₃) δ 2.30 (3 H, s, CH₃), 5.10 (2 H, s, CH₂), 7.03 (1 H, d, J = 8 Hz, H-5), 7.20 (5 H, m, benzyl aromatics), 7.26 (1 H, d, J = 8 Hz, H-6). Anal. (C₁₄H₁₂N₂O₅) C, H, N.

3-(Benzyloxy)-6-methyl-1,2-phenylenediamine (7) ($\mathbf{R}' = \mathbf{H}$). A solution of 6 ($\mathbf{R}' = \mathbf{H}$, 7 g, 0.03 mol) in EtOH (300 mL) was hydrogenated over Raney nickel (3 g) at room temperature and pressure until hydrogen uptake ceased. The catalyst was removed by filtration through Celite and the filtrate evaporated to dryness in vacuo to afford the title compound as an oil (5.07 g, 91%): ¹H NMR (CDCl₃) δ 2.10 (3 H, s, CH₃), 3.36 (4 H, br, exchangeable NH₂), 4.96 (2 H, s, CH₂), 6.38 (1 H, d, J = 8 Hz, H-5), 6.50 (1 H, d, J = 8 Hz, H-6), 7.30 (5 H, m, benzyl aromatics).

2,3-Dinitrophenol Benzyl Ether. The title compound was prepared from 2,3-dinitrophenol²¹ in 90% yield by a procedure identical with that used for the preparation of **6** and crystallized from CHCl₃-hexane: mp 100–101 °C. Anal. ($C_{13}H_{10}N_2O_5$) C, H, N.

2,3-Diaminophenol Benzyl Ether. The title compound was prepared in quantitative yield by catalytic hydrogenation of the dinitro analogue over Raney nickel, in a way similar to that used for the preparation of 7, and used without purification.

4-(Benzyloxy)-1*H*-benzimidazoles (Table I). Method A. The diamine 7 (R' = H) and an appropriately substituted carboxylic acid (3-6 equiv) were heated together at 120-130 °C for 1-6 h with rapid stirring. After cooling, the mixture was taken up into CH₂Cl₂ and washed successively with 10% aqueous Na₂CO₃ (twice) and H₂O. The organic phase was dried (MgSO₄) and evaporated to dryness in vacuo to afford the crude 4-(benzyloxy)-1*H*-benzimidazole, which was purified by recrystallization: ¹H NMR (DMSO-d₆) δ 2.36-2.50 (3 H, s, CH₃), 5.12-5.32 (2 H, s, OCH₂), 6.63-6.74 (1 H, d, J = 8 Hz, H-5), 6.80-6.91 (1 H, d, J = 8 Hz, H-6), 7.31-7.51 (5 H, m, benzyl aromatics), 12.33-13.50 (1 H, br, exchangeable NH).

Method B. A mixture of the diamine 7 (R' = H) and an appropriately substituted benzaldehyde (2–3 equiv) in xylene was heated under reflux for 3–6 h and allowed to cool overnight. The resulting solid was collected by filtration, washed with Et₂O, and recrystallized from a suitable solvent to afford the 4-(benzyl-oxy)-1*H*-benzimidazole in moderate yield.

1*H*-Benzimidazol-4-ols (Table II). Method C. A solution of the 4-(benzyloxy)-1*H*-benzimidazole in EtOH was hydrogenated over 10% palladium on charcoal at room temperature and pressure until hydrogen uptake ceased. The catalyst was removed by filtration through Kieselguhr and washed with EtOH and the filtrate evaporated to dryness in vacuo. Crystallization of the resulting solid afforded the expected product in high yield.

Alternatively the crude product could be converted into its hydrochloride salt with ethanolic hydrogen chloride and this salt recrystallized from a suitable solvent: ¹H NMR (DMSO- d_6) δ 2.47–2.61 (3 H, s, CH₃), 6.84–6.92 (1 H, d, J = 8 Hz, H-5), 7.06–7.12 (1 H, d, J = 8 Hz, H-6), 10.72–10.80 (1 H, br s, exchangeable OH), 13.00–16.50 (2 H, br s, exchangeable NH₂⁺).

Method D. The 4-(benzyloxy)-1*H*-benzimidazole was heated under reflux in 5 M aqueous HCl for 24 h, the solution allowed to cool, and the resulting precipitate collected by filtration. The solid was washed with H_2O and dried in vacuo to give the essentially pure 1*H*-benzimidazol-4-ol as its hydrochloride salt in high yield. Recrystallization where necessary afforded material of analytical purity.

5-tert-Butyl-1H-benzimidazol-4-ols (Table II). Method E. The dinitrophenol 5 $[R' = C(CH_3)_3]$ in EtOH was hydrogenated over 10% palladium on charcoal at room temperature and pressure to afford the corresponding diamino compound, which was used without purification. This diamine and a suitable carboxylic acid (4 equiv) were heated together at 120–130 °C under nitrogen for 1–2 h, and the mixture was cooled and extracted with CHCl₃. The organic solution was washed with 10% aqueous Na₂CO₃ and H₂O and dried (Na₂SO₄). Evaporation of the solvent in vacuo afforded the expected benzimidazol-4-ol, which was purified by recrystallization.

2-(4-Chlorobenzyl)-7-methyl-1*H*-benzimidazol-4-ol Hydrobromide (34). A suspension of 53 (4.0 g, 14 mmol) in 48% aqueous HBr (400 mL) under nitrogen was heated at 130 °C for 5 h, cooled, and evaporated in vacuo. The residue was crystallized from EtOH-Et₂O to give 3.95 g (80%) of 34 as a white solid: mp 275-277 °C; ¹H NMR (DMSO- d_6) δ 2.45 (3 H, s, CH₃), 4.50 (2 H, s, CH₂), 6.85 (1 H, d, J = 8 Hz, H-5), 7.10 (1 H, d, J = 8 Hz, H-6), 7.50 (4 H, s, aromatic H), 10.60 (1 H, br, exchangeable OH). Anal. (C₁₅H₁₃ClN₂O-HBr) C, H, N.

2-(Phenylmethyl)-1*H*-benzimidazol-4-ol (51). The title compound was prepared in two steps from 3-(benzyloxy)-1,2-phenylenediamine and phenylacetic acid by using methods A and C: mp 172-174 °C (EtOH-Et₂O). Anal. ($C_{14}H_{12}N_{2}O$) C, H, N.

2-[(4-Methoxyphenyl)methyl]-1*H*-benzimidazol-4-ol Hydrochloride (52). The title compound was prepared in two steps from 3-(benzyloxy)-1,2-phenylenediamine and 4-methoxyphenylacetic acid by using methods A and C: mp 194-196 °C (MeOH-Et₂O). Anal. ($C_{15}H_{14}N_2O_2$ ·HCl) C, H, N.

2-[(4-Chlorophenyl)methyl]-4-methoxy-7-methyl-1Hbenzimidazole (53). The title compound was prepared in 67% yield according to method A from 3-methoxy-6-methyl-1,2phenylenediamine²⁰ and 4-chlorophenylacetic acid: mp 195–197 °C (CHCl₃-pentane); ¹H NMR (DMSO- d_6) δ 2.35 (3 H, s, CH₃), 3.85 (3 H, s, OCH₃), 4.12 (2 H, s, CH₂), 6.55 (1 H, d, J = 8 Hz, H-5), 6.85 (1 H, d, J = 8 Hz, H-6), 7.35 (4 H, s, aromatic H), 12.35 (1 H, br, exchangeable NH). Anal. (C₁₆H₁₅ClN₂O) C, H, N, Cl.

4-(Benzyloxy)-7-methyl-2-n-propyl-1H-benzimidazole (55). The title compound was prepared in 46% yield by the reaction of 7 (R' = H) with butyric acid according to method A: mp 192-194 °C (EtOH-EtOAc); ¹H NMR (CHCl₃ + DMSO-d₆) δ 0.91 (3 H, t, J = 8 Hz, CH₂CH₃), 1.75 (2 H, m, CH₂CH₂CH₃), 2.45 (3 H, s, Ar CH₃), 2.80 (2 H, t, J. = 8 Hz, CH₂CH₂CH₃), 5.15 (2 H, s, OCH₂), 6.60 (1 H, d, J = 8 Hz, H-5), 6.85 (1 H, d, J = 8 Hz, H-6), 7.30 (5 H, m, benzyl aromatics). Anal. (C₁₈H₂₀N₂O) C, H, N.

7-Methyl-2-*n***-propyl-1***H***-benzimidazol-4-ol Hydrochloride (54). The title compound was prepared from 55 in 64% yield by using method C: mp 204–206 °C (EtOH–EtOAc–Et₂O); ¹H NMR (DMSO-d_6) \delta 0.95 (3 H, t, J = 8 Hz, CH₂CH₃), 1.90 (2 H, m, CH₂CH₂CH₃), 2.50 (3 H, s, Ar CH₃), 3.10 (2 H, t, J = 8 Hz, CH₂CH₂CH₃), 6.85 (1 H, d, J = 8 Hz, H-5), 7.05 (1 H, d, J = 8 Hz, H-6), 10.80 (1 H, br, exchangeable OH). Anal. (C₁₁N₁₄N₂-O-HCl) C, H, N, Cl.**

7-Methyl-2-(trifluoromethyl)-1H-benzimidazol-4-ol Hydrochloride (56). The diamine 7 ($\mathbf{R'} = \mathbf{H}$) (3.88 g, 17 mmol) was heated under reflux in trifluoroacetic acid (25 mL) for 4 h, allowed to cool, and evaporated to dryness in vacuo. The residue was partitioned between saturated aqueous NaHCO₃ solution (100 mL) and CH₂Cl₂ (100 mL), the phases were separated, and the organic phase was washed with H_2O . The dried (MgSO₄) organic phase was evaporated to dryness in vacuo to afford the O-benzyl derivative of 56, which was used without purification. This intermediate was dissolved in EtOH (150 mL) and hydrogenated over 10% palladium on carbon at room temperature and atmospheric pressure until hydrogen uptake ceased. The catalyst was removed by filtration and washed with EtOH, and the combined filtrates were acidified with hydrogen chloride gas. Removal of the solvent in vacuo afforded 1.52 g (41%) of 56: mp 193-4 °C dec (EtOH). Anal. $(C_9H_7F_3N_2O \cdot HCl \cdot 0.25H_2O)$. C, H, N.

5-tert -Butyl-7-methyl-2-(trifluoromethyl)-1*H*-benzimidazol-4-ol (57). The title compound was prepared in 47% yield according to method E by using trifluoroacetic acid: mp 140-141 °C (Et₂O-pentane). Anal. ($C_{13}H_{15}F_3N_2O$) C, H, N.

⁽²⁰⁾ Dadsworth, M. F.; Kenner, J. J. Chem. Soc. 1927, 580.

⁽²¹⁾ Bantlin, A. Ber. Dtsch. Chem. Ges. 1878, 11, 2104.

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RBL-1 5-Lipoxygenase Assay. 5-Lipoxygenase enzyme was prepared as a 10000g supernatant from RBL-1 cells by the method of Jakschik et al.²² The supernatant was diluted with homogenization buffer to the equivalent of $1.5-2.5 \times 10^7$ cells/mL and made 2 mM with respect to CaCl₂. Aliquots of 0.5 mL were then dispensed into tubes and incubated at 29 °C with 5 μ L of dimethyl sulfoxide or the test compound in dimethyl sulfoxide at the desired concentration for 2 min. Then [1-14C]arachidonic acid was added in buffer to give a final concentration of 6.3 μ M and 0.2 μ Ci per incubation, and the reaction was continued at 29 °C for 2 min. The reaction was terminated by the addition of 1 mL of acetone and cooling on ice. Ice-cold saline (0.5 mL) and 100 μ L of 2 M formic acid were added, and the mixture was extracted with 2 \times 2 mL of CHCl₃. The resulting extract was stored under nitrogen at -20 °C until analysis by TLC on silica gel G using the solvent system toluene-dioxane-acetic acid (65:35:1, v/v). Activity was measured as the percentage of the total radioactivity found in 5-HETE and 5,12-diHETE, and inhibition was calculated as (1 -D/C × 100, where D is the activity in the presence of compound and C is the control activity. IC_{50} values were computed by linear regression analysis using two or more concentrations (each a minimum of three determinations) causing inhibition significantly different from controls (p < 0.05) and spanning the 50% value.

Rat Peritoneal Anaphylaxis. Passive peritoneal anaphylaxis (PPA) was carried out in the rat as described previously.¹⁴ Antiserum to ovalbumin (Sigma grade V) was raised in rats by using either Bordetella pertussis vaccine (Burroughs Wellcome, London) or pertussis vaccine adsorbed on alum (Lister Institute, Elstree) as an adjuvant. Rats were given intraperitoneal injections of 2 mL of a 1:5 dilution of the rat antiserum in isotonic saline. Two hours later, 0.3 mL of a 5% solution of Pontamine Sky Blue (G. T. Gurr, London) in isotonic saline was injected intravenously, followed immediately by an intraperitoneal injection of 1 mL of a solution of test compound in saline or in saline containing 1-2%dimethyl sulfoxide. Control rats received 1 mL of saline or saline containing 1-2% dimethyl sulfoxide. Each dose of a compound or of control was given to groups of five to seven rats, and treatments were randomized. The concentrations of drug administered were adjusted so that the required concentration was that in the 6 mL of fluid injected into the peritoneal cavity. After 2.5 min the rats were given an intraperitoneal injection of 5 mL of a Tyrode solution containing 0.4 mL of ovalbumin and 6 units/mL heparin. Exactly 5 min after challenge, the rats were stunned and bled, and the peritoneal fluids were collected and assayed for SRS-A as previously described.¹⁴

Rat Pleural Exudate Model. The method used was based on that of Ackerman et al.¹⁵ Female rats (Charles River Wistar, 175–225 g) were lightly anesthetized (Hypnorm) and received injections of 0.2 mL of a 2% solution of λ carrageenan into the

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pleural cavity. Seventy-two hours later the animals were killed by an overdose of pentobarbitone, the pleural cavity was exposed, and 4 mL of a solution containing 0.375% phenol red and EDTA in saline was flushed into the cavity. After gentle massage, the fluid was harvested and the inflammatory exudate was measured by dye dilution using a spectrophotometer. The leukocyte concentration was estimated by the fluorimetric assay of DNA.²³ Compounds were administered orally 1 h before carrageenan, and 24 and 48 h subsequently, suspended in 0.7% methylcellulose solution.

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Registry No. 1, 2409-55-4; 2, 103059-07-0; 3, 103059-06-9; 4, 103059-08-1; 5, 103059-09-2; 5-diamine, 103059-10-5; 6, 103080-29-1; 7, 103080-30-4; 10, 103059-03-6; 11, 103059-01-4; 12, 103059-02-5; 13, 110191-32-7; 14, 110191-33-8; 15, 103059-05-8; 16, 110191-34-9; 17, 110191-35-0; 18, 110191-36-1; 19, 110191-37-2; **20**, 110191-38-3; **21**, 110191-39-4; **22**, 110191-40-7; **23**, 110191-41-8; 24, 110191-42-9; 25, 110191-43-0; 26, 110191-44-1; 27, 110191-45-2; 28, 110191-46-3; 29, 103059-40-1; 29·HCl, 103059-14-9; 30, 103059-11-6; 31, 110191-47-4; 32, 103059-39-8; 32-HCl, 103059-13-8; 33, 103059-19-4; 34·HBr, 103080-32-6; 35, 103406-47-9; 35·HCl, 103059-35-4; 36, 103059-43-4; 36·HCl, 103059-21-8; 37, 110191-48-5; 37.HCl, 110191-49-6; 38, 103059-56-9; 38.HCl, 103059-34-3; 39, 103059-57-0; 39·HCl, 103059-36-5; 40, 103059-53-6; ,40·HCl, 103059-32-1; 41, 103059-58-1; 41·HCl, 103059-37-6; 42, 103059-55-8; 42-HCl, 103080-35-9; 43, 103059-54-7; 43-HCl, 103059-33-2; 44, 110191-50-9; 44-HCl, 110191-51-0; 45, 110191-52-1; 45-HCl, 110191-53-2; 46, 103059-46-7; 46-HCl, 103059-24-1; 47, 103059-41-2; 47.HCl, 103059-16-1; 48, 103059-51-4; 48.HCl, 103080-34-8; 49.HCl, 103059-31-0; 50, 103059-15-0; 51, 103059-25-2; 52, 103059-26-3; **53**, 103059-12-7; **54**, 103059-20-7; **55**, 103059-04-7; **56**, 108381-55-1; 56 (o-benzyl deriv.), 110191-54-3; 57, 108381-56-2; C₆H₅CH₂COOH, 103-82-2; C₆H₅(CH₂)₂COOH, 501-52-0; 4-CH₃C₆H₄CH₂COOH, 622-47-9; 4-CH₃OC₆H₄COOH, 100-09-4; 4-CH₃OC₆H₄CH₂COOH, 104-01-8; 3-CH₃OC₆H₄CH₂COOH, 1798-09-0; 2-CH₃OC₆H₄CH₂COOH, 93-25-4; 4-C₂H₅OC₆H₄CH₂COOH, 4919-33-9, 4-HOC₆H₄CH₂COOH, 156-38-7, 3, 4-(CH₃O)₂C₆H₃CH₂COOH, 93-40-3; 3,4,5-(CH_3O)₃C₆H₂CH₂COOH, 951-82-6; $C_{10}H_7CH_2COOH$, 86-87-3; 2- $\check{C}_{10}H_7CH_2COOH$, 581-96-4; C_6H_5C -HO, 100-52-7; 3-CF₃C₆H₄, 454-89-7; 4-N(CH₃)₂C₆H₄CHO, 100-10-7; 4-methyl-2,3-dinitrophenol, 68191-07-1; 2,3-dinitrophenol benzyl ether, 103080-31-5; 2,3-dinitrophenol, 66-56-8; 2,3-diaminophenol benzyl ether, 89521-55-1; 3-methoxy-6-methyl-1,2-phenylenediamine, 101251-28-9; 4-chlorophenylacetic acid, 1878-66-6; butyric acid, 107-92-6; trifluoroacetic acid, 76-05-1; 5-lipoxygenase, 80619-02-9; 2-thiopheneacetic acid, 1918-77-0; 3-pyridineacetic acid, 501-81-5; 2-furancarboxaldehyde, 98-01-1.

(23) Karsten, U.; Wollenberger, A. Anal. Biochem. 1977, 77, 464.

⁽²²⁾ Jakschik, B. A.; Sun, F. F.; Lee, L. M.; Steinhoff, M. M. Biochem. Biophys. Res. Commun. 1980, 95, 103.