

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters 16 (2006) 4567-4570

Bioorganic & Medicinal Chemistry Letters

## Potent 4-amino-5-azaindole factor VIIa inhibitors

Huiyong Hu,\* Aleksandr Kolesnikov, Jennifer R. Riggs, Kieron E. Wesson,Robin Stephens, Ellen M. Leahy, William D. Shrader, Paul A. Sprengeler,Michael J. Green, Ellen Sanford, Margaret Nguyen, Erik Gjerstad,Ronnel Cabuslay and Wendy B. Young

Celera Genomics, 180 Kimball Way, South San Francisco, CA 94080, USA

Received 19 April 2006; revised 3 June 2006; accepted 6 June 2006 Available online 21 June 2006

Abstract—The 4-amino-5-azaindole as an amidino-benzimidazole replacement is described. A series of potent and selective analogs were discovered and showed desirable *ex vivo* efficacy as measured by PT. © 2006 Elsevier Ltd. All rights reserved.

The development of novel anticoagulants with improved therapeutic indices is an attractive goal in the pharmaceutical industry. The activated factor VIIa-tissue factor complex (fVIIa·TF), which exists in the extrinsic pathway of the coagulation cascade, has been an appealing molecular target for the treatment of various thrombosis related disorders.<sup>1–4</sup>

We have previously reported on the discovery and development of small molecule fVIIa TF complex inhibitors.<sup>5-7</sup> Many of these described inhibitors contain an amidino moiety, which forms a salt bridge with Asp 189 in the S1 pocket of fVIIa providing increased bonding and potency. The amidino compound 1 (Fig. 1) has a 0.013  $\mu$ M inhibition K<sub>i</sub> for fVIIa TF and good selectivity (>200-fold) versus primary antitargets fXa, thrombin, and trypsin. The strongly basic amidino group, however, is considered to be a major limitation to oral bioavailability. We have studied the oral bioavailability of various amidine and amidine prodrugs in our biaryl scaffold.<sup>8</sup> These compounds suffer from low oral bioavailability due to low absorption or low conversion of prodrug to parent amidine. In an effort to replace the 5-amidino-benzimidazole, we explored less basic P1 elements including 5-azaindoles9,10 and 4-chloro-5azaindoles<sup>10</sup> as well as others.<sup>9</sup> The 5-azaindole compound 2 (Fig. 1) has a potency of 0.22 µM for fVIIa and >10-fold selectivity against the antitargets. Most recently, we identified the 4-amino-5-azaindole (1*H*-pyr-rolo[3,2-*c*]pyridin-4-ylamine) moiety as another viable amidine replacement. Compound **3**, which was made as a direct comparison of **2**, has further improved potency (fVIIa  $K_i = 0.081 \mu$ M) and selectivity (>400-fold against antitargets). The 4-amino-5-azaindole analogs retained desirable potency<sup>10</sup> and *ex vivo* efficacy in human plasma as monitored by the coagulation assays PT (prothrombin time) and aPTT (activated partial thromboplastin time).<sup>11</sup> In this communication, we disclose our discovery of analogs with good *ex vivo* efficacy based on the 4-amino-5-azaindole.

We have discovered in our 5-amidino-benzimidazole analogs that substituents bearing an acid moiety at the C5' position of the central aryl ring improved potency against fVIIa by interacting with Lys 192 of the enzyme.<sup>5</sup> Starting from 3, we installed various acid substitutions at the C5' position in order to achieve more desirable potency (compounds 4 and 5, Table 1). The C5' benzoic acid 4 gave the best potency against fVIIa, as well as ex vivo efficacy, as measured by the fVIIa TF dependent clotting assay PT. However, compound 4 was also quite active in the fVIIa TF independent clotting assay aPTT, possibly by inhibiting proteases along the intrinsic coagulation pathway. Compound 5, although less potent compared to 4, has moderately differentiated the PT and aPTT. To target more specifically the fVIIa TF complex, which is associated with the extrinsic and common coagulation pathway, we choose the potent compound 5 as a starting point to further optimize our analogs for an improved PT profile.

*Keywords*: Factor VIIa; Tissue factor; Anticoagulant; Serine protease; Amidine; Amidine replacement; PT; aPTT; 4-amino-5-azaindole; 1*H*-pyrrolo[3,2-*c*]pyridin-4-ylamine.

<sup>\*</sup> Corresponding author. Tel.: +1 408 835 2435; e-mail: huiyong.hu@yahoo.com

<sup>0960-894</sup>X/\$ - see front matter @ 2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2006.06.016



rile 8 followed by hydrolysis of the nitrile, deprotection of

the methyl ether, and finally esterification provided ester 9. Compound 9 was selectively *ortho*-formylated with

paraformaldehyde and MgCl<sub>2</sub>.<sup>13</sup> The resulting salicylal-

dehyde was brominated with N-bromosuccinamide in

DMF, followed by benzyl protection of the phenol to af-

ford ether 10. Suzuki coupling between bromide 10 and

the boronic acid **11** led to biaryl compound **12**. Subsequent treatment of aldehyde **12** with the diazophospho-

nate Ohira reagent<sup>14,15</sup> generated alkyne **13**. The

4-chloro-5-azaindole ring was established via a Sonagashira reaction between 13 and mesylate  $14^{16}$  to form a tran-

sient biaryl alkyne, which cyclized spontaneously to form the *N*-mesyl indole. The indole was subsequently treated

with NaOH/MeOH to induce cleavage of the sulfonamide

and hydrolysis of the ester providing compound 15. The

chloro-substituted indole 15 was treated with NH<sub>4</sub>OAc

in melting phenol to displace the 4-chloro group providing

the 4-amino-5-azaindole. The boc and benzyl protecting

groups were removed upon treatment with 6 N HCl(aq)

followed by hydrogenation with Pearlman's catalyst to afford **16**. The amine **16** was treated with 2,6-difluoroisocy-

anate **17** under basic conditions followed by purification on reverse phase HPLC provided **7** as a mono HCl salt.

Figure 1. Potency for fVIIa TF and selected antitargets in amidino and non-amidino scaffolds.

In our effort to improve the potency and selectivity of the 5-azaindole series, we had identified substituted phenylureas as the optimal S1' substitution.<sup>9</sup> In the 5-azaindole series, the para-benzoic acid-substituted phenylurea gave desirable potency and decreased lipophilicity, which is believed to lower binding to serum albumin and therefore improve the PT. The same SAR transfers to the current series where the corresponding compound 6 (Table 2) retained potency against fVIIa and ex vivo efficacy with a greater differentiation between PT and aPTT. The 2,6-difluorophenylurea moiety was also found to provide further improvement to fVIIa potency and selectivity in 5-azaindole series.9 Accordingly, compound 7 showed an increase in potency against fVIIa by  $\sim$ 10-fold as well as selectivity against other trypsin family enzymes. The PT profile was further improved to 2.4  $\mu$ M, while aPTT was greater than 20  $\mu$ M. This 4amino-5-azaindole series marks an improvement in the PT and the differentiation between the intrinsic and extrinsic coagulation pathways when compared to the 5-azaindole series.

Synthesis of analog 7 is outlined in Scheme 1). Methylation of commercially available 4-methoxyphenylacetonit-

Table 1. SAR at the C5'-position of central aryl ring<sup>12</sup>

NH2 NH2 H H HO H H O H

Compound	R	fVIIa·TF $K_i$ ( $\mu$ M)	2× PT (μM)	$2 \times aPTT (\mu M)$	Selectivity for fVIIa versus		
					Thrombin <sup>a</sup>	fXa <sup>b</sup>	Trypsin <sup>c</sup>
3	Н	0.081	12	22	$1.2 \times 10^{3}$	$6.7 \times 10^{2}$	$4.7 \times 10^{2}$
4	COOH	0.0013	4.5	3.7	$4.0 \times 10^{4}$	$5.0 \times 10^{3}$	$3.5 \times 10^{3}$
5	$C(CH_3)_2CO_2H$	0.015	6.4	17	$>1.0 \times 10^{4}$	$2.8 \times 10^{3}$	$3.2 \times 10^{3}$

<sup>a</sup> Thrombin  $K_i$  ( $\mu$ M)/fVIIa·TF  $K_i$  ( $\mu$ M) = fold selectivity.

<sup>b</sup> fXa  $K_i$  ( $\mu$ M)/fVIIa·TF  $K_i$  ( $\mu$ M) = fold selectivity.

<sup>c</sup> Trypsin  $K_i$  ( $\mu$ M)/fVIIa·TF  $K_i$  ( $\mu$ M) = fold selectivity.

Table 2. SAR on distal aryl ring<sup>12</sup>



Compound	R	fVIIa·TF $K_i$ ( $\mu M$ )	2× PT (μM)	$2 \times aPTT (\mu M)$	Selectivity for fVIIa versus		
					Thrombin	fXa	Trypsin
5	PH CH	0.015	6.4	17	>1.0×10 <sup>4</sup>	$2.8 \times 10^{3}$	$3.2 \times 10^{3}$
6	H <sup>5</sup> 3 <sup>°</sup> N OH	0.020	5.8	>20	>7.5×10 <sup>3</sup>	$5.5 \times 10^{3}$	$5.5 \times 10^{3}$
7	H F Ö	0.0026	2.4	>20	>5.8×10 <sup>4</sup>	$2.8 \times 10^4$	$2.9 \times 10^{4}$

In conclusion, we have identified potent and selective fVIIa<sup>·</sup>TF inhibitors in a novel non-amidino scaffold which differentiates between the intrinsic and extrinsic coagulation pathways. This series was dropped in lieu of pursuing a more attractive series and absorption data were never obtained. We would like to suggest that from a potency standpoint, the 4-amino-5-azaindole is a suitable surrogate for the benzamidine and that others con-



Scheme 1. Reagents and conditions: (a) *t*-BuOK, MeI, THF, rt; (b) KOH, ethylene glycol/water (5:1), 150 °C; (c) Py·HCl, 180 °C; (d) SOCl<sub>2</sub>, MeOH; (e) MgCl<sub>2</sub>, anhydrous paraformaldehyde, Et<sub>3</sub>N, CH<sub>3</sub>CN, reflux; (f) NBS, DMF; (g) BnBr, DIEA, CH<sub>3</sub>CN; (h) 11, Pd(PPh<sub>3</sub>)<sub>4</sub>, K<sub>2</sub>CO<sub>3</sub>, DME, reflux; (i) Ohira's reagent, K<sub>2</sub>CO<sub>3</sub>, MeOH; (j) 14, PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub>, CuI, Et<sub>3</sub>N, CH<sub>3</sub>CN, 80 °C; (k) 50% NaOH, MeOH, 60 °C; (l) NH<sub>4</sub>OAc, PhOH, 105 °C; (m) 6 N HCl, reflux; (n) 20% Pd(OH)<sub>2</sub> on carbon (Pearlman's catalyst)/H<sub>2</sub>, EtOH; (o) 17, Et<sub>3</sub>N, DMF.

sider such a moiety in their drug development programs, especially in the challenging arena of developing fVIIa inhibitors.

## **References and notes**

- 1. Harker, L. A.; Hanson, S. R.; Wilcox, J. N.; Kelly, A. B. *Haemostasis* **1996**, *26*, 76.
- Himber, J.; Kirchhofer, D.; Riederer, M.; Tschopp, T. B.; Steiner, B.; Roux, S. P. *Thromb. Haemost.* 1997, 78, 1142.
- Suleymanov, O. D.; Szalony, J. A.; Salyers, A. K.; Lachance, R. M.; Parlow, J. J.; South, M. S.; Wood, R. S.; Nicholson, N. S. J. Pharmacol. Exp. Ther. 2003, 306, 1115.
- Olivero, A. G.; Eigenbrot, C.; Goldsmith, R.; Robarge, K.; Artis, D. R.; Flygare, J.; Rawson, T.; Sutherlin, D. P.; Kadkhodayan, S.; Beresini, M.; Elliott, L. O.; DeGuzman, G. G.; Banner, D. W.; Ultsch, M.; Marzec, U.; Hanson, S. R.; Refino, C.; Bunting, S.; Kirchhofer, D. J. Biol. Chem. 2005, 280, 9160.
- Young, W. B.; Kolesnikov, A.; Rai, R.; Sprengeler, P. A.; Leahy, E. M.; Shrader, W. D.; Sangalang, J.; Burgess-Henry, J.; Spencer, J.; Elrod, K.; Cregar, L. *Bioorg. Med. Chem. Lett.* 2001, 11, 2253.
- Young, W. B.; Mordenti, J.; Torkelson, S.; Hanson, S. R.; Marzec, U. M.; Shrader, W. D.; Rai, R.; Kolesnikov, A.; Liu, L.; Hu, H.; Leahy, E.; Sprengeler, P.; Katz, B.; Janc, J. W. Bioorg. Med. Chem. Lett. 2006, 16, 2034.
- Shrader, W. D.; Kolesnikov, A.; Burgess-Henry, J.; Rai, R.; Hu, H.; Torkelson, S.; Young, W. B.; Sprengeler, P.; Katz, B.; Yu, C.; Cabuslay, R.; Sanford, E.; Janc, J. *Bioorg. Med. Chem. Lett.* 2006, 16, 1596.
- Riggs, J. R.; Kolesnikov, A.; Hendrix, J.; Young, W. B.; Shrader, W. D.; Stephens, S.; Liu, L.; Pan, L.; Mordenti, J.; Green, M. J.; Sukbuntherng, J. *Bioorg. Med. Chem. Lett.* 2006, 16, 2224.
- (a) Riggs, J. R.; Hu, H.; Kolesnikov, A.; Leahy, E. M.; Wesson, K. E.; Shrader, W. D.; Vijaykumar, D.; Wahl, T. A.; Tong, Z.; Sprengeler, P. A.; Green, M. J.; Yu, C.; Katz, B. A.; Sanford, E.; Nguyen, M.; Cabuslay, R.; Young, W. B. *Bioorg. Med. Chem. Lett.* 2006, *16*, 3197; (b) Rai, R.; Kolesnikov, A.; Sprengeler, P. A.; Torkelson, S.; Ton, T.; Katz, B. A.; Yu, C.; Hendrix, J.; Shrader, W. D.; Stephens, S.; Cabuslay, R.; Sanford, E.; Young, W. B. *Bioorg. Med. Chem. Lett.* 2006, *16*, 2270.
- 10. The 5-azaindole, which has a calculated  $pK_a \sim 7.8$  (ACD Labs/pKa DB), is thought to be partially protonated in the

assays (pH 7.4) and forms water bridged hydrogen bonds with Asp 189 of the enzyme to afford desired nanomolar potency against fVIIa. The 4-chloro-5-azaindole P1, which has a calculated  $pK_a \sim 5.7$ , is not protonated and therefore has little interaction with Asp 189, which is consistent with the loss of potency against fVIIa (micro-molar  $K_i$  for fVIIa.TF). The 4-amino-5-azaindole P1, which has a calculated  $pK_a \sim 9.9$ , is mostly protonated and forms a stronger interaction with Asp 189 through water bridged hydrogen bonds. In addition, the 4-amino group is also thought to interact with G219 of the enzyme. Both interactions are considered to contribute to the improved potency.

- 11. Bajaj, S. P.; Joist, J. H. Semin. Thromb. Hemost. 1999, 25, 407.
- 12. Inhibition assays for factor Xa and thrombin were performed as described (Cregar, L.; Elrod, K. C.; Putnam, D.; Moore, W. R. Arch. Biochem. Biophys. 1999, 366, 125), with the pH adjusted to 7.4. The trypsin and fVIIa assays were performed and analyzed as in the above reference with the following additional details. Factor VIIa (Enzyme Research) was incubated at 7 nM and CH<sub>3</sub>SO<sub>2</sub>-D-CHA-But-Arg-pNA (Centerchem) was used as the substrate. The buffer for the factor VIIa assay was supplemented with 11 nM relipidated tissue factor and 5 mM CaCl<sub>2</sub>. Trypsin (Athens Research Institute) was incubated at 10 nM with variable concentrations of inhibitor in 50 mM Tris (pH 7.4), 150 mM NaCl, 1.5 mM EDTA, 0.05% Tween 20, and 10% DMSO. The reaction was initiated with substrate, Tosyl-Gly-Pro-Lys-pNA (Centerchem), supplied at the  $K_{\rm m}$  (25 µM). Coagulation assays (PT and aPTT) were carried out at 37 °C in human plasma using a Beckman Coulter ACL100 in accordance with the manufacturer's instructions.
- 13. Hofslokken, N. U.; Skattebol, J. Acta Chem. Scand. 1999, 53, 258.
- 14. Ohira, S. Synthetic Commun. 1989, 19, 561.
- 15. Roth, G. J.; Liepold, B.; Müller, S. G.; Bestmann, H. J. *Synthesis* **2004**, 59.
- 16. The synthesis of intermediate 14 is shown in Scheme 2. Commercially available 4-amino-2-chloropyridine 18 was iodinated to afford a mixture of iodopyridines 19 and 20 ( $\sim$ 6:4). Compound 20 can be easily isolated by chromatography on silica. Treatment of 20 with methanesulfonyl chloride led to a mixture of monomesylate 14 and bis-*N*mesylated byproduct. Subsequent treatment with NaO-H(aq)/THF converted the bismesylate to monomesylate 14.



Scheme 2. Reagents and conditions: (a) ICl, KOAc/HOAc, 60-70 °C; (b) MsCl, Et<sub>3</sub>N, DCM, 0 °C; (c) 10% NaOH, THF.