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Synthesis of Potent and Highly Selective Inhibitors of Human Tryptase

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Abstract—The serine protease tryptase has been implicated in allergic and inflammatory diseases and associated with asthma. The synthesis and SAR of a series of N1-activated-4-carboxy azetidinones are described, resulting in identification of BMS-363131 (2) as a potent inhibitor of human tryptase (IC₅₀ < 1.7 nM) with high selectivity (> 3000-fold) for tryptase versus related serine proteases including trypsin.

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Tryptase is a serine-like protease that is the major protein component found in mast cells. Upon activation, mast cells degranulate, releasing tryptase, histamine, chymase, and other mediators of allergic and inflammatory conditions. Tryptase has been implicated in allergic and inflammatory diseases as well as autoimmune diseases and is believed to play an important role in inflammatory, bronchoconstrictive, and remodeling processes associated with asthma.^{1–3} It is therefore believed that inhibition of tryptase may offer a therapeutic benefit for the treatment of asthma. Our preceding report⁴ described the identification of BMS-262084 (1) as a potent inhibitor of tryptase (IC₅₀=4 nM) with moderate to good selectivity against related serine proteases but not for trypsin.

In ovalbumin-sensitized guinea pig models, intratracheally dosed BMS-262084 demonstrated efficacy in preventing allergen-induced bronchoconstriction and in protecting against inflammatory cell infiltration into the lung. It was recognized that a significant portion of any inhaled drug would be swallowed; consequently, good selectivity against the gastric protease trypsin was considered highly desirable. In this Letter, we report on our efforts to improve upon BMS-262084 by exploring the effects of conformationally constrained guanidino groups at C-3 and new substituents at N-1 of the azetidinone nucleus. These investigations led to the discovery of BMS-363131 (2) as a potent inhibitor of tryptase having excellent selectivity versus other serine proteases including trypsin and having improved hydrolytic stability.



The general Scheme for preparation of compounds **9b–f** in Table 1 is outlined in Scheme 1. Alkylation of (*S*)-azetidinone 4^5 with iodides 3 gave compounds 5, which were converted in two steps to 6. Removal of the Boc group and then guanylation gave the bis-Cbz protected guanidine compounds 7. Subsequent acylation with chlorocarbonylpiperazine 8a and hydrogenation provided (3*R*,4*S*) compounds 9b–f.

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 Table 1.
 Tryptase inhibition, selectivities and chemical stability for compounds 1 and 9

^aMixture of diastereomers at C-6.

^bSingle diastereomer at C-6, compounds **9e** and **9f** are the isolated diastereomers of **9d**.



Scheme 1. (a) LDA, THF; (b) TBAF, THF; (c) BnBr, NaHCO₃, DMF; (d) TFA, CH₂Cl₂; (e) 1-[C(NCbz)(NHCbz)]pyrazole, TEA, DMF or MeSC(NCbz)(NHCbz), Hg(OAc)₂, TEA, DMF; (f) TEA, DMAP, DMF; (g) Pd/C, H₂, dioxane, aq 1 N HCl.

The iodo compounds **3a–c** employed in Scheme 1 were prepared from the intermediate Boc protected amino alcohols **11**, **13**, and **15** (Scheme 2), which could be obtained from the commercially available amino acid, amino alcohol, or amino ester. Chlorocarbonyl-piperazines (**8a–g**) used in either Scheme 1 or Scheme 4 were prepared as shown in Scheme 3. Compounds **8a–e** were prepared from Boc-piperazine (**16**) by acylation, removal of the Boc group, and finally treatment with phosgene. The synthesis of compounds **8f–g** involved



Scheme 2. (a) Procedures described in ref 7; (b) I_2 , PPh₃, imidazole, CH₂Cl₂; (c) Pd/C, H₂, MeOH or *i*PrOH; (d) Boc₂O, THF; (e) LAH, THF.

the formation of the carbamoylchloride of Boc-piperazine (16) followed by sequential treatment with an amine, removal of the Boc group, and treatment with phosgene.

The synthesis of diastereomeric mixtures 18a-f and the single diastereomer BMS-363131 (2), which are listed in Table 2, are outlined in Scheme 4. The mixtures 18a-f were prepared by acylation of diastereomeric mixtures 7a with chlorocarbonylpiperazines 8b-g and deprotection. BMS-363131 (2), however, was prepared from the single diastereomer 21 by acylation with 8b and deprotection. Compound 21 was derived from (*R*)-ethyl



Scheme 3. (a) HO₂C-R, EDAC, DIEA, CH_2Cl_2 ; (b) TFA, CH_2Cl_2 ; (c) Cl_2CO . CH_2Cl_2 , TEA or NaHCO₃; (d) CIOC-R, TEA, DMF; (e) $H_2NR_1(CH_2)_4Ph R_1 = H \text{ or } Me$, NaHCO₃ or TEA, CH_2Cl_2 .



Scheme 4. (a) TEA, cat. DMAP, DMF, 82%; (b) Pd/C, H₂, dioxane, aq 1 N HCl, (c) polyvinylpyridine resin, H₂O, 92% 2 steps; (d) Boc₂O, CH₂Cl₂, 87%; (e) LAH, THF, 87%; (f) I₂, PPh₃, imidazole, CH₂Cl₂, 77%; (g) 4, LDA, THF, $-70 \rightarrow -20$ °C; (h) TBAF, THF; (i) BnBr, NaHCO₃, DMF, 76% 3 steps; (j) TFA, CH₂Cl₂; (k) 1-[C(NCbz)(NHCbz)]pyrazole, TEA, DMF, 73% 2 steps. Yields are for the synthesis of BMS-363131 (2).

nipecotate (19), which along with the (S)-enantiomer was obtained by resolution of racemic ethyl nipecotate.⁶ Boc protection of 19, followed by reduction and iodination gave compound 20. Alkylation of (S)-azetidinone 4 with 20 and sequential desilylation, benzylation, removal of the Boc group, and guanylation provided intermediate 21. BMS-363130 (22), which has the (S)configuration at C-6 in contrast to BMS-363131 (2) which has the (R)-configuration at C-6, was prepared in the same manner as BMS-363131, however, starting from (S)-ethyl nipecotate in place of 19.

Guanidinylpropyl compounds **9a** (Table 1) and **23** (Table 2) were prepared using chlorocarbonylpiperazines **8a** and **8b**, respectively, following methodology described in the previous Letter to prepare BMS-262084.

The tryptase inhibition, selectivities, and chemical stability for the new conformationally constrained guanidino compounds are shown in Tables 1 and 2. Table 1 displays several constrained guanidino analogues of guanidinylpropyl compound **9a**. Compound **9a** and BMS-262084 (1), which differ in their N-1 substituent, were equally potent against tryptase ($IC_{50} = 4 \text{ nM}$) and exhibited similar hydrolytic stability profiles. However, the overall selectivity profile of 9a offered an improved template to investigate constrained guanidine analogues. The amidinoazetidinyl analogue 9b showed potent activity against tryptase (IC₅₀ = 12 nM) but no significant improvement in selectivity against plasmin, thrombin, urokinase (uPA), tissue plasminogen activator (tPA), or trypsin. The amidinopyrrolidinyl analogue 9c, although potent against tryptase, similarly showed poor selectivity against trypsin. Activity against tryptase was substantially reduced for the unsymmetrical amidinopiperidinyl diastereomeric mixture 9d (diastereomeric at the piperidinylmethyl juncture) and the single diastereomers 9e and 9f. However, 9d displayed greatly improved selectivity for tryptase over trypsin (258-fold), as well as enhanced aqueous stability $(t_{1/2}>48$ h at pH 9). The diastereomers **9e** and **9f**, comprising the mixture 9d, were similarly selective.

SAR in the amidinopiperidinyl series was studied further by examining substituents on N-4 of the piperazine





in 9d. This led to a series of diastereomeric mixtures (18a–f) of highly potent inhibitors of tryptase with excellent selectivity against related serine proteases including trypsin as shown in Table 2. The diastereomeric mixture 18a was comparable to the guanidinyl-propyl analogue 3 in potency versus tryptase but exceedingly more selective against plasmin, uPA, and trypsin. Additionally, 18a exhibited improved hydrolytic stability at pH 9 ($t_{1/2} > 17.5$ h) compared to BMS-262084 (1) in Table 1 ($t_{1/2} = 8.0$ h). The other amidino-piperidinyl diastereomeric mixtures in Table 2 displayed potent activity versus tryptase and high selectivity against other serine proteases including trypsin (most > 1000-fold).

Potency versus tryptase and selectivity for the two single diastereomers BMS-363131 (2) and BMS-363130 (22) that comprise the mixture **18a** are also shown in Table 2. Each of the diastereomers had an $IC_{50} < 1.7$ nM against tryptase with selectivity > 3,000-fold versus trypsin and > 10,000-fold versus plasmin, thrombin, uPA, and tPA. BMS-363131 was selected over BMS-363130 for further evaluation following comparisons of other biological properties of the two compounds. In the ovalbumin-sensitized guinea pig model of lung inflammation,⁴ intratracheal dosing of BMS-363131 effected a reduction in inflammatory cell count in the lungs compared to control.

In summary our investigation of the SAR at N-1 and C-3 of the azetidinone nucleus of BMS-262084 (1) has shown that conformationally constrained guanidinyl groups at C-3 and extended substituents appended to the piperazine at N-1 provide potent inhibitors of typtase with highly improved selectivity versus other serine proteases including trypsin. One of the most potent and selective compounds, BMS-363131 (2), was shown to be efficacious in a guinea pig model of asthma.

References and Notes

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