

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry 14 (2006) 5099-5109

Bioorganic & Medicinal Chemistry

# Thrombomodulin induction in cultured human endothelial cells by 9-*cis*-locked retinoic acid analogues

Shiro Ikegami,<sup>a,\*</sup> Takamasa Iimori,<sup>a</sup> Minoru Sudo,<sup>a</sup> Maroka Kitsukawa,<sup>a</sup> Alireza Foroumadi,<sup>a</sup> Takeshi Yonemura,<sup>a</sup> Hideyo Takahashi,<sup>a</sup> Keiichiro Kizaki<sup>a</sup> and Hidemi Ishii<sup>b</sup>

<sup>a</sup>Faculty of Pharmaceutical Sciences, Teikyo University, Sagamiko, Kanagawa 199-0195, Japan <sup>b</sup>Showa Pharmaceutical University, Higashi-Tamagawagakuen, Machida, Tokyo 194-8543, Japan

> Received 1 March 2006; revised 3 April 2006; accepted 4 April 2006 Available online 19 May 2006

Abstract—9-cis-Retinoic acid (RA) analogues devised to lock the 9-cis double bond by ring formation were synthesized using two stereoselective carbon–carbon bond formation reactions as key steps. The palladium-mediated Suzuki reaction was adopted to construct a 7*E*-double bond (RA numbering) and the Horner–Emmons olefination was employed for stereoselective 11*E*-double bond (RA numbering) formation. The synthesized 9-cis-RA analogues that are locked by five-membered ring systems (cyclopentene, dihydrofuran, and dihydrothiophene) were shown to have comparable thrombomodulin induction activities to that of 9-cis RA. Conformational analysis of these compounds showed their similarity to 9-cis RA in the spatial orientation of the side chain and the terminal carboxy group.

© 2006 Elsevier Ltd. All rights reserved.

# 1. Introduction

Retinoic acid (RA) is well known to play a critical role in many aspects of cell proliferation and differentiation, and has proved useful for the treatment of dermatologic diseases and certain cancers.<sup>1</sup> We have demonstrated that all-trans-RA (ATRA) causes an increase in thrombomodulin (TM) antigen in human umbilical vein endothelial cells (HUVECs) in vitro.<sup>2,3</sup> TM is an endothelial membrane surface glycoprotein and represents one of the most important antithrombotic factors in the anticoagulant system. TM forms a 1:1 stoichiometric complex with thrombin and then thrombin loses its procoagulant functions such as fibrin formation. Additionally, the TM-thrombin complex markedly promotes activation of protein C, which proteolytically degrades coagulation factors.<sup>4,5</sup> Since TM functions as an antithrombotic agent, RA has the ability to change the coagulant balance by controlling TM expression.

Keywords: Retinoic acid; 9-cis-Retinoic acid; Suzuki coupling; Thrombomodulin.

Recently, much attention has been paid to 9-*cis*-RA, because it has been identified as a second endogenous RA that binds both retinoic acid receptors (RARs) and retinoid X receptors (RXRs) with nanomolar affinity.<sup>6–8</sup> This finding prompted us to investigate the structure-activity relationships of RA analogues to TM induction. Figure 1 shows our preliminary observations of TM induction by geometric isomers of RAs (ATRA, 9-*cis*-RA, 11-*cis*-RA, and 13-*cis*-RA). The TM expression depended on the amount of all of Ras,<sup>9</sup> but a significant increase was observed at 0.1  $\mu$ M 9-*cis*-RA, which was most potent among examined. Thus, our attention was centered around the structural modification which fixes the *cis*-double bond of 9-*cis*-RA.

# 2. Results and discussion

# 2.1. Molecular design

Our design of 9-*cis*-RA analogues presented herein was based on locking its 9-*cis*-double bond by introducing an appropriate ring system.<sup>10</sup>

As shown in Figure 2, two modes of ring formation can be applied to lock the 9-*cis*-double bond. Mode A forms

<sup>\*</sup> Corresponding author. Tel./fax: +81 42 685 3729; e-mail: shi-ike@ pharm.teikyo-u.ac.jp

<sup>0968-0896/\$ -</sup> see front matter @ 2006 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmc.2006.04.043



Figure 1. Thrombomodulin induction in human endothelial cells by geometric isomers of RAs.



Figure 2. Two modes of ring formation of 9-cis-locked RA.

a ring system inside the molecule, and mode B forms one outside. Some groups used mode A ring formation to lock the 11-cis-double bond in their studies on the mechanism of vision in which cis-trans isomerization of retinal tissue plays an important role.<sup>11-14</sup> Successful results were obtained in those studies, although mode A ring formation was considered to alter the three-dimensional skeleton of the parent cis-RA. Molecular mechanics calculations have been widely employed to investigate stable conformations of biologically interesting molecules, and Dawson's group has adopted this technique to discuss the functionally active conformation of 9-cis-RA based on comparison with RAR- and RXR-selective retinoids.<sup>15,16</sup> Mohamadi et al. conducted a similar computational analysis<sup>17</sup> of 9-cis-locked RAs 1a and 2a having five-membered ring systems as representative molecules for our design. Monte Carlo conformational searches<sup>18</sup> of **1a** and **2a** showed that they have different conformational properties. The most stable conformer of 9-cis-RA, which was selected as the functionally active conformer based on Jong et al.'s assumption,<sup>15</sup> is superimposed on the corresponding similar conformers of 1a or 2a to visualize their

differences (polytube model in Fig. 3, colored: 9-cis-RA and black: 9-cis-locked analogue).

Apparently, ring formation in **1a** causes distinct locations of the side chain and the terminal carboxyl group.



Figure 3. Molecular design of 9-cis-locked RA by inducing a ring system. Left, 1a superimposed on the most stable conformer of 9-cis-RA; right, 2a superimposed on the most stable conformer of 9-cis-RA.

On the contrary, mode B ring formation (2a) retains these locations nearly unchanged. Employment of a six-membered or seven-membered ring system resulted in analogous conformational properties. Katsuta et al. have reported the synthesis of 9-cis-RA analogues employing mode A ring formation (six-membered ring) and showed that it was less effective than 9-cis-RA.<sup>19</sup> Their result is consistent with our computational consideration because the carboxyl group of their derivative was also calculated to be differently positioned from that of 9-cis-RA. Since 2a was found to have a similar conformational property based on the inspection of this computational molecular modeling, we focused our attention on the 'outer locked' 9-cis-RAs (mode B ring formation). Recently, the cyclohexene derivative 2b (Ro 25-6603), which is one of our target molecules, has been reported to be an RXR-selective ligand.<sup>20,21</sup> Similarly, analogues of 9-cis-RA incorporating an alicyclic ring, which included our target molecules 2a-c, have been synthesized and their binding affinities for the RXR receptor were determined.<sup>22</sup> Conformationally defined 6-s-trans-RA analogues employing a similar ring formation have also been reported.<sup>23</sup>

In the first stage of this study, we prepared cycloalkane and benzene derivatives 2a-d. After performing biological evaluations showing that the cyclopentene derivative 2a was most effective (vide infra), we next examined the incorporation of heteroatoms into fivemembered ring systems (2e-h).

# 2.2. Chemistry

9-*cis*-Locked RA analogues **2a**-**h** were synthesized using two stereoselective carbon–carbon bond formation reactions as key steps<sup>22</sup> (Scheme 1).

In the first step, the Suzuki reaction of alkenyl triflates (**3a**-c, e, and g) or aryl iodides (**3d** and f) with alkenylborane **4** was employed to construct the 7*E*-double bond (RA numbering). This palladium-catalyzed cross-coupling has been known to proceed with complete retention of stereochemistry of the double bond,<sup>24,25</sup> and completely stereochemically defined coupling products **5a**-g were obtained in high yield (90%-quantitative). The esters **5a**-g thus obtained were converted to alcohols **6a**-g using an appropriate reducing reagent.



Scheme 1. Reagents: (a) LiAlH<sub>4</sub>, THF (for 6a-d, f); DIBAL, CH<sub>2</sub>Cl<sub>2</sub>, (for 6e, g); (b) MnO<sub>2</sub>, pet. ether (for 7a-d); (COCl)<sub>2</sub>, DMSO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, (for 7e-g); (COCl)<sub>2</sub>, DMSO, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, then MnO<sub>2</sub>, pet. ether (for 7h from 6g); (c) NaOH, aq EtOH.

In the case of dihydrofuran and dihydrothiophene derivatives (6e and g), DIBAL was used in place of LiAlH<sub>4</sub> because ring cleavage of these heterocycles was observed. After conversion of 6a-g to aldehydes 7a-g in MnO<sub>2</sub> or Swern oxidations, Horner-Emmons olefination resulted in a stereoselective coupling reaction to form an 11E-double bond (RA numbering) in 90-98% yield.<sup>26</sup> Thiophene aldehyde 7h was prepared by oxidative aromatization of 7g and was similarly converted into 8h. Although a small amount (ca. <5% estimated by <sup>1</sup>H NMR) of stereoisomers was formed in the olefination, they were easily removed by recrystallization at the final stage. Hydrolysis of esters 8a-h by NaOH in aqueous EtOH afforded the desired acids 2a-h. In this straightforward synthesis, RA analogues were prepared stereoselectively in high overall yield (65–81%).

#### 2.3. Biological activity

HUVECs were treated with various concentrations of ATRA, 9-*cis*-RA, and synthesized RA analogues **2a**–**d** for 24 h, and TM antigen levels in cell lysates were measured in an enzyme immunoassay.<sup>27</sup>

As shown in Figure 4, the effects of 9-cis-RA and cyclopentene derivative **2a** were dose-dependent and the TM antigen levels increased to 150% of the baseline level at the dose of RAs of 0.001  $\mu$ M and to 180% at 0.01  $\mu$ M. Cyclohexene derivative **2b** showed a similar activity to 9-cis-RA at 0.001  $\mu$ M, but lower activity was observed at higher concentrations (1–10  $\mu$ M). The other derivatives **2c** and **d** were less potent in this assay.

The effects of heterocyclic analogues 2e-h were also measured at the concentrations of 0.001  $\mu$ M, 0.1  $\mu$ M, and 10  $\mu$ M (Fig. 5). Dihydrofuran and dihydrothiophene derivatives 2e and g increased TM antigen levels to 200% of the baseline level at 0.1  $\mu$ M, which was comparable to the increase induced by 2a. Aromatization of heterocycles (2f and h) significantly lowered TM induction potency.

## 2.4. Discussion

Based on the result that 9-cis-RA induced TM most effectively, we designed RA analogues that retain the conformational property of 9-cis-RA. The computational conformational searches of RA analogues showed that **2a–c** could have a low-energy conformation similar to the 'active conformation' of 9-cis-RA. Incorporation of a five-membered ring system especially modifies the spatial orientation of the side chain relative to 9-cis-RA, as shown in Figure 2. On the other hand, a similar conformation of the aromatic analogue 2d was calculated to locate ca. +1 kcal/mol above the global minimum, and the distinct conformation was found to be stable. This conformational analysis suggested a relation between the conformational properties and the activity of TM induction. The five-membered ring system (in 2a, e, and g), which mimicked the most stable conformation of 9-cis-RA, could be used as an effective lock with retention of biological activity. The low potency of TM induction by the aromatic analogues 2d, f, and **h** was also consistent with the conformational properties mentioned above. Although it was difficult to determine the relation between ring size and activity, it appears plausible that the lower activity of 2b and c was the result of changes in steric bulk caused by introduction of the six- or seven-membered ring system.

Since the analogues described herein are designed to mimic 9-*cis*-RA, which activates both RARs and RXRs, it is conceivable that they are pan-agonists (activating both RARs and RXRs). However, recent reports on RAR- and RXR-selective ligands,<sup>22,23</sup> in which the cyclohexene derivative **2b** was shown to be highly



Figure 4. TM induction by 9-cis-locked RA analogues (2a-d).



Figure 5. TM induction by 9-cis-locked RA analogues (2e-h).

RXR selective, have suggested that the 9-*cis*-locked analogues are also RXR selective. Although there are many reports on the RXR selectivity of 9-*cis*-RA,<sup>28,29</sup> we have not clarified which receptor(s), RAR and/or RXR, is involved in the expression of TM induction; a study concerning the precise role of RAs in the TM induction is now underway.

### 3. Conclusion

Molecular design aided by computational chemistry efficiently creates 9-*cis*-locked RA analogues that retain the 'active conformation' of the parent 9-*cis*-RA using appropriate ring formation. The analogues thus designed were synthesized stereoselectively and, as expected, induced TM with potency comparable to that of 9-*cis*-RA. It is also noteworthy that the synthesis of the analogues is very efficient (65–88% overall yield) and thus provides a large quantity of the compounds for various types of biological evaluation.

## 3.1. Experimental

**3.1.1. General.** Melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. IR spectra were recorded on a JASCO FT/IR-8000 spectrometer. <sup>1</sup>H NMR spectra were recorded on a JEOL GSX-400 spectrometer (400 MHz). High and low mass spectra were conducted on an JEOL SX-102A spectrometer. Elemental analysis was performed by Perkin-Elmer Series II CHNS/O analyzer 2400. Tetrahydrofuran and benzene were distilled from sodium metal prior to use. Methylene chloride was distilled from CaH<sub>2</sub>. Column chromatography was performed by using BW-820 (Fuji Silysia) or Wakogel C-200, and thin-layer chromatography was carried out on 0.25-mm E. Merck precoated silica-gel glass plate (Art. 5715).

**3.1.2. Computer modeling.** Monte Carlo conformational searches<sup>18</sup> were performed using MacroModel 4.0<sup>17</sup> with the MM2\* derivative of the MM2 forcefield.

3.1.3. Ethyl 2-[(E)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenvilcyclopent-1-enecarboxylate (5a).<sup>22</sup> To a solution of Pd(PPh<sub>3</sub>)<sub>4</sub> 28 mg (0.024 mmol) in benzene (2 ml) was added enol triflate 3a 172 mg (0.60 mmol) in 3 ml of benzene. After stirring for 15 min under Ar, 0.9 M benzene solution of alkenylborane  $4^{28}$  1.0 ml (0.90 mmol) and 4.0 M KOH aqueous solution 0.23 ml (0.90 mmol) were added at room temperature. The mixture was heated to reflux for 1 h, and the residual organoborane was oxidized by 3 M NaOH (0.1 ml) and 30% H<sub>2</sub>O<sub>2</sub> (0.1 ml) for 1 h at room temperature. The reaction mixture was filtered through Celite, and the filtrate was extracted with ether. The organic layer was washed with water and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated and the crude residue was purified by silica gel column chromatography (hexane/ethyl acetate 40:1 as an eluent) to afford 5a 167 mg (97% yield) as a colorless oil. IR (neat) 2930, 1701, 1227, 1107 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.07 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.30 (3H, t, J = 7.0 Hz, COOCH<sub>2</sub>CH<sub>3</sub>), 1.47 (2H, m, CH<sub>2</sub>), 1.62 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 1.88 (2H, m, CH<sub>2</sub>), 2.04 (2H, t, J = 6.3 Hz, CH<sub>2</sub>), 2.73 (4H, t, J = 7.6 Hz,  $2(CH_2)$ ), 4.21 (2H, q, J = 7.0 Hz,  $COOCH_2CH_3$ ), 6.21 (1H, d, J = 16.5 Hz, C=CH), 7.34 (1H, d, J = 16.5 Hz, C=CH). MS (EI) m/z 288 (M<sup>+</sup>).

Similar reactions of **3b**, **3c**, **3e**, and **3g** gave the following compounds.

**3.1.4. Ethyl 2-[(***E***)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]cyclohex-1-enecarboxylate (5b).<sup>22</sup> Yield: 98%. IR (neat) 2930, 1709, 1208 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) \delta 1.02 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.30 (3H, t, J = 7.0 Hz, COOCH<sub>2</sub>CH<sub>3</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.61 (2H, m, CH<sub>2</sub>), 1.67 (4H, m, 2(CH<sub>2</sub>)), 1.72 (3H, d, J = 0.6 Hz, CH<sub>3</sub>), 2.01 (2H, t, J = 6.2 Hz, CH<sub>2</sub>), 2.39**  (4H, m, 2(CH<sub>2</sub>)), 4.21 (2H, q, J = 7.0 Hz, COOCH<sub>2</sub>CH<sub>3</sub>), 6.32 (1H, d, J = 16.3 Hz, C=CH), 6.83 (1H, d, J = 16.3 Hz, C=CH). MS (EI) m/z 302 (M<sup>+</sup>).

**3.1.5. Ethyl 2-[(***E***)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]cyclohept-1-enecarboxylate (5c).<sup>22</sup> Yield: 95%. IR (neat) 2924, 1703, 1200 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) \delta 1.02 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.29 (3H, t, J = 7.1 Hz, COOCH<sub>2</sub>CH<sub>3</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.51– 1.64 (6H, m, 3(CH<sub>2</sub>)), 1.71 (3H, d, J = 0.6 Hz, CH<sub>3</sub>), 1.76 (2H, m, CH<sub>2</sub>), 2.01 (2H, t, J = 6.3 Hz, CH<sub>2</sub>), 2.53 (4H, m, 2(CH<sub>2</sub>)), 4.20 (2H, q, J = 7.1 Hz, COOCH<sub>2</sub>CH<sub>3</sub>), 6.33 (1H, d, J = 16.2 Hz, C=CH), 6.74 (1H, d, J = 16.2 Hz, C=CH). MS (EI) m/z 316 (M<sup>+</sup>).** 

**3.1.6.** Methyl **4-**[*(E*)-2-(2,6,6-trimethylcyclohex-1enyl)ethenyl]-2,5-dihydrofuran-3-carboxylate (5e). Yield: 93%. IR (neat) 2930, 1715, 1237 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.05 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.47 (2H, m, CH<sub>2</sub>), 1.62 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.2 Hz, CH<sub>2</sub>), 3.77 (3H, s, COOCH<sub>3</sub>), 4.89 (2H, t, *J* = 4.2 Hz, OCH<sub>2</sub>), 4.99 (2H, t, *J* = 4.2 Hz, OCH<sub>2</sub>), 6.26 (1H, d, *J* = 16.6 Hz, C=CH), 7.19 (1H, d, *J* = 16.6 Hz, C=CH). MS (EI) *m/z* 276 (M<sup>+</sup>).

**3.1.7.** Methyl 4-[(*E*)-2-(2,6,6-trimethylcyclohex-1enyl)ethenyl]-2,5-dihydrothiophen-3-carboxylate (5g). Yield: 97%. IR (neat) 2928, 1715, 1599, 1435, 1219, 1103 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.05 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.47 (2H, m, CH<sub>2</sub>), 1.62 (2H, m, CH<sub>2</sub>), 1.76 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 3.77 (3H, s, COOCH<sub>3</sub>), 4.06 (2H, m, SCH<sub>2</sub>), 4.13 (2H, m, SCH<sub>2</sub>), 6.41 (1H, dd, *J* = 16.5, 0.6 Hz, C=CH), 7.43 (1H, d, *J* = 16.5 Hz, C=CH). MS (EI) *m*/*z* 292 (M<sup>+</sup>).

3.1.8. Methvl 2-[(E)-2-(2,6,6-trimethylcyclohex-1envl)ethenvllbenzoate (5d). To a solution of  $Pd(PPh_3)_4$ 17 mg (0.015 mmol) in benzene (1 ml) was added methyl o-iodobenzoate 3d 78 mg (0.30 mmol) in 1 ml of benzene. After stirring for 15 min under Ar, 0.9 M benzene solution of alkenylborane  $4^{24}$  0.50 ml (0.45 mmol) and 4.0 M KOH aqueous solution 0.34 ml (1.4 mmol) were added at room temperature. The mixture was heated to reflux for 2 h, and the residual organoborane was oxidized by 3 M NaOH (0.1 ml) and  $30\% \text{ H}_2\text{O}_2$  (0.1 ml) for 1 h at room temperature. The reaction mixture was filtered through Celite, and the filtrate was extracted with ether. The organic layer was washed with water and brine, and dried over MgSO<sub>4</sub>. The solvent was evaporated and the crude residue was purified by silica gel column chromatography (hexane/ethyl acetate 30:1 as an eluent) to afford **5d** 82 mg (96% yield) as a colorless oil. IR (neat) 2924, 1723, 1244, 1076 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.10 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.50 (2H, m, CH<sub>2</sub>), 1.65 (2H, m, CH<sub>2</sub>), 1.80 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 2.04 (2H, t, J = 6.3 Hz, CH<sub>2</sub>), 3.89  $(3H, s, COOCH_3)$ , 6.57 (1H, dd, J = 16.2, 0.9 Hz)C=CH), 7.06 (1H, d, J = 16.2 Hz, C=CH), 7.27 (1H, dt, J = 1.1, 7.6 Hz, Ar), 7.47 (1H, dt, J = 1.1, 7.6 Hz, Ar), 7.61 (1H, dd, J = 1.1, 7.6 Hz, Ar), 7.86 (1H, dd, J = 1.1, 7.6 Hz, Ar). MS (EI) m/z 284 (M<sup>+</sup>).

Similar reactions of 3f gave the following compound.

**3.1.9.** Methyl **4-**[*(E*)-2-(2,6,6-trimethylcyclohex-1enyl)ethenyl]furan-3-carboxylate (5f). Yield: 98%. IR (neat) 2928, 1728, 1148, 1094 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.06 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.63 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 2.03 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 3.83 (3H, s, COOCH<sub>3</sub>), 6.49 (1H, d, *J* = 16.6, C=CH), 6,55 (1H, d, *J* = 16.6 Hz, C=CH), 7.55 (1H, d, *J* = 1.7 Hz, Ar), 7.97 (1H, d, *J* = 1.7 Hz, Ar). MS (EI) *m*/*z* 274 (M<sup>+</sup>).

3.1.10. 2-[(E)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenvllcvclopent-1-enemethanol (6a).<sup>22</sup> To a suspension of LiAlH<sub>4</sub> 65 mg (1.7 mmol) in THF (5 ml) was added dropwise ester 5a 119 mg (0.41 mmol) in THF (3 ml) at 0 °C and the reaction mixture was stirred for 30 min. Saturated aqueous Na<sub>2</sub>SO<sub>4</sub> solution (1 ml) was slowly added to this mixture, and precipitate was removed by filtration through Celite. The filtrate was dried over MgSO<sub>4</sub> and concentrated in vacuo. Alcohol 6a 92 mg (90% yield) was obtained as a colorless oil after purification by silica gel column chromatography (hexane/ethyl acetate 8:1 as an eluent) IR (neat) 3312, 2926, 1456, 1030 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 1.01 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.61 (2H, m, CH<sub>2</sub>), 1.71 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 1.90 (2H, m, CH<sub>2</sub>), 2.01 (2H, t, J = 6.1 Hz, CH<sub>2</sub>), 2.58 (4H, t, J = 7.5 Hz, 2(CH<sub>2</sub>)), 4.33 (2H, s, CH<sub>2</sub>OH), 6.05 (1H, d, J = 15.8 Hz, C=CH), 6.35 (1H, d, J = 158 Hz, C=CH). MS (EI) m/z 246 (M<sup>+</sup>).

Similar reactions of **5b–5d** and **5f** gave the following compounds.

**3.1.11. 2-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethen**yl]cyclohex-1-enemethanol (6b)**.<sup>22</sup> Yield: 90%. IR (neat) 3227, 2924, 1458 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.01 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.61 (2H, m, CH<sub>2</sub>), 1.66 (4H, m, 2(CH<sub>2</sub>)), 1.70 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 2.00 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.26 (4H, m, 2(CH<sub>2</sub>)), 4.23 (2H, s, CH<sub>2</sub>OH), 6.11 (1H, dd, *J* = 16.2, 0.6 Hz, C=CH), 6.48 (1H, d, *J* = 16.2 Hz, C=CH). MS (EI) *m*/*z* 260 (M<sup>+</sup>).

**3.1.12. 2-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethen**yl]cyclohept-1-enemethanol** (**6c**).<sup>22</sup> Yield: 94%. IR (neat) 3252, 2923, 1451 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.01 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.45–1.55 (6H, m, 3(CH<sub>2</sub>)), 1.62 (2H, m, CH<sub>2</sub>), 1.70 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 1.78 (2H, m, CH<sub>2</sub>), 2.02 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.42 (4H, m, 2(CH<sub>2</sub>)), 4.26 (2H, s, CH<sub>2</sub>OH), 6.11 (1H, d, *J* = 15.9 Hz, C=CH), 6.38 (1H, d, *J* = 15.9 Hz, C=CH). MS (EI) *m*/*z* 256 (M<sup>+</sup>-H<sub>2</sub>O).

**3.1.13. 2-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethenyl]benzyl alcohol (6d). Yield: 95%. IR (neat) 3330, 2926, 1455, 1099 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 1.07 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.50 (2H, m, CH<sub>2</sub>), 1.65 (2H, m, CH<sub>2</sub>), 1.78 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 2.04 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 4.76 (2H, s, CH<sub>2</sub>OH), 6.58 (1H, dd, *J* = 16.2, 0.9 Hz, C=CH), 6.65 (1H, d, *J* = 16.2 Hz, C=CH), 7.24 (1H, dt, *J* = 1.4, 7.4 Hz, Ar), 7.30 (1H,

5105

dt, J = 1.4, 7.4 Hz, Ar), 7.37 (1H, dd, J = 1.4, 7.4 Hz, Ar), 7.54 (1H, dd, J = 1.4, 7.4 Hz, Ar). MS (EI) m/z 256 (M<sup>+</sup>).

**3.1.14. 4-**[*(E)*-**2-(2,6,6-Trimethylcyclohex-1-enyl)ethenyl]furan-3-methanol (6f).** Yield: 92%. IR (neat) 3330, 2928, 1458, 1055 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.05 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.63 (2H, m, CH<sub>2</sub>), 1.75 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 2.02 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 4.64 (2H, s, CH<sub>2</sub>OH), 6.17 (1H, d, *J* = 16.4, C=CH), 6.48 (1H, d, *J* = 16.4 Hz, C=CH), 7.39 (1H, d, *J* = 1.5 Hz, Ar), 7.45 (1H, d, *J* = 1.5 Hz, Ar), MS (EI) *m*/*z* 246 (M<sup>+</sup>).

3.1.15. 4-[(E)-2-(2,6,6-Trimethylcyclohex-1-enyl)ethenyl]-2,5-dihydrofuran-3-methanol (6e). To a solution of ester **5e** 104 mg (0.015 mmol) in 3 ml  $CH_2Cl_2$  was added 1.76 M DIBAL in hexane 0.64 ml (1.13 mmol) at -78 °C and the reaction mixture was stirred for 30 min at the same temperature. Methanol 0.3 ml and saturated aqueous Rochelle salt 4 ml were sequentially added to this mixture. After being stirred for 90 min at room temperature, the two-phase mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with brine and dried over MgSO<sub>4</sub>. The solvent was evaporated and the crude residue was purified by silica gel column chromatography (hexane/ethyl acetate 2:1 as an eluent) to afford 5e 90 mg (97% yield) as a colorless oil. IR (neat) 3407, 2928, 1456, 1362,  $1071 \text{ cm}^{-1}$ . <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.00 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.61 (2H, m, CH<sub>2</sub>), 1.71 (3H, s, CH<sub>3</sub>), 2.01  $(2H, t, J = 6.3 \text{ Hz}, CH_2), 4.40 (2H, s, CH_2OH), 4.84$  $(2H, d, J = 3.4 \text{ Hz}, \text{ OCH}_2), 4.86 (2H, d, J = 3.4 \text{ Hz},$ OCH<sub>2</sub>), 5.91 (1H, d, J = 16.3 Hz, C=CH), 6.24 (1H, d, J = 16.3 Hz, C=CH). MS (EI) m/z 248 (M<sup>+</sup>).

Similar reactions of 5g gave the following compound.

**3.1.16. 4-**[*(E)*-**2-**(**2,6,6-Trimethylcyclohex-1-enyl)ethenyl]-<b>2,5-dihydrothiophen-3-methanol** (**6g**). Yield: 97%. IR (neat) 3395, 2930, 1456, 1109 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.00 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.46 (2H, m, CH<sub>2</sub>), 1.62 (2H, m, CH<sub>2</sub>), 1.70 (3H, s, CH<sub>3</sub>), 2.01 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 3.99 (4H, m, CH<sub>2</sub>SCH<sub>2</sub>), 4.37 (2H, s, CH<sub>2</sub>OH), 6.07 (1H, d, *J* = 16.2 Hz, C=CH), 6.33 (1H, d, *J* = 16.2 Hz, C=CH). MS (EI) *m*/*z* 264 (M<sup>+</sup>).

**3.1.17. 2-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethenyl]cyclopent-1-enecarbaldehyde (7a). To a solution of alcohol **6a** 126 mg (0.51 mmol) in 20 ml of petroleum ether (boiling range 30–60 °C) was added 898 mg of MnO<sub>2</sub>. After being stirred for 3.5 h at room temperature, the reaction mixture was filtered through Celite and the filtrate was concentrated in vacuo to afford aldehyde **7a** 119 mg (95%) as a yellow oil. IR (neat) 2930, 1659, 1611, 1208 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.05 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.75 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 1.92 (2H, m, CH<sub>2</sub>), 2.06 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.68 (2H, t, *J* = 7.5 Hz, CH<sub>2</sub>), 2.82 (2H, m, CH<sub>2</sub>), 6.52 (1H, dd, *J* = 15.9, 0.9 Hz, C=CH), 6.98 (1H, d, *J* = 15.9 Hz, C=CH), 10.19 (1H, s, CHO). MS (EI) *m/z* 244 (M<sup>+</sup>). Similar reactions of **6b–d** gave the following compounds.

**3.1.18. 2-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethenyl]cyclohex-1-enecarbaldehyde (7b). Yield: 96%. IR (neat) 2932, 1661, 1154 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.03 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.66–1.72 (6H, m, 3(CH<sub>2</sub>)), 1.73 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 2.04 (2H, t, J = 6.3 Hz, CH<sub>2</sub>), 2.32 (2H, t, J = 6.1 Hz, CH<sub>2</sub>), 2.49 (2H, m, CH<sub>2</sub>), 6.46 (1H, dd, J = 15.9, 0.9 Hz, C=CH), 6.96 (1H, d, J = 15.9 Hz, C=CH), 10.31 (1H, s, CHO). MS (EI) m/z 258 (M<sup>+</sup>).

**3.1.19. 2-**[*(E)*-**2-**(**2,6,6-Trimethylcyclohex-1-enyl)ethenyl]cyclohept-1-enecarbaldehyde (7c).** Yield: 86%. IR (neat) 2924, 1659, 1165 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.43–1.50 (4H, m, 2(CH<sub>2</sub>)), 1.57–1.66 (4H, m, CH<sub>2</sub> and CH<sub>2</sub>), 1.74 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 1.80 (2H, m, CH<sub>2</sub>), 2.04 (2H, t, J = 6.4 Hz, CH<sub>2</sub>), 2.58 (4H, m, 2(CH<sub>2</sub>)), 6.40 (1H, dd, J = 15.9, 0.9 Hz, C=CH), 6.77 (1H, d, J = 15.9 Hz, C=CH), 10.13 (1H, s, CHO). MS (EI) m/z 272 (M<sup>+</sup>).

**3.1.20.** 2-[(*E*)-2-(2,6,6-Trimethylcyclohex-1-enyl)ethenyl]benzaldehyde (7d). Yield: 92%. IR (neat) 2928, 1698, 1595, 1188 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.09 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.51 (2H, m, CH<sub>2</sub>), 1.66 (2H, m, CH<sub>2</sub>), 1.81 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 2.06 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 6.60 (1H, dd, *J* = 16.1, 0.9 Hz, C=CH), 7.17 (1H, d, *J* = 16.1 Hz, C=CH), 7.38 (1H, dt, *J* = 7.6, 1.3 Hz, Ar), 7.55 (1H, dt, *J* = 7.6, 1.3 Hz, Ar), 7.59 (1H, dd, *J* = 7.6, 1.3 Hz, Ar), 7.84 (1H, dd, *J* = 7.6, 1.3 Hz, Ar), 10.33 (1H, s, CHO). MS (EI) *m*/z 254 (M<sup>+</sup>).

3.1.21. 4-[(E)-2-(2,6,6-Trimethylcyclohex-1-enyl)ethenyl]-2,5-dihydrofuran-3-carbaldehyde (7e). To a solution of oxalyl chloride 0.11 ml (1.3 mmol) in  $CH_2Cl_2$  (4 ml) was added DMSO 0.18 ml (2.5 mmol) at -78 °C under Ar and the mixture was stirred for 3 min. To this solution alcohol **6e** 240 mg (0.97 mmol) in  $CH_2Cl_2$  (3 ml) was added and stirring was continued for an additional 15 min. Triethylamine 0.68 ml (4.9 mmol) was added, and the reaction mixture was stirred for 5 min and allowed to warm to room temperature. Water 10 ml was added and the aqueous layer was reextracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layer was washed with brine and dried (MgSO<sub>4</sub>). Purification on silica gel column chromatography (hexane/ethyl acetate 10:1 as an eluent) afforded 7e 218 mg (92%) as a colorless oil. IR (neat) 2932, 1659, 1613,  $1225 \text{ cm}^{-1}$ . <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.06 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 2.07  $(2H, t, J = 6.3 Hz, CH_2), 4.93 (2H, t, J = 3.9 Hz,$ OCH<sub>2</sub>), 5.05 (2H, t, J = 3.9 Hz, OCH<sub>2</sub>), 6.38 (1H, d, J = 16.2 Hz, C = CH), 6.85 (1H, d, J = 16.2 Hz, C = CH), 10.11 (1H, s, CHO). MS (EI) *m*/*z* 246 (M<sup>+</sup>).

Similar reactions of **6a** and **6g** gave the following compounds.

**3.1.22. 4-**[(*E*)-**2-**(**2,6,6-Trimethylcyclohex-1-enyl)ethenyl]furan-3-carbaldehyde** (**7f).** Yield: 92%. IR (neat) 2928, 1694, 1530, 1146 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.06 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.63 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 2.03 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 6.48 (1H, d, *J* = 16.6 Hz, C=CH), 6.70 (1H, d, *J* = 16.6 Hz, C=CH), 7.57 (1H, s, Ar), 8.01 (1H, d, *J* = 1.6 Hz, Ar), 10.01 (1H, s, CHO). MS (EI) *m*/*z* 244 (M<sup>+</sup>).

**3.1.23. 4-**[*(E)*-**2-**(**2**,**6**,**6**-Trimethylcyclohex-1-enyl)ethenyl]-2,**5**-dihydrothiophen-3-carbaldehyde (7g). Yield: 88%. A yellow crystalline solid, mp 56–58 °C IR (nujol) 2932, 1665, 1456, 1173 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 1.05 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.76 (3H, s, CH<sub>3</sub>), 2.07 (2H, t, *J* = 6.4 Hz, CH<sub>2</sub>), 4.02 (2H, t, *J* = 3.7 Hz, SCH<sub>2</sub>), 4.22 (2H, t, *J* = 3.7 Hz, SCH<sub>2</sub>), 6.52 (1H, d, *J* = 16.0 Hz, C=CH), 6.99 (1H, d, *J* = 16.0 Hz, C=CH), 10.17 (1H, s, CHO). MS (EI) *m*/*z* 262 (M<sup>+</sup>).

3.1.24. 4-I(E)-2-(2.6.6-Trimethylcyclohex-1-envl)ethenvllthiophen-3-carbaldehvde (7h). To a stirred solution of aldehyde 7g 102 mg (0.39 mmol) in 15 ml of petroleum ether (boiling range 30-60 °C) was added 998 mg MnO<sub>2</sub>. After being stirred for 6 h at room temperature, the reaction mixture was filtered though Celite and the filtrate was concentrated in vacuo. Purification on silica gel column chromatography (hexane/ethyl acetate 10:1 as an eluent) afforded **7h** 89 mg (89%) as a yellow oil. IR (neat) 2928, 1686, 1435, 911 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.08 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.79 (3H, s, CH<sub>3</sub>), 2.04  $(2H, t, J = 6.3 \text{ Hz}, CH_2), 6.59 (1H, d, J = 16.4 \text{ Hz},$ C=CH), 6.93 (1H, d, J = 16.4 Hz, C=CH), 7.33 (1H, dd, J = 32, 0.8 Hz, Ar), 8.09 (1H, d, J = 3.2 Hz, Ar), 10.03 (1H, s, CHO). MS (EI) m/z 260 (M<sup>+</sup>).

3.1.25. Methyl (E,E)-3-methyl-5-[2-[(E)-2-(2,6,6-trimethvlcvclohex-1-envl)ethenvl|cvclopent-1-envl|penta-2,4-dienoate (8a). To LDA in THF (3 ml) prepared from 0.11 ml (0.75 mmol) of diisopropylamine and 0.51 ml of 1.5 M BuLi solution in hexane (0.75 mmol) was added the phosphate<sup>26</sup> 188 mg (0.75 mmol) at -78 °C under Ar and the reaction mixture was stirred for 15 min. A solution of aldehyde 7a 119 mg (0.38 mmol) in THF (2 ml) was then added at the same temperature, and the reaction mixture was allowed to warm to room temperature and stirred for further 15 min in the dark. Saturated aqueous NH<sub>4</sub>Cl solution was added and the mixture was extracted with ether, washed with brine, and dried (MgSO<sub>4</sub>). The solvent was removed, and chromatography of the residue on silica gel (hexane/ ethyl acetate 30:1 as an eluent) afforded methyl ester 8a 150 mg (91%) as a yellow crystalline solid, mp 77–79 °C. IR (nujol) 2924, 1719, 1154 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.75 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 1.93 (2H, m, CH<sub>2</sub>), 2.05 (2H, t, J = 6.3 Hz, CH<sub>2</sub>), 2.37 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 2.66 (4H, m, 2(CH<sub>2</sub>)), 3.71 (3H, s, COOCH<sub>3</sub>), 5.81 (1H, s, C=CH), 6.19 (1H, d, J = 15.8 Hz, C=CH), 6.20 (1H, d, J = 15.8 Hz, C = CH, 6.58 (1H, d, J = 15.8 Hz,C=CH), 7.12 (1H, d, J = 15.8 Hz, C=CH). MS (EI) m/z 340 (M<sup>+</sup>). HRMS Calcd for C<sub>23</sub>H<sub>32</sub>O<sub>2</sub>: 340.2403. Found: 340.2409.

Similar reactions of **7b–7h** gave the following compounds.

**3.1.26.** Methyl (*E,E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl]enta-2,4-dienoate (8b). Yield: 91%. A yellow oil. IR (nujol) 2926, 1715, 1605, 1154 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.69 (4H, m, 2(CH<sub>2</sub>)), 1.75 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 2.04 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 2.32 (2H, m, CH<sub>2</sub>), 2.36 (3H, d, *J* = 1.2 Hz, CH<sub>3</sub>), 2.40 (2H, m, CH<sub>2</sub>), 3.71 (3H, s, COOCH<sub>3</sub>), 5.81 (1H, s, C=CH), 6.23 (1H, d, *J* = 15.8 Hz, C=CH), 6.28 (1H, d, *J* = 15.8 Hz, C=CH), 6.40 (1H, d, *J* = 15.8 Hz, C=CH). MS (EI) *m*/*z* 354 (M<sup>+</sup>).

**3.1.27.** Methyl (*E,E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]cyclohept-1-enyl]penta-2,4-dienoate (8c). Yield: 92%. A yellow oil. IR (nujol) 2924, 1715, 1605, 1154 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.47–1.54 (6H, m, 3(CH<sub>2</sub>)), 1.64 (2H, m, CH<sub>2</sub>), 1.76 (3H, s, CH<sub>3</sub>), 1.77 (2H, m, CH<sub>2</sub>), 2.05 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.36 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 2.53 (4H, m, 2(CH<sub>2</sub>)), 3.71 (3H, s, COOCH<sub>3</sub>), 5.81 (1H, s, C=CH), 6.23 (1H, d, *J* = 15.9 Hz, C=CH), 6.30 (1H, d, *J* = 15.9 Hz, C=CH), 7.27 (1H, d, *J* = 15.9 Hz, C=CH). MS (EI) *m*/*z* 360 (M<sup>+</sup>).

**3.1.28.** Methyl (*E*,*E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]phenyl]penta-2,4-dienoate (8d). Yield: 92%. A yellow-green oil. IR (nujol) 2926, 1719, 1456, 1156 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.08 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.51 (2H, m, CH<sub>2</sub>), 1.65 (2H, m, CH<sub>2</sub>), 1.80 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 2.40 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 3.73 (3H, s, COOCH<sub>3</sub>), 5.90 (1H, s, C=CH), 6.49 (1H, dd, *J* = 16.2, 0.9 Hz, C=CH), 6.62 (1H, d, *J* = 16.2 Hz, C=CH), 6.70 (1H, d, *J* = 15.9 Hz, C=CH), 7.22–7.30 (3H, m, C=CH, Ar), 7.46 (1H, dd, *J* = 1.5, 7.6 Hz, Ar), 7.50 (1H, d, *J* = 7.6 Hz, Ar). MS (EI) *m/z* 350 (M<sup>+</sup>).

**3.1.29.** Methyl (*E,E*)-3-methyl-5-[2-](*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]2,5-dihydrofuran-3-yl]penta-2,4-dienoate (8e). Yield: 94%. A yellow crystalline solid, mp 78–81 °C. IR (nujol) 2926, 1717, 1595, 1154 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.03 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.63 (2H, m, CH<sub>2</sub>), 1.76 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.36 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 3.72 (3H, s, COOCH<sub>3</sub>), 4.89 (2H, d, *J* = 3.5 Hz, OCH<sub>2</sub>), 4.93 (2H, d, *J* = 3.5 Hz, OCH<sub>2</sub>), 5.82 (1H, s, C=CH), 6.02 (1H, d, *J* = 16.2 Hz, C=CH), 6.03 (1H, d, *J* = 16.2 Hz, C=CH), 6.44 (1H, d, *J* = 16.2 Hz, C=CH), 6.94 (1H, d, *J* = 16.2 Hz, C=CH). MS (EI) *m*/z 342 (M<sup>+</sup>). HRMS Calcd for C<sub>22</sub>H<sub>30</sub>O<sub>3</sub>: 342.2195. Found: 342.2196.

**3.1.30.** Methyl (*E,E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]furan-3-yl]penta-2,4-dienoate (8f). Yield: 96%. A yellow oil. IR (neat) 2928, 1717, 1613, 1237, 1157 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.06 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.77 (3H, s, CH<sub>3</sub>), 2.04 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.37 (3H, s,

CH<sub>3</sub>), 3.72 (3H, s, COOCH<sub>3</sub>), 5.80 (1H, s, C=CH), 6.17 (1H, d, J = 16.5 Hz, C=CH), 6.38 (1H, d, J = 16.5 Hz, C=CH), 6.61 (1H, d, J = 16.2 Hz, C=CH), 6.84 (1H, d, J = 16.2 Hz, C=CH), 7.45 (1H, s, Ar), 7.55 (1H, s, Ar). MS (EI) m/z 340 (M<sup>+</sup>).

**3.1.31.** Methyl (*E,E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]-2,5-dihydrothiophen-3-yl]penta-2,4-dienoate (8g). Yield: 94%. A yellow crystalline solid, mp 145–146 °C. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.03 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.75 (3H, s, CH<sub>2</sub>), 2.05 (2H, t, *J* = 6.0 Hz, CH<sub>2</sub>), 2.37 (3H, d, *J* = 0.9 Hz, CH<sub>2</sub>), 3.72 (3H, s, COOCH<sub>3</sub>), 4.01 (2H, m, SCH<sub>2</sub>), 4.06 (2H, m, SCH<sub>2</sub>), 5.84 (1H, s, C=CH), 6.20 (2H, d, *J* = 16.0 Hz, C=CH), 6.56 (1H, d, *J* = 16.0 Hz, C=CH). MS (EI) *m*/*z* 358 (M<sup>+</sup>). HRMS Calcd for C<sub>22</sub>H<sub>30</sub>O<sub>2</sub>S: 358.1997. Found: 358.1996.

**3.1.32.** Methyl (*E,E*)-3-methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]-thiophen-3-yl]penta-2,4-dienoate (8h). Yield: 93%. A yellow oil. IR (neat) 2928, 1717, 1613, 1237, 1156 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.07 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.50 (2H, m, CH<sub>2</sub>), 1.65 (2H, m, CH<sub>2</sub>), 1.78 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.39 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 3.72 (3H, s, COOCH<sub>3</sub>), 5.86 (1H, s, H-14), 6.36 (1H, d, *J* = 16.2 Hz, C=CH), 6.48 (1H, d, *J* = 16.2 Hz, C=CH), 6.68 (1H, d, *J* = 16.0 Hz, C=CH), 6.98 (1H, d, *J* = 16.0 Hz, C=CH), 7.24 (1H, d, *J* = 3.2 Hz, Ar), 7.39 (1H, d, *J* = 3.2 Hz, Ar). MS (EI) *m/z* 356 (M<sup>+</sup>).

3.1.33. (E,E)-3-Methyl-5-[2-](E)-2-(2,6,6-trimethylcyclohex-1-envl)ethenvl]cyclopent-1-envl]penta-2,4-dienoic acid  $(2a)^{22}$ . To a stirred solution of ester 8a 162 mg (0.48 mmol) in 3 ml of ethanol was added 3.0 ml of 6.0 M KOH in the dark under Ar, and the reaction mixture was heated to 60 °C for 1 h. After hydrolysis was complete (by TLC), ethanol was evaporated in vacuo, and the mixture was cooled to 0 °C and acidified with 10% aqueous HCl. The organics were extracted with ether, washed with brine, dried over MgSO<sub>4</sub>, and concentrated in vacuo. The product was purified by column chromatography on silica gel (CH2Cl2/ether 5:1) to give 149 mg of desired acid 2a (89%) as a yellow solid. Recrystallization from CH<sub>2</sub>Cl<sub>2</sub>/hexane afforded yellow needles, mp 193–194 °C. IR (nujol) 3445, 2924, 1684, 1156 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>3</sub>), 1.76 (3H, s, CH<sub>3</sub>), 1.94 (2H, m, CH<sub>2</sub>), 2.05 (2H, t, J = 6.0 Hz, CH<sub>2</sub>), 2.38 (3H, s, CH<sub>2</sub>), 2.67 (4H, m, 2(CH<sub>2</sub>)), 5.84 (1H, s, C=CH), 6.20 (1H, d, J = 15.8 Hz, C=CH) 6.23 (1H, d, J = 15.8 Hz, C=CH), 6.59 (1H, d, J = 15.8 Hz, C=CH), 7.16 (1H, d, J = 15.8 Hz, C=CH). Anal. Calcd for C<sub>22</sub>H<sub>30</sub>O<sub>2</sub>/0.75H<sub>2</sub>O: C, 77.72; H, 9.34. Found: C, 77.75; H, 9.34.

Similar reactions of **8b-h** gave the following compounds.

**3.1.34.** (*E,E*)-**3-Methyl-5-[2-[**(*E*)-**2-(2,6,6-trimethylcyclohex-1-enyl]ethenyl]cyclohex-1-enyl]penta-2,4-dienoic acid** (**2b**).<sup>22</sup> Yield: 99%. Yellow needles, mp 165–167 °C (CH<sub>2</sub>Cl<sub>2</sub>/hexane). IR (nujol) 3445, 2924, 1682,

1651 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.70 (4H, m, 2(CH<sub>2</sub>)), 1.75 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, J = 6.1 Hz, CH<sub>2</sub>), 2.33 (2H, m, CH<sub>2</sub>), 2.37 (3H, d, J = 0.9 Hz, CH<sub>3</sub>), 2.40 (2H, m, CH<sub>2</sub>), 5.83 (1H, s, C=CH), 6.24 (1H, d, J = 15.8 Hz, C=CH), 6.31 (1H, d, J = 15.8 Hz, C=CH), 6.31 (1H, d, J = 15.8 Hz, C=CH), 7.38 (1H, d, J = 15.8 Hz, C=CH). MS (EI) *m*/*z* 340 (M<sup>+</sup>). Anal. Calcd for C<sub>23</sub>H<sub>32</sub>O<sub>2</sub>/0.75H<sub>2</sub>O: C, 78.03; H, 9.54. Found: C, 77.65; H, 9.21.

**3.1.35.** (*E*,*E*)-3-Methyl-5-[2-](*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]cyclohept-1-enyl]penta-2,4-dienoic acid (2c)<sup>22</sup>. Yield: 96%. Yellow needles, mp 143–145 °C (CH<sub>2</sub>Cl<sub>2</sub>/hexane). IR (nujol) 3445, 2924, 1684 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.47–1.55 (6H, m, 3C(CH<sub>3</sub>)<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.76 (3H, s, CH<sub>3</sub>), 1.78 (2H, m, CH<sub>2</sub>), 2.05 (2H, t, *J* = 6.1 Hz, CH<sub>2</sub>), 2.37 (3H, s, CH<sub>3</sub>), 2.53 (4H, m, 2(CH<sub>2</sub>)), 5.83 (1H, s, C=CH) 6.24 (1H, d, *J* = 15.8 Hz, C=CH), 6.33 (1H, d, *J* = 15.8 Hz, C=CH), 6.62 (1H, d, *J* = 15.8 Hz, C=CH). MS (EI) *m/z* 354 (M<sup>+</sup>). Anal. Calcd for C<sub>24</sub>H<sub>34</sub>O<sub>2</sub>/0.5H<sub>2</sub>O: C, 79.29; H, 9.70. Found: C, 78.90; H, 9.65.

**3.1.36.** (*E*,*E*)-3-Methyl-5-[2-](*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]phenyl]penta-2,4-dienoic acid (2d). Yield: 98%. Yellow plates, mp 176–178 °C (CH<sub>2</sub>Cl<sub>2</sub>/hexane). IR (neat) 3445, 2924, 1682, 1458 cm<sup>-1.</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.09 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.51 (2H, m, CH<sub>2</sub>), 1.66 (2H, m, CH<sub>2</sub>), 1.81 (3H, s, CH<sub>3</sub>), 2.06 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 2.42 (3H, s, CH<sub>3</sub>), 5.93 (1H, s, C=CH), 6.50 (1H, d, *J* = 16.2 Hz, C=CH), 6.62 (1H, d, *J* = 16.2 Hz, C=CH), 6.62 (1H, d, *J* = 16.2 Hz, C=CH), 7.23–7.32 (2H, m, Ar), 7.32 (1H, d, *J* = 15.9 Hz, C=CH), 7.47 (1H, dd, *J* = 7.6, 1.0 Hz, Ar), 7.52 (1H, dd, *J* = 7.6, 1.0 Hz, Ar). MS (EI) *m*/*z* 336 (M<sup>+</sup>). Anal. Calcd for C<sub>23</sub>H<sub>28</sub>O<sub>2</sub>: C, 82.10; H, 8.39. Found: C, 81.84; H, 8.41.

**3.1.37.** (*E*,*E*)-3-Methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]-2,5-dihydrofuran-3-yl]penta-2,4-dienoic acid (2e). Yield: 94%. Yellow needles, mp 224–226 °C (MeOH). IR (neat) 2926, 1694, 1591, 1458, 1181 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.03 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.48 (2H, m, CH<sub>2</sub>), 1.63 (2H, m, CH<sub>2</sub>), 1.76 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.0 Hz, CH<sub>2</sub>), 2.37 (3H, s, CH<sub>3</sub>), 4.91 (2H, d, *J* = 3.5 Hz, OCH<sub>2</sub>), 4.94 (2H, d, *J* = 3.5 Hz, OCH<sub>2</sub>), 5.85 (1H, s, C=CH) 6.02 (2H, d, *J* = 16.0 Hz, 2(C=CH)), 6.44 (1H, d, *J* = 16.0 Hz, C=CH), 6.98 (1H, d, *J* = 16.0 Hz, C=CH). MS (EI) *m*/*z* 328 (M<sup>+</sup>). Anal. Calcd for C<sub>21</sub>H<sub>28</sub>O<sub>3</sub>: C, 82.10; H, 8.39. Found: C, 76.47; H, 8.73.

**3.1.38.** (*E*,*E*)-3-Methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]furan-3-yl]penta-2,4-dienoic acid (2f). Yield: 97%. Colorless needles, mp 175–177 °C (MeOH). IR (nujol) 2928, 1692, 1599, 1574, 1456, 1061 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.06 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.78 (3H, s, CH<sub>3</sub>), 2.04 (2H, t, *J* = 6.0 Hz, CH<sub>2</sub>), 2.38 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 5.83 (1H, s, C=CH), 6.17 (1H, d, *J* = 16.2 Hz, C=CH), 6.39 (1H, d, *J* = 16.2 Hz, C=CH) 6.40 (1H, d, J = 16.0 Hz, C=CH), 6.88 (1H, d, J = 16.0 Hz, C=CH), 7.46 (1H, s, Ar), 7.57 (1H, d, J = 1.2 Hz, Ar). MS (EI) m/z 326 (M<sup>+</sup>). Anal. Calcd for C<sub>21</sub>H<sub>26</sub>O<sub>3</sub>: C, 77.27; H, 8.03. Found: C, 77.11; H, 7.96.

**3.1.39.** (*E*,*E*)-3-Methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]-2,5-dihydrothiophen-3-yl]penta-2,4-dienoic acid (2g). Yield: 96%. Yellow needles, mp 231–234 °C (AcOEt). IR (nujol) 3422, 2924, 1688, 1590, 1458, 1197 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.04 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.49 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.75 (3H, s, CH<sub>3</sub>), 2.07 (2H, t, *J* = 6.3 Hz, CH<sub>2</sub>), 2.38 (3H, d, *J* = 0.6 Hz, CH<sub>3</sub>), 4.01 (2H, d, *J* = 3.4 Hz, SCH<sub>2</sub>), 4.07 (2H, d, *J* = 3.4 Hz, SCH<sub>2</sub>), 5.87 (1H, s, C=CH) 6.21 (1H, d, *J* = 16.0 Hz, C=CH), 6.56 (1H, d, *J* = 16.0 Hz, C=CH), 7.12 (1H, d, *J* = 16.0 Hz, C=CH). MS (EI) *m/z* 344 (M<sup>+</sup>). Anal. Calcd for C<sub>21</sub>H<sub>28</sub>O<sub>2</sub>S: C, 73.21; H, 8.19; S, 9.31. Found: C, 72.96; H, 8.38; S, 9.32.

**3.1.40.** (*E*,*E*)-3-Methyl-5-[2-[(*E*)-2-(2,6,6-trimethylcyclohex-1-enyl)ethenyl]thiophen-3-yl]penta-2,4-dienoic acid (2h). Yield: 97%. Colorless needles, mp 172–174 °C (AcOEt/hexane). IR (neat) 2926, 1684, 1595, 1256, 1176 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  1.08 (6H, s, C(CH<sub>3</sub>)<sub>2</sub>), 1.50 (2H, m, CH<sub>2</sub>), 1.64 (2H, m, CH<sub>2</sub>), 1.79 (3H, s, CH<sub>3</sub>), 2.05 (2H, t, *J* = 6.2 Hz, CH<sub>2</sub>), 2.40 (3H, d, *J* = 0.9 Hz, CH<sub>3</sub>), 5.89 (1H, s, C=CH), 6.36 (1H, d, *J* = 16.2 Hz, C=CH), 6.48 (1H, d, *J* = 16.2 Hz, C=CH) 6.71 (1H, d, *J* = 16.2 Hz, C=CH), 7.02 (1H, d, *J* = 16.2 Hz, C=CH), 7.25 (1H, d, *J* = 3.4 Hz, Ar), 7.39 (1H, d, *J* = 3.4 Hz, Ar). MS (EI) *m*/*z* 356 (M<sup>+</sup>). Anal. Calcd for C<sub>21</sub>H<sub>26</sub>O<sub>2</sub>S: C, 73.64; H, 7.65; S, 9.36. Found: C, 73.57; H, 7.82; S, 9.16.

# 3.2. Measurement of thrombomodulin antigen

9-cis-RA analogues were dissolved in dimethyl sulfoxide to a stock concentration of 10 mM and stored at -80 °C. All experiments were performed in subdued light, and tubes containing RA analogues were covered with aluminum foil. Controls were run using the same concentration of dimethyl sulfoxide as present and this concentration of diluent had no effect.

Endothelial cells were isolated from human umbilical cord veins, and were cultured in Dulbecco's modified Eagle's medium (Flow Laboratories) supplemented with 20% fetal calf serum (FCS; Filtron), 20 µg/ml endothelial cell growth supplement (Collaborative Research), 100 µg/ml heparin (Sigma), 50 µg/ml penicillin, and 50 µg/ml streptomycin in a humidified atmosphere of 5% CO<sub>2</sub> at 37 °C as described previously.<sup>3</sup> Cells were used for the experiments within three passages. The confluent cells on 96 well gelatin-coated plate (Falcon; Becton Dickinson) were incubated in above culture medium containing retinoids for 24 h. After the incubation, conditioned medium was discarded and the monolayer was used for the assay of thrombomodulin antigen.

Cells were washed three times with Hanks' balanced salt solution (pH 7.4), and then solubilized for 1 h at 4 °C

with 50 mM Tris–HCl (pH 7.4) containing 0.15 M NaCl, 0.5% Triton X-100, and 1 mM benzamidine hydrochloride. Triton X-100-insoluble material was removed by centrifugation (18,000g for 15 min at 4 °C), and TM antigen levels in the cell lysates were measured by an enzyme immunoassay using mouse monoclonal anti-human TM antibodies as previously described. Purified human placental TM was used as a standard.

# Acknowledgments

We are grateful to Misses K. Ichikawa, J. Shimode, and J. Nonobe for spectroscopic measurements. This study was performed through Special Coordination Funds of the Science and Technology Agency of the Japanese Government. Partial financial support from the Ministry of Education, Science, Sports, and Culture of Japan is also acknowledged.

### **References and notes**

- 1. Sporn, M. B.; Roberts, A. B.; Goodman, D.S. Eds.; *The Retinoids*, 2nd ed.; Raven: New York, 1994.
- Horie, S.; Kizaki, K.; Kazama, M. Biochem. J. 1992, 281, 149.
- 3. Ishii, H.; Horie, S.; Kizaki, K.; Kazama, M. Blood 1992, 80, 2556.
- 4. Esmon, C. T. Science 1987, 235, 1348.
- 5. Dittman, W. A.; Majerus, P. W. Blood 1990, 75, 329.
- Heyman, R. A.; Mangelsdorf, D. J.; Dyck, J. A.; Stein, R. B.; Eichele, G.; Evans, R. M.; Thaller, C. *Cell* **1992**, *68*, 397.
- Levin, A. A.; Sturzenbecker, L. J.; Kazmer, S.; Bosakowski, T.; Huselton, C.; Allenby, G.; Speck, J.; Kratzeisen, C.; Rosenberger, M.; Lovey, A.; Grippo, J. F. *Nature* 1992, 355, 359.
- Allenby, G.; Janocha, R.; Kazmer, S.; Speck, J.; Grippo, J. F.; Levin, A. A. J. Biol. Chem. 1994, 269, 16689.
- Horie, S.; Ishii, H.; Matsumoto, F.; Kusano, M.; Kizaki, K.; Matsuda, J.; Kazama, M. J. Biol. Chem. 2001, 276, 2440–2450.
- Ikegami, S.; Iimori, T.; Kazama, M.; Ishii, H.; Japanese Kokai Tokkyo Koho 1997, JP 09059207; *Chem. Abstr.* 1997, 126, 305659.
- Akita, H.; Tanis, S. P.; Adam, M.; Balogh-Nair, V.; Nakanishi, K. J. Am. Chem. Soc. 1980, 102, 6370.
- Mao, B.; Tsuda, M.; Ebrey, T. G.; Akita, H.; Balogh-Nair, V.; Nakanishi, K. *Biophys. J.* **1981**, *35*, 543.
- Ito, M.; Kodama, A.; Tsukida, K.; Fukada, Y.; Shichida, Y.; Yoshizawa, T. Chem. Pharm. Bull. 1982, 30, 1913.
- 14. Fukada, Y.; Shichida, Y.; Yoshizawa, T.; Ito, M.; Kodama, A.; Tsukida, K. *Biochemistry* **1984**, *23*, 5826.
- Jong, L.; Lehmann, J. M.; Hobbs, P. D.; Harlev, E.; Huffman, J. C.; Pfahl, M.; Dawson, M. I. J. Med. Chem. 1993, 36, 2605.
- Dawson, M. I.; Jong, L.; Hobbs, P. D.; Cameron, J. F.; Chao, W.-R.; Pfahl, M.; Lee, M.-O.; Shroot, B.; Pfahl, M. *J. Med. Chem.* **1995**, *38*, 3368.
- Mohamadi, F.; Richards, N. G. J.; Guida, W. C.; Liskamp, R.; Lipton, M.; Caufield, C.; Chang, G.; Hendrickson, T.; Still, W. C. J. Comput. Chem. 1990, 11, 440.
- Chang, G.; Guida, W. C.; Still, W. C. J. Am. Chem. Soc. 1989, 111, 4379.
- Katsuta, Y.; Aoyama, Y.; Osone, H.; Wada, A.; Uchiyama, S.; Kitamoto, T.; Masushige, S.; Kato, S.; Ito, M. *Chem. Pharm. Bull.* **1994**, *42*, 2659.

- 20. Brooks, S. C.; Kazmer, S.; Levin, A. A.; Yen, A. Blood 1996, 87, 227.
- Rusten, L. S.; Dybedal, I.; Blomhoff, H. K.; Blomhoff, R.; Smeland, E. B.; Jacobsen, S. E. W. *Blood* 1996, *87*, 1728.
- 22. Paz Otero, M.; Torrado, A.; Pazos, Y.; Sussman, F.; de Lera, A. R. J. Org. Chem. 2002, 67, 5876.
- Vaezi, M. F.; Alam, M.; Sani, B. P.; Rogers, T. S.; Simpson-Herren, L.; Wille, J. J.; Hill, D. L.; Doran, T. I.; Brouillette, W. J.; Muccio, D. D. J. Med. Chem. 1994, 37, 4499.
- 24. Miyaura, N.; Suzuki, A. J. Chem. Soc., Chem. Commun. 1979, 866.

- 25. Yanagi, T.; Oh-e, T.; Miyaura, N.; Suzuki, A. Bull. Chem. Soc. Jpn. 1989, 62, 3892.
- Safaryn, J. E.; Chiarello, J.; Chen, K.-M.; Joullié, M. M. Tetrahedron 1986, 42, 2635.
- Ishii, H.; Nakano, N.; Tsubouchi, J.; Ishikawa, T.; Uchiyama, H.; Hiraishi, S.; Tahara, C.; Miyajima, Y.; Kazama, M. *Thromb. Haemost.* **1990**, *63*, 157.
- Pazos, Y.; Iglesias, B.; de Lera, A. R. J. Org. Chem. 2001, 66, 8483.
- Cavasotto, C.; Liu, G.; James, S. Y.; Hobbs, P. D.; Peterson, V. J.; Bhattacharya, A. A.; Kolluri, S. K.; Zhang, X.-K.; Leid, M.; Abagyan, R.; Liddington, R. C.; Dawson, M. I. J. Med. Chem. 2004, 47, 4360.