



# A Novel Class of Inhibitors for Steroid 5 $\alpha$ -Reductase: Synthesis and Evaluation of Umbelliferone Derivatives

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**Abstract**—A series of umbelliferone derivatives was prepared and their 5 $\alpha$ -reductase type 1 inhibitory activities were evaluated in cell culture systems. Our studies have identified a new series of potent 5 $\alpha$ -reductase type 1 inhibitors and provided the basis for further development for the treatment of human endocrine disorders associated with overproduction of DHT by 5 $\alpha$ -reductase type 1. The preliminary structure–activity relationship was described to elucidate the essential structural requirements. © 2001 Elsevier Science Ltd. All rights reserved.

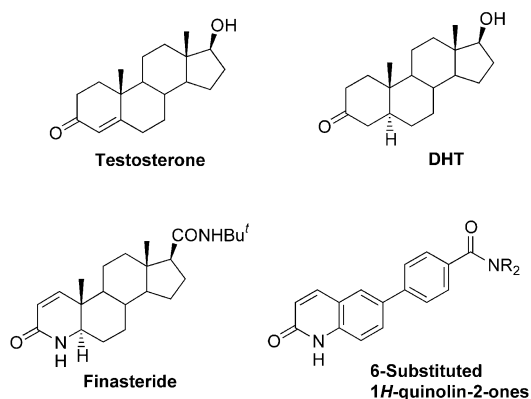
## Introduction

Steroid 5 $\alpha$ -reductase (5 $\alpha$ R) is an NADPH-dependent enzyme that catalyzes the reduction of testosterone (T) into the more potent androgen, dihydrotestosterone (DHT) (Fig. 1).<sup>1</sup> Two different 5 $\alpha$ -reductase isozymes, type 1 and type 2 (5 $\alpha$ R-1 and 5 $\alpha$ R-2) have been cloned, expressed, and characterized.<sup>1,2</sup> These two isozymes are differently distributed in the human tissues. While 5 $\alpha$ R-1 is present mainly in hair follicles, sebaceous glands of the skin (including the scalp) and liver, 5 $\alpha$ R-2 is found predominantly in the prostate, genital skin, seminal vesicles, epididymis, and liver.<sup>2,3</sup> It was reported that many human endocrine diseases such as acne, male pattern baldness, alopecia in men and hirsutism in women are due to elevated levels of DHT.<sup>4</sup>

A number of different classes of steroidal compounds that inhibit 5 $\alpha$ R have been designed and synthesized as therapeutic agents for the treatment of DHT-related pharmacological disorders.<sup>5,6</sup> In designing these inhibitors, mimics of the proposed enolate intermediate of the natural substrate in the reduction were usually employed.<sup>6,7</sup> This approach led to the introduction of finasteride<sup>8</sup> (Proscar<sup>®</sup>), a 4-azasteroid inhibiting mainly 5 $\alpha$ R-2 (Fig. 1). Recently, a number of simpler, non-

steroidal inhibitors have also been designed based on the structures of the related steroidal inhibitors. Selected examples of these nonsteroidal inhibitors are benzophenone carboxylic acids,<sup>9</sup> indole derivatives,<sup>6</sup> benzoquinolizinones<sup>10</sup> and hydrophenanthren-2-ones.<sup>11</sup> More recently, the evaluation of 6-substituted 1*H*-quinolin-2-ones (Fig. 1) as a new class of nonsteroidal inhibitors of 5 $\alpha$ R was reported by Hartmann.<sup>12</sup>

In the course of our study to find a potent 5 $\alpha$ R-1 inhibitor, we focused our attention on nonsteroidal inhibitors to avoid some of the well-known side effects of steroidal drugs. On the basis of the transition state



**Figure 1.** Chemical structures of testosterone, dihydrotestosterone (DHT), finasteride and 6-substituted 1*H*-quinolin-2-ones.

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paradigm along with Hartmann's results,<sup>12</sup> we opted for the synthesis and biochemical evaluation of the B-ring substituted coumarin derivatives. To the best of our knowledge, no coumarin derivatives have been reported as inhibitors of 5 $\alpha$ R. The coumarin skeleton can be considered as a mimetic of the proposed transition state of the natural substrate as well as bioisostere of quinolin-2-one. We designed and prepared a series of derivatives of umbelliferone (7-hydroxycoumarin), and screened their 5 $\alpha$ R inhibitory activities. Herein, we report our preliminary results of these studies.

### Synthesis

The synthesis of derivatives of umbelliferone **1** was outlined in Scheme 1. Propargyl ether **2** was obtained by heating the solution of umbelliferone **1** and propargyl bromide together with potassium carbonate and potassium iodide in DMF. Compound **3** was prepared according to a method of Pettus et al.<sup>13</sup> Reduction of **3** under H<sub>2</sub> gas in the presence of Lindlar catalyst and quinoline in EtOAc provided compound **4**.

8-Allyl-7-hydroxycoumarin **7** was a key intermediate which can easily be obtained from umbelliferone **1** in two steps. Treatment of umbelliferone **1** with allyl bromide in H<sub>2</sub>O/CH<sub>2</sub>Cl<sub>2</sub> two-phase solution in the presence of sodium hydroxide and tetrabutylammonium iodide afforded **5** in 82% yield, which was then heated with *p*-xylene in sealed tube at 260 °C to afford mainly the C-8 Claisen rearrangement product **7** together with its C-6 isomer **6** in a ratio of 9:1.<sup>14</sup>

Compound **7** was condensed with phenylisocyanate in the presence of pyridine in CH<sub>2</sub>Cl<sub>2</sub> at room temperature to furnish **8**. Stirring a solution of **7**, benzoyl chloride and pyridine in CH<sub>2</sub>Cl<sub>2</sub> at 0 °C gave the corresponding ester **9**. Acetylation of **7** using acetyl chloride and pyridine in THF at room temperature afforded ester **10**. Compounds **11**, **12**, **14**, and **15** were synthesized

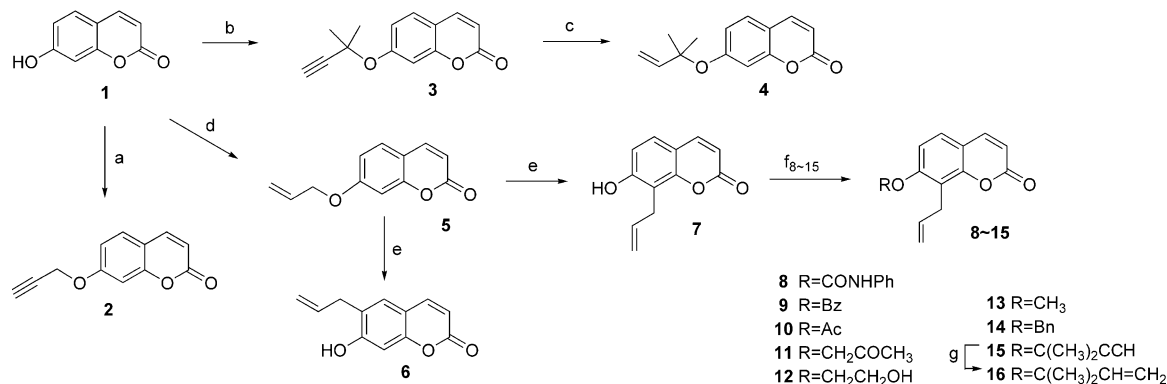
according to a general procedure<sup>15</sup> using **7** and its corresponding alkylhalide or benzylhalide together with potassium carbonate in DMF or acetone. Methylation of coumarin **7** with dimethyl sulfate in aqueous ethanol solution resulted in **13**. Hydrogenation of **15** was conducted in the same manner for compound **4** to afford **16**.

### Enzyme Inhibitory Activity and Discussion

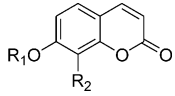
LNCaP cells are the androgen-sensitive human prostatic cancer cell line widely available for use in preclinical investigations<sup>16</sup> and it has been reported that 5 $\alpha$ R-1 is expressed in LNCaP cells but not 5 $\alpha$ R-2.<sup>17</sup> The existence of androgen sensitivity as well as 5 $\alpha$ -reductase activity in LNCaP cells offered us the opportunity to study the effects of inhibitors in the enzymatic and in vitro functional LNCaP responses to androgens in a single human cell line.

The compounds were evaluated for their steroid 5 $\alpha$ R-1 inhibitory activities in cell culture system using LNCaP cells according to a documented procedure with some modification.<sup>18</sup> Inhibitory effects were represented with the concentration ( $\mu$ g/mL) giving 50% inhibition (IC<sub>50</sub>) relative to the control (0.5% DMSO). In the case of highly active compounds, IC<sub>50</sub> values are also presented as  $\mu$ M. The obtained results were summarized in Table 1.

Umbelliferone **1** and 7-alkoxycoumarins **2**, **3**, and **5** do not display any inhibitory activity. However, 1',1'-dimethylallyloxycoumarin **4** shows potent inhibitory activity (IC<sub>50</sub> = 1.3  $\mu$ M) for the 5 $\alpha$ R-1. The considerably different activity between **1**–**3**, **5**, and **4** is possibly a result of conformational effects of geminal dimethyl group (the Thorpe–Ingold effect).<sup>19</sup> This observation led us to investigate the compounds **6** and **7** which are the Claisen rearrangement products of **5**. To our delight, 8-allyl-7-hydroxycoumarin **7** exhibits a potent inhibitory activity (IC<sub>50</sub> = 0.99  $\mu$ M) against 5 $\alpha$ R-1. The compound



**Scheme 1.** Reagents and conditions: (a) propargyl bromide, K<sub>2</sub>CO<sub>3</sub>, KI, DMF, 80 °C, 2 h, 98%; (b) (CF<sub>3</sub>CO)<sub>2</sub>O, (CH<sub>3</sub>)<sub>2</sub>C(OH)C $\equiv$ CH, DBU, then **1**, CuCl<sub>2</sub>·H<sub>2</sub>O, DBU, rt, 2 h, 61%; (c) H<sub>2</sub>, quinoline, Lindlar cat., EtOAc, rt, 1 h, 73%; (d) allyl bromide, NaOH, KI, cat. (*n*-Bu)<sub>4</sub>NI, H<sub>2</sub>O, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 h, 82%; (e) *p*-xylene, 260 °C, 17 h (90% for **7**, 10% for **6**); (f<sub>8</sub>) C<sub>6</sub>H<sub>5</sub>N=C=O, pyr, CH<sub>2</sub>Cl<sub>2</sub>, rt, 2 days, 85%; (f<sub>9</sub>) C<sub>6</sub>H<sub>5</sub>COCl, pyr, CH<sub>2</sub>Cl<sub>2</sub>, 0 °C, 5 h, 87%; (f<sub>10</sub>) CH<sub>3</sub>COCl, pyr, THF, rt, overnight, 89%; (f<sub>11</sub>) CH<sub>3</sub>COCH<sub>2</sub>Cl, K<sub>2</sub>CO<sub>3</sub>, acetone, reflux, 4 h, 84%; (f<sub>12</sub>) BrCH<sub>2</sub>CH<sub>2</sub>OH, K<sub>2</sub>CO<sub>3</sub>, acetone, reflux, 1 day, 91%; (f<sub>13</sub>) (CH<sub>3</sub>)<sub>2</sub>SO<sub>4</sub>, NaOH, C<sub>2</sub>H<sub>5</sub>OH, H<sub>2</sub>O, rt, 5 h, 63%; (f<sub>14</sub>) BnBr, K<sub>2</sub>CO<sub>3</sub>, DMF, 60 °C, 3 h, 98%; (f<sub>15</sub>) CHCC(CH<sub>3</sub>)<sub>2</sub>Cl, K<sub>2</sub>CO<sub>3</sub>, DMF, 90 °C, 26 h, 56%; (g) H<sub>2</sub>, quinoline, Lindlar cat., EtOAc, rt, 5 h, 90%.

**Table 1.** Steroid 5 $\alpha$ -reductase type 1 inhibitory activities in cell culture systems using LNCaP cells


| Compound    | R <sub>1</sub>                                      | R <sub>2</sub>                     | IC <sub>50</sub> <sup>a</sup><br>( $\mu$ g/mL) [ $\mu$ M] |
|-------------|-----------------------------------------------------|------------------------------------|-----------------------------------------------------------|
| Finasteride |                                                     |                                    | 19.8 [53]                                                 |
| <b>1</b>    | H                                                   | H                                  | >20                                                       |
| <b>2</b>    | CH $\equiv$ CCH <sub>2</sub>                        | H                                  | >20                                                       |
| <b>3</b>    | CH $\equiv$ CC(CH <sub>3</sub> ) <sub>2</sub>       | H                                  | >20                                                       |
| <b>4</b>    | CH <sub>2</sub> =CHC(CH <sub>3</sub> ) <sub>2</sub> | H                                  | 0.3 [1.3]                                                 |
| <b>5</b>    | CH <sub>2</sub> =CHCH <sub>2</sub>                  | H                                  | >20                                                       |
| <b>6</b>    | (see Scheme 1)                                      | H                                  | 1.7 [8.4]                                                 |
| <b>7</b>    | H                                                   | CH <sub>2</sub> =CHCH <sub>2</sub> | 0.2 [0.99]                                                |
| <b>8</b>    | C <sub>6</sub> H <sub>5</sub> NHCO                  | CH <sub>2</sub> =CHCH <sub>2</sub> | 0.2 [0.62]                                                |
| <b>9</b>    | C <sub>6</sub> H <sub>5</sub> CO                    | CH <sub>2</sub> =CHCH <sub>2</sub> | 0.15 [0.49]                                               |
| <b>10</b>   | CH <sub>3</sub> CO                                  | CH <sub>2</sub> =CHCH <sub>2</sub> | 0.2 [0.82]                                                |
| <b>11</b>   | CH <sub>3</sub> COCH <sub>2</sub>                   | CH <sub>2</sub> =CHCH <sub>2</sub> | >20                                                       |
| <b>12</b>   | HOCH <sub>2</sub> CH <sub>2</sub>                   | CH <sub>2</sub> =CHCH <sub>2</sub> | >20                                                       |
| <b>13</b>   | CH <sub>3</sub>                                     | CH <sub>2</sub> =CHCH <sub>2</sub> | >4                                                        |
| <b>14</b>   | C <sub>6</sub> H <sub>5</sub> CH <sub>2</sub>       | CH <sub>2</sub> =CHCH <sub>2</sub> | >4                                                        |
| <b>15</b>   | HC $\equiv$ CC(CH <sub>3</sub> ) <sub>2</sub>       | CH <sub>2</sub> =CHCH <sub>2</sub> | >4                                                        |
| <b>16</b>   | H <sub>2</sub> C=CHC(CH <sub>3</sub> ) <sub>2</sub> | CH <sub>2</sub> =CHCH <sub>2</sub> | 13.6 [50.3]                                               |

<sup>a</sup>Duplicate samples were treated at nontoxic doses to obtain IC<sub>50</sub> values.

**6**, 6-allyl-7-hydroxycoumarin (IC<sub>50</sub> = 8.4  $\mu$ M) is also more active than finasteride (IC<sub>50</sub> = 53  $\mu$ M) against 5 $\alpha$ R-1, but less active than the corresponding C-8 isomer **7** by about 10-fold.

In the investigation of the substituent effects on the 7-hydroxyl group of **7**, the introduction of carbonyl groups (such as in compounds **8**, **9**, and **10**) resulted in a slight enhancement of 5 $\alpha$ R-1 inhibitory. However, these compounds are about 100 times more potent than finasteride. The introduction of alkyl groups into the 7-hydroxyl position was not suitable for increasing inhibitory potency (**13–16**). Replacement of the hydrophobic alkyl group with an acetyl group (**11**) or ethanol group (**12**) also caused a drop in potency. From these results, the existence of a carbonyl group-containing side chain at the C-7 hydroxyl position of 8-allylcoumarin appears to be desirable for potent inhibitory activity against 5 $\alpha$ R-1.

In conclusion, we have designed and evaluated the B-ring substituted umbelliferone derivatives as 5 $\alpha$ R-1 inhibitors. Several compounds showed potent inhibitory activity towards 5 $\alpha$ R-1 isozyme. This new series of potent 5 $\alpha$ R-1 inhibitors could be leads for the development of a drug for the treatment of human endocrine disorders associated with overproduction of DHT by 5 $\alpha$ R-1. Further studies for more potent inhibitors based on the above findings are in process in our laboratory.

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#### References and Notes

- Andersson, S.; Russell, D. W. *Proc. Natl. Acad. Sci. U.S.A.* **1990**, *87*, 3640.
- Russell, D. W.; Wilson, J. D. *Annu. Rev. Biochem.* **1994**, *63*, 25.
- Thigpen, A. E.; Silver, R. I.; Graul, A. J. *Clin. Invest.* **1993**, *92*, 903.
- (a) For reviews see: Abell, A. D.; Henderson, B. R. *Curr. Med. Chem.* **1995**, *2*, 583. (b) Holt, D. A.; Levy, M. A.; Metcalf, B. W. In *Advances in Medicinal Chemistry*; Maryanoff, B. E., Maryanoff, C. A. Eds.; JAI: London, 1993; Vol. 2, pp 1–29.
- For a review of comparative QSAR analysis, see: Kurup, A.; Garg, R.; Hansch, C. *Chem. Rev.* **2000**, *100*, 909.
- Igarashi, S.; Inami, H.; Hara, H.; Fujii, M.; Koutoku, H.; Oritani, H.; Mase, T. *Chem. Pharm. Bull.* **2000**, *48*, 382 and references therein.
- Ahmed, S.; Denison, S. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 409 and references therein.
- Rasmusson, G. H.; Reynolds, G. F.; Utne, T.; Jobson, R. B.; Primka, R. L.; Berman, C.; Brooks, J. R. *J. Med. Chem.* **1984**, *27*, 1690.
- Holt, D. A.; Yamashita, D. S.; Konialian-Beck, A. L.; Luengo, Y. I.; Abell, A. D.; Bergsma, D. J.; Levy, M. A. *J. Med. Chem.* **1995**, *38*, 13.
- Guarna, A.; Machetti, F.; Occhiato, E. G.; Scarpi, D. *J. Med. Chem.* **2000**, *43*, 3718.
- Abell, A. D. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2871.
- Baston, E.; Paluszczak, A.; Hartmann, R. W. *Eur. J. Med. Chem.* **2000**, *35*, 931.
- Pettus, Th. R. R.; Inoue, M.; Chen, X.-T.; Danishefsky, S. J. *J. Am. Chem. Soc.* **2000**, *122*, 6160.
- Cairns, N.; Harwood, L. M.; Astles, D. P.; Orr, A. J. *Chem. Soc., Perkin Trans. 1* **1994**, 3095.
- Wulff, H.; Rauer, H.; During, T.; Hanselmann, C.; Ruff, K.; Wrisch, A.; Grissmer, S.; Hansel, W. *J. Med. Chem.* **1998**, *41*, 4542.
- Horoszewicz, J. S.; Leong, S. S.; Kawinski, E.; Karr, J. P.; Rosenthal, H.; Chu, M. T.; Mirand, E. A.; Murphy, G. P. *Cancer Res.* **1983**, *43*, 1908.
- Negri-Cesi, P.; Poletti, A.; Colciago, A.; Magni, P.; Motta, M. *Prostate* **1998**, *34*, 283.
- (a) Reichert, W.; Jose, J.; Hartmann, R. W. *Arch. Pharm. (Weinheim)* **2000**, *333*, 201. (b) 5 $\alpha$ -Reductase assay: LNCaP cells were used for 5 $\alpha$ R-1 enzyme assay. In brief, cells were seeded at a density of 2 $\times$ 10<sup>5</sup> cells/mL/well in 24-well plates and cultured for 24 h in media containing 5% hormone-free fetal bovine serum. Then cells were treated with [<sup>3</sup>H]-testosterone and test samples (final 0.5% DMSO). After an additional 18 h of incubation, the analysis of [<sup>3</sup>H]-T and [<sup>3</sup>H]-DHT within the medium was carried out by thin-layer chromatography (TLC). The incubated medium was collected in a microcentrifuge tube and [<sup>3</sup>H]-T and [<sup>3</sup>H]-DHT in medium were extracted with equal volume of ethyl acetate via vigorous vortex. The portion of the ethyl acetate extract was dried in vacuo and resuspended in a 10  $\mu$ L ethyl acetate. Then, 10  $\mu$ L ethyl acetate was subjected to TLC with silica gel (Silica 60W, Merck, Darmstadt) using chloroform/methanol (96:4, v/v) as the solvent system. The measurement of [<sup>3</sup>H]-T and [<sup>3</sup>H]-DHT was performed by using an image plate reader (BAS-1500, Fuji Film, Tokyo). The conversion from T to DHT was calculated from the ratio of the radioactivity of DHT to the sum of the radioactivity of T and DHT. Inhibitory effects were represented with the concentration ( $\mu$ g/mL) giving 50% inhibition (IC<sub>50</sub>) relative to the control (0.5% DMSO).
- (a) Keese, R.; Meyer, M. *Tetrahedron* **1993**, *49*, 2055. (b) Kirby, A. J. *Adv. Phys. Org. Chem.* **1980**, *17*, 183. (c) Mandolini, L. *Adv. Phys. Org. Chem.* **1986**, *22*, 1.