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2-Aryl-3-methyloctahydrophenanthrene-2,3,7-triols as Potent Dissociated Glucocorticoid Receptor Agonists[†]

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Supporting Information

ABSTRACT: A significant improvement in agonist activity of the previously described 2-aryloctahydrophenanthrene-2,3,7-triol series of dissociated glucocorticoid receptor agonists (DAGRs) was achieved by modifying the substitution at C3 from (S)-3-hydroxy to (R)-3-hydroxy-3-methyl. The IC₅₀ of the prototype **13** in the efficacy assay measuring repression of IL-1 induced MMP-13 expression was 3.5 nM, exhibiting 87% of the maximal effect of dexamethasone (DEX). It displayed a dissociated



profile by exhibiting 42% of the maximal effect of DEX in a mouse mammary tumor virus (MMTV) luciferase reporter transactivation assay. Compound 13 and analogues containing heterocyclic replacements for the C2 phenyl and modified B rings showed high repression of TNF α production in human whole blood, with IC₅₀ values (43–167 nM) approaching the level of DEX (21 nM). On the basis of X-ray structures and force field calculations, the overall potency of this series was attributed to a favorable conformation of the C2 α phenyl, induced by the neighboring C3 α methyl.

INTRODUCTION

Glucocorticoid receptor (GR) agonists,¹ such as prednisone (PRED) and dexamethasone (DEX), are among the most effective oral treatments for inflammatory and autoimmune diseases, including rheumatoid arthritis (RA), Crohn's disease, colitis, and multiple sclerosis. They have expanded use in the treatment of asthma and atopic dermatitis as inhalants and topical ointments, respectively, and are also used for treating transplant rejection. Despite their extraordinary efficacy in treating inflammatory diseases, chronic administration of GR agonists results in debilitating side effects.² These include diabetes, bone loss, skin thinning, muscle atrophy, and cataracts. Moreover, GR agonists can elicit off-target pharmacology depending upon cross-reactivity with other nuclear hormone receptors, particularly the mineralocorticoid (MR), progesterone (PR), androgen (AR), and estrogen (ER α , $ER\beta$) receptors.

During the past decade new structural classes of GR agonists have emerged with the potential to have improved side effect profiles, as compared with classical steroid agonists.^{3a-d,4a,b} The underlying hypothesis for designing these compounds, which we have called "dissociated agonists of the GR" (DAGRs),⁵ is evidence that the anti-inflammatory and side-effect activities of ligand-activated GRs occur via dissociated mechanisms transrepression (TR) and transactivation (TA).^{6a,b} According to this hypothesis, the TR pathway is associated with the antiinflammatory activity of GR agonists and leads to inhibition of transcription factors for genes, including NF κ B and AP1, that drive the expression of genes that encode pro-inflammatory proteins, such as IL-1, IL-8, TNF α , and MMP13. On the other hand, the TA pathway activates genes involved in metabolism and endocrine function. In this pathway, the dimeric form of the ligand-activated GR functions as an endogenous transcription factor by binding to specific DNA sequences in the promoter region, called glucocorticoid response elements (GREs). Examples of GRE-driven target genes that may contribute to adverse metabolic and bone effects include tyrosine amino transferase (TAT) and DKK1.^{7a,b}

The TR/TA hypothesis, although providing a rationale for discovering safer glucocorticoids, has been criticized as an oversimplification of the complex and unraveling process of gene expression regulation by GR.8 This is based on recent evidence that is not consistent and even refutes the hypothesis, and that the reporter genes used to measure dissociation are too simplified and unable to effectively target therapeutically relevant genes. Furthermore, targeting functional antagonism rather than TA has been described as an alternative approach of predicting the side effects of GR agonists.⁹ Despite this controversy, the screening strategy used herein relies on a measure of intrinsic dissociation, TR/TA in the same SW1353 chondrosarcoma cell background, and having only native GR and associated transcription machinery. This has led to the discovery of a novel pharmacophore that demonstrated a spectrum of TR/TA activities that differed from those of classical GR agonists.⁵ Arguably, compounds displaying such dissociated activities represent potential candidates to test for improved safety/efficacy profiles in human. In fact, a recent phase 2 clinical study with DAGR PF-04171327 has shown

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Table 1. Representative Octahydrophenanthrene-2,7-diol Analogues⁵



						Transrepression ^{a,b}	Transactivation
	R ₁	R ₂	R ₃	R ₄	GR Binding	MMP-13	MMTV ^a
					$IC_{50} (nM)^a$	IC ₅₀ (nM)	% max dex
						(% max dex)	
1	Н	1-propynyl	Н	Allyl	6.2	14 (79)	9
2	Н	1-propynyl	Н	Et	5.6	14 (86) ^c	6
3	Н	1-propynyl	ОН	Et	35	63 (79)	13
4	Н	Ph	ОН	Et	2.5	140 (67) ^{c,d}	6
5	N	Ph	ОН	Et	1.5	38 (78) ^{c,e}	6
6	Н	Ph	Н	Et	21	>5000	1

 ${}^{a}n = 1$ (dose response curves run in triplicate), except where otherwise noted. ${}^{b}IC_{50}$ concentration of test compound that inhibits IL-1 induced MMP-13 production by 50%. ^cInhibition reversed by co-incubation with RU-486. ${}^{d}n = 7$. ${}^{e}n = 2$ (IC₅₀ values = 25 and 50 nM).

similar efficacy to 10 mg of PRED but with effects on bone formation and glucose markers at 5 mg of PRED.¹⁰

In our first paper, we described the discovery of a novel series of DAGRs containing the octahydrophenanthrene-2,7-diol pharmacophore (1-5, Table 1).⁵ These analogues functioned as agonists in in vitro assays of TR (inhibition of IL-1 induced IL-8 and MMP-13 expression) but possessed reduced capacity for TA in an assay measuring activation of a mouse mammary tumor virus (MMTV) reporter construct, as compared to DEX. In addition, we outlined our early optimization efforts on this series focusing on replacing the allyl (R₄) and particularly the propynyl (R₂) substituents with those more likely to be chemically stable and less likely to produce toxic metabolites. The culmination of this effort was the identification of compounds 4 and 5, in which the C4a allyl and C2 1-propynyl groups of 1 were replaced with ethyl and phenyl, respectively.

In the discovery of phenyl as a replacement for propynyl at the C2 α position, a key structural modification was the introduction of the C3 α hydroxyl group. This change was beneficial in three ways: the lipophilicity (clogP) of the octahydrophenanthrene template was reduced; receptor selectivity was increased by reducing binding to ER; and, most importantly, agonist activity of C2 α phenyl analogues, which was otherwise lacking, (e.g., **6**), was restored. We speculated that the ability of the C3 α hydroxyl to impart agonist activity to analogues with a C2 α phenyl is due to a neighboring steric interaction, which stabilizes a particular range of rotamers of the C2 α phenyl that is favorable for TR. This conjecture was based, in part, on the fact that introduction of a C3 α hydroxyl in the C2 α 1-propynyl series (i.e., **3** vs **2**) did not improve agonist potency. In this case, no steric effect is possible because the propynyl is cylindrically symmetric, and hence all rotation is degenerate.

On the basis of our working hypothesis regarding the role of the C3 α hydroxyl group in determining the functional agonist potency of C2 α phenyl analogues, we sought to explore further structural modifications at C3 α with the immediate goal of increasing potency against TR, while also minimizing GR signaling via TA. To accomplish this, we first needed to develop a more efficient and versatile synthesis to allow wider variation of substitutions at C3.¹¹ We now report a new and efficient synthesis of C2 α phenyl analogues containing *alkyl* substitution at C3 α , leading to new DAGRs with significantly improved functional potency in both cell-free and whole blood assays, which approaches the level of DEX. We also provide further support for our hypothesis regarding the relationship between

Scheme 1^a



^a(a) PhCHO, NaOEt, EtOH (81%); (b) PhMgBr, CeCl₃, THF, −40 °C (96%); (c) p-NO₂C₆H₄COCl, 1 N NaOH, 0 °C (100%); (d) (i) O₃, CH₂Cl₂, MeOH, −78 °C, (ii) DMS, −78 °C to rt, (iii) NaOH, THF, 0 °C (62%, **11**; 11%, **12**); (e) NaBH₄, EtOH, THF (95%); (f) MeLi·LiI, THF, −78 °C (72%, **1**; 12%, **14**); (g) EtMgBr or vinylLi, THF, −78 °C (19%, **15**; 18%, **16**; 49%, **17**).

the torsion angle of the C2 α phenyl and agonist potency, based on X-ray crystallography and force field calculations.

RESULTS AND DISCUSSION

Chemistry. Our revised synthetic strategy was based on converting the previously described ketone 7⁵ to keto alcohol **11**, a key penultimate intermediate from which new C3 α substituents would be introduced directly by organometallic addition to the ketone (Scheme 1). Vital to this approach was the placement of a group at C3 of 7 that would preserve stereoelectronically favored, axial addition of the C2 α phenyl to C2 and also serve as a synthon for the C3 ketone of **11**. A planar benzylidine group (i.e., **8**) seemed well-suited for this dual purpose, because it does not contain an α -substituent to interfere with axial attack at C2 and can be easily cleaved by ozonolysis to produce a ketone.

Ketone 7 was condensed with benzaldehyde under thermodynamic conditions to give the desired benzylidine derivative 8. Addition of excess phenylmagnesium bromide to 8 under Luche conditions occurred stereospecifically from the axial direction to afford the allylic alcohol 9. The corresponding C2 epimer, the product of equatorial attack, was not detected. Initial attempts to obtain 11 directly from 9 by ozonolysis were accompanied by substantial oxidative degradation of the electron rich phenol (A ring). However, performing the ozonolysis on the p-nitrobenzoyl (PNB) derivative 10, followed by in situ saponification of the PNB group, produced the key intermediate 11 in 62% overall yield from 9. Diketone 12, the product of benzylic oxidation, was also isolated in 11% yield. This material proved valuable for the synthesis of B ring analogues, as will be described. The structure of 11 was confirmed by conversion to known analogue 4 by a highly stereoselective directed reduction¹² of the ketone with NaBH₄.

A small amount (5%) of the corresponding C3 epimer (16) was detected by NMR. The structure of 4 was confirmed by X-ray crystallography.

The first analogues were prepared by reacting 11 with MeLi to afford a mixture of C-ring diols 13 and 14 in a ratio of 6:1. These compounds were easily separated by flash chromatography, and their structures were confirmed by X-ray crystallography. In contrast to the NaBH₄ reduction of 11, nucleophilic addition of MeLi occurred predominantly in equatorial fashion, anti to the $\mathrm{C}2\beta$ hydroxyl and syn to the bulkier C2 α phenyl. This result is consistent with the model of Rosenberg et al.,¹³ in which nucleophilic addition to a cyclohexanone containing an adjacent equatorial dipole group is favored by a trajectory opposite to the negative end of the dipole. In the case of **11**, if deprotonation of the C2 β -hydroxyl had occurred prior to the addition of MeLi, this effect would be amplified because the negative end of the resulting $C-O^{-}$ dipole would be fully charged. Another factor favoring equatorial addition of MeLi is the avoidance of the 1,3-diaxial interaction with the angular ethyl.

The encouraging activity of analogue 13 led us to increase the size of the C3 α substituent by reacting 11 with ethylmagnesium bromide and vinyllithium, respectively, to afford analogues 15 and 17. In these cases, none of the trans diols were detected, presumably due to the enhanced 1,3-diaxial interaction between the larger incoming nucleophile and the angular ethyl group at C4a. In the reaction with ethylmagnesium bromide, the reduced secondary alcohol 16 (corresponding to the C3 epimer of 4) was also isolated as the product of β -hydride elimination of the Grignard reagent.^{14a,b} The stereospecificity of the reaction is rationalized by a cyclic transition state involving equatorial addition of hydride, to minimize the 1,3-diaxial interaction with the angular ethyl (Figure 1).



Figure 1. Formation of 16 from 11 by reaction with EtMgBr.

Replacement of the C2 α phenyl of analogue 13 by 2-pyridyl was accomplished analogously, although it was first necessary to protect the phenol as the *tert*-butyldimethylsilyl (TBDMS) ether derivative 18 (Scheme 2). Addition of 2-pyridyllithium, generated by transmetalation of 2-bromopyridine, to 18 afforded carbinol 19, which was converted to PNB ester 20. Ozonolysis followed by in situ deprotection gave keto alcohol 21, and subsequent addition of MeLi furnished the desired 2-pyridyl analogue 22. The corresponding 3- and 4-pyridyl analogues 23 and 24, respectively, were prepared analogously.

To more efficiently vary the C2 α phenyl of analogue 13, our basic route was modified by reversing the order of installation of the C2 α and C3 α substituents, respectively, to obtain 29 as an alternative key penultimate intermediate (Scheme 3). Thus, enone 8 was doubly protected by a two-step process involving ketalization to give 25 and subsequent formation of the PNB ester 26. Ozonolysis of 26 followed by in situ saponification of the PNB group gave the monoprotected α -diketone 27. The desired penultimate intermediate 29 was then obtained by addition of methyllithium to give carbinol 28, followed by acidic hydrolysis of the ketal. The stereospecificity of the methyllithium addition from the α -face was explained by the avoidance of the 1,3-diaxial interaction with the angular ethyl.

Reaction of 29 with eight organometallic reagents produced analogues 30-37, respectively. Although the reactions were not

Scheme 2^{a}

generally stereospecific, in most cases the desired 2α -alkyl epimer could be separated by flash chromatography. When separation was not possible, the analogues were tested as mixtures with the ratios determined by NMR. The unexpected erosion in axial selectivity to give mixtures of 2α and 2β alkyl products (vis-à-vis 8) was explained by initial complexation of the organometallic reagent with the axial 3β hydroxyl leading to equatorial attack. This route provided the theoretically interesting 2α -propynyl analogue 36, which otherwise would be difficult to obtain by the initial route due to potential oxidation of the triple bond in the ozonolysis step (Scheme 1). Finally, the corresponding 2-thiazolyl analogue 39 was prepared by protection of 29 as the TBDMS protected phenol derivative 38, followed by addition of 2-lithiothiazole.

Two analogues of 13 containing modifications in the B-ring were initially obtained by treating diketone 12 with excess methyllithium (Scheme 4). Addition occurred completely at the C3 ketone and partially at the C9 ketone (with concomitant dehydration) to provide two pairs of diastereomers epimeric at C3: C9 ketones 40 and 41 and 9-methyl- Δ^9 olefins 42 and 43, respectively. The four products were easily separable by reverse phase HPLC, and we were able to obtain pure samples of the desired 9-keto and 9-methyl- Δ^9 analogues, 40 and 42, respectively, corresponding to the C3 stereochemistry of 13.

The corresponding 9β -hydroxy and Δ^9 analogues, 47 and 51, respectively, were synthesized from 40 (Scheme 5). First, a more efficient method was developed to prepare 40, which involved direct benzylic oxidation of 44, the PNB derivative of 13, by ozonolysis. Formation of the benzoate derivative 45 and subsequent reduction with NaBH₄ gave the 9β -hydroxy derivative 46,¹⁵ which was saponified to afford analogue 47. For the synthesis of 51, the diol of 45 was first protected as the *p*-methoxyphenyl (PMP) acetal to give 48. The trans relationship between the PMP group in the acetal ring and the angular Me and Ph groups was tentatively assigned by assuming thermodynamic control. Reduction of 48 with NaBH₄ furnished alcohol 49,¹⁵ which underwent dehydration with



^{*a*}(a) TBDMSCl, imidazole, CH₂Cl₂; (b) (i) bromopyridine, *n*-BuLi, THF, -78 °C, (ii) enone **18** addition, warm to 0 °C, (iii) TBAF; (c) p-NO₂C₆H₄COCl, 1 N NaOH, 0 °C; (d) (i) O₃, CH₂Cl₂, MeOH, -78 °C, (ii) DMS, -78 °C to rt, (iii) NaOH, THF, 0 °C; (e) MeLi, dimethoxyethane, THF, -78 °C.

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Scheme 3^d



^{*a*}Contains 25% of β -isomer; ^{*b*}Contains 50% of β -isomer; ^{*c*}Contains 20% of β -isomer. ^{*d*}(a) HOCH₂CH₂OH, TsOH, toluene, Δ (74%); (b) PNBCl, 1 N NaOH, acetone, 0 °C (74%); (c) (i) O₃, MeOH, CH₂Cl₂, -78 °C, (ii) DMS, (iii) NaOH, THF (98%); (d) MeLi-LiBr, THF, -78 °C (97%); (e) 6 N HCl (87%); (f) RLi or RMgBr, -78 to -30 °C (43%, **30**; 18%, **31**; 6%, **32**; 44%, **33**; 24%, **34**; 28%, **35**; 13%, **36**; 15%, **37**); (g) TBDMSCl, imidazole, CH₂Cl₂ (68%); (h) (i) 2-bromothiazole, *n*-BuLi, THF, -78 °C, (ii) TBAF, THF (43%).

Scheme 4^{*a*}



^a(a) MeLi·LiBr, THF, toluene, 0 °C (28%, 40; 3%, 41; 25%, 42; 5%, 43).

Burgess reagent to give **50**. Oxidative cleavage of the PMP acetal and subsequent saponification of the benzoate then afforded the desired Δ^9 analogue **51**.

Discussion. Our first objective was to improve the agonist, or TR, activity of analogue 4, as measured by repression of IL-1 induced MMP-13 expression while maintaining good dissociation with respect to TA in the MMTV assay. According to our hypothesis, this was to be accomplished by replacing the C3 α hydroxyl of 4 with bulkier groups capable of enhancing the steric interaction with the adjacent C2 α phenyl, thus stabilizing a new set of C2 α phenyl rotamers that are more beneficial toward TR. Our new synthesis readily provided analogues with alkyl substitution at C3 α and a hydroxyl at C3 β .

The first analogue containing a methyl at C3 α (13) showed vastly improved agonist activity over the earlier lead 4 (Table 2). The IC₅₀ of 13 in the MMP-13 assay improved from 140

nM for 4 to 3.5 nM, exhibiting 87% of the maximal effect of DEX. The corresponding analogue 16 lacking a substituent at C3 α was inactive as an agonist, despite excellent GR binding (IC₅₀ = 3.8 nM), lending support to our hypothesis regarding the steric effect of the C3 α substituent on attenuating agonist potency. Analogue 13 retained the dissociative profile of the octahydrophenanthrene-2,7-diol series, exhibiting half the maximal effect of DEX (42%) in the MMTV assay as compared to the MMP-13 assay. This level of dissociation, however, was not as great as that of 4.

Replacement of the C3 α methyl group of 13 with ethyl (i.e., 15) resulted in ~2-fold weaker activity in the binding and MMP-13 assays (IC₅₀ = 6 nM). No other alkyl substituents at the C3 α position were explored, as it was assumed that methyl was optimal and that further increases in size would not be productive. The vinyl analogue 17 was devoid of MMP-13

Scheme 5^{*a*}



^{*a*}(a) PNBCl, acetone, NaOH (aq); (b) (i) O₃, CH₂Cl₂, MeOH, -30 °C, (ii) DMS, rt; (iii) NaOH; (c) benzoyl chloride, acetone, NaOH; (d) NaBH₄, MeOH, THF; (e) NaOH, H₂O, THF; (f) 4-(MeO)C₆H₄CHO, TsOH, benzene, Δ ; (g) NaBH₄, MeOH, -78 to 0 °C; (h) Burgess salt, benzene, Δ ; (i) (i) DDQ, CH₂Cl₂, H₂O, (ii) KOH, EtOH, H₂O.

Table 2. Analogues with variation at C	Table	. Ana	logues	with	V	ariation	at	C
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	R_1	R_2	GR binding IC_{50} (nM) ^a	transrepression ^{<i>a</i>} MMP-13 IC ₅₀ (nM) (% max DEX)	transactivation ^a MMTV (% max DEX)					
13	OH	Me	0.6	$3.5 (87)^b$	42					
16	OH	Н	3.8	>5000 ^b						
15	OH	Et	1.2	6 (78)	60					
17	OH	vinyl	5.2	>5000						
14	Me	OH	38	>5000						
$a^{n}n = 1$ (dose response curves run in triplicate), except where otherwise noted. $b^{n}n = 2$.										

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activity despite good binding (IC₅₀ = 5.2 nM). Although similar in length to ethyl, the planar vinyl group is effectively smaller because the α -carbon has one fewer hydrogen to hinder the rotation of the C2 α phenyl (-CH=CH₂ vs -CH₂CH₃). We presume this induces a different rotamer population that is less beneficial toward agonist activity, as compared to 13. Analogue 14 (C3 epimer of 13) containing a C3 α hydroxyl was also inactive as an agonist. Comparison of the activity profile of 14 versus 4, also containing a C3 α hydroxyl, suggests that decreased binding resulting from the introduction of the methyl at the C3 β -position (IC₅₀ = 38 vs 2.5 nM) was a major factor in the abolishment of MMP-13 activity. With the discovery of the potent analogue 13, we next explored varying the C2 α phenyl to see if further potency increases could be achieved (Table 3). A secondary objective was to lower lipophilicity. Preserving the optimal C2 substitution present in 13, several analogues containing *p*-tolyl, heteroaryl, alkyl, vinyl, and alkynyl substituents at C2 α , respectively, were examined. The *p*-tolyl analogue 37 was 2–3-fold weaker in the binding and MMP-13 assays than 13, demonstrating that extending the length of the C2 α substituent beyond phenyl is not beneficial. In the pyridyl series (22–24), only the 2-pyridyl analogue 22 showed high potency in the MMP-13 assay (IC₅₀ = 1.5 nM), which was 2-fold more potent

Table 3. Analogues with Variation at $C2\alpha$



	R	GR binding IC_{50} (nM)	transrepression ^{<i>a</i>} MMP-13 IC ₅₀ (nM) (% max DEX)	transactivation ^a MMTV % max DEX	clogP
37	<i>p</i> -tolyl	1.3	12 (74)	13	
22	2-Pyr	1.5	1.5 (95)	49	3.0
23	3-Pyr	22	80 (92)	31	
24	4-Pyr	9.1	100 (69)	11	
39	2-Thiaz	0.79	1.5 (93)	53	2.9
30	Me	211	NT		
31	Et^{b}	47	>5000		
32	Pr	20.1	>5000		
33	allyl	35.4	>5000		
34	vinyl ^c	12.1	650 (60)	7	
35	(E)-1-propenyl ^d	2.1	6 (95)	37	
36	propynyl	1.5	1.9 (89)	43	
$a_{n} = 1$	(dose response cu	rves run in triplicate). ^b 3	:1 mixture of diastereomers. ^c 1:1 mixture of diaste	reomers. ^d 4:1 mixture of diastereom	ers.

Table 4. Analogues with B-Ring Variation



	A–B	R	GR binding IC_{50}^{a} (nM)	transrepression ^{<i>a</i>} MMP-13 IC ₅₀ (nM) (% max DEX)	transactivation a MMTV (% max DEX)	clogP
40	COCH ₂	Н	2.5	$5 (87)^b$	41	2.1
47	$CH(OH)CH_2$	Н	79	>5000		
51	СН=СН	Н	0.3	4.0 (89)	32	4.2
42	C(Me) = CH	Н	0.67	$0.6 (94)^b$	52	1.8
45	COCH ₂	Bz	1.7	28 (91)	39	
a. 1	(1			b_{1}		

 ${}^{a}n = 1$ (dose response curves run in triplicate), except where otherwise noted. ${}^{b}n = 2$.

than 13. It was also less lipophilic by 1.5 clogP units. The 2thiazolyl analogue 39 displayed a similar profile with an excellent combination of agonist potency and relatively low clogP. Both analogues were dissociated, exhibiting about half the maximal effect of DEX in the MMTV assay, as compared to the MMP-13 assay.

None of the analogues with alkyl replacements for the C2 α phenyl, **30–33**, were active in the MMP-13 assay. The vinyl analogue **34** was weakly active; however, adding a terminal carbon to give the (*E*)-propenyl analogue **35** restored MMP-13 activity to the single-digit nanomolar level (IC₅₀ = 6 nM). These results demonstrate that, for optimal activity, the C2 α substituent must be attached to C2 by an sp² (or sp) carbon and have the length of phenyl (or propynyl).

The C2 α propynyl analogue **36** was synthesized to confirm our hypothesis that the main effect of the C2 α substituent on agonist potency is to induce a favorable conformation of the adjacent C2 α substituent. As predicted, no significant improvement in agonist activity was observed between **36** and the corresponding propynyl analogue **2** lacking substitution at C2, because rotation about the bond connecting C2 and the propynyl substituent is degenerate. The minor 2-fold improvement in MMP-13 activity (IC₅₀ = 6 vs 14 nM) was attributed to increased GR binding due to the C3 β hydroxyl (IC₅₀ = 1.5 vs 5.6 nM). A similar effect was observed between **6** (IC₅₀ = 21 nM) and the corresponding C3 β hydroxyl analogue **16** (IC₅₀ = 3.8 nM).

Modification of the B-ring was explored with the similar goals of improving agonist potency and lowering clogP (Table 4). Compared to 13, the 9-keto analogue 40 was only slightly weaker in the MMP-13 assay (IC₅₀ = 5 nM) but had a considerably lower clogP of 2.1. The corresponding 9-hydroxy analogue 47 was inactive. Placement of an olefinic linkage between C9 and C10 was well tolerated. The corresponding Δ^9 analogue 51 had activity similar to that of 13, and the change was associated with a modest improvement in dissociation in the MMTV assay (32% max DEX). Moreover, the 9-methyl- Δ^9 analogue 42 was the most potent analogue in the octahydrophenanthrene-2,7-diol series yet discovered with an IC₅₀ of 0.6 nM in the MMP-13 assay. However, this compound was slightly less dissociated than 13 (52% max DEX). Finally, the benzoate derivative 45, tested as an initial probe of the SAR of the C7 phenolic position, was less active in the MMP-13 assay than the corresponding free phenol 40. Nevertheless, the IC₅₀ of 28 nM was respectable and dissociation was retained. This result, together with that of the alkylated phenol derivative 5 from the previous series (Table 1), indicates that further attenuation of activity is possible by modification of the phenol.

An additional in vitro assay was established to measure the efficacy of key analogues in human whole blood (HWB), a more clinically relevant milieu. In this assay, the end point was

the inhibition of TNF α release induced by lipopolysaccharide (LPS) with DEX used as the positive control. We also tested DAGR analogue **52** (Figure 2) from the trifluoromethylcarbinol series discovered at Schering.^{16a,b}



Figure 2. Structure of DAGR 52 from the trifluoromethylcarbinol series. 16a,b

Among the octahydrophenanthrene-2,7-diols, there was a general decline in TR activity in going from the cell-free to the HWB assay, presumably due to protein binding (Table 5). This varied from 14-fold in the case of **40** to 110-fold in the case of **22**. The decline, also seen for DEX (12-fold), did not generally correlate with clogP, although the difference in clogP values was at most not more than 1.8 log units. The most potent analogue was **42** with an IC₅₀ of 43 nM, which was only 2-fold weaker than the "gold standard" DEX. The trifluoromethylcarbinol DAGR **52** was weaker than any of the octahydrophenanthrene-2,7-diols in the HWB assay with an IC₅₀ of 1000 nM. The large 130-fold decline in activity compared to the cell-free assay was perhaps due to high protein binding, as reflected by the high clogP of 6.7. This compound was also less dissociated than the octahydrophenanthrene-2,7-diols.

Binding assays were performed to determine the affinities of analogue 13, the prototype of the octahydrophenanthrene-2,7diol series, and the trifluoromethylcarbinol DAGR 52 for the AR, PR, ER α , and ER β receptors (Table 6). Analogue 13 was very specific for GR, exhibiting >500-fold selectivity versus AR and PR and even greater selectivity versus ER α and ER β . On the other hand, 52 was nonselective versus PR but was 100-fold selective versus AR and >10⁵-selective versus ER α and ER β .

Finally, we obtained structural information to support our hypothesis regarding the conformation of the C2 α phenyl and its effect on agonist efficacy. First, on the basis of the corresponding X-ray structures, the solid state torsion angles of the C2 α phenyl (or 2-pyridyl)¹⁷ were determined for key analogues (Table 7). Among those with the same B ring (13, 22, 4, 14, and 16), high agonist potency was associated with narrow torsion angles, -2° and 8° , in which the C2 α phenyl and C2 α hydroxyl groups were nearly eclipsed (Figure 3). Potency declined as the torsion widened to 44° and was

Table 6. Affinity of 13 and 52 for Five Steroid Receptors

	binding IC_{50}^{a} (nM)							
	GR	AR	PR	ERα	ERβ			
13	0.6	300	321	>3200	>3200			
52^b	0.4	40	0.2	>10 ⁵	>10 ⁵			
'n = 1 acemate	(dose respo	onse curves	run in t	triplicate). ^b Te	sted as the			

abolished beyond 60°. In the case of the B-ring analogues 42 and 40, high potency was associated with somewhat larger torsion angles, 13° and 37° , than that of 13.

Second, Merck Molecular Force Field 94 (MMFF94)¹⁸ calculations were performed on two analogues, 4 and 13, to obtain a dynamic perspective of the relationship between the torsion angle and agonist activity (Figure 4). These analogues differed only in substitution at C3. The energy profile of analogue 4 containing a C3 α hydroxyl showed a steep energy well corresponding to a population of preferred (lowest energy) torsion angles ranging from 40° to 60°. This was consistent with the 44° torsion angle observed in the X-ray structure of 4. On the other hand, the range of preferred torsion angles for 13 containing a C3 α methyl (and C3 β hydroxyl), ±5–50°, was broader and shifted toward the eclipsed conformation of 0°. This range of preferred torsion angles was consistent with that observed in the X-ray structures of the more potent analogues (13, 22, 42, and 40) containing a C3 α methyl.

In summary, both the X-ray data and force field calculations showed a correlation between low torsion angles and high agonist potency, consistent with our original hypothesis. On the basis of the X-ray structures, the optimal torsion angle was between -2° and 37° , although a larger maximum angle of 50° was allowed on the basis of the force field calculations. When the solid state torsion angle was 44° , a transition toward weaker activity occurred, and activity was abolished when the angle reached 60° . These results will be useful for designing more structurally novel analogues.

CONCLUSION

We have shown that simple inversion of the C3 α hydroxyl of DAGR 4 and placement of a C3 α methyl group result in a significant increase in TR efficacy in vitro. The prototypical analogue 13, as well as those containing heterocyclic replacements for the C2 α phenyl (22, 39) and modifications in the B ring (40, 42, 51), had low single-digit nanomolar IC₅₀ values in the cell-free MMP-13 inhibition efficacy assay. Efficacy in HWB, as measured by inhibition TNF α release, was high with three analogues displaying IC₅₀ values <100 nM, or within 2–4-

Table 5. Cell-free and Human Whole Blood TR Activity of Key Analogues Compared to Dexamethasone

	GR binding IC ₅₀ (nM)	transrepression MMP-13 IC ₅₀ (nM) (% max DEX)	transactivation MMTV (% max DEX)	transrepression HWB-TNF α IC ₅₀ ^{<i>a</i>} (nM) (% max DEX)	clogP
13	0.6	3.5 (87)	42	109 (85)	4.5
22	1.5	1.5 (95)	49	167 (79)	3.0
39	0.79	1.5 (93)	53	83 (88)	2.9
40	2.5	5 (87)	41	72.5 (74)	3.5
42	0.67	0.6 (94)	52	43 (82)	4.7
51	0.3	4.0 (89)	32	400 (60)	4.2
52	3.2	7.5 (98)	66	1000 (52)	6.7
DEX	3.5	1.8	100	21 (100)	

 ${}^{a}n = 1$ (dose response curves run in triplicate).

Table	7.	Correlation	between	the S	olid	State	Torsion	Angle	of	the	$C2\alpha$	Phenyl	l and	TR	Efficac	ş
																,



				но	A ^B	
	\mathbb{R}^1	R ²	R ³	A–B	torsion angle (deg)	transrepression MMP-13 IC ₅₀ (nM)
13	Ph	Me	OH	CH ₂ CH ₂	-2	3.5
22	2-Pyr	Me	OH	CH ₂ CH ₂	8	1.5
4	Ph	OH	Н	CH ₂ CH ₂	44	140
14	Ph	OH	Me	CH ₂ CH ₂	60	>5000
16	Ph	Н	OH	CH ₂ CH ₂	77	>5000
42	Ph	Me	OH	$C(Me) = CH_2$	13	0.6
40	Ph	Me	ОН	COCH ₂	37	5



Figure 3. Overlay of the X-ray structures of **13**, **22**, **4**, **14**, and **16**, in counterclockwise order of the orientation of the C2 α phenyl, respectively. The color code represents analogues with high (green), moderate (yellow), and minimal (red) agonist potency.

fold of the level of DEX ($IC_{50} = 21$ nM). These compounds maintained the dissociated profile of a DAGR, exhibiting 1.8– 2.8-fold less activity in the MMTV assay relative to DEX (max). Analogue 13 showed minimal affinity for the AR, PR, ER α , and ER β receptors. Compared to a potent DAGR analogue 52 from the structurally distinct trifluoromethylcarbinol series, the series based on 2-phenyl-3-methyloctahydrophenanthrene-2,3,7-triol 13 has clear advantages with respect to in vitro dissociation, activity in HWB, and affinity for the AR and PR receptors. On the basis of the X-ray structures of key analogues and MMFF94 force field calculations, high agonist potency was correlated with a low torsion angle of the C2 α phenyl. Preliminary SAR indicated that potency and physical–chemical parameters can be further attenuated by modification of the C3 phenol, and this has guided subsequent efforts.

Due to its high agonist potency and spectrum of activity, this series is a promising starting point for the identification of



Figure 4. Energy profiles (MMFF94) of **4** and **13** with respect to the torsion angle of the C2 α phenyl, defined as 0° when the phenyl is eclipsed to the C2 α hydroxyl.

developmental candidates through optimization of side-effect dissociation. In fact, potent analogues containing substitutions at the angular ethyl (C4b) and C7 phenolic positions with >99% antagonist activity have been reported and display dissociative activity of target genes in human cell lines.¹⁹

EXPERIMENTAL SECTION

Chemistry. Reagents were purchased commercially and used without further purification unless otherwise indicated. Reactions were performed under a nitrogen atmosphere, magnetically stirred, and run at room temperature except when otherwise indicated. Solutions of sodium bicarbonate, ammonium chloride, and sodium chloride (brine) were aqueous. Analytical thin-layer chromatography (TLC) was carried out using Merck silica gel 60 F₂₅₄ plates on which compounds were visualized using ultraviolet (UV) light and ceric ammonium molybdate stain. Flash chromatography under medium pressure was performed with Baker silica gel (40 μ m) packed manually in glass columns or using a Biotage Flash 40 with 12M, 40S, or 40M columns. In addition, automated flash chromatography was performed using an Isco CombiFlash. Preparative reverse phase chromatography was performed using a Waters Symmetry C8 column

¹H NMR spectra were recorded in CDCl₃, CD₃OD, or DMSO- d_6 solutions on a Varian Unity 400 or 500 MHz spectrometer. Significant chemical shifts are reported in parts per million (δ) relative to residual CHCl₃ (7.24 ppm), DMSO (2.49 ppm), or CD₃OD (3.31 ppm) as

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internal reference, respectively. Coupling constants (J) are reported in hertz (Hz). The peak shapes are denoted as follows: s, singlet; d, doublet; t, triplet; m, multiplet; br, broad. Multiplets can consist of overlapping peaks. Low-resolution liquid chromatography mass spectra (LC-MS) were recorded by positive and negative electrospray ionization using a Waters/Micromass ESI/MS model ZMD/LCZ mass spectrometer equipped with a Gilson 215 liquid handling system and a Hewlett-Packard 1100 diode array detector. Elemental analyses were performed by Schwarzkopf Microanalytical Laboratory (Flushing, NY, USA) and Quantitative Technologies, Inc. (Whitehouse, NJ, USA) and are within $\pm 0.4\%$ of the calculated values unless stated otherwise. X-ray crystal structures were collected on a Bruker APEX or Siemens P4 diffractometer.

(3*E*,4a*R*,10a*R*)-3-Benzylidene-4a-ethyl-7-hydroxy-3,4,4a,9,10,10a-hexahydro-1*H*-phenanthren-2-one (8).



A dry 1 L round-bottom flask was charged with ethanol (500 mL) and cooled to 0 $^\circ\text{C}.$ Sodium chunks (3.30 g, 144 mmol) were added in portions, and the mixture was stirred for 1 h. Ketone 7 (14.1 g, 57.5 mmol) was added and the solution stirred for 1 h. A solution of benzaldehyde (6.41 g, 60.4 mmol) in ethanol (50 mL) was added, and the resulting dark red mixture was stirred for 5 h. Additional benzaldehyde (0.91 g, 8.6 mmol) was added, and stirring was continued overnight. The pH was adjusted to 1-2 by the addition of aqueous 1 M hydrochloric acid, and the mixture was diluted with water (1.25 L) and stirred for 1 h. The yellow precipitate was collected by filtration, and the solid (21 g) was stirred in diethyl ether (400 mL) as a suspension for 1 h. The solid was filtered to afford 15.4 g (81%) of title compound 8 as a yellow solid, mp 264-266 °C. An analytical sample was prepared in a previous run by trituration with methanol/ THF (1:1), mp 265–267 °C. ¹H NMR (CDCl₃): δ 0.53 (3H, t, J = 7.5), 1.42-1.55 (2H, m), 1.64-1.72 (1H, m), 1.78-1.90 (1H, m), 2.26-2.34 (1H, m), 2.43-2.51 (2H, m), 2.67 (1H, dd, J = 5.6 and 18.9), 2.82–2.96 (2H, m), 3.68 (1H, d, J = 15.8), 4.70 (1H, br s), 6.59 (1H, d, J = 2.5), 6.64 (1H, dd, J = 2.9 and 8.3), 7.11 (1H, d, J = 8.3), 7.33–7.39 (1H, m), 7.43–7.44 (4H, m), 7.67 (1H, d, J = 2.1). LC-MS: m/z 333.4 [M + H]⁺. Anal. Calcd for C₂₃H₂₄O₂: C, 83.10; H, 7.28. Found: C, 82.83; H, 7.52.

(25,3E,4aR,10aR)-3-Benzylidene-4a-ethyl-2-phenyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,7-diol (9).



A dry, three-neck, 2 L round-bottom flask was charged with CeCl₂ (31.5 g, 128 mmol) and THF (800 mL). The suspension was stirred for 45 min and then cooled to -78 °C. A solution of 1.0 M phenylmagnesium bromide in THF (128 mL, 128 mmol) was added, and the mixture was stirred for 1.5 h. A suspension of ketone 8 (8.50 g, 25.6 mmol) in THF (250 mL) was added via a wide-bore cannula (-70 °C to -55 °C) and then stirred at -40 °C for 2.5 h. A solution of 10% aqueous acetic acid (0.15 L) was added followed by water (400 mL), and the mixture was allowed to warm to room temperature. The aqueous layer was separated and extracted with ethyl acetate $(1 \times 500$ mL, 1×250 mL). The combined organic layers were washed with saturated sodium bicarbonate (250 mL) and brine (300 mL), dried (MgSO₄), and concentrated to give a greenish brown paste (12.8 g). This material was combined with 1.9 g (4.6 mmol) of crude material prepared similarly in a previous run, and the contents were purified by flash chromatography (400 g silica gel) using ether/hexanes (30, 50, 70%; 2 L each) as eluent to afford 11.95 g (96%, adjusted) of title compound 9 as a beige foam. ¹H NMR (CDCl₃): δ 0.11 (3H, t, J =

7.6), 1.13–1.26 (1H, m), 1.57–1.90 (6H, m), 2.05 (1H, t, J = 12.7), 2.57 (1H, d, J = 13.3), 2.74–2.82 (2H, m), 3.63 (1H, d, J = 13.7), 6.43–6.48 (2H, m), 6.75 (1H, d, J = 8.7), 7.21–7.45 (10 H, m), 7.58 (2H, J = 7.9). LC-MS: m/z 393.3 [M + H – H₂O]⁺, 409.4 [M – 1]⁻, 455.4 [M – 1 + H₂CO₂]⁻.

(4bR,75,8aR,E)-6-Benzylidene-4b-ethyl-7-hydroxy-7-phenyl-4b,5,6,7,8,8a,9,10-octahydrophenanthren-2-yl 4-Nitrobenzoate (10).



In a 1 L round-bottom flask, a stirred solution of phenol 9 (11.96 g, 29.1 3 mmol) in acetone (400 mL) was chilled to 0 °C and treated with 1 N aqueous sodium hydroxide (30.5 mL, 30.5 mmol). After 20 min, a solution of 4-nitrobenzoyl chloride (6.48 g, 34.9 mmol) in acetone (50 mL) was added, and the mixture was stirred for 1 h. Additional 1 N aqueous sodium hydroxide (18.3 mL, 18.3 mmol) and 4-nitrobenzoyl chloride (3.60 g, 19.4 mmol) were added in portions as the course of the reaction was monitored by pH (final pH 7-8). The resulting suspension was filtered, and the filtrate was concentrated under vacuum to a volume of approximately 100 mL. Saturated aqueous NaHCO₃ solution (250 mL) was added, and the organic phase was extracted with ethyl acetate $(1 \times 400 \text{ mL and } 2 \times 200 \text{ mL})$. The combined organic layers were washed with brine, dried $(MgSO_4)$, and concentrated to afford 16.5 g (theory = 16.3) of title compound 10 as a yellow foam. ¹H NMR (CDCl₃): δ 0.14 (3H, t, J = 7.6), 1.20– 1.32 (1H, m), 1.63–1.95 (6H, m), 2.03–2.13 (1H, m), 2.61 (1H, d, J = 12.9), 2.85–2.90 (2H, m), 3.68 (1H, d, J = 13.3), 6.81 (1H, dd, J = 2.3 and 8.5), 6.85 (1H, s), 6.96 (1H, d, J = 8.3), 7.22-7.48 (9H, m), 7.59 (2H, d, J = 7.9), 8.30 (4H, s). LC-MS: m/z 542.3 [M + H - H_2O^{+} .

(2*R*,4a*R*,10a*R*)-4a-Ethyl-2,7-dihydroxy-2-phenyl-1,2,4,4a,10,10a-hexahydrophenanthren-3(9*H*)-one (11) and (2*R*,4a*R*,10a*S*)-4a-Ethyl-2,7-dihydroxy-2-phenyl-1,2,4,4a,10,10a-hexahydrophenanthrene-3,9-dione (12).



A 1 L round-bottom flask was charged with olefin 10 (10.4 g, 18.6 mmol), methanol (300 mL), and dichloromethane (300 mL). After the mixture had cooled to -78 °C, ozone was bubbled into it until a dark blue color persisted. The flask was capped and the solution stirred for 3 h, occasionally purging with ozone to maintain a blue color. The mixture was purged with nitrogen for 5 min, and excess dimethyl sulfide (27.5 mL, 23.1 g, 372 mmol) was added. After warming to room temperature over the course of 1 h, the mixture was concentrated to give an oil (10.9 g), which was dissolved in methanol (500 mL) and cooled in an ice bath. A solution of 1 N aqueous sodium hydroxide (37.2 mL, 37.2 mmol) was added, and the mixture was stirred for 2 h at 0 °C. The mixture was poured onto saturated sodium bicarbonate solution (500 mL), and the organic layer was separated. The aqueous layer was extracted with ethyl acetate (1 \times 500 mL, 1 \times 300 mL). The combined organic layers were washed with saturated sodium bicarbonate solution (1 \times 100 mL), brine (1 \times 300 mL), dried $(MgSO_4)$, and concentrated to give an oil (6.7 g). This material was purified by flash chromatography (350 g silica gel) loading using a minimum amount of ethyl ether (25 mL) and eluting with ethyl ether/ hexanes (40, 50, 60, 100%) followed by 100% ethyl acetate. The less polar fractions were evaporated to give 3.9 g (62%) of title ketone 11 as a white solid, mp 97–98 °C. ¹H NMR (CDCl₃): δ 0.76 (3H, t, J =

7.5), 1.39–1.50 (2H, m), 1.58 (2H, br s), 1.82–1.92 (2H, m), 2.06 (1H, t, J = 13.9), 2.18–2.30 (1H, m), 2.35 (1H, d, J = 12.8), 2.84–2.92 (3H, m), 3.21 (1H, J = 12.8), 6.50–6.57 (2H, m), 6.81 (1H, d, J = 8.3), 7.24–7.36 (5H, m). LC-MS: m/z 319.4 [M + 1 – H₂O]⁺, 381.4 [M – 1 + H₂CO₂]⁻. Anal. Calcd for C₂₂H₂₄O₃: C, 78.54; H, 7.16. Found: C, 78.50; H 7.46. The more polar fractions were evaporated to give 0.7 g (11%) of title diketone **12** as a white solid. ¹H NMR (CDCl₃): δ 0.73 (3H, t, J = 7.3), 1.39–1.50 (1H, m), 1.46–1.82 (3H, overlapping m and br s), 2.17 (1H, t, J = 13.5), 2.46 (1H, d, J = 12.8), 2.69–2.83 (3H, m), 2.89 (1H, d, J = 13.3), 3.28 (1H, d, J = 12.9), 6.93–7.02 (2H, m), 7.21–7.40 (5H, m), 7.49 (1H, d, J = 2.5). LC-MS: m/z 349.3 [M – 1]⁻, 395.4 [M – H + H₂CO₂]⁻.

(2*R*,3*S*,4a*R*,10a*R*)-4a-Ethyl-2-phenyl-1,2,3,4,4a,9,10,10a-oc-tahydrophenanthrene-2,3,7-triol (4).



A 30 mL round-bottom flask was charged with ketone 11 (75 mg, 0.22 mmol), THF (5 mL), and ethanol (10 mL). Solid NaBH₄ (17 mg, 0.50 mmol) was added, and the mixture was stirred for 2 h. Additional NaBH₄ (15 mg, 0.40 mmol) was added, and stirring was continued for 2 h. A solution of 1 N hydrochloric acid (10 mL) and water (40 mL) were added, and the mixture was extracted with ethyl acetate (3×60) mL). The combined organic layers were washed with brine, dried $(MgSO_4)$, and concentrated to afford a white solid (78 mg, 100%). The ¹H NMR (CDCl₃) spectrum showed two isomers (95:5) with the major isomer corresponding to title compound 4. The minor isomer corresponded to compound 16 (vide infra). The structure of 4 was confirmed by single-crystal X-ray analysis of another sample which had been purified by chromatography and crystallized out of CDCl₃ as star needles, mp 191–192 °C. ¹H NMR (DMSO- d_6): δ 0.68 (3H, t, J = 7.3), 1.09-1.12 (1H, m), 1.25-1.49 (2H, m), 1.56-1.70 (1H, m), 1.72–1.80 (2H, m), 1.96 (1H, dd, J = 2.9 and 13.7), 2.39 (1H, dd, J = 3.7 and 13.3), 2.54-2.74 (2H, m), 3.83-3.89 (1H, m), 4.82 (1H, s), 4.86 (1H, d, J = 3.3), 4.91 (1H, s), 6.36 (1H, d, J = 2.5), 6.42 (1H, dd, J = 2.7 and 8.4), 6.84 (1H, d, J = 8.3), 7.08-7.12 (1H, m), 7.15-7.21 (2H, m), 7.70 (2H, d, J = 7.5), 8.95 (1H, s). LC-MS: *m*/*z* 321.3 [M + $H - H_2O^{+}$

(2*R*, 3*R*, 4a*R*, 10a*R*) - 4a - Ethyl - 3 - methyl - 2 - phenyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (13) and (2*R*, 3*S*, 4a*R*, 10a*R*) - 4a - Ethyl - 3 - methyl - 2 - phenyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (14).



A 1- L round-bottom flask was charged with ketone 11 (6.95 g, 20.7 mmol) and THF (700 mL). The solution was cooled to 0 °C, and a solution of 1 M methyllithium-lithium iodide complex in diethyl ether (100 mL, 100 mmol) was added dropwise. The mixture was allowed to warm to room temperature and then stirred overnight. To the resulting light green mixture was added a solution of 10% aqueous acetic acid (100 mL) followed by water (200 mL). The organic layer was separated and the aqueous layer extracted with ethyl acetate (2 \times 300 mL). The combined organic layers were washed with saturated sodium bicarbonate solution and brine, dried (MgSO₄), and concentrated. The oily residue (8.2 g) was purified by flash chromatography (dry loaded onto 400 g of silica gel) eluting with ethyl ether/hexanes (30-100% gradient in 10% increments). Evaporation of the less polar fractions afforded 5.2 g (72%) title compound 13 as a white solid. ¹H NMR (CD₃OD): δ 0.78 (3H, t, *J* = 7.1,7.5), 1.12 (3H, s), 1.15-1.31 (1H, m), 1.33-1.40 (1H, m), 1.42 (1H, d, J = 12.8), 1.75 - 1.92 (3H, m), 2.23 - 2.37 (2H, m), 2.68 - 2.80(3H, m), 6.50 (1H, s), 6.53 (1H, d, J = 8.7), 6.99 (1H, d, J = 8.3), 7.12-7.18 (1H, m), 7.19-7.24 (2H, m), 7.59 (2H, d, J = 7.9). LC-MS:

m/z 351.4 [M - H]⁻, 381.4 [M - OH + H₂CO₂]⁻. The analytical sample was prepared from a previous sample by crystallization from ethyl ether/hexanes, mp 197-199 °C. Anal. Calcd for C23H28O3: C, 78.38; H, 8.01. Found: C, 78.03; H, 7.91. The structure was confirmed by single-crystal X-ray analysis of a sample that had been crystallized first from chloroform and then recrystallized from ethyl ether/hexanes. Evaporation of the more polar fractions gave title compound 14 as a white solid, 0.90 g (12%). ¹H NMR (DMSO- d_6): δ 0.65 (3H, t, J =7.6), 1.10-1.22 (2H, m), 1.22 (3H, s), 1.28-1.35 (1H, m), 1.55-1.72 (3H, m), 1.78–1.91 (3H, m), 2.52–2.69 (2H, m), 4.12 (1H, s), 4.73 (1H, s), 6.31 (1H, d, J = 2.5), 6.42 (1H, dd, J = 2.5 and 8.7), 6.82 (1H, d, *J* = 8.3), 7.05 (1H, t, *J* = 7.3), 7.14 (2H, t, *J* = 7.4), 7.68 (2H, d, *J* = 7.5), 8.89 (1H, s). LC-MS: m/z 351.4 $[M - H]^{-}$, 397.4 [M - H + H_2CO_2 ⁻. The analytical sample was prepared from a previous sample by recrystallization from ethyl ether/hexanes, mp 228-229 °C. Anal. Calcd for C₂₃H₂₈O₃: C, 78.38; H, 8.01. Found: C, 78.17; H, 8.29. The structure was confirmed by single-crystal X-ray analysis.

(2R,3R,4aR,10aR)-3,4a-Diethyl-2-phenyl-1,2,3,4,4a,9,10,10aoctahydrophenanthrene-2,3,7-triol (15) and (2R,3R,4aR,10aR)-4a-Ethyl-2-phenyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (16).



A dry 20 mL round-bottom flask was charged with ketone 11 (100 mg, 0.30 mmol), lithium chloride (94 mg, 2.2 mmol), and toluene (5 mL). A solution of 1 M ethylmagnesium bromide in THF (2.2 mL, 2.2 mmol) was added, and the mixture was stirred overnight at room temperature. The suspension was treated with 10% aqueous acetic acid and then combined with additional material from a previous reaction run identically (0.45 mmol of 11). Water (30 mL) was added, and the mixture was extracted with ethyl acetate $(3 \times 30 \text{ mL})$. The combined organic layers were washed with saturated sodium bicarbonate and brine, dried (MgSO₄), and concentrated to give an oil (0.29 g). Purification was performed by flash chromatography (40 g silica gel) eluting with ethyl acetate/hexanes (10, 25, and 35%, 400 mL each). The less polar fractions were pooled and concentrated to afford 51 mg (19%) of title analogue 15 as a foam. ¹H NMR (DMSO- d_6): δ 0.67 (3H, t, J = 7.1), 0.79 (3H, t, J = 7.6), 1.09–1.36 (5H, m), 1.49 (1H, d, J = 15.0, 1.62–1.77 (2H, m), 2.10–2.23 (2H, m), 2.58–2.74 (3H, m), 3.67 (1H, s), 5.22 (1H, s), 6.36 (1H, d, J = 2.5), 6.45 (1H, dd, J = 2.5 and 8.3), 6.91 (1H, d, J = 8.7), 7.09 (1H, t, J = 7.3), 7.18 (2H, t, J = 7.5), 7.49 (2H, d, J = 7.5), 8.93 (1H, s). LC-MS: *m*/*z* 365.4 [M – H]⁻, 411.4 $[M - H + H_2CO_2]^-$. The more polar fractions were pooled and concentrated to afford 45 mg (18%) of title alcohol 16 as a foam, which was crystallized from ethyl ether/hexanes, mp 207-209 °C. ¹H NMR (CDCl₃ + drop of CD₃OD): δ 0.83 (3H, t, *J* = 7.3), 1.19–1.23 (3H, m), 1.22–1.30 (1H, m), 1.42 (1H, d, J = 13.5), 1.61–1.72 (2H, J)m), 1.88–2.00 (1H, m), 2.07 (1H, d, J = 13.5), 2.20–2.33 (2H, m), 2.75-2.88 (3H, m), 4.56 (1H, br s), 6.50-6.54 (2H, m), 6.94 (1H, d, J = 8.3), 7.25–7.35 (3H, m), 7.52 (2H, d, J = 7.3). LC-MS: *m*/*z* 321.4 $[M + H - H_2O]^+$, 383.4 $[M - 1 + H_2CO_2]^-$. The structure of 16 was confirmed by single-crystal X-ray analysis.

(2 R, 3 S, 4 a R, 1 0 a R) - 4 a - E t h y l - 2 - p h e n y l - 3 - v i n y l -1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (17).



To a stirred solution of 1 M vinyl bromide in THF (9 mL, 9 mmol) at -78 °C was added a solution of 1.7 M *tert*-butyllithium in hexane (10 mL, 17 mmol). After stirring for 1 h, the mixture was warmed to 0 °C, and a solution of ketone **11** (0.30 g, 0.89 mmol) in THF (10 mL) was added. The mixture was allowed to warm to room temperature and

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then stirred overnight. The pH was adjusted to 5 by the addition of 10% aqueous acetic acid, and water (30 mL) was added. The mixture was extracted with ethyl acetate (3 × 30 mL), and the combined organic layers were washed with brine, dried (Na₂SO₄), and concentrated. The oily residue (0.4 g) was purified by flash chromatography (40 g of silica gel) eluting with ethyl ether/hexanes (20–100%) to afford 160 mg (49%) of title compound 17 as an oil. ¹H NMR (CDCl₃): δ 0.76 (3H, t, *J* = 7.3), 1.15–1.27 (1H, m), 1.29–1.35 (1H, m), 1.48 (1H, dd, *J* = 1.6 and 13.0), 1.74–1.84 (2H, m), 1.97 (1H, d, *J* = 15.0), 2.22–2.32 (2H, m), 2.65–2.74 (3H, m), 3.73 (3H, br s), 5.02 (1H, dd, *J* = 1.3 and 10.6), 5.37 (1H, dd, *J* = 1.6 and 17.1), 5.94 (1H, dd, *J* = 10.9 and 17.1), 6.46 (1H, d, *J* = 2.6), 6.53 (1H, dd, *J* = 2.6 and 8.3), 6.97 (1H, d, *J* = 8.8), 7.12 (1H, t, *J* = 7.3), 7.17 (2H, t, *J* = 7.5), 7.51 (2H, d, *J* = 7.8). LC-MS: *m/z* 347.1 [M + H – H₂O]⁺, 363.1 [M – H]⁻ and 409.0 [M – H + H₂CO₂]⁻.

(4aR, 10aR, E)-3-Benzylidene-7-((*tert*-butyldimethylsilyl)oxy)-4a-ethyl-1,4,4a,9,10,10a-hexahydrophenanthren-2(3*H*)-one (18).



To a solution of enone 7 (10.0 g, 30.0 mmol) in dichloromethane (300 mL) was added imidazole (3.47 g, 51.0 mmol) and tertbutyldimethylsilyl chloride (7.69 g, 51.2 mmol). The mixture was stirred overnight at room temperature. An additional 0.33 equiv each of imidazole and tert-butyldimethylsilyl chloride was added, and stirring was continued for 8 h. A catalytic amount of 4dimethylaminopyridine was added, and the mixture was stirred overnight. Aqueous 0.5 N citric acid solution was added, and the organic layer was separated. The aqueous layer was extracted with dichloromethane, and the combined organic layers were washed with water and brine, dried (MgSO₄), and concentrated. The residue was triturated with ether to give 1.53 g of recovered 7. The mother liquor was concentrated, and the residue was purified by flash chromatography using an Isco CombiFlash eluting with ethyl acetate/hexanes (0-30%). The desired fractions were pooled and concentrated to give 6.59 g (49%) of title compound 18 as a waxy solid. ¹H NMR (CDCl₃) δ 0.18 (6H, s), 0.52 (3H, t, J = 7.5), 0.96 (9H, s), 1.43-1.51 (2H, m), 1.62-1.71 (1H, m), 1.78-1.91 (1H, m), 2.28-2.35 (1H, m), 2.43-2.51 (2H, m), 2.66 (1H, dd, J = 5.8 and 18.7), 2.79-2.96 (2H, m), 3.68 (1H, d, J = 15.8), 6.58 (1H, d, J = 2.5), 6.62 (1H, dd, J = 2.7 and 8.5), 7.06 (1H, d, J = 8.7), 7.35-7.42 (1H, m), 7.42-7.45 (4H, m), 7.65 (1H, d, J = 2.9). LC-MS: m/z 447.1 [M + H]⁺

(25,4aR,10aR,E)-3-Benzylidene-4a-ethyl-2-(pyridin-2-yl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,7-diol (19).



A solution of 2-bromopyridine (23.3 g, 148 mmol) in THF (500 mL) was chilled to -78 °C and treated dropwise with a solution of 2.5 M butyllithium (58.0 mL, 145 mmol) in hexanes. When the addition was complete, the mixture was stirred for an additional 30 min, and a solution of intermediate **18** (6.59 g, 14.8 mmol) in THF (150 mL) was added. Stirring at -78 °C was continued for 4 h, and the mixture was quenched by the addition of water (200 mL). The aqueous layer was extracted twice with ethyl acetate, and the combined extracts were washed with water and brine, dried (MgSO₄), and concentrated to give 13.3 g of a red oil. This material was dissolved in THF (230 mL), cooled to 0 °C, and treated with a solution of 1.0 M

tetrabutylammonium fluoride (23 mL, 23 mmol) in THF. After stirring for 4 h, the mixture was filtered through a plug of silica gel and concentrated. The residue was purified by flash chromatography using an Isco CombiFlash eluting with ethyl acetate/hexanes (30–65%) followed by an ethyl acetate flush. The desired fractions were pooled and concentrated to give 3.76 g (62%) of title compound **19** as a yellow solid. ¹H NMR (CD₃OD): δ 0.09 (3H, t, *J* = 7.5), 1.16–1.25 (1H, m), 1.56–1.81 (5H, m), 1.88 (1H, t, *J* = 12.5), 2.70–2.76 (2H, m), 2.87 (1H, d, *J* = 10.8), 3.68 (1H, d, *J* = 13.7), 6.36–6.40 (2H, m), 6.66 (1H, d, *J* = 8.3), 7.19–7.27 (3H, m), 7.32–7.42 (4H, m), 7.64 (1H, d, *J* = 7.9), 7.74–7.79 (1H, m), 8.51–8.52 (1H, m). LC-MS: *m*/*z* 412.3 [M + H]⁺.

(4b*R*,7*S*,8a*R*,*E*)-6-Benzylidene-4b-ethyl-7-hydroxy-7-(pyridin-2-yl)-4b,5,6,7,8,8a,9,10-octahydrophenanthren-2-yl 4-Ni-trobenzoate (20).



A suspension of intermediate **19** (2.27 g, 5.52 mmol) in acetone (60 mL) was cooled to 0 °C and treated with a solution of 1 M sodium hydroxide (5.50 mL, 5.50 mmol). After the solids dissolved, *p*-nitrobenzoyl chloride (1.19 g, 6.41 mmol) was added. The mixture was stirred for 2.5 h and then diluted with saturated sodium bicarbonate solution and extracted twice with EtOAc. The combined organic layers were washed with brine, dried (MgSO₄), and concentrated to give 2.83 g (92%) of title compound **20** as a white foam. ¹H NMR (CDCl₃): δ 0.24 (3H, t, *J* = 7.5), 1.27–1.33 (1H, m), 1.65 (1H, br s), 1.70–1.78 (3H, m), 2.83–2.91 (3H, m), 2.84 (1H, dd, *J* = 2.6 and 12.9), 2.85–2.91 (2H, m), 3.79 (1H, d, *J* = 13.7), 6.88 (1H, dd, *J* = 2.6 and 8.3), 6.91 (1H, d, *J* = 2.4), 7.04 (1H, d, *J* = 8.3), 7.22–7.45 (7H, m), 7.58 (1H, d, *J* = 7.9), 7.69–7.71 (1H, m), 8.34 (4H, s), 8.63 (1H, dd, *J* = 0.7 and 1.7). LC-MS: *m/z* 560.9 [M + H]⁺.

(2R,4aR,10aR)-4a-Ethyl-2,7-dihydroxy-2-(pyridin-2-yl)-1,4,4a,9,10,10a-hexahydrophenanthren-3(2H)-one (21).



Compound 20 (3.70 g, 6.60 mmol) was dissolved in a mixture of dichloromethane (200 mL) and methanol (100 mL), and the contents were chilled to -78 °C and treated with a solution of 6 N hydrochloric acid (1.25 mL, 7.5 mmol). Ozone was bubbled into the mixture until saturated, as indicated by the persistence of a blue color. Stirring was continued until TLC analysis indicated the consumption of starting material. The mixture was purged with oxygen, and then dimethyl sulfide (10 mL) was added. The mixture was allowed to warm to room temperature and concentrated. The solid residue was dissolved in THF (40 mL), and a solution of 1 N sodium hydroxide (20 mL, 20 mmol) was added. The mixture was stirred overnight, neutralized with 1 N hydrochloric acid solution, and extracted twice with ethyl acetate. The combined organic layers were washed with saturated sodium bicarbonate solution and brine, dried (MgSO₄), and concentrated to give 2.4 g of a white solid. This material was triturated with ether/ hexanes to give 1.38 g (62%) of title compound 21 as a white solid. Additional product (0.58 g, 26%) was obtained by chromatography of the concentrated mother liquor using an Isco CombiFlash eluting with ethyl acetate/hexanes (10–70%). ¹H NMR (CDCl₃): δ 0.76 (3H, t, J = 7.5), 1.40-1.50 (2H, m), 1.65 (1H, br s), 1.78-1.90 (2H, m), 2.08 (1H, t, J = 13.7), 2.38–2.51 (1H, m), 2.74 (1H, d, J = 12.8), 2.78–2.87 (3H, m), 3.32 (1H, d, J = 12.8), 4.65 (1H, br s), 6.56 (1H, s), 6.59 (1H, d, J = 2.9), 6.89 (1H, d, J = 8.7), 7.18–7.21 (1H, m), 7.51 (1H, d,

J = 7.9), 7.69–7.74 (1H, m), 8.48 (1H, dd, J = 0.8 and 5.0). LC-MS: m/z 338.3 [M + H]⁺.

(2R, 3R, 4aR, 10aR)-4a-Ethyl-3-methyl-2-(pyridin-2-yl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (22).



To a solution of intermediate 21 (579 mg, 1.72 mmol) in dimethoxyethane (100 mL) chilled to -15 °C was added dropwise a solution of 1.6 M methyllithium (13 mL, 21 mmol) in ether. The cooling bath was removed, and the mixture was allowed to warm to room temperature. The progress of the reaction was monitored by LC-MS. After stirring overnight, the mixture was quenched with saturated sodium bicarbonate solution, diluted with water, and extracted with ethyl acetate. The combined extracts were filtered to remove undissolved solids, washed with brine, dried (MgSO₄), and concentrated to give a tan foam. This material was purified by flash chromatography using an Isco CombiFlash eluting with ethyl acetate/ hexanes (10-50%). The desired fractions were pooled and concentrated to give 321 mg (53%) of title compound 22 as white foam. This material was crystallized from ethyl acetate/hexanes to give 133 mg of white crystals suitable for X-ray analysis. The structure was confirmed by single-crystal X-ray analysis. ¹H NMR (CDCl₃): δ 0.83 (3H, s), 0.85 (3H, t, J = 7.5), 1.24–1.33 (1H, m), 1.50 (1H, dd, J = 3.1 and 13.5), 1.52–1.57 (1H, m), 1.83 (1H, d, J = 14.5), 1.87–1.96 (1H, m), 2.21-2.28 (1H, m), 2.35-2.45 (2H, m), 2.79 (1H, d, J = 15.0). 2.87–2.93 (2H, m), 3.32 (1H, d, J = 1.6), 5.42 (1H, br s), 6.62 (1H, d, J = 2.6), 6.65 (1H, dd, J = 2.9 and 8.5), 7.12 (1H, d, J = 8.8), 7.22-7.25 (1H, m), 7.56 (1H, d, J = 8.3), 7.61–7.64 (1H, m), 8.51–8.52 (1H, m). LC-MS: m/z 354.4 [M + H]⁺. The more polar fractions were pooled and concentrated to give 100 mg (17%) of recovered 21.

(2*R*,3*R*,4a*R*,10a*R*)-4a-Ethyl-3-methyl-2-(pyridin-3-yl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (23).



Analogue 23 was synthesized analogously to analogue 22 starting with 3-bromopyridine and intermediate 18. LC-MS: m/z 354.3 [M + H]⁺. (2R,3R,4aR,10aR)-4a-Ethyl-3-methyl-2-(pyridin-4-yl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (24).



Analogue **24** was synthesized analogously to analogue **22** starting with 4-bromopyridine and intermediate **18**. ¹H NMR (CD₃OD): δ 0.79 (3H, t, *J* = 7.1.7.5), 1.06 (3H, s), 1.20–1.30 (1H, m), 1.38–1.43 (1H, m), 1.41 (1H, d, *J* = 10.8), 1.83–1.91 (3H, m), 2.26–2.37 (2H, m), 2.76–2.82 (3H, m), 6.47 (1H, d, *J* = 2.9), 6.53 (1H, dd, *J* = 2.5 and 8.3), 7.00 (1H, d, *J* = 8.3), 7.66 (2H, dd, *J* = 1.7 and 5.0), 8.34 (2H, dd, *J* = 1.5 and 4.8). LC-MS: *m*/*z* 354.4 [M + H]⁺.

(3' E, 4 a' R, 10 a' R) - 3' - B e n z y l i d e n e - 4 a' - e t h y l - 3',4',4a',9',10',10a'-hexahydro-1'*H*-spiro[[1,3]dioxolane-2,2'-phenanthren]-7'-ol (25).



A mixture of enone 8 (19.0 g, 57.2 mmol), ethylene glycol (16.0 mL, 17.8 g, 286 mmol), and p-toluenesulfonic acid monohydrate (0.5 g, 2.6 mmol) in toluene (1.4 L) was heated to reflux with water separation using a Dean-Stark trap apparatus. After 12 h, the cooled mixture was washed with saturated sodium bicarbonate solution $(1 \times 500 \text{ mL})$ and brine (2 \times 500 mL), dried (K₂CO₃), and concentrated. The residue was triturated with dichloromethane (40 mL) to afford 13.8 g (64%) of title compound 25 as beige solid. The mother liquor was concentrated and purified by flash chromatography (200g of silica gel) eluting with ethyl acetate/hexanes (10-50%) to afford an additional 3.0 g (14%) of 25. ¹H NMR (CD₃OD): δ 0.07 (3H, t, J = 7.5), 1.09-1.16 (1H, m), 1.35-1.49 (1H, m), 1.52-1.65 (1H, m), 1.69-1.80 (2H, m), 1.85 (1H, t, J = 13.1), 2.02 (1H, d, J = 13.3), 2.06-2.20 (1H, m), 2.72-2.85 (2H, m), 3.55 (1H, d, J = 13.7), 3.81-4.08 (4H, m), 6.44 (1H, d, J = 2.5), 6.46 (2H, s), 6.77 (1H, d, J = 9.1), 6.84 (1H, s), 7.19–7.40 (5H, m). LC-MS: m/z 377.3 [M + H]⁺.

(3' E, 4a' R, 10a' R) - 3' - B e n z y l i d e n e - 4a' - e t h y l -3',4',4a',9',10',10a'-hexahydro-1'*H*-spiro[[1,3]dioxolane-2,2'phenanthrene]-7'-yl 4-Nitrobenzoate (26).



To a stirred solution of ketal 25 (8.70 g, 23.1 mmol) in THF (250 mL) chilled to $-78\ ^\circ C$ was added dropwise a solution of 2.5 M nbutyllithium in hexanes (10.1 mL, 25.2 mmol) over 15 min. After an additional 15 min of stirring, p-nitrobenzoyl chloride (5.14 g, 27.7 mmol) was added in one portion. The red mixture was allowed to warm to room temperature for 1 h and then quenched with saturated ammonium chloride solution. The mixture was extracted with ethyl acetate (1×200 mL, 1×100 mL). The combined organic layers were washed with brine, dried (Na2SO4), and concentrated. The resulting foam was crystallized from acetone (50 mL) and filtered to afford 8.9 g (74%) of title compound 26 as a white solid. ¹H NMR (CDCl₃): δ 0.15 (3H, t, J = 7.3), 1.12–1.26 (1H, m), 1.45–1.59 (1H, m), 1.62– 1.70 (1H, m), 1.72–1.86 (2H, m), 1.95 (1H, t, J = 13.3), 2.16–2.29 (2H, m), 2.84–3.01 (2H, m), 3.57 (1H, d, J = 13.3), 3,85–3.92 (1H, m), 3.99-4.12 (3H, m), 6.86-6.92 (3H, m), 7.07 (1H, d, J = 8.3), 7.24-7.41 (5H, m), 8.33 (4H, s).

(4a'*R*,10a'*R*)-4a'-Ethyl-7'-hydroxy-4',4a',10',10a'-tetrahydro-1'*H*-spiro[[1,3]dioxolane-2,2'-phenanthren]-3'(9'*H*)-one (27).



A solution of olefin 26 (22.2 g, 42.2 mmol) in a mixture of dichloromethane (440 mL) and methanol (220 mL) was chilled to -78 °C. A stream of ozone was bubbled into the mixture for 15 min, at which time it became blue. Ozone treatment was continued for an additional 15 min, and the excess ozone was purged by bubbling nitrogen into the mixture. Excess dimethyl sulfide (40 mL) was added, and the mixture was slowly allowed to warm to room temperature. The solution was concentrated overnight under a stream of nitrogen. The residual solid was triturated with hexanes (500 mL) and filtered to give beige crystals, which were dissolved in THF (780 mL). The mixture was cooled to 0 °C, treated with a solution of 1 N sodium hydroxide (100 mL, 100 mmol), and then stirred for 5 h. The aqueous layer was separated and extracted with ethyl acetate (1 \times 300 mL, 1 \times 200 mL). The combined organic layers were washed with brine (2 \times 200 mL), dried (MgSO₄), and concentrated to afford 12.5 g (98%) of title compound 27 as a foam. ¹H NMR (CDCl₃): δ 0.72 (3H, t, J =

7.5), 1.35–1.43 (2H, m), 1.68–1.88 (2H, m), 1.97 (1H, dd, J = 4.2 and 14.1), 2.09 (1H, apparent t, J = 13.9), 2.33–2.44 (1H, m), 2.67 (1H, dd, J = 0.8, 13.3), 2.81–2.98 (2H, m), 3.10 (1H, d, J = 13.3), 3.73–3.82 (1H, m), 3.92–3.98 (1H, m), 4.05–4.15 (2H, m), 4.99 (1H, br s), 6.59–6.61 (2H, m), 6.92 (1H, d, J = 8.7). LC-MS: m/z 303.4 [M + H]⁺, 301.3 [M – H]⁻, 347.3 [M – H + H₂CO₂]⁻.

(3'*R*,4a'*R*,10a'*R*)-4a'-Ethyl-3'-methyl-3',4',4a',9',10',10a'hexahydro-1'*H*-spiro[[1,3]dioxolane-2,2'-phenanthrene]-3',7'diol (28).



To a solution of ketone 27 (12.5 g, 41.3 mmol) in THF (1 L) chilled to -78 °C was added a solution of 1.5 M methyllithium-lithium bromide complex in diethyl ether (138 mL, 207 mmol) slowly via canula over 15 min. After 1 h, the suspension was allowed to warm to room temperature, and stirring was continued for 12 h. Saturated ammonium chloride solution (200 mL) was added, and the organic layer was separated and saved. The aqueous layer was extracted with ethyl acetate (1×500 mL, 1×200 mL). The combined organic layers were washed with brine (3 \times 500 mL), dried (Na₂SO₄), and concentrated to afford 12.3 g (97%) of title compound 28 as a beige solid. ¹H NMR (CDCl₃): δ 0.68 (3H, t, J = 6.6), 1.09 (3H, s), 1.08– 1.19 (1H, m), 1.30–1.35 (1H, m), 1.39 (1H, d, J = 14.5), 1.48–1.52 (1H, m), 1.72–1.93 (2H, m), 1.97–2.12 (2H, m), 2.42 (1H, d, J = 14.1), 2.70-2.80 (2H, m), 3.85-3.96 (4H, m), 6.43-6.51 (2H, m), 6.89 (1H, d, J = 8.3). LC-MS: m/z 301.4 $[M + H - H_2O]^+$, 317.4 [M- H]-.

(3*R*, 4a*R*, 10a*R*)-4a-Ethyl-3, 7-dihydroxy-3-methyl-3,4,4a,9,10,10a-hexahydrophenanthren-2(1*H*)-one (29).



A solution of ketal 28 (12.7 g, 40 mmol) in THF (500 mL) was diluted with a solution of 2 N hydrochloric acid (350 mL), and the mixture was heated to reflux for 2 h. After cooling, the organic layer was separated and saved, and the aqueous layer was extracted with ethyl acetate (1×300 mL, 1×150 mL). The combined organic layers were washed with saturated sodium bicarbonate solution (1×300) mL) and brine $(2 \times 300 \text{ mL})$, dried (Na_2SO_4) , and concentrated. The brown residue was crystallized from ether to give 2.4 g of title compound 29 as a beige solid. The mother liquor was redissolved in ether and stored in the refrigerator to give 2.3 g of a second crop and 1.3 g of a third crop of 29. The mother liquor from the third crop was purified by flash chromatography (300 g of silica gel) eluting with ethyl ether/hexanes (50-100%). During the charging process, 0.3 g of 29 crystallized out of solution and was recovered. The desired fractions were pooled and concentrated to afford 3.3 g of 29; total yield = 9.6 g (87%). ¹H NMR (CDCl₃): δ 0.76 (3H, t, J = 7.3), 1.36 (3H, s), 1.38– 1.55 (2H, m), 1.64–1.72 (1H, m), 1.82–1.91 (1H, m), 1.92 (1H, dd, J = 1.2 and 14.5), 2.30-2.40 (1H, m), 2.48-2.66 (2H, m), 2.79 (1H, d, *J* = 14.5), 2.82–2.90 (2H, m), 6.55 (1H, d, *J* = 2.5), 6.62 (1H, dd, *J* = 2.7 and 8.5), 7.00 (1H, d, J = 8.3). LC-MS: m/z 275.3 [M + H]⁺, 273.3 $[M - H]^{-}$, 319.4 $[M - H + H_2CO_2]^{-}$

Representative Procedure A: (25,3*R*,4a*R*,10a*R*)-4a-Ethyl-2,3dimethyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7triol (30).



In a 20 mL parallel synthesis reaction vessel, lithium chloride (46 mg, 1.1 mmol) was dried at 150 °C under high vacuum for 30 min. The flask was flushed with nitrogen and chilled to -30 °C. A solution of ketone 29 (50 mg, 0.18 mmol) in THF (10 mL) was added followed by a solution of 1 M methyllithium-lithium iodide complex in ethyl ether (1.1 mL, 1.1 mmol). After stirring for 1 h, the mixture was allowed to warm to room temperature and then stirred for an additional 12 h. The reaction was quenched by the addition of 1 N hydrochloric acid solution (5 mL), diluted with water (15 mL), and extracted with ethyl acetate (2 \times 20 mL). The combined organic layers were washed with brine, dried (Na2SO4), and concentrated. The residue was purified by flash chromatography using a Biotage 12 M and eluting with ethyl acetate/hexanes (20-40%). The desired fractions were pooled and concentrated to afford 21 mg (43%) of title compound 30 as a solid. ¹H NMR (CDCl₂): δ 0.79 (3H, t, I =7.5), 1.25 (3H, s), 1.26 (3H, s), 1.26-1.30 (1H, m), 1.36-1.42 (2H, m), 1.58-1.68 (1H, m), 1.73-1.82 (1H, m), 1.84-1.93 (1H, m), 2.04–2.20 (2H, m), 2.55 (1H, d, J = 14.5), 2.85–2.91 (2H, m), 5.30 (1H, br s), 6.55–6.61 (2H, m), 7.00 (1H, d, J = 8.3). LC-MS: m/z273.7 $[M + H - H_2O]^+$.

Representative Procedure B: (2*S*,3*R*,4*aR*,10*aR*)-2,4a-Diethyl-3-methyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (31).



In a 20 mL parallel synthesis reaction vessel, lithium chloride (46 mg, 1.1 mmol) was dried at 150 $^\circ C$ under high vacuum for 30 min. The flask was flushed with nitrogen and chilled to -30 °C. A solution of ketone 29 (50 mg, 0.18 mmol) in THF (10 mL) was added followed by a solution of 1 M ethylmagnesium bromide in THF (1.1 mL, 1.1 mmol). After stirring for 1 h, the mixture was allowed to warm to room temperature and then stirred for an additional 12 h. The reaction mixture was quenched by the addition of a solution 1 N hydrochloric acid (5 mL), diluted with water (15 mL), and extracted with ethyl acetate (2 \times 20 mL). The combined organic layers were washed with brine, dried (Na_2SO_4) , and concentrated. The residue was purified by preparative HPLC using a Waters Symmetry C-8 30 mm × 50 mm column and eluting with acetonitrile/water (30-90% containing 0.1% formic acid) over 15 min (flow rate = 25 mL/min). The desired fractions were pooled and concentrated to afford 10 mg (18%) of title compound 31 (containing 25% of the 2β -ethyl epimer). ¹H NMR (CDCl₃): δ 0.69–0.74 (3H, two overlapping triplets), 0.84 and 0.91 (3H total, two triplets, I = 7.7, ratio 1:3), 1.13 and 1.22 (3H total, two singlets, ratio 1:3), 1.14-1.26 (1H, m), 1.30-2.18 (10.25H, m), 2.24 (0.75H, d, J = 14.5), 2.78–2.88 (2H, m), 6.49–6.55 (2H, m), 6.93 and 6.94 (1H, two doublets, J = 8.1, ratio 1:3). LC-MS: m/z 303.3 [M - $H^{-}, 349.3 [M - H + H_2CO_2]^{-}.$

(2 *S*, 3 *R*, 4 a *R*, 1 0 a *R*) - 4 a - E thyl-3 - m e thyl-2 - propyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (32).



Analogue **32** was obtained following procedure B using a solution of 2.5 M propylmagnesium bromide in hexane; yield = 6%. The 2β -propyl epimer was also isolated in 10% yield. ¹H NMR (CDCl₃): δ 0.73 (3H, t, *J* = 7.3), 0.85 (3H, t, *J* = 6.7), 1.15 (3H, s), 1.11–1.19 (1H, m), 1.20–1.31 (2H, m), 1.32–1.47 (3H, m), 1.50–1.61 (3H, m), 1.74–1.87 (2H, m), 2.08–2.17 (1H, m), 2.47 (1H, d, *J* = 14.5), 2.80–3.00 (2H, m overlapping with water peak), 6.48–6.56 (2H, m), 6.93 (1H, d, *J* = 8.3). LC-MS: m/z 301.1 [M + H – H₂O]⁺, 317.1 [M – H]⁻, 363.0 [M – H + H₂CO₂]⁻.

(2 S, 3 R, 4 a R, 1 0 a R) - 2 - A ||y|-4 a - e t h y|-3 - m e t h y|-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (33).



Analogue **33** was obtained following procedure B using a solution of 1 M allylmagnesium bromide in ethyl ether; yield = 44%. The 2β -allyl epimer was also isolated in 9% yield. ¹H NMR (CDCl₃): δ 0.62 (3H, t, J = 7.5), 1.06–1.12 (1H, m), 1.14 (3H, s), 1.27 (1H, dd, J = 2.9 and 13.7), 1.35–1.47 (1H, m), 1.53 (1H, d, J = 14.1), 1.63–1.79 (2H, m), 1.82–1.95 (2H, m), 2.13 (1H, d, J = 14.2), 2.23–2.39 (2H, m), 2.68–2.74 (2H, m), 4.96–5.05 (2H, m), 5.78–5.88 (1H, m), 6.38–6.45 (2H, m), 6.83 (1H, d, J = 8.3). LC-MS: m/z 299.1 [M + H – H₂O]⁺, 315.1 [M – H]⁻, 361.0 [M – H + H₂CO₂]⁻.

(2 R, 3 R, 4 a R, 10 a R) - 4 a - Ethyl - 3 - methyl - 2 - vinyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (34).



Analogue 34 was obtained following procedure B using a solution of 1 M vinylmagnesium bromide in THF; yield = 24% as a 1:1 mixture of 2α and β epimers. ¹H NMR (CDCl₃): δ 0.70–0.75 (3H, two overlapping triplets), 1.06 and 1.15 (3H total, two singlets), 1.12–1.24 (1H, m), 1.29–1.37 (1H, m), 1.48–1.55 (1H, m), 1.61 (0.5H, d, *J* = 14.1), 1.75–1.84 (1.5H, m), 2.00–2.18 (3H, m), 2.28 (0.5H, d, *J* = 14.5), 2.52 (0.5H, d, *J* = 14.9), 2.77–2.86 (2H, m), 5.04 (0.5H, d, *J* = 11.2), 5.18 (0.5H, d, *J* = 10.8), 5.29 (0.5H, d, *J* = 17.4), 5.36 (0.5H, d, *J* = 17.0), 6.09–6.20 (1H, m), 6.48–6.52 (2H, m), 6.93 (1H, d, *J* = 8.3). LC-MS: m/z 301 [M – H]⁻.

(2R,3R,4aR,10aR)-4a-Ethyl-3-methyl-2-((E)-prop-1-enyl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (35).



Analogue **35** was obtained following procedure B using a solution of 0.5 M (*E*)-propenylmagnesium bromide in THF; yield = 28% as a 4:1 mixture of 2α - and 2β -(*E*)-prop-1-enyl epimers. ¹H NMR (CDCl₃): δ 0.78 (3H, t, *J* = 7.3), 1.12 and 1.20 (3H total, two s), 1.17–1.25 (1H, m), 1.36–1.44 (3H, m), 1.67 (3H, d, *J* = 5.4), 1.75–1.97 (3H, m), 2.04–2.23 (2H, m), 2.35 (0.2H, d, *J* = 14.5), 2.56 (0.8H, d, *J* = 14.9), 2.82–2.88 (2H, m), 5.53–5.55 (0.2H, m), 5.75–5.87 (1.8H, m), 6.55–6.62 (2H, m), 6.98–7.03 (2H, overlapping doublets). LC-MS: m/z 299.3 [M + H – H₂O]⁺, 315.2 [M – H]⁻, 361.3 [M – H + H₂CO₂]⁻.

(2*R*, 3*R*, 4a*R*, 10a*R*)-4a-Ethyl-3-methyl-2-(prop-1-ynyl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (36).



Analogue **36** was obtained following procedure B using a solution of 0.5 M propynylmagnesium bromide in THF; yield = 13%. The 3 β -propargyl epimer was also isolated in 36% yield. ¹H NMR (CDCl₃): δ 0.0.77 (3H, t, *J* = 7.6), 1.11–1.23 (1H, m), 1.38 (3H, s), 1.52–1.71 (4H, m), 1.77 (3H, m), 2.04–2.15 (3H, m), 2.52 (1H, d, *J* = 14.5), 2.85–2.94 (2H, m), 6.52–6.61 (2H, m), 7.08 (1H, d, *J* = 7.9). LC-MS: *m*/*z* 297.3 [M + H – H₂O]⁺, 313.3 [M – H]⁻, 359.3 [M – H + H₂CO₂]⁻.

(2*R*, 3*R*, 4a*R*, 10aR)-4a-Ethyl-3-methyl-2-*p*-tolyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (37).



Analogue **37** was obtained following procedure A using a solution of 1 M *p*-tolylmagnesium bromide in THF; yield = 15% after trituration with ether, mp 213–214 °C. ¹H NMR (CDCl₃): δ 0.78 (3H, t, *J* = 7,6), 1.11 (3H, s), 1.15–1.30 (1H, m), 1.31–1.43 (1H, m), 1.48 (1H, dd, *J* = 2.5 and 12.9), 1.75–1.93 (4H, m), 2.15–2.25 (1H, m), 2.25 (3H, s), 2.69–2.78 (3H, m), 6.51 (1H, d, *J* = 2.5), 6.58 (1H, dd, *J* = 2.5 and 8.3), 6.98–7.05 (3H, m), 7.45 (2H, *J* = 8.3). LC-MS: *m*/*z* 349.3 [M + H – H₂O]⁺, 365.3 [M – H]⁻, 411.2 [M – H + H₂CO₂]⁻.

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(3*R*,4a*R*,10a*R*)-7-(*tert*-Butyldimethylsilyloxy)-4a-ethyl-3-hydroxy-3-methyl-3,4,4a,9,10,10a-hexahydrophenanthren-2(1*H*)-one (38).



A solution of 2.02 g (7.35 mmol) of ketone 29 in dichloromethane (20 mL) was chilled to 0 $^{\circ}\mathrm{C}$ and treated sequentially with imidazole (550 mg, 8.08 mmol) and a solution of 1.0 M TBDMSCl in dichloromethane (8.0 mL, 8.0 mmol). A precipitate formed almost immediately, and the ice bath was removed. After 1.25 h of stirring, the reaction was diluted with water and the layers were separated. The organic layer was dried (MgSO₄) and evaporated. The residual oil was purified by flash chromatography using a Biotage 40 Samplet (40S column) eluting with ethyl acetate/heptanes (0-50%). The desired fractions were pooled and concentrated to give 1.94 g (68%) of title compound 38 as a semisolid. ¹H NMR (DMSO- d^6): δ 0.12 (6H, s), 0.73 (3H, t, J = 8), 0.90 (9H, s), 1.14 (3H, s), 1.18–1.29 (1H, m), 1.32 (1H, d, J = 12), 1.52–1.60 (1H, m), 1.75–1.94 (2H, m), 2.06 (1H, dd, J = 1 and 12), 2.19–2.27 (1H, m), 2.71 (1H, d, J = 12), 2.78 (2H, br t), 2.93 (1H, t, J = 12), 5.20 (1H, s), 6.49 (1H, s), 6.53 (1H, d, J = 8), 6.98 (1H, d, J = 8). LC-MS: m/z 371.2 $[M + H - H_2O]^+$.

(2*S*,3*R*,4a*R*,10a*R*)-4a-Ethyl-3-methyl-2-(thiazol-2-yl)-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7-triol (39).



A solution of 1.7 M tert-butyllithium in pentane (31 mL, 53 mmol) was added dropwise to a prechilled $(-78 \ ^\circ C)$ solution of 2bromothiazole (8.5 g, 52 mmol) in ethyl ether (100 mL). When the addition was complete, the mixture was stirred for 5 min and a solution of ketone 38 (4.00 g, 10.3 mmol) in ethyl ether (100 mL) was added dropwise over 50 min. The mixture was stirred for an additional 2.5 h and then quenched with ammonium chloride solution. After warming to room temperature, the organic layer was separated, and the aqueous layer extracted with ethyl ether. The combined organic layers were dried (Na2SO4) and evaporated. The residual oil was purified by flash chromatography using a Biotage 40S column eluting with ethyl acetate/heptanes (0-10%). The desired fractions were pooled and concentrated to afford 2.36 g (48%) of the desired carbinol as a foam. The impure fractions were evaporated and repurified by flash chromatography eluting with ethyl acetate/heptanes (0-25%) to give an additional 1.84 g (38%) of carbinol. Total yield = 4.2 g (86%). LC-MS: m/z 474.1 [M + H]⁺.

A solution of 1.88 g (3.97 mmol) of the above material in THF (40 mL) was treated with a solution of 1 M TBAF in THF (4.80 mL, 4.80 mmol). After stirring for 5 min, analysis by LC-MS indicated that the

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reaction was complete. This mixture was concentrated and purified by flash chromatography using a Biotage 40S column eluting with ethyl acetate/heptanes (0-100%) followed by flushing with dichloromethane/methanol. The desired fractions were pooled and concentrated, and the residue was triturated with ethyl ether/heptanes to afford 1.37 g of a solid, which was a mixture of 2α and 2β isomers by LC-MS. The material was recrystallized from acetonitrile-water at room temperature to afford 537 mg (38%) of title compound 39. An additional 172 mg (12%) of 39 was obtained similarly by recrystallization of the concentrated mother liquor. Total yield = 705 mg (50%). ¹H NMR (DMSO- d^6): δ 0.71 (3H, t, J = 7.1), 0.80 (3H, s), 0.98–1.08 (1H, m), 1.29–1.43 (2H, m), 1.58–1.77 (1H, m), 2.06-2.26 (3H, m), 2.34 (1H, d, J = 14.1), 2.48-2.56 (1H, m), 2.62-2.72 (2H, m), 4.19 (1H, s), 5.70 (1H, s), 6.40 (1H, d, J = 2.5), 6.44 (1H, dd, J = 2.5 and 8.4), 6.89 (1H, d, J = 8.6), 7.47 (1H, d, J = 3.3), 7.56 (1H, d, J = 3.3), 8.89 (1H, s). LC-MS: m/z 360.1 [M + H]⁺.

(2R,3R,4aR,10aS)-4a-Ethyl-2,3,7-trihydroxy-3-methyl-2-phenyl-2,3,4,4a,10,10a-hexahydrophenanthren-9(1*H*)-one (40).



(2*R*,3*S*,4a*R*,10a*S*)-4a-Ethyl-2,3,7-trihydroxy-3-methyl-2-phenyl-2,3,4,4a,10,10a-hexahydrophenanthren-9(1*H*)-one (41).



(2*R*, 3*R*, 4a*R*, 10a*S*)-4a-Ethyl-3, 9-dimethyl-2-phenyl-1, 2, 3, 4, 4a, 10a-hexahydrophenanthrene-2, 3, 7-triol (42).



(2*R*, 3*S*, 4a*R*, 10a*S*)-4a-Ethyl-3, 9-dimethyl-2-phenyl-1, 2, 3, 4, 4a, 10a-hexahydrophenanthrene-2, 3, 7-triol (43).



Intermediate 12 (100 mg, 0.285 mmol) was dissolved in a 1:1 mixture of THF and toluene (10 mL) and treated with a solution of 1.5 M methyllithium–lithium bromide complex (1.90 mL, 0.285 mmol) in diethyl ether. The mixture was stirred for 2 h at room temperature, and the excess methyllithium was quenched by the addition of 1 N hydrochloric acid solution. The mixture was extracted with ethyl acetate (3×30 mL), and the combined extracts were washed with brine, dried (Na₂SO₄), and concentrated to give 129 mg of an oil. This material was purified by preparative reverse phase HPLC: Waters Symmetry C-8 column (30 mm \times 50 mm) eluting with 30–70% acetonitrile in water (each containing 0.1% formic acid) over 15 min at a flow rate of 25 mL/min.

Fraction 2 was concentrated to give 3.5 mg (3%) of analogue 41 as a white solid. ¹H NMR (CD₃OD): δ 0.77 (3H, t, *J* = 7.3), 1.27–1.33 (1H, m), 1.43 (3H, s), 1.97 (1H, dd, *J* = 3.7 and 14.1), 2.08–2.15 (1H, m), 2.18–2.29 (3H, m), 2.40–2.53 (1H, m), 2.57–2.68 (2H, m), 6.98 (1H, dd, *J* = 2.6 and 8.5), 7.12–7.17 (1H, m), 7.20–7.29 (4H, m),

7.77–7.79 (2H, m). LC-MS: m/z 367.3 [M + H]⁺, 365.3 [M – H]⁻, 411.3 [M – H + H₂CO₂]⁻.

Fraction 4 was concentrated to give 5.0 mg (5%) of analogue **43** as a white solid. ¹H NMR (CD₃OD): δ 0.59 (3H, t, *J* = 7.5), 1.47 (3H, s), 1.51–1.61 (1H, m), 1.69–1.77 (1H, m), 1.89–1.93 (4H, m), 2.22 (1H, t, *J* = 13.7), 2.34 (1H, d, *J* = 14.1), 2.43–2.51 (1H, m), 2.50 (1H, d, *J* = 14.1), 5.31 (1H, s), 6.60 (1H, dd, *J* = 2.5 and 8.3), 6.66 (1H, d, *J* = 2.5), 6.95 (1H, d, *J* = 8.3), 7.10–7.14 (1H, m), 7.20–7.24 (2H, m), 7.81 (2H, d, *J* = 8.1). LC-MS: *m/z* 347.3 [M + H – H₂O]⁺, 363.3 [M – H]⁻, 409.3 [M – H + H₂CO₂]⁻.

Fraction 5 was concentrated to give 26 mg (25%) of analogue 42 as a white solid. ¹H NMR (CD₃OD): δ 0.56 (3H, t, *J* = 7.5), 1.18 (3H, s), 1.45–1.51 (2H, m), 1.92 (3H, s), 2.10 (1H, d, *J* = 15.0), 2.15–2.27 (1H, m), 2.49–2.60 (1H, m), 2.50 (1H, d, *J* = 11.6), 2.62 (1H, d, *J* = 14.5), 5.25 (1H, s), 6.63 (1H, dd, *J* = 2.5 and 8.3), 6.68 (1H, d, *J* = 2.5), 6.98 (1H, d, *J* = 8.3), 7.13–7.17 (1H, m), 7.23 (2H, t, *J* = 7.7), 7.63 (2H, d, *J* = 8.1). LC-MS: *m/z* 347.3 [M + H – H₂O]⁺, 363.3 [M – H]⁻, 409.3 [M – H + H₂CO₂]⁻. The structure was confirmed by single-crystal X-ray analysis on crystals prepared by recrystallization from ethyl acetate–hexanes, mp 257–260 °C.

Fraction 8 was concentrated to give 26 mg (28%) of analogue **40** as a white solid. ¹H NMR (CD₃OD): δ 0.75 (3H, t, *J* = 7.3), 1.15–1.20 (1H, m), 1.17 (3H, s), 1.46 (1H, d, *J* = 10.4), 2.05 (1H, d, *J* = 15.0), 2.15 (1H, dd, *J* = 3.9 and 18.9), 2.38–2.45 (2H, m), 2.56–2.65 (2H, m), 2.80 (1H, d, *J* = 15.0), 7.00 (1H, dd, *J* = 2.9 and 8.7), 7.11–7.14 (1H, m), 7.15–7.24 (3H, m), 7.31 (1H, d, *J* = 2.9), 7.61 (2H, d, *J* = 8.1). LC-MS: *m*/*z* 367.3 [M + H]⁺, 365.3 [M – H]⁻, 411.3 [M – H + H₂CO₂]⁻. The structure was confirmed by single-crystal X-ray analysis on crystals prepared by recrystallization from ethanol–water, mp 148–150 °C.

(4bR,6R,7R,8aR)-4b-Ethyl-6,7-dihydroxy-6-methyl-7-phenyl-4b,5,6,7,8,8a,9,10-octahydrophenanthren-2-yl 4-Nitrobenzoate (44).



To a solution of 13 (300 mg, 0.851 mmol) in acetone (50 mL) was added a solution of 1 N sodium hydroxide (0.890 mL, 0.890 mmol) followed by p-nitrobenzoyl chloride (189 mg, 1.02 mmol). The mixture was stirred for 1 h, and additional p-nitrobenzoyl chloride (90 mg, 0.48 mmol) was added. Additional 1 N sodium hydroxide solution was added periodically to maintain a neutral pH. The mixture was concentrated, diluted with water (75 mL), and extracted with ethyl acetate (2 \times 75 mL). The combined extracts were washed with saturated sodium bicarbonate solution and brine, dried (MgSO₄), and concentrated to give 447 mg (>100%) of title compound 44 as a foam. This material was used in the next step without further purification. ¹H NMR (CDCl₃): δ 0.84 (3H, t, J = 7.6), 1.18 (3H, s), 1.26–1.32 (1H, m), 1.42-1.52 (1H, m), 1.59 (1H, dd, J = 2.7 and 13.1), 1.84-2.04(3H, m), 2.27–2.38 (2H, m), 2.83 (1H, d, J = 15.0), 2.87–2.92 (2H, m), 6.93 (1H, d, J = 2.5), 6.98 (1H, dd, J = 2.7 and 8.5), 7.19–7.31 (4H, m), 7.61 (2H, d, J = 7.5), 8.35 (4H, s). LC-MS: *m*/*z* 546.4 [M - $H + H_2CO_2$

(2R,3R,4aR,10aS)-4a-Ethyl-2,3,7-trihydroxy-3-methyl-2-phenyl-2,3,4,4a,10,10a-hexahydrophenanthren-9(1*H*)-one (40).



A solution of PNP ester 44 (0.851 mmol) dissolved in a mixture of 1:1 dichloromethane and methanol (125 mL) was chilled to -30 °C. Ozone was bubbled into the mixture until a blue color persisted for 1

h. The mixture was purged with nitrogen, and excess dimethyl sulfide (ca. 2 mL) was added using a Pasteur pipet. After warming to room temperature and stirring overnight, the mixture was concentrated and the residual oil was dissolved in THF (50 mL). A solution of 1 N sodium hydroxide (2.1 mL, 2.1 mmol) was added and, after 1 h of stirring, additional 1 N sodium hydroxide solution (3.0 mL, 3.0 mmol) was added and stirring was continued for 1 h. Saturated sodium bicarbonate solution (100 mL) was added, and the organic layer was separated and saved. The aqueous layer was extracted with ethyl acetate (100 mL), and the combined organic layers were washed with saturated sodium bicarbonate solution and brine, dried (MgSO₄), and concentrated. The residue was purified by flash chromatography using a Biotage Flash 40 S eluting with ethyl acetate/hexanes (30-40%). The appropriate fractions were pooled and concentrated to afford 190 mg (61%) of title compound 40. ¹H NMR (CDCl₃): δ 0.76 (3H, t, J = 7.6), 1.22 (3H, s), 1.15–1.26 (1H, m), 1.55 (1H, d, J = 10.3), 2.03 (1H, d, J = 15.3), 2.25 (1H, dd, J = 3.9 and 18.9), 2.41–2.48 (2H, m), 2.50-2.57 (1H, m), 2.61-2.73 (1H, m), 2.81 (1H, d, J = 15.0), 2.95 (1H, br s), 3.13 (1H, br s), 7.07 (1H, dd, J = 2.9 and 8.3), 7.15–7.28 (4H, m), 7.55-7.59 (3H, m).

(4bR,6R,7R,8aS)-4b-Ethyl-6,7-dihydroxy-6-methyl-10-oxo-7-phenyl-4b,5,6,7,8,8a,9,10-octahydrophenanthren-2-yl Benzoate (45).



To a solution of phenol ${\bf 40}~(1.374~{\rm g},\,3.749~{\rm mmol})$ in acetone (50 mL) was added a solution of 1 N sodium hydroxide (3.94 mL, 3.94 mmol). After stirring for 5 min, benzoyl chloride (0.457 mL, 408 mg, 3.94 mmol) was added, and the mixture was stirred for 2.5 h at room temperature. Saturated ammonium chloride solution (5 mL) was added, and the mixture was partially evaporated to remove acetone. The residue was partitioned between saturated ammonium chloride solution (50 mL) and ether (50 mL). The ether layer was separated and the aqueous layer further extracted with ether (50 mL). The combined ether layers were washed with saturated sodium bicarbonate solution (50 mL) and brine (50 mL), dried (Na₂SO₄), and concentrated to give 1.62 g (92%) of title compound 45 as a yellowish white solid. ¹H NMR (CDCl₃): δ 0.85 (3H, t, J = 7.3), 1.27 (3H, s), 1.23-1.36 (2H, m), 1.40-1.90 (1H, br s), 1.64 (1H, d, J =11.9), 2.13 (1H, d, J = 14.5), 2.35 (1H, dd, J = 4.1 and 18.7), 2.46-2.77 (4H, m), 2.90 (1H, d, J = 14.5), 7.26-7.70 (10H, m), 7.86 (1H, d, J = 2.1), 8.22 (2H, d, J = 8.3). LC-MS: m/z 471.4 [M + H]⁺, 469.4 $[M - H]^{-}$, 515.4 $[M - H + H_2CO_2]^{-}$.

(4bR,6R,7R,8aR,10R)-4b-Ethyl-6,7,10-trihydroxy-6-methyl-7phenyl-4b,5,6,7,8,8a,9,10-octahydrophenanthren-2-yl Benzoate (46).



A solution of ketone **45** (249 mg, 0.529 mmol) in methanol (10 mL) was cooled to 0 °C and treated with sodium borohydride (30 mg, 0.80 mmol). The mixture was stirred for 2 h, and additional sodium borohydride (15 mg, 0.40 mmol) was added. The ice bath was removed, and the mixture was stirred for 2 h. Saturated ammonium chloride solution was added, and the mixture was extracted with ethyl acetate (3 × 25 mL). The combined extracts were washed with saturated sodium bicarbonate solution (1 × 25 mL) and brine (1 × 25 mL), dried (Na₂SO₄), and evaporated to give 235 mg of a light brown foam. This material was purified by flash chromatography using a Biotage Flash 40 eluting with 2.75:1 hexanes/ethyl acetate. The

appropriate fractions were pooled and concentrated to give 89 mg (36%) of title compound 46 as a white foam. ¹H NMR (CDCl₃): δ 0.92 (3H, t, *J* = 6.7,7.3), 1.21 (3H, s), 1.43–1.52 (1H, m), 1.56 (1H, d, *J* = 13.0), 1.72 (2–3H, br s), 1.87–1.92 (1H, m), 1.97–2.11 (2H, m), 2.39–2.48 (2H, m), 2.86 (1H, d, *J* = 15.0), 4.83 (1H, t, *J* = 8.3), 7.11–7.13 (1H, m), 7.28–7.38 (5H, m), 7.52–7.55 (2H, m), 7.62–7.68 (3H, m), 8.21 (2H, d, *J* = 8.3). LC-MS: *m*/*z* 471.3 [M – H]⁻, 517.3 [M – H + H₂CO₃]⁻.

(2*R*, 3*R*, 4a*R*, 9*R*, 10a*R*)-4a-Ethyl-3-methyl-2-phenyl-1,2,3,4,4a,9,10,10a-octahydrophenanthrene-2,3,7,9-tetraol (47).



A solution of benzoate **46** (29 mg, 0.061 mmol) in THF (2 mL) was treated with 1 N sodium hydroxide solution (2.0 mL, 2.0 mL), and the mixture was stirred vigorously overnight. The mixture was poured onto saturated ammonium chloride solution (20 mL) and extracted with ethyl acetate (2 × 20 mL). The combined extracts were washed with saturated sodium bicarbonate solution and brine, dried (NaSO₄), and concentrated to give 23 mg (100%) of title compound **47** as an off-white solid. ¹H NMR (CDCl₃): δ 0.85 (3H, t, *J* = 7.6), 1.17 (3H, s), 1.32–1.42 (1H, m), 1.48–1.80 (4H br m), 1.52 (1H, dd, *J* = 2.1 and 12.9), 1.79–1.86 (1H, m), 1.88–2.00 (2H, m), 2.31–2.41 (2H, m), 2.78 (1H, d, *J* = 15.0), 4.73 (1H, t, *J* = 8.3), 6.72 (1H, dd, *J* = 2.7 and 8.5), 6.97 (1H, d, *J* = 2.9), 7.07 (1H, d, *J* = 8.3), 7.20–7.31 (3H, m), 7.58 (2H, d, *J* = 7.1). LC-MS: *m/z* 333.5 [M + H – 2H₂O]⁺, 367.5 [M – H]⁻, 413.5 [M – H + H₂CO₂]⁻.

(6a*S*,7a*R*,9*R*,10a*R*,11a*R*)-11a-Ethyl-9-(4-methoxyphenyl)-10a-methyl-5-oxo-7a-phenyl-5,6,6a,7,7a,10a,11,11aoctahydrophenanthro[2,3-*d*][1,3]dioxol-3-yl Benzoate (48).



To a solution of intermediate 45 (1.78 g, 3.78 mmol) in benzene (50 mL) was added p-anisaldehyde dimethylacetal (6.45 mL, 6.89 g, 37.8 mmol) followed by p-toluenesulfonic acid (10 mg). After 5 min of stirring, the mixture became blue and was stirred for 7 h at room temperature. The mixture was poured onto saturated sodium bicarbonate solution (75 mL), and the organic layer was separated and saved. The aqueous layer was extracted with ether $(2 \times 50 \text{ mL})$, and the combined organic layers were washed with brine, dried (Na_2SO_4) , and concentrated. The residual oil was purified by flash chromatography using a Biotage Flash 40 eluting with 10:1 to 5:1 hexanes in ether to give 1.39 g (62%) of title compound 48 as a yellow-white solid. ¹H NMR (CDCl₃): δ 0.87 (3H, t, J = 7.1), 1.04 (3H, s), 1.38–1.51 (1H, m), 1.79 (1H, dd, J = 3.1 and 14.3), 1.87 (1H, d, J = 15.0), 2.39–2.53 (3H, m), 2.69–2.81 (2H, m), 3.17 (1H, d, J = 15.0), 3.82 (3H, s), 6.22 (1H, s), 6.91-6.94 (2H, m), 7.24-7.54 (11H, m), 7.63–7.67 (1H, m), 7.90 (1H, d, J = 2.5), 8.20 (2H, dd, J = 1.2 and 8.3).

(5*R*,6a*R*,7a*R*,9*R*,10a*R*,11a*R*)-11a-Ethyl-5-hydroxy-9-(4-methoxyphenyl)-10a-methyl-7a-phenyl-5,6,6a,7,7a,10a,11,11aoctahydrophenanthro[2,3-*d*][1,3]dioxol-3-yl Benzoate (49).



Ketone 48 (994 mg, 1.69 mmol) was dissolved in a mixture of methanol (20 mL) and THF (10 mL), chilled to -78 °C, and treated with sodium borohydride (76.6 mg, 2.02 mmol). The mixture was slowly allowed to warm to 0 °C and then stirred for 1.5 h. The mixture was poured onto saturated ammonium chloride solution (5 mL), and the contents were partially evaporated to remove methanol and THF. The aqueous residue was partitioned between ether (40 mL) and saturated ammonium chloride solution (40 mL). The organic layer was separated and saved and the aqueous layer extracted with ether (40 mL). The combined organic layers were washed with brine, dried (Na_2SO_4) , and concentrated to give 1.03 g (theory = 997 mg) of title compound 49 as a yellow solid. This material was used directly in the next step without further purification. ¹H NMR (CDCl₃): δ 0.93 (3H, t, J = 7.1), 1.01 (3H, s), 1.14–1.28 (2H, m), 1.68–1.72 (1H, m), 1.76 (1H, d, J = 10.4), 1.94–2.05 (2H, m), 2.23–2.40 (2H, m), 2.46 (1H, t, J = 13.9, 3.12 (1H, d, J = 15.4), 3.81 (3H, s), 4.92 (1H, t, J = 8.7), 6.18 (1H, s), 6.92 (2H, d, J = 8.7), 7.11 (1H, dd, J = 2.5 and 8.3), 7.25-7.37 (4H, m), 7.42 (1H, d, J = 2.5), 7.49-7.53 (6H, m), 7.62-7.66 (1H, m), 8.20 (2H, d, I = 7.1).

(6a*S*,7a*R*,9*R*,10a*R*,11a*R*)-11a-Ethyl-9-(4-methoxyphenyl)-10a - methyl - 7a - phenyl - 6a, 7, 7a, 10a, 11, 11a hexahydrophenanthro[2,3-*d*][1,3]dioxol-3-yl Benzoate (50).



A mixture of alcohol 49 (1.69 mmol), dry benzene (15 mL), and Burgess reagent (483 mg, 2.03 mmol) was heated to 60 °C for 3 h. Additional Burgess reagent (75 mg, 0.31 mmol) was added, and heating was continued for 3 h. The mixture was cooled and diluted with brine. The organic layer was separated and saved, and the aqueous layer was extracted with ether $(2 \times 35 \text{ mL})$. The combined organic layers were washed with brine, dried (Na2SO4), and concentrated to give a yellow solid. This material was purified by flash chromatography using a Biotage Flash 40 eluting with 15:1 hexane/ethyl acetate to give 610 mg (63%) of title compound 50 as a sticky yellow foam. ¹H NMR (CDCl₃): δ 0.68 (3H, t, J = 7.6), 1.07 (3H, s), 1.68–1.79 (1H, m), 1.87 (1H, dd, J = 3.5 and 13.9), 1.96 (1H, d, J = 15.0), 2.00-2.11 (1H, m), 2.65 (1H, t, J = 14.5), 3.01-3.15 (2H, m), 3.81 (3H, s), 5.62 (1H, d, J = 9.1), 6.19 (1H, s), 6.46 (1H, dd, J = 2.9 and 9.1), 6.91–6.96 (3H, m), 7.07 (1H, dd, J = 2.5 and 8.3), 7.25– 7.35 (4H, m), 7.49–7.58 (6H, m), 7.64 (1H, t, J = 7.5), 8.20 (2H, d, J = 7.1).

(2*R*, 3*R*, 4*aR*, 10*aS*)-4*a*-Ethyl-3-methyl-2-phenyl-1,2,3,4,4a,10a-hexahydrophenanthrene-2,3,7-triol (51).



To a solution of acetal 50 (708 mg, 1.24 mmol) in dichloromethane (40 mL) was added water (3 mL) and 2,3-dichloro-5,6-dicyano-1,4benzoquinone (589 mg, 2.58 mmol). The mixture was stirred for 3 h at room temperature and then diluted with saturated sodium bicarbonate solution (50 mL). The organic layer was separated and the aqueous layer extracted with dichloromethane (50 mL). The combined organic layers were washed with brine, dried (Na_2SO_4) , and concentrated to give 727 mg (99%) of a light brown solid. This material was dissolved in a solution of 10% KOH in 1:1 ethanol/water (30 mL), and the mixture was heated to 60 °C for 5 h. After cooling to room temperature, the mixture was diluted with saturated ammonium chloride solution (100 mL) and extracted with ethyl acetate (3×75) mL). The combined extracts were washed with saturated sodium bicarbonate solution (100 mL) and brine, dried (Na₂SO₄), and concentrated to give 450 mg (theory = 435 mg) of analogue 51 as a yellow solid, which contained a small amount of residual solvent. ¹H NMR (CD₃OD): δ 0.60 (3H, t, *J* = 7.3), 1.22 (3H, s), 1.55 (1H, dd, *J* = 2.6 and 12.4), 1.58–1.64 (1H, m), 2.15 (1H, d, J = 15.0), 2.22–2.30 (1H, m), 2.53-2.58 (1H, m), 2.63-2.70 (2H, m), 5.48 (1H, dd, J = 2.3 and 9.6), 6.30 (1H, dd, J = 3.1 and 9.3), 6.49 (1H, d, J = 3.1), 6.64 (1H, dd, J = 2.6 and 8.3), 7.01 (1H, d, J = 8.3), 7.19 (1H, t, J = 7.3), 7.25–7.29 (2H, m), 7.67 (2H, d, J = 7.8). LC-MS: m/z 349.4 [M – H^{-} , 395.4 $[M - H + H_2CO_2]^{-}$.

Receptor Binding Assays. Compound affinities for human GR and AR were assessed as previously described.²⁰ Human PR, ER α , and ER β were obtained from PanVera (Invitrogen). The ER α and ER β binding assays were run in a similar manner to that described for AR.²⁰ The PR assay was run according to the protocol supplied by PanVera (Invitrogen).

Whole Cell Functional Assay: IL-1 Stimulated MMP-13 Production. SW1353 human chondrosarcoma cells were plated at confluence into 96-well plates in Dulbecco's modified Eagle medium (DMEM) with 10% fetal bovine serum. After 24 h, medium was removed and replaced with 200 μ L/well serum-free DMEM containing 1 mg/L insulin, 2g/L lactalbumin hydrosylate, and 0.5 mg/L ascorbate. The medium was removed 16 h later and replaced with 150 μ L/well fresh serum-free medium containing ±20 ng/mL IL-1 β , ± 5 nM dexamethasone, ± test compound. After 24 h of incubation at 37 °C under 5% CO₂, 125 μ L sample from each well was removed under asceptic conditions for MMP-13 production analysis (Amersham Bio-Trak MMP-13 ELISA) using the manufacturer's protocol.²¹

Whole Cell Functional Assay: MMTV. Clonal SW1353 human chondrosarcoma cells stably transfected with pMAM-neo-luciferase mouse mammary tumor virus (MMTV) reporter construct were plated near confluence into 96-well plates in DMEM with 10% fetal bovine serum. After 24 h, medium was removed and replaced with phenol red-free, serum-free DMEM supplemented with 2 mM L-glutamine, 1 mg/L insulin, 0.5 mg/L ascorbate, and 2 g/L lactalbumin hydrosylate. Treatments were \pm dexamethasone or compound. After 16 h of incubation at 37 °C under 5% CO₂, 100 µL of LucLite reagent (Packard) was added to each well. After 5 min of incubation in the dark, luminescence was detected in a TopCount microplate scintillation counter.²¹

Whole Blood Functional Assay: IL-1 Stimulated TNF α Production. Heparinized blood was collected from medication-free human volunteers. The following 10× mixture was added to 96-well U-bottom plates: RPMI/200 mM 4-(2-hydroxyethyl)-1-piperazinee-thanesulfonic acid (HEPES; Invitrogen) containing drug or DMSO vehicle (1.0%) and LPS (1000 ng/mL, Sigma L-2637, *Escherichia coli* 0111:B4). Human whole blood was then added and gently mixed with the final concentration of DMSO being 0.1% and LPS 100 ng/mL.

The samples were incubated for 4 h at 37 °C in a humidified 5% CO₂ tissue culture incubator. Plasma was collected following centrifugation at 4 °C. Plasma was immediately diluted 1:5 in TNF α ELISA RD6 diluent (R&D Systems). TNF α levels were determined by ELISA (1:20 final dilution) after calibration to a standard curve as indicated by the manufacturer.

ASSOCIATED CONTENT

S Supporting Information

X-ray crystallographic parameters and ¹H NMR spectra. This material is available free of charge via the Internet at http:// pubs.acs.org.

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Author Contributions

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Notes

The authors declare no competing financial interest.

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DEDICATION

[†]Dedicated to Dr. M. Ross Johnson for his valued contributions to medicinal chemistry and mentorship to many colleagues.

ABBREVIATIONS

AP-1, activator protein-1; DAGR, dissociated agonist of the glucocorticoid receptor; DEX, dexamethasone; DKK1, dick-kopf-related protein 1; ER, estrogen receptor; GR, glucocorticoid receptor; RA, rheumatoid arthritis; GRE, glucocorticoid response element; HWB, human whole blood; IL-1, interleukin-1; MMFF94, Merck Molecular Force Field 94; MMP-13, matrix metalloproteinase 13; MMTV, mouse mammary tumor virus; MR, mineralocorticoid receptor; NF κ B, nuclear factor kappa B; PR, progesterone receptor; AR, androgen receptor; PRED, prednisone; TA, transactivation; TAT, tyrosine amino transferase; TNF α , tumor necrosis factor alpha; TR, transrepression

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