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The surface properties of amine oxides with a fluoroether chain

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ABSTRACT

Persistent organic pollutants (POPs) includes long-chained fluorosurfactants, for instance, perfluorooctanonate (PFOA) and perfluorooctanesulfonate (PFOS). In order to find out alternative fluorosurfactants, we synthesized eight amine oxides with a fluoroether chain. Their surface properties were evaluated and compared with perfluoroalkyl and hydrocarbon analogues. The surface tensions at critical micelle concentration ($\gamma_{\rm Cmc}$) for the eight amine oxides with a fluoroether chain were at a range from 15.5 to 23.0 mN/m, and $\gamma_{\rm Cmc}$ of four fluoroether amine oxides were below 17 mN/m comparable to perfluorooctyl analogues (16.4 mN/m) and much lower than perfluorohexyl (20.5 mN/m) and hydrocarbon analogues (24.2 and 24.7 mN/m). The critical micelle concentration (cmc) for the eight amine oxides with a fluoroether chain were 3 to 535×10^{-4} mol/L. The cmc of four fluoroether amine oxides were 3 to 21×10^{-4} mol/L (0.2 to 1.0 g/L) comparable to perfluorooctyl analogue (36.9 g/L) and hydrocarbon analogues (4 and 10×10^{-4} mol/L) and much lower than perfluorohexyl analogues (36.9 g/L). The surface excesses, the limiting molecule areas and the free energies of micellization of amine oxides were calculated. Fluoroether surfactants are promising alternatives for PFOA and PFOS.

1. Introduction

Amine oxides are amphoteric surfactants, showing nonionic characteristics in neutral or alkaline solutions, and cationic characteristics in acidic solutions. Amine oxide surfactants have good foaming properties, wettability, thickening, low toxic and biodegradability, and they cause less irritation to the skin [1]. Because of these properties, amine oxide surfactants are widely used in detergents, shampoos, cosmetics and textile auxiliaries. Amine oxide surfactants are also used as foam boosters and stabilizers. On the other hand, fluorinated surfactants usually have high thermal and chemical stability. Furthermore, fluorinated surfactants have higher surface activities at lower critical micelle concentrations (cmc) compared to the traditional surfactants. Therefore, fluorinated surfactants play a special role in many applications such as electroplating, fire-fighting foams and repellents [2].

However, according to the Stockholm Convention, long-chained perfluoroalkyl substances such as perfluorooctanonate (PFOA) and perfluorooctanesulfonate (PFOS) are classified as organic persistent pollutants (POPs) because they are toxic, persistent, bioaccumulative and able to transfer to long distance in environment [3]. Although shortening fluorinated segments were proposed to reduce toxicity, the cmc and the surface tension (γ) will increase with the shortening of the chain. Fluorosurfactants with high surface activities and environmentally friendly properties are in a high demand in order to replace the compounds with long-chained perfluoroalkyl groups. We have been focusing on the risk evaluation of several kinds of emerging fluorosurfactants including their occurrence in environment and livings and also their toxicities to aquatic organisms and mammals [4-8]. Assisted by the biological and environmental research, we designed gemini cationic surfactants with flexible perfluorinated-ether chains last year [9]. We wish that newly-designed structures with a fluoroether chain bring more understanding about structural effect on not only surface property but also bioaccumulative ability. While the alkyl amine oxide surfactants are well researched,-very few research on fluorinated amine oxides has been reported [10]. The first fluoroether amine oxide dates back to 50 years ago [10e]. Commercially available fluoroether

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Received 1 February 2021; Received in revised form 8 April 2021; Accepted 8 April 2021 Available online 10 April 2021 0022-1139/© 2021 Elsevier B.V. All rights reserved. surfactants, such as GenX and ADONA, are anionic surfactants using in fluoropolymer manufacture. Amerduri et al. have developed CH₂-containing alternatives based on PVDF [10f,g]. Comparative structure-property correlations for amine-oxide surfactants with fluoroether and fluoroalkyl tail structures are needed.

In this article, we report our recent study on perfluoroether amine oxides and their performance together with a comparison with perfluoroalkyl amine oxides and aliphatic amine oxides.

2. Result and discussion

We synthesized tertiary amines from fluorinated esters, carbonyl fluorides, carbonyl chlorides or acids. The tertiary amines were oxidized to provide amino oxides for surface properties studies. (Scheme 1)

After we obtained the amine oxides surfactants, the surface tensions in gradient concentration were measured. The surface tensions at cmc (γ_{cmc}), cmc, the surface excesses (Γ_{max}), the limiting molecule areas (A_{min}) and the free energies of micellization (ΔG^{o}) of amine oxides were calculated and depicted in Table 1.

The surface excess Γ_{max} (also written as Γ_{cmc}) and the limiting molecule area A_{min} (also written as A_{cmc}) were used to characterize the behavior of surfactant molecules at air-water interface. Γ_{max} is defined as the concentration of surfactant molecules in a surface plane relative to

that at a similar plane in the bulk at cmc. A_{min} corresponds to the area per surfactant molecule at air-water interface at cmc [2]. We calculated Γ_{max} and A_{min} by using the Gibbs equation

$$\Gamma_{\rm max} = -\frac{1}{2.303 \times nRT} \times \frac{d\gamma}{d {\rm log}c}$$

and the equation

$$A_{\min} = \frac{10^{14}}{N_A \Gamma_{\max}}$$

The "n" was taken as 1 for amine oxide surfactants because they were looked as a nondissociating molecule in a neutral aqueous solution.

The free energies of micellization (ΔG^{0}) were calculated based on the equation $\Delta G^{0} = RT lncmc$.

In the Table 1 and Fig. 1, the γ_{cmc} of compounds were in an order as C61-Oxide 21 > C62-Oxide 24 > C72-Oxide 22 > C73-Oxide 25 > C82-Oxide 23. The same order was found with cmc of these five surfactants. C61-Oxide 21 has been reported with a surface tension of 28.5 mN/m in 1% aqueous solution [10e], which is higher than our result (18.4 mN/m). The surface tensions and cmc were determined by hydrophobic fluorinated chains if hydrophilic groups were same. The number of fluorinated carbons was prominent on the reducing of γ and



Scheme 1. The synthesis of amine oxides with fluoroether, perfluoroalkyl and aliphatic groups.

Table 1

The surface properties of amine oxides.

Compound		γ_{cmc} (mN/m)	cmc		$\Gamma_{\rm max}$ (× 10 ⁻¹⁰ mol/cm ²)	A _{min} (nm ²)	Δ G ^o (kJ/mol)	Purity ^a	T (°C)
			$(\times 10^{-4} \text{ mol/L})$	(g/L)					
21	C61-Oxide	18.4	159	6.9	2.8	0.59	-10	98%	23
22	C72-Oxide	16.8	21	1.0	3.3	0.51	-15	96%	22
23	C82-Oxide	15.5	3	0.2	2.8	0.60	-20	92%	22
24	C62-Oxide	17.0	88	3.9	3.3	0.50	-12	96%	23
25	C73-Oxide	15.8	14	0.7	0.8	2.15	-16	98%	22
26	C6-Oxide	20.5	891	36.9	2.4	0.68	-6	96%	24
27	C8-Oxide	16.4	7	0.4	2.8	0.59	-18	96%	24
28	C72-O-Oxide-E	23.0	472	22.8	2.5	0.68	-8	94%	23
29	C72-O-Oxide	21.4	535	26.6	1.0	1.63	-7	98%	22
30	C72-Oxide-E	16.6	19	0.9	2.8	0.59	-15	99%	24
32	CH12-Oxide	24.2	4	0.1	3.4	0.50	-19	95%	22
33	C12-Oxide	24.7	10	0.2	2.4	0.69	-17	99%	22

^a The purity were analyzed by the NMR spectroscopy (see supplementary material)



Fig. 1. The γ -logC curves for fluoroether amine oxides.

cmc. Increasing oxygen number on the hydrophobic fluorinated segments reduced γ and cmc as well. Introducing oxygen atoms into fluorinated chains increased the hydrophobic properties of fluorosurfactants [11]. Γ_{max} and A_{min} were calculated by the slope of the γ -logC curves. The slope of **C73-Oxide 25** was smaller than others and its Γ_{max} was smallest and A_{min} was largest. This could be explained by the high flexibility of the oxygen-rich hydrophobic fluorinated chain within **C73-Oxide 25** molecule [9]. The Γ_{max} and A_{min} of the other four surfactants (**C61-Oxide 21, C62-Oxide 24, C72-Oxide 22** and **C82-Oxide 23**) were very close. The trend of ΔG° was based on the trend of cmc. The order of ΔG° was the same as that for cmc.

In the Table 1 and Fig. 2, we compared fluoroether C72-Oxide 22 with fluorinated amine oxide surfactants (C6-Oxide 26 and C8-Oxide 27) and aliphatic amine oxide surfactants (CH12-Oxide 32 and C12-Oxide 33). The surface properties of C72-Oxide 22 and C8-Oxide 27 were close. The γ_{cmc} for C72-Oxide 22 and C8-Oxide 27 were 16.8 and 16.4 mN/m and the cmc for C72-Oxide 22 and C8-Oxide 27 were 21 and 7×10^{-4} mol/L (1.0 and 0.4 g/L), respectively. Surface tension was determined majorly by the fluorinated segment. The perfluoroalkyl chain CF₃(CF₂)₆- behaved comparable to the fluoroether chain CF₃OCF (CF₃)CF₂OCF(CF₃)-. One fluorinated carbon worked roughly as two oxygen atoms within the fluoroether chain. Shortening two carbons in the perfluoroalkyl chain gave C6-Oxide 26, resulting a sharp increase of $\gamma_{\rm cmc}$ (20.5 mN/m) and cmc (36.9 g/L). Although the $\gamma_{\rm cmc}$ of CF₃(CF₂)₄amine oxide 26 was lower than those of the two hydrocarbon surfactants CH12-Oxide 32 and C12-Oxide 33, the cmc of aliphatic CH12-Oxide 32 and C12-Oxide 33 were closer to those of fluorinated C72-Oxide 22 and C8-Oxide 27 probably due to the similar hydrophilic-lipophilic balance (HLB) and lower than that of C6-Oxide 26. Increasing carbon chain length in compounds 36 CH18-oxide with a $C_{17}H_{35}$ chain resulted



Fig. 2. The γ -logC curves for fluoroether amine oxide 22 compared with perfluoroalkyl and aliphatic amine oxides.

in a extremely low solubility and failed to give a low surface tension. The γ of hydrocarbon **32** and **33** were higher than 24 mN/m, much higher than that of fluorinated surfactant. The low polarizability and weak dispersion force for perfluorinated carbon and ether chains with an enough length can reduce not only surface tension but also cmc [2,12]. The cmc relates to the efficiency and the surface tension relates to effectiveness [2]. Only fluorinated surfactants can satisfy both requirements to reach low surface tension and low cmc.

An illustration in Scheme 2 may help to understand the air-water interfacial behavior of the surfactants bearing with various fluorinated chains. The surface tensions were determined majorly by the length of perfluoroalkyl or perfluoroether chains. When we discuss the term "length" here, both backbond and branched chains were considered. As we discussed above, the effect of two ether bonds on surface tension will be vaguely equal to that of a difluoromethylene group. So, the surface tensions were consistent with the apparent lengths when the surfactant molecules were packing tightly in the air-water interfaces.

In Scheme 2, the flexibility of fluorinated chains resulting from the ether bonds were illustrated. As we all know, silicone surfactants enhance their surface activities by the multiple flexible ether bonds [2b]. Fluoroether chains can enhance the surface activities by the same mechanism. The most extraordinary example is **C73-Oxide 25** with the limiting molecular area of 2.15 nm², much larger than others. We contributed the wide A_{min} of **C73-Oxide 25** to the three flexible ether bonds. While flexibility has an essential influence on A_{min} , the bulk of fluorinated groups change the area covered by surfactant molecules when they are saturated and packing tightly in the interface. The A_{min} of **C62-Oxide 24**, **C72-Oxide 22**, **C8-Oxide 27** and **C82-Oxide 23** were



Scheme 2. An illustration to explain the effect of perfluoroalkyl and fluoroether chain on the surface tension and the limiting molecular area.

reasonable according to this explanation. The large limiting molecular areas of **C61-Oxide 21** and **C6-Oxide 26** may be attributed to the lower surface excess Γ_{max} for higher solubilities of short-chained fluorinated surfactants in water.

In the Table 1 and Fig. 3, we examined linkage effect by comparing fluoroether C72-Oxide 22 with ester linkage amine oxides C72-O-Oxide 29 and C72-O-Oxide-E 28 and shorter linkage amine oxides (C72-Oxide-E 30). Ester linkages increased cmc and γ_{cmc} dramatically. By shortening propylene (-CH2CH2CH2-) to ethylene (-CH2CH2-), slightly change of C72-Oxide-E 30 in γ_{cmc} (16.6 mN/m) and cmc (0.9 g/L) were found. Due to the lone-pair electron resonance of nitrogen atom and carbonyl group, amide bonds are normally rigid compared with ester bonds [12]. The surfactants with amide bonds are less flexible and consequently have lower entropies compared with the surfactants with ester bonds, and tend to be pushed out of the bulk aqueous environment to the air-water interface. Thus, the surface excess of C72-Oxide 22 was larger than that of C72-O-Oxide 29, and the limiting molecular area of C72-Oxide 22 was smaller than that of C72-O-Oxide 29 (Table 1). We also observed higher solubility of C72-O-Oxide 29 and C72-O-Oxide-E 28 than that of C72-Oxide 22 and C72-Oxide-E 30. The high surface tension and cmc of ester surfactants 28 and 29 could also be attributed to their solubilities of monomers inside the bulk aqueous solution.



Fig. 3. The γ -logC curves for amine oxides with different alkyl chain lengths and different linkage groups.

3. Conclusion

We have synthesized thirteen surfactants including fluoroether, perfluoroalkyl and aliphatic amine oxides. Their surface properties were examined and compared. The γ_{cmc} and cmc of fluoroether surfactants can reach as low as 15.5 mN/m and 0.2 g/L. By adjusting oxygen and carbon number of fluoroether chains, we can provide surfactants with properties comparable to PFOA-based amino oxides. Oxygen atoms within fluoroether chain, which can reduce surface tension and cmc, are hydrophobic. Fluorinated surfactants are irreplaceable for their properties of both low surface tension and cmc. Modification of surfactants by fluoroether chains provide a alternative strategy for fluorinated POPs.

4. Experimental section

4.1. General information

Fluorinated starting materials were provided by Sanming Hexafluo Chemicals Co., LTD. (see SI), and all other reagents were of AR grade quality and used without further purification. Chemical shifts are expressed in parts per million (ppm) with respect to the residual solvent peak. Chemical shifts for ¹⁹F NMR are reported in ppm downfield from fluorotrichloromethane (CFCl₃). ¹³C NMR was broad-band decoupled from hydrogen nuclei. Coupling constants are reported as hertz (Hz). Signal shapes and splitting patterns are indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. The resonances corresponding the N-H protons are not included in the analysis – possibly due to exchange broadening. HRMS (ESI) data were tested on a Water Micromass GCT Premier. Surface tensions were obtained with KRÜSS(DLSB-5L/-20) surface tension meter by Wilhelmy platinum plate method. Specific test information is recorded in supplementary material.

4.2. Experimental procedures and characterization data for compounds

N-(3-(dimethylamino)propyl)-2,3,3,3-tetrafluoro-2-(perfluoropropoxy)propanamide (**11**) **C61-Amine** [9]

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL). Then, compound $CF_3CF_2CF_2OCF(CF_3)COOCH_3$ (1, 20 mmol, 6.88 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary

evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was a light-yellow liquid (8.04 g, 97%) [9]. ¹H NMR (400 MHz, CD₃OD) δ 3.37 – 3.27 (m, 2H), 2.33 (t, *J* = 7.6 Hz, 2H), 2.20 (s, 6H), 1.75 – 1.67 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -81.9 (dm, *J* = 145.9 Hz, 1F), -82.9 (t, *J* = 7.1 Hz, 3F), -84.6 (d, *J* = 1.9 Hz, 3F), -86.2 (dm, *J* = 149.3 Hz 1F), -131.2 (s, 2F), -134.0 (dd, *J* = 19.6, 6.8 Hz, 1F). IR (film) ν/cm^{-1} : 3342.1, 2954.7, 2869.8, 2830.6, 2790.3, 1715.9, 1533.1, 1470.5, 1343.1, 1322.2, 1296.4, 1235.4, 1200.1, 1163.3, 1133.1, 1107.8, 1062.5, 1041.6, 991.0, 809.1, 747.3, 719.8, 628.6, 534.8.

N-(3-(dimethylamino)propyl)-2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-

hexafluoro-2-(trifluoromethoxy)propoxy)propanamide (12) C72-Amine

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL). Then, compound CF₃OCF(CF₃)CF₂OCF(CF₃)COOCH₃ (2, 20 mmol, 8.20 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was a light-yellow liquid (8.84 g, 92%). ¹H NMR (400 MHz, CD₃OD) δ 3.38 – 3.30 (m, 2H), 2.34 (t, *J* = 7.6 Hz, 2H), 2.22 (s, 6H), 1.77 – 1.69 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -55.0 (m, 3F), -80.8 (m, 1F), -81.5 (m, 3F), -84.1 (dd, J = 6.4, 1.5 Hz, 3F), -84.5 (m, 1F), -133.9 (m, 1F), -147.8 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 157.8 (m), 56.6, 43.9, 38.1, 25.9, carbons corresponding to the CF₃OCF(CF3)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{12}H_{14}F_{13}N_2O_3~[M+H]^+$: 481.0791 found: 481.0793. IR (film) ν/cm^{-1} : 3342.6, 2955.0, 2869.7, 2830.6, 2790.8, 1724.6, 1533.7, 1471.0, 1239.0, 1161.4, 1106.3, 1077.6, 1062.6, 1041.8, 982.1, 892.7, 809.2, 765.6, 739.1, 683.9, 643.8, 534.3.

N-(3-(dimethylamino)propyl)-2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-

hexafluoro-2-(perfluoroethoxy)propoxy)propanamide (13) C82-Amine To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL). Then, compound CF₃CF₂OCF(CF₃)CF₂OCF(CF₃)COOCH₃ (3, 20 mmol, 9.20 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a light-yellow liquid (9.76 g, 92%). ¹H NMR (400 MHz, CD₃OD) δ 3.38 – 3.32 (m, 2H), 2.34 (t, J = 7.4 Hz, 2H), 2.22 (s, 6H), 1.77 – 1.69 (m, 2H). $^{19}{\rm F}$ NMR (376 MHz, CD₃OD) δ -81.3 (m, 3F), -84.2 (m, 3F), -80.7 – -84.7 (m, 2F), -87.0 (m, 2F), -88.3 (d, J = 15.8 Hz, 3F), -133.8 (dd, J = 19.6, 7.1 Hz, 1F), -146.4 (t, J = 22.0 Hz, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 157.8 (d, J = 26 Hz), 56.6, 43.9, 38.1, 25.9, carbons corresponding to the CF₃CF₂OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{13}H_{14}F_{15}N_2O_3 \ \mbox{[M} \ + \ \mbox{H}\mbox{]}^+$: 531.0759, found: 531.0754. IR (film) v/cm⁻¹: 3342.8, 2955.5, 2870.6, 2831.5, 2790.8, 1716.5, 1533.7, 1470.9, 1305.5, 1235.2, 1153.5, 1125.3, 1090.3, 1041.7, 981.8, 916.4, 825.6, 805.8, 765.2, 743.5, 720.6, 691.2, 646.0, 523.5.

N-(3-(dimethylamino)propyl)-2,3,3,3-tetrafluoro-2-(1,1,2,2-tetrafluoro-2-(trifluoromethoxy)ethoxy)propanamide (14) C62-Amine [9]

To a 100 mL three-necked round-bottom flask containing a magnetic

stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL). Then, compound CF₃OCF₂CF₂OCF(CF₃)COOCH₃ (4, 20 mmol, 7.20 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a light-yellow liquid (8.26 g, 96%).^[9] This compound was obtained as a light-yellow liquid. ¹H NMR (400 MHz, CD₃OD) δ 3.34 – 3.30 (m, 2H), 2.33 (t, J = 7.6 Hz, 2H), 2.21 (s, 6H), 1.75 – 1.68 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -57.0 (t, J = 9.0 Hz, 3F), -84.1 (d, J = 1.9 Hz, 3F), -86.2 (dd, *J* = 146.6, 18.4 Hz, 1F), -90.3 (dd, *J* = 146.6, 8.3 Hz, 1F), -91.9 (m, 2F), -134.1 (dd, J = 18.1, 7.9 Hz, 1F). IR (film) ν/cm^{-1} : 3342.5, 2954.7, 2869.9, 2830.6, 2790.3, 1723.5, 1533.6, 1470.4, 1285.2, 1224.0, 1149.8, 1114.1, 1061.8, 1041.5, 988.2, 902.2, 795.8, 765.5, 720.7, 681.3, 616.5.

2-(2-(difluoro(trifluoromethoxy)methoxy)-1,1,2,2-tetrafluoroethoxy)-N-(3-(dimethylamino)propyl)-2,3,3,3-tetrafluoropropanamide (**15**) **C73-Amine** [9]

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and tert-butyl methyl ether (10 mL). Then, compound CF₃OCF₂OCF₂CF₂OCF(CF₃)COF (5, 20 mmol, 8.28 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 4 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a light-yellow liquid (9.71 g, 94%) [9]. ¹H NMR (400 MHz, CD₃OD) δ 3.40 -3.32 (m, 2H), 2.42 (t, J = 7.6 Hz, 2H), 2.28 (s, 6H), 1.82 - 1.75 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -55.3 (m, 2F), -58.7 (t, J = 8.8 Hz, 3F), -84.1 (d, J = 1.5 Hz, 3F), -86.7 (m, 1F), -90.1 (m, 1F), -91.7 (q, J = 21.1, 10.5 Hz, 2F) -134.0 (dd, J = 18.4, 7.9 Hz, 1F). IR (film) ν/cm^{-1} : 3336.3, 2956.5, 2871.2, 2832.1, 1714.9, 1534.2, 1471.0, 1390.5, 1309.5, 1219.9, 1108.1, 1040.7, 989.3, 964.1, 869.4, 794.7, 765.3, 720.6, 647.1.

N-(3-(dimethylamino)propyl)-2,2,3,3,4,4,5,5,6,6,6-undeca-fluorohexanamide (16) C6-Amine [13]

3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL) was added into a 100 mL three necked round-bottom flask. The mixture was stirred evenly at room temperature. Then, compound CF₃CF₂CF₂CF₂CF₂COOH (**6**, 20 mmol, 6.28 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. The solvent was removed by rotary evaporation under vacuum. The final product was obtained as a lightyellow liquid (7.65 g, 96%). ¹H NMR (400 MHz, CD₃OD) δ 2.92 (t, *J* = 6.8 Hz, 2H), 2.44 (t, *J* = 7.2 Hz, 2H), 2.24 (s, 6H), 1.79 – 1.72 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -82.5 (m, 3F), -118.0 (m, 2F), -123.8 (m, 4F), -127.5 (m, 2F). IR (film) ν/cm^{-1} : 3429.6, 2959.5, 2835.0, 1685.8, 1471.6, 1394.6, 1350.1, 1237.8, 1204.7, 1146.6, 1106.8, 1083.0, 867.9, 827.5, 807.9, 747.6, 732.5, 713.8, 655.7, 566.3, 530.4.

N-(3-(dimethylamino)propyl)-2,2,3,3,4,4,5,5,6,6,7,7,8,8,8-pentadecafluorooctanamide (**17**) **C8-Amine** [14]

3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL) was added into 100 ml three necked round-bottom flask. Stir the mixture evenly at room temperature. Then, compound $CF_3(CF_2)_6COOH$ (7, 20 mmol, 8.28 g, 1.0 eq.) was slowly dropped in and the mixture were stirred at room temperature for 24 h. The solvent was removed by rotary evaporation under vacuum. The final product was obtained as a white solid (8.99 g, 94%). ¹H NMR (400 MHz, CD₃OD) δ 2.90 (t, *J* = 7.2 Hz, 2H), 2.42 (t, *J* = 7.2 Hz, 2H), 2.22 (s, 6H), 1.74 (m, 2H). $^{19}{\rm F}$ NMR (376 MHz, CD₃OD) δ -82.5 (m, 3F), -118.0 (m, 2F), -122.7 (m, 2F), -123.1 (m, 2F), -123.6 (m, 2F), -123.9 (m, 2F), -127.4 (m, 2F). IR (film) $\nu/{\rm cm}^{-1}$: 2960.6, 2836.1, 1686.1, 1472.9, 1394.2, 1360.4, 1240.9, 1206.3, 1149.0, 1102.1, 1013.1, 812.9, 746.6, 721.9, 700.0, 663.8, 641.5, 560.8, 530.4.

2-(dimethylamino)ethyl2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexa-

fluoro-2-(trifluoromethoxy)propoxy)propanoate (**18**) **C72-O-Amine-E** To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 2-(dimethylamino)ethan-1-ol (80 mmol, 7.13 g, 4 eq.), methanol (10 mL). Then, compound CF₃OCF(CF₃)CF₂OCF(CF₃)

COF (8, 20 mmol, 7.96 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Ethyl acetate (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (8.04 g, 86%). ¹H NMR (400 MHz, CD_3OD) δ 3.67 (t, J = 5.6 Hz, 2H), 2.57 (t, J = 6.0 Hz, 2H), 2.34 (s, 6H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.8 (m, 3F), -80.4 – -84.9 (m, 2F), -83.9 (m, 3F), -126.5 (m, 1F), -147.8 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 162.4 (m), 61.9, 59.6, 45.4, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-EI (m/z) calcd for C₁₁H₁₀F₁₃NO₄ [M – H]⁻: 467.0397, found: 467.0393. IR (film) v/cm⁻¹: 3675.9, 3412.4, 2839.4, 1697.4, 1464.5, 1403.7, 1355.8, 1232.3, 1157.9, 1040.3, 980.7, 892.7, 820.1, 771.4, 738.9, 682.9, 653.0, 619.6, 537.3.

2-(dimethylamino)propyl2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propoxy)propanoate (**19**) **C72-O-Amine**

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-(dimethylamino)propan-1-ol (100 mmol, 10.3 g, 4 eq.), methanol (10 mL). Then, compound CF₃OCF(CF₃)CF₂OCF(CF₃) COF (8, 25 mmol, 9.95 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Ethyl acetate (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (10.11 g, 84%). ¹H NMR (400 MHz, CD₃OD) δ 3.57 (t, *J* = 6.4 Hz, 2H), 2.45 (t, *J* = 7.6 Hz, 2H), 2.26 (s, 6H), 1.74 – 1.65 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.9 (m, 3F), -81.6 - -84.2 (m, 2F), -83.9 (m, 3F), -126.6 (m, 1F), -147.9 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 162.3 (d, J = 26 Hz), 61.4, 57.8, 45.3, 30.8, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)group cannot be identified due to C-F coupling. HRMS-EI (m/z) calcd for $C_{12}H_{12}F_{13}NO_4 [M + H]^+$: 481.0553, found: 481.0551. IR (film) ν/cm^{-1} : 1695.4, 1667.6, 1411.6, 1232.5, 1159.1, 1042.2, 981.2, 892.9, 821.8, 771.5, 739.3, 683.4, 654.7, 539.2.

N-(2-(dimethylamino)ethyl)-2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propoxy)propanamide (20) C72-Amine-E

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added N,N-dimethylethane-1,2-diamine (20 mmol, 1.76 g, 1 eq.), triethylamine (30 mmol, 3.04 g, 1.5 eq.) and methanol (10 mL). Then, compound CF₃OCF(CF₃)CF₂OCF(CF₃)COOCH₃ (**2**, 20 mmol, 8.20 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product obtained as a light-yellow liquid (8.77 g, 94%). ¹H NMR (400 MHz, CD₃OD) δ 3.42 (t, *J* = 6.4 Hz, 2H), 2.47 (t, *J* = 7.6 Hz, 2H), 2.26 (s, 6H). ¹⁹F NMR (376 MHz, CD₃OD) δ -55.0 (m, 3F), -81.0 (m, 3F), -80.4 – -84.9 (m, 2F), -84.3 (m, 3F), -134.0 (m, 1F), -147.8 (m, 1F). ¹³C NMR (100 MHz, CD₃OD); δ

159.5 (d, J = 26 Hz), 58.3, 45.4, 38.8, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for C₁₁H₁₀F₁₃N₂O₃ [M – H]⁻: 465.0489, found: 465.0488. IR (film) ν/cm^{-1} : 3350.1, 2955.1, 2869.2, 2830.1, 2781.8, 1716.0, 1520.0, 1463.6, 1356.2, 1235.5, 1162.2, 1107.5, 1077.4, 1054.8, 981.9, 893.0, 846.7, 808.9, 773.3, 739.1, 684.1, 619.6, 536.0.

N,N-dimethyl-3-(2,3,3,3-tetrafluoro-2-(perfluoropropoxy)propanamido)propan-1-amine oxide (**21**) **C61-Oxide** [10e]

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF3CF2CF2OCF(CF3)C(O)NHCH2CH2CH2N(CH3)2 (11, 5 mmol, 2.07 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 12 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.09 g, 97%). ¹H NMR (400 MHz, CD₃OD) δ 3.42 (t, J = 6.8 Hz,2H), 3.32 (m, 2H), 3.16 (s, 6H), 2.15 – 2.07 (m, 2H). ¹⁹F NMR (376 MHz, CD_3OD) δ -81.8 (dm, J = 149.7 Hz, 1F), -82.9 (t, J = 7.5 Hz, 3F), -84.0 (d, J = 1.9 Hz, 3F), -86.2 (dm, J = 149.3 Hz 1F), -131.1 (m, 2F), -134.0 (dd, J = 19.2, 6.4 Hz, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 159.5 (d, J = 27Hz), 69.2, 58.6 (d, J = 3 Hz), 38.8, 24.2, carbons corresponding to the CF₃CF₂CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{11}H_{14}F_{11}N_2O_3$ [M + H]⁺: 431.0823, found: 431.0813. IR (film) v/cm⁻¹: 3261.9, 2810.1, 1709.2, 1544.3, 1477.0, 1451.4, 1323.0, 1234.7, 1165.6, 1134.9, 1072.0, 991.6, 927.8, 886.1, 809.2, 747.9, 721.0, 630.3, 536.6.

N,N-dimethyl-3-(2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propoxy)propanamido)propan-1-amine oxide (22) C72-Oxide

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF3OCF(CF3)CF2OCF(CF3)C(O)NHCH2CH2CH2N(CH3)2 (12, 5 mmol, 2.40 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.43 g, 98%). ¹H NMR (400 MHz, CD₃OD) δ 3.43 (t, J = 6.8Hz,2H), 3.31 (m, 2H), 3.15 (s, 6H), 2.14 – 2.06 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.0 (m, 3F), -80.3 - -84.9 (m, 2F), -84.4 (m, 3F), -133.9 (m, 1F), -147.8 (m, 1F). ^{13}C NMR (100 MHz, CD_3OD): δ 159.5 (d, J = 26 Hz), 69.2, 58.7 (d, J = 4 Hz), 38.8, 24.2, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{12}H_{14}F_{13}N_2O_4$ [M + H]⁺: 497.0741, found: 497.0736. IR (film) $\nu/$ cm⁻¹: 3226.1, 2804.3, 1709.6, 1541.4, 1455.9, 1161.7, 1070.9, 1026.3, 981.8, 892.8, 809.2, 773.4, 739.0, 684.0, 644.5, 535.5.

N,N-dimethyl-3-(2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(perfluoroethoxy)propoxy)propanamido)propan-1-amine oxide (23) C82-Oxide

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF₃CF₂OCF(CF₃)CF₂OCF(CF₃)C(O)NHCH₂CH₂CH₂N(CH₃)₂ (13, 5 mmol, 2.65 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 12 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.68 g, 98%). ¹H NMR (400 MHz, CD₃OD) δ 3.39 (t, J = 7.2 Hz, 2H), 3.30 (m, 2H), 3.14 (s, 6H), 2.13 – 2.04 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -81.5 (m, 3F), -80.7 – -84.8 (m, 2F), -84.1 (m, 3F), -87.0 (m, 2F), -88.2 (d, J = 13.9 Hz, 3F), -133.8 (dd, J = 19.9, 7.1 Hz, 1F), -146.4 (t, J = 21.8 Hz, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 159.4 (d, J = 26 Hz), 69.2, 58.6 (d, J = 3 Hz), 38.7, 24.2, carbons corresponding to the CF₃CF₂OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{13}H_{14}F_{15}N_2O_4~[M~+~H]^+\!\!:$ 547.0709, found: 547.0693. IR (film) v/cm⁻¹: 3250.5, 2806.1, 1708.4, 1541.1, 1476.8, 1455.1, 1307.1, 1234.9, 1153.6, 1090.3, 1070.0, 1026.2, 981.7, 927.9, 885.5, 825.7, 805.9, 765.3, 743.6, 721.4, 691.5, 645.0, 521.9.

N,N-dimethyl-3-(2,3,3,3-tetrafluoro-2-(1,1,2,2-tetrafluoro-2-(tri-

fluoromethoxy)
ethoxy)propanamido)propan-1-amine oxide ${\bf (24)}$ C62-Oxide

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF₃OCF₂CF₂OCF(CF₃)C(O)NHCH₂CH₂CH₂N(CH₃)₂ (14, 5 mmol, 2.15 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 12 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.14 g, 96%). ¹H NMR (400 MHz, CD₃OD) δ 3.41 (t, J = 7.2 Hz, 2H), 3.31 (m, 2H), 3.16 (s, 6H), 2.14 – 2.07 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -57.0 (t, J = 9.6 Hz, 3F), -84.0 (d, J = 2.3 Hz, 3F), -86.3 (dd, J = 146.6, 18.4 Hz, 1F), -90.2 (dd, J = 146.6, 8.3 Hz, 1F), -91.9 (m, 2F), -134.0 (dd, J = 18.4, 7.9 Hz, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 159.6 (d, J = 27 Hz), 69.2, 58.7 (d, J = 3 Hz), 38.8, 24.0, carbons corresponding to the CF₃OCF₂CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{11}H_{14}F_{11}N_2O_4$ [M + H]⁺: 447.0772, found: 447.0761. IR (film) ν/cm^{-1} : 3254.2, 2810.0, 1709.0, 1541.3, 1452.0, 1398.6, 1223.4, 1150.1, 1068.4, 1026.8, 984.6, 967.7, 903.0, 795.8, 721.9, 681.3, 616.9.

1,1,1,3,3,5,5,6,6,8-decafluoro-N,N-dimethyl-9-oxo-8-(tri-

fluoromethyl)-2,4,7-trioxa-10-azatridecan-13-amine oxide (25) C73-Oxide

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF₃OCF₂OCF₂CF₂OCF(CF₃)C(O)NHCH₂CH₂CH₂N(CH₃)₂ (15, 5 mmol, 2.48 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 $^\circ\text{C}$ for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.48 g, 97%). ¹H NMR (400 MHz, CD₃OD) δ 3.44 – 3.34 (m, 4H), 3.21 (s, 6H), 2.14 - 2.04 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -55.2 (m, 2F), -58.7 (t, J = 9.0 Hz, 3F), -84.1 (d, J = 1.9 Hz, 3F), -86.6 (dd, J = 146.3, 18.4 Hz, 1F), -90.2 (dd, J = 146.3, 7.9 Hz, 1F), -91.7 (m. 2F), -134.0 (dd, J = 18.8, 8.7 Hz, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 159.7 (d, J = 27 Hz), 68.9, 58.1 (d, J = 3 Hz), 38.6, 24.1, carbons corresponding to the CF₃OCF₂OCF₂CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{12}H_{14}F_{13}N_2O_5~[M~+~H]^+\!\!:$ 513.0690, found: 513.0677. IR (film) $\nu/$ cm⁻¹: 3265.4, 2812.7, 1708.2, 1541.2, 1478.5, 1450.9, 1392.3, 1312.0, 1216.0, 1111.3, 1035.5, 964.5, 870.3, 794.5, 721.0, 646.8.

N,N-dimethyl-3-(2,2,3,3,4,4,5,5,6,6,6-undecafluorohexanamido) propan-1-amine oxide (26) C6-Oxide [13]

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF3CF2CF2CF2CF2C(O)NHCH2CH2CH2N(CH3)2 (16, 5 mmol, 1.99 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 $^{\circ}$ C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final compound product was obtained as a white viscous liquid (1.97 g, 95%). ¹H NMR (400 MHz, CD₃OD) δ 3.51 (t, J = 6.4 Hz, 2H), 3.19 (s, 6H), 2.98 (t, *J* = 6.4 Hz, 2H), 2.18 – 2.09 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -82.5 (m, 3F), -118.1 (m, 2F), -123.8 (m, 4F), -127.5 (m, 2F). ¹³C NMR (100 MHz, CD₃OD): δ 162.9 (t, J = 24 Hz), 70.1, 58.8, 39.0, 23.6, carbons corresponding to the CF₃CF₂CF₂CF₂CF₂group cannot be identified due to C-F coupling. IR (film) ν/cm^{-1} : 3058.3, n 1686.2, 1479.1, 1458.8, 1395.4, 1350.5, 1235.6, 1204.6, 1149.5, 1106.9, 1084.1, 1061.8, 994.7, 953.5, 868.5, 827.4, 808.7, 748.0, 732.7, 714.4, 656.0, 566.4, 531.3.

N,N-dimethyl-3-(2,2,3,3,4,4,5,5,6,6,7,7,8,8,8-pentadeca-

fluorooctanamido)propan-1-amine oxide (27) C8-Oxide [14]

To a 100 mL round-bottom flask containing a magnetic stirring bar

was added CF₃CF₃CF₃CF₂CF₂CF₂CF₂CC₂C(O)NHCH₂CH₂CH₂CH₂N(CH₃)₂ (17, 5 mmol, 2.49 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (2.42 g, 97%). ¹H NMR (400 MHz, CD₃OD) δ 3.48 (t, *J* = 6.8 Hz, 2H), 3.17 (s, 6H), 2.92 (t, *J* = 6.8 Hz, 2H), 2.08 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -82.5 (m, 3F), -118.1 (m, 2F), -122.8 (m, 2F), -123.2 (m, 2F), -123.7 (m, 2F), -123.9 (m, 2F), -127.5 (m, 2F). IR (film) *v*/cm⁻¹: 3034.9, 1659.9, 1456.4, 1397.9, 1363.0, 1323.1, 1238.3, 1204.2, 1146.2, 1104.5, 1017.9, 815.9, 736.3, 722.7, 666.1, 642.0, 560.5, 530.5.

N,N-dimethyl-2-((2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propaxy)propanoyl)oxy)ethan-1-amine oxide (28) C72-O-Oxide-E

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF₃OCF(CF₃)CF₂OCF(CF₃)C(O)OCH₂CH₂N(CH₃)₂ (18, 5 mmol, 2.34 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (2.32 g, 96%). ¹H NMR (400 MHz, CD₃OD) δ 4.01 (m, 2H), 3.47 (m, 2H), 3.26 (s, 6H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.5 (m, 3F), -81.6 - -84.1 (m, 2F), -83.7 (m, 3F), -126.5 (m, 1F), -147.8 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 162.4 (m), 72.6, 59.2, 57.5, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-EI (m/z) calcd for C11H10F13NO5 $[M]^+$: 483.0346, found: 483.0348. IR (film) ν/cm^{-1} : 3427.5, 2820.1, 1689.7, 1401.2, 1235.3, 1158.5, 1092.6, 1040.1, 981.7, 893.2, 820.5, 771.4, 739.4.

N,N-dimethyl-3-((2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propaxy)propanoyl)oxy)propan-1-amine oxide (29) C72-O-Oxide

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF3OCF(CF3)CF2OCF(CF3)C(O)OCH2CH2CH2N(CH3)2 (19, 5 mmol, 2.41 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (2.44 g, 98%). ¹H NMR (400 MHz, CD₃OD) δ 3.62 (t, J = 6.0 Hz, 2H), 3.41 (t, J = 7.6 Hz, 2H), 3.17 (s, 6H), 2.08 – 1.98 (m, 2H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.5 (m, 3F), -81.6 - -84.2 (m, 2F), -83.8 (m, 3F), -126.6 (m, 1F), -147.9 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 162.3 (m), 70.0, 60.4, 58.5, 28.0, carbons corresponding to the CF₃OCF(CF₃)CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. IR (film) v/cm⁻¹: 3237.6, 2807.7, 1694.3, 1405.6, 1232.5, 1157.6, 1092.1, 1039.6, 980.8, 892.5, 820.0, 770.9, 738.8, 682.9, 653.5, 537.3.

N,N-dimethyl-2-(2,3,3,3-tetrafluoro-2-(1,1,2,3,3,3-hexafluoro-2-(trifluoromethoxy)propoxy)propanamido)ethan-1-amine oxide (30) C72-Oxide-E

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CF₃OCF(CF₃)CF₂OCF(CF₃)C(O)NHCH₂CH₂N(CH₃)₂ (**20**, 5 mmol, 2.33 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 18 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous liquid (2.41 g, 98%). ¹H NMR (400 MHz, CD₃OD) δ 3.79 (t, *J* = 6.4 Hz, 2H), 3.44 (t, *J* = 8.0 Hz, 2H), 3.17 (s, 6H). ¹⁹F NMR (376 MHz, CD₃OD) δ -54.9 (m, 3F), -81.5 (m, 3F), -84.0 – -84.9 (m, 2F), -84.4 (m, 3F), -134.0 (m, 1F), -147.8 (m, 1F). ¹³C NMR (100 MHz, CD₃OD): δ 159.5 (m), 68.1, 59.0 (d, *J* = 4 Hz), 36.1, carbons corresponding to the CF₃OCF(CF₃)

CF₂OCF(CF₃)- group cannot be identified due to C-F coupling. HRMS-ESI (m/z) calcd for $C_{11}H_{10}F_{13}N_2O_4$ [M – H]⁻: 481.0438, found: 481.0438. IR (film) ν /cm⁻¹: 3208.5, 2799.1, 1715.5, 1541.4, 1478.1, 1456.5, 1235.3, 1161.9, 1073.4, 1043.9, 982.1, 893.1, 809.1, 773.6, 739.5, 684.1, 620.8, 535.0, 469.9.

N-(3-(dimethylamino)propyl)dodecanamide (31) CH12-Amine [1a] To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar was added 3-dimethylaminopropylamine (20 mmol, 2.04 g, 1 eq.), chloroform (10 mL). Then, compound C₁₁H₂₃COCl (9, 20 mmol, 4.38 g, 1.0 eq.) was slowly dropped in and the mixture was stirred at room temperature for 24 h. After the reaction the solvent was removed by rotary evaporation under vacuum. Dichloromethane (50 mL) was added to the residue. The resulting mixture was washed with saturated brine and water. The organic phase was separated and then dried over anhydrous sodium sulfate. After filtration, the solvent was removed by rotary evaporation under vacuum. The final product was obtained as a white solid (5.12 g, 90%). ¹H NMR (400 MHz, CD₃OD) δ 3.18 (t, J = 6.8Hz, 2H), 2.36 (t, J = 7.6 Hz, 2H), 2.25 (s, 6H), 2.16 (t, J = 7.2 Hz, 2H), 1.75 - 1.53 (m, 4H), 1.36 - 1.21 (m, 18H), 0.89 (t, J = 6.4 Hz, 3H). IR (film) v/cm⁻¹: 3294.1, 2925.0, 2854.2, 2815.3, 2764.3, 1645.5, 1556.4, 1464.8, 1383.9, 1263.3, 1154.9, 1099.6, 1042.0, 721.7.

3-dodecanamido-N,N-dimethylpropan-1-amine oxide (32) CH12-Oxide [1a]

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CH₃(CH₂)₁₀C(O)NHCH₂CH₂CH₂N(CH₃)₂ (**31**, 5 mmol, 1.42 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 24 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (1.46 g, 97%). ¹H NMR (400 MHz, CD₃OD) δ 3.35 – 3.23 (m, 4H), 3.15 (s, 6H), 2.18 (t, *J* = 7.6 Hz, 2H), 2.08 – 1.98 (m, 2H), 1.64 – 1.54 (m, 2H), 1.36 – 1.21 (m, 18H), 0.89 (t, *J* = 6.4 Hz, 3H). IR (film) ν /cm⁻¹: 3331.2, 3103.9, 2915.3, 2848.2, 1643.7, 1545.9, 1472.6, 1462.5, 1425.1, 1403.3, 1375.1, 1360.4, 1328.4, 1306.9, 1273.5, 1250.3, 1232.8, 1224.3, 1210.3, 1121.0, 1066.1, 1027.6, 1009.5, 976.6, 935.5, 864.6, 786.2, 770.5, 754.2, 729.2, 719.6, 689.7, 615.0, 561.6, 514.0, 489.8, 427.8.

N,N-dimethyldodecan-1-amine oxide (33) C12-Oxide [1b]

To a 100 mL round-bottom flask containing a magnetic stirring bar was added $CH_3(CH_2)_{11}N(CH_3)_2$ (**10**, 5 mmol, 1.07 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (15 mmol, 1.70 g, 3.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 24 h. The organic extract was concentrated by rotary evaporation under vacuum. The final product was obtained as a white viscous solid (1.10 g, 96%). ¹H NMR (400 MHz, CD₃OD) δ 3.23 (t, *J* = 8.4 Hz, 2H), 3.11 (s, 6H), 1.85 – 1.75 (m, 2H), 1.40 – 1.20 (m, 18H), 0.86 (t, *J* = 6.8 Hz, 3H). IR (film) ν/cm^{-1} : 3372.9, 2921.8, 2852.3, 1652.2, 1467.3, 1378.0, 965.7, 924.8, 773.8, 721.0.

N-(3-(dimethylamino)propyl)stearamide (35) CH18-Amine [15]

To a 100 mL three-necked round-bottom flask containing a magnetic stirring bar and N₂ were added C₁₇H₃₅COOH (20 mmol, 6.04 g, 1.0 eq.), and toluene (25 mL). Then, compound 3-dimethylaminopropylamine (60 mmol, 6.12 g, 3 eq.) was slowly dropped into the mixture at 75 °C, and the reaction was heated to 130 °C for 24 h. After removing the solvent by rotary evaporation under vacuum. The residue was recrystallized over acetone for twice. The final product was obtained as a white solid (5.24 g, 71%). ¹H NMR (400 MHz, CD₃OD) δ 3.18 (t, *J* = 6.8 Hz, 2H), 2.33 (t, *J* = 8 Hz, 2H), 2.26 (s, 6H), 1.72 – 1.54 (t, *J* = 7.6 Hz, 4H), 1.38 – 1.21 (m, 24H), 0.89 (t, *J* = 6.4 Hz, 3H). IR (film) *v*/cm⁻¹: 3310.4, 2917.6, 2849.3, 2814.2, 2763.0, 1640.0, 1557.4, 1472.1, 1395.0, 1376.5, 1258.7, 1241.4, 1186.4, 1155.0, 1042.4, 942.3, 729.6, 718.9.

N,N-dimethyl-3-stearamidopropan-1-amine oxide (36) CH18-Oxide [15].

To a 100 mL round-bottom flask containing a magnetic stirring bar was added CH₃(CH₂)₁₆C(O)NHCH₂CH₂CH₂N(CH₃)₂ (**35**, 5 mmol, 1.84 g, 1 eq.), and ethanol (50 mL). The reaction mixture was stirred evenly and heated to 50 °C. Then, hydrogen peroxide (20 mmol, 2.28 g, 4.0 eq.) was slowly dropped in and the mixture was stirred at 50 °C for 12 h. The organic extract was concentrated by rotary evaporation under vacuum. The final compound product was obtained as a white solid (1.87 g, 97%). ¹H NMR (400 MHz, CD₃OD) δ 3.31 – 3.21 (m, 4H), 3.15 (s, 6H), 2.16 (t, *J* = 8.0 Hz, 2H), 2.05 – 1.98 (m, 2H), 1.64 – 1.53 (m, 2H), 1.38-1.16 (m, 28H), 0.88 (t, *J* = 6.8 Hz, 3H). IR (film) ν/cm^{-1} : 3325.6, 2916.7, 2849.0, 1643.7, 1550.8, 1467.5, 1378.6, 1274.1, 1258.8, 1242.3, 1223.7, 1208.8, 1123.1, 1066.0, 1010.3, 936.3, 869.4, 720.1, 616.7, 488.3, 427.1.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi

Declaration of Competing Interest

The authors report no declarations of interest.

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Supplementary materials

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References

 (a) R. Wang, Y. Li, Q. Li, Synthesis and properties of dodecyldiethoxylamine oxide, J. Surfact. Deterg. 16 (2013) 509–514, https://doi.org/10.1007/s11743-013-1460-6;

(b) M.J. Rosen, D. Friedman, M. Gross, A surface tension study of the interaction of dimethyldodecylamine oxide with potassium dodecanesulfonate in dilute aqueous solution, J. Phys. Chem. 68 (1964) 3219–3225, https://doi.org/10.1021/ j100793a023;

(c) A. Fabozzi, R. Vitiello, I.R. Krauss, M. Iuliano, G.D. Tommaso, A. Amoresano, G. Pinto, L. Paduano, C. Jones, M.D. Serio, Synthesis, surface properties, and self-aggregation behavior of a branched N,N-dimethylalkylamine oxide surfactant, J. Surfact. Deterg. 22 (2019) 115–124, https://doi.org/10.1002/jsde.12205.

- [2] (a)E. Kissa, Fluorinated Surfactants and Repellents, Marcel Dekker, Inc., New York, 2001. (b) A. Czajka, G. Hazell, J. Eastoe, Surfactants at the design limit, Langmuir 31 (2015) 8205–8217, https://doi.org/10.1021/acs.langmuir.5b00336.
- [3] (a) Z. Wang, J.C. DeWitt, C.P. Higgins, I.T. Cousins, A never-ending story of perand polyfluoroalkyl substances (PFASs)? Environ. Sci. Technol. 51 (2017) 2508–2518, https://doi.org/10.1021/acs.est.6b04806.(b)The information about new POPs including PFOA and PFOS under the Stockholm Convention is provided at the web site: http://www.pops.int/TheConvention/ThePOPs/TheNewPOPs/ tabid/2511/Default.aspx. (c)P.C. Bed. Technick and the function of the stockholm Convention and the story of the

(c)R.C. Buck, Toxicology data for alternative "short-chain" fluorinated substances. In: J.C. DeWitt (eds) Toxicological effects of perfluoroalkyl and polyfluoroalkyl substances. Molecular and integrative toxicology. Humana Press, Cham, 2015, http s://www.toxicology.org/groups/ss/RASS/docs/20151209_SOT_Webinar_Chemou rs Presentation RCB final.pdf.

- [4] J. Chen, H. Li, J. Yao, H. Guo, H. Zhang, Y. Guo, N. Sheng, J. Wang, J. Dai, Chronic exposure to PFO4DA and PFO5DoDA, two perfluoroalkyl ether carboxylic acids (PFECAs), suppresses hepatic stress signals and disturbs glucose and lipid metabolism in male mice, J. Hazard. Mater. 411 (2021), 124963, https://doi.org/ 10.1016/j.jhazmat.2020.124963.
- [5] (a) N. Sheng, J. Wang, Y. Guo, J. Wang, J. Dai, Interactions of perfluorooctanesulfonate and 6:2 chlorinated polyfluorinated ether sulfonate with human serum albumin: a comparative study, Chem. Res. Toxicol. 33 (2020) 1478–1486, https://doi.org/10.1021/acs.chemrestox.0c00075;
 (b) J. Wang, G. Shi, J. Yao, N. Sheng, R. Cui, Z. Su, Y. Guo, J. Dai, Perf luoropolyether carboxylic acids (novel alternatives to PFOA) impair zebrafish posterior swim bladder development via thyroid hormone disruption, Environ. Int. 134 (2020), 105317, https://doi.org/10.1016/j.envint.2019.105317;
 (c) J. Yao, Y. Pan, N. Sheng, Z. Su, Y. Guo, J. Dai, Novel perfluoroalkyl ether carboxylic acids (PFECAs) and sulfonic acids (PFESAs): occurrence and association with serum biochemical parameters in residents living near a fluorochemical plant in China, Environ. Sci. Technol. 54 (2020) 13389–13398, https://doi.org/10.1021/acs.est.0c02888.

[6] (a) H. Guo, J. Wang, J. Yao, S. Sun, N. Sheng, X. Zhang, X. Guo, Y. Guo, Y. Sun, J. Dai, Comparative hepatotoxicity of novel PFOA alternatives (perfluoropolyether carboxylic acids) on male mice, Environ. Sci. Technol. 53 (2019) 3929–3937, https://doi.org/10.1021/acs.est.9b00148;

(b) G. Shi, Q. Cui, H. Zhang, R. Cui, Y. Guo, J. Dai, Accumulation, biotransformation, and endocrine disruption effects of fluorotelomer surfactant mixtures on zebrafish, Chem. Res. Toxicol. 32 (2019) 1432–1440, https://doi.org/10.1021/acs.chemrestox.9b00127;

(c) G. Shi, Q. Cui, J. Wang, H. Guo, Y. Pan, N. Sheng, Y. Guo, J. Dai, Chronic exposure to 6:2 chlorinated polyfluorinated ether sulfonate acid (F-53B) induced hepatotoxic effects in adult zebrafish and disrupted the PPAR signaling pathway in their offspring, Environ. Pollut. 249 (2019) 550–559, https://doi.org/10.1016/j. envpol.2019.03.032;

(d) G. Shi, J. Wang, H. Guo, N. Sheng, Q. Cui, Y. Pan, Y. Guo, Y. Sun, J. Dai, Parental exposure to 6:2 chlorinated polyfluorinated ether sulfonate (F-53B) induced transgenerational thyroid hormone disruption in zebrafish, Sci. Total Environ. 665 (2019) 855–863, https://doi.org/10.1016/j.scitotenv.2019.02.198.

[7] (a) N. Sheng, R. Cui, J. Wang, Y. Guo, J. Wang, J. Dai, Cytotoxicity of novel fluorinated alternatives to long-Chain perfluoroalkyl substances to human liver cell line and their binding capacity to human liver fatty acid binding protein, Arch. Toxicol. 92 (2018) 359–369, https://doi.org/10.1007/s00204-017-2055-1;
(b) Q. Cui, Y. Pan, H. Zhang, N. Sheng, J. Wang, Y. Guo, J. Dai, Occurrence and tissue distribution of novel perfluoroether carboxylic and sulfonic acids and legacy per/polyfluoroalkyl substances in black-spotted frog (*Pelophylax nigromaculatus*), Environ. Sci. Technol. 52 (2018) 982–990, https://doi.org/10.1021/acs.est 7M03662*

(c) N. Sheng, Y. Pan, Y. Guo, Y. Sun, J. Dai, Hepatotoxic effects of hexafluoropropylene oxide trimer acid (HFPO-TA), a novel perfluorooctanoic acid (PFOA) alternative, on mice, Environ. Sci. Technol. 52 (2018) 8005–8015, https://doi.org/10.1021/acs.est.8b01714;

(d) G. Shi, H. Guo, N. Sheng, Q. Cui, Y. Pan, J. Wang, Y. Guo, J. Dai, Twogenerational reproductive toxicity assessment of 6:2 chlorinated polyfluorinated ether sulfonate (F-53B, a novel alternative to perfluorooctane sulfonate) in zebrafish, Environ. Pollut. 243 (2018) 1517–1527, https://doi.org/10.1016/j. envpol.2018.09.120;

(e) Y. Pan, H. Zhang, Q. Cui, N. Sheng, L.Y.W. Yeung, Y. Sun, Y. Guo, J. Dai, Worldwide distribution of novel perfluoroether carboxylic and sulfonic acids in surface water, Environ. Sci. Technol. 52 (2018) 7621–7629, https://doi.org/ 10.1021/acs.est.8b00829;

(f) H. Zhang, X. Zhou, N. Sheng, R. Cui, Q. Cui, H. Guo, Y. Guo, Y. Sun, J. Dai, Subchronic hepatotoxicity effects of 6:2 chlorinated polyfluorinated ether sulfonate (6:2 Cl-PFESA), a novel perfluorooctanesulfonate (PFOS) alternative, on adult male mice, Environ. Sci. Technol. 52 (2018) 12809–12818, https://doi.org/10.1021/ acs.est.8b04368;

(g) G. Shi, Y. Xie, Y. Guo, J. Dai, 6:2 Fluorotelomer sulfonamide alkylbetaine (6:2 FTAB), a novel perfluorooctane sulfonate alternative, induced developmental toxicity in zebrafish embryos, Aquat. Toxicol. 195 (2018) 24–32, https://doi.org/10.1016/j.aquatox.2017.12.002.

[8] (a) G. Shi, Q. Cui, Y. Pan, N. Sheng, Y. Guo, J. Dai, 6:2 Fluorotelomer carboxylic acid (6:2 FTCA) exposure induces developmental toxicity and inhibits the formation of erythrocytes during zebrafish embryogenesis, Aquat. Toxicol. 190 (2017) 53–61, https://doi.org/10.1016/j.aquatox.2017.06.023;
(b) N. Sheng, X. Zhou, F. Zheng, Y. Pan, X. Guo, Y. Guo, Y. Sun, J. Dai, Comparative hepatotoxicity of 6:2 fluorotelomer carboxylic acid and 6:2

fluorotelomer sulfonic acid, two fluorinated alternatives to long-chain perfluoroalkyl acids, on adult male mice, Arch. Toxicol. 91 (2017) 2909–2919, https://doi.org/10.1007/s00204-016-1917-2;

(c) Y. Pan, H. Zhang, Q. Cui, N. Sheng, L.Y.W. Yeung, Y. Guo, Y. Sun, J. Dai, First report on the occurrence and bioaccumulation of hexafluoropropylene oxide trimer acid: an emerging concern, Environ. Sci. Technol. 51 (2017) 9553–9560, https://doi.org/10.1021/acs.est.7b02259;

(d) Y. Pan, Y. Zhu, T. Zheng, Q. Cui, S.L. Buka, B. Zhang, Y. Guo, W. Xia, L.W. Y. Yeung, Y. Li, A. Zhou, L. Qiu, H. Liu, M. Jiang, C. Wu, S. Xu, J. Dai, Novel chlorinated polyfluorinated ether sulfonates and legacy per-/polyfluoroalkyl substances: placental transfer and relationship with serum albumin and glomerular filtration rate, Environ. Sci. Technol. 51 (2017) 634–644, https://doi.org/10.1021/ acs.est.6b04590;

(e) G. Shi, Q. Cui, Y. Pan, N. Sheng, S. Sun, Y. Guo, J. Dai, 6:2 Chlorinated polyfluorinated ether sulfonate, a PFOS alternative, induces embryotoxicity and disrupts cardiac development in zebrafish embryos, Aquat. Toxicol. 185 (2017) 67–75, https://doi.org/10.1016/j.aquatox.2017.02.002.

- [9] F. Du, Y. Guo, M. Huang, Q. Chen, H. Yang, W. Xie, W. Cao, C. Wu, M. Wang, Gemini cationic surfactants with flexible perfluorinated-ether chains, J. Fluorine Chem. 239 (2020), 109632, https://doi.org/10.1016/j.jfluchem.2020.109632.
- 10] (a) D. Zhang, M. Sha, R. Pan, X. Lin, P. Xing, B. Jiang, Synthesis and properties study of novel fluorinated surfactants with perfluorinated branched ether chain, J. Fluorine Chem. 219 (2019) 62–69, https://doi.org/10.1016/j. ifluchem 2018 11 001.

(b) C. Lin, R. Pan, P. Xing, B. Jiang, Synthesis and combined properties of novel fluorinated cationic surfactants derived from hexafluoropropylene dimer, Chin. Chem. Lett. 29 (2018) 1613–1616, https://doi.org/10.1016/j.cclet.2018.04.017; (c) T. Coope, K. Moloy, A. Yake, V. Petrov, C. Taylor, M. Hung, S. Peng, Fluorinated sulfamido amphoteric surfactants, J. Fluorine Chem. 161 (2014) 41–50, https://doi.org/10.1016/j.jfluchem.2014.01.022;

(d) Y. Chaudier, F. Zito, P. Barthélémy, D. Stroebel, B. Améduri, J.L. Popot,
B. Pucci, Synthesis and preliminary biochemical assessment of ethyl-terminated perfluoroalkylamine oxide surfactants, Bioorg. and Med. Chem. Lett. 12 (2002) 1587–1590, https://doi.org/10.1016/S0960-894X(02)00242-1;
(e) P.L. Bartlett, Perfluoroalkyl ether amidoamine oxides, US3547995A, 1970. (f)
H. Hori, H. Tanaka, T. Tsuge, R. Honma, S. Banerjee, B. Ameduri, Decomposition of fluoroelastomer: poly(vinylidene fluoride-ter-hexafluoropropylene-ter-tetrafluoroethylene) terpolymer in subcritical water, Eur. Polym. J. 94 (2017) 322–331, https://doi.org/10.1016/j.eurpolymj.2017.05.042;
(g) H. Hori, H. Tanaka, K. Watanabe, T. Tsuge, T. Sakamoto, A. Manseri,

(g) H. Holl, H. Hallaka, K. Watalabe, T. Isuge, T. Sakalnob, A. Malsell,
B. Ameduri, Hydrogen peroxide induced efficient mineralization of poly(vinylidene fluoride) and related copolymers in subcritical water, Ind. Eng. Chem. Res. 54 (2015) 8650–8658, https://doi.org/10.1021/acs.iecr.5b01716;
(h) Z. Wang, I.T. Cousins, M. Scheringer, K. Hungerbühler, Fluorinated alternatives to long-chain perfifluoroalkyl carboxylic acids (PFCAs),

perflfluoroalkane sulfonic acids (PFSAs) and their potential precursors, Environ. Int. 60 (2013) 242–248, https://doi.org/10.1016/j.envint.2013.08.021.

- [11] X. Song, R. Vestergren, Y. Shi, J. Huang, J. Huang, Emissions, transport, and fate of emerging per- and polyfluoroalkyl substances from one of the major fluoropolymer manufacturing facilities in China, Environ. Sci. Technol. 52 (2018) 9694–9703, https://doi.org/10.1021/acs.est.7b06657.
- [12] D. O'Hagan, Understanding organofluorine chemistry. An introduction to the C-F bond, Chem. Soc. Rev. 37 (2008) 308–319, https://doi.org/10.1039/B711844A.
- [13] S. Yuan, S. Huang, L. Dong, Z. Liu, Y. Lv, Y. Ren, C. Zhao, Z. Liu, Z. Xu, Z. Wang, C. Liu, X. Cong, K. Dong, P. Wang, Y. Yu, J. He, Q. Cui, J. Wang, Y. Li, Method for preparing high-carbon branched-chain secondary alcohol, CN110240538A, 2019.
- [14] A.K. Price, A.N. Fenster, Novel amidoamine oxides, US3828085A, 1974.
- [15] V.E. Limanov, A.E. Épshtein, M.I. Yakushkin, M.Yu. Telegin, V.I. Kotov, T. A. Fedorova, E.K. Skvortsova, Synthesis and bactericidal activity of some quaternary N-acylaminopropylammonium salts, Pharm. Chem. J. 15 (1981) 85–87.