

Available online at www.sciencedirect.com



Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 15 (2005) 3580-3583

Total synthesis and antiangiogenic activity of cyclopentane analogues of fumagillol

Byeong-Seon Jeong,^{a,†} Nam Song Choi,^a Soon Kil Ahn,^{a,*} Hoon Bae,^{b,‡} Hak Sung Kim^c and Deukjoon Kim^b

^aNew Drug Research Laboratories, Chong Kun Dang Research Institute, Cheonan 330-831, Republic of Korea ^bCollege of Pharmacy, Seoul National University, Seoul 151-742, Republic of Korea ^cCollege of Pharmacy, Wonkwang University, Iksan 570-749, Republic of Korea

Received 14 April 2005; revised 12 May 2005; accepted 14 May 2005

Abstract—Novel cyclopentane analogues of fumagillol were synthesized and their endothelial cell proliferation inhibitory activities were evaluated. The cyclopentane-fumagillol derivatives were synthesized from (-)-2,3-O-isopropylidene-D-erythronolactone via stereoselective glycolate Claisen rearrangement and intramolecular ester enolate alkylation as key steps. © 2005 Elsevier Ltd. All rights reserved.

Angiogenesis is the phenomenon of generating a new capillary vessel, which is one of the normal physiological functions and one of the pathological functions caused by various diseases as well. It has deep connection with growth and transfer of solid cancer, rheumatic arthritis, diabetic retinopathy, psoriasis, etc.¹ Since the Folkman research group suggested a novel concept of treating solid cancer by inhibiting angiogenesis in 1971,² clinical importance of therapeutic agents controlling angiogenesis has been emphasized, and much endeavor on angiogenesis seems to have been bestowed on it.

Fumagillin (1), a natural product isolated from Aspergillus fumigatus,³ was found by the Folkman group in 1990⁴ to strongly inhibit endothelial cell proliferation. Many semi-synthetic analogues have been synthesized from fumagillol (2), the hydrolysis product of fumagillin.⁴ Among these analogues, TNP-470 (3)^{4a} and CKD-732 (4)^{4c,d} are currently employed in clinical trials for the treatment of a variety of cancers (Fig. 1). The mechanism of action of these agents, including fumagillin, is proposed to inhibit angiogenesis by selective inhibition



Figure 1.

of methionine aminopeptidase type 2 (MetAP-2) through covalent bonding to the His231 residue of MetAP-2 by opening the spiro-epoxide.⁵

We have reported the first asymmetric total synthesis of fumagillol, featuring the chelation-controlled glycolate Claisen rearrangement and the intramolecular ester enolate alkylation as the key steps.^{6,7} As an ongoing project to expand structural diversity, the cyclopentane ring shown in **5** was introduced, instead of the naturally occurring cyclohexane skeleton of fumagillol (Fig. 1). We anticipated that this approach could lead to novel angiogenesis inhibitors since cyclopentane analogue **5** is not available from fermentation and it possesses a high level of homology to the fumagillol structure. It has a spiro-epoxide and an isoamyl side chain, which are proposed to be essential for activity and for

Keywords: Fumagillol; Angiogenesis.

^{*} Corresponding author. Tel.: +82 41 529 3107; fax: +82 41 558 3004; e-mail: skahn@ckdpharm.com

[†]Present address: Department of Chemistry, Vanderbilt University, Nashville, TN 37235, USA.

[‡]Present address: Department of Chemistry and Biochemistry, Florida State University, Tallahassee, FL 32306, USA.

hydrophobic interaction with MetAP-2, respectively. Moreover, the stereochemistry of the substituents in the cyclopentane ring is identical to that of fumagillol.

Our synthesis, which is described in Scheme 1,⁸ began with commercially available (-)-2,3-O-isopropylidene-D-erythronolactone 6, which was converted to the corresponding lactol with DIBAL-H, followed by the Wittig reaction with (carbomethoxyethylidene)triphenyl-phosphorane to afford the α,β -unsaturated ester 7 in 87% yield for the two steps. After protection of the alcohol with the TBDPS group, the methyl ester was transformed to the aldehyde 8 by the sequence of reduction with DIBAL-H and MnO₂ oxidation (83% overall yield for the three steps). Addition of isoamylmagnesium bromide to the aldehyde 8 produced a 3:1 mixture of allylic alcohols 9 and 9' where each compound could be isolated by silica column chromatography in 95% total yield. The absolute configuration of the newly generated stereocenter in 9 and 9' was confirmed by spectroscopic analysis of the compounds 13 and 13' (vide infra). The allylic glycolate ester 10 could be prepared using either DCC coupling from 9 or Mitsunobu reaction from 9'in good yield. The γ , δ -unsaturated glycolate 11 was efficiently obtained as a single diastereomer by 1,3-chirality transfer through a chelation-controlled chair-like transition state of the Burke-Fujisawa-Kallmerten modificathe Ireland–Claisen rearrangement.^{9,10} of tion Intramolecular ester enolate alkylation of the tosylate 12, which was converted from 11 by desilylation and tosylation, was performed with KHMDS at 0 °C in a short time to produce the desired cyclopentane-carboxvlate 13 in 66% yield with a 10:1 mixture of inseparable diastereomers. The excellent diastereomeric excess implies that this cyclization may proceed through an allylic strain-controlled 'H-eclipsed' transition state.¹⁰

The absolute configuration of 13 was verified by chemical transformation and NMR analysis. An observation of the formation of the lactones (15 or 15') in high yield in the case of treatment of the diol 14 with DBU is reliable evidence that expected stereo-chemistry of the C(a) position in 13 is correct. For the stereochemistry at the C(b) position, ¹H NMR studies were conducted.



Scheme 1. Reagents and conditions: (a) DIBAL-H, CH_2Cl_2 , $-78 \degree C$, 3 h, 92%; (b) $Ph_3P=C(Me)CO_2Me$, toluene, reflux, 1 h, 95%; (c) TBDPSCl, imidazole, DMF, rt, 3 h, 95%; (d) DIBAL-H, CH_2Cl_2 , $-78 \degree C$, 4 h, 96%; (e) MnO_2 , CH_2Cl_2 , rt, 20 h, 91%; (f) isoamylMgBr, THF, $-78 \degree C$, 1 h, 70% of 9 and 25% of 9'; (g) BnOCH₂CO₂H, DCC, DMAP, CH_2Cl_2 , rt, 3 h, 77%; (g') BnOCH₂CO₂H, PPh₃, DIAD, THF, 0 °C, 30 min then rt, 3 h, 82%; (h) (i) LiHMDS, TMSCI/Et₃N, THF, $-78 \degree C$, 1 h then rt, 10 h, (ii) CH_2N_2 , Et_2O , 0 °C, 5 min, 70% (for 2 steps); (i) TBAF, THF, rt, 12 h, 80%; (j) TsCl, pyridine, CHCl₃, 4 °C, 20 h, 98%; (k) KHMDS, THF, 0 °C, 10 min, 66% (10:1 mixture of inseparable diastereomers); (l) PPTS, MeOH, reflux, 20 h, 98%; (m) DBU, CH₃CN, reflux, 6 h, 96% (15:15' = 1:1); (m') NaOMe, MeOH, rt, 6 h, 95%; (n) Ag₂O, MeI, reflux, 4 h, 80%; (o) Li, *liq*. NH₃, $-78 \degree C$, 3 h, 62%; (p) TsCl, Et₃N, DMAP, CH₂Cl₂, rt, 18 h, 80%; (q) K₂CO₃, MeOH, rt, 2 h, 90%; (r) *m*-CPBA, NaHCO₃, CH₂Cl₂, 0 °C, 4 h, 94% (ca. 3:1 of inseparable diastereomeric mixture); (s) chloroacetyl isocyanate, Et₃N, DMAP, CH₂Cl₂, 0 °C, 71% (18% of **20** and 53% of **20**').

Upon irradiation of the H(b) in 13, nuclear Overhauser effect (NOE) was observed to be 1.4% for the H(c), while NOE in the diastereomer 13' prepared from 9' was 5.6%. These results nicely support the fact that H(b) and H(c) in 13 have a trans-relationship (Fig. 2).

Although several methods for the selective methylation of C(c)-alcohol in 14, which was prepared from 13 by treatment with PPTS in methanol, with/without protection of the other hydroxyl group were attempted, we failed to obtain the desired mono-methoxy product in good yield. Lactonization turned out to be the best protection to give a 1:1 mixture of separable regioisomers (15 and 15'), with the undesired isomer 15' being recycled. Then, methylation of the hydroxyl group in 15 was successfully performed with Ag₂O/MeI reaction system to give 16 in 80% yield. Removal of the benzylprotecting group in 16 along with the concurrent reduction of the lactone was accomplished using excess lithium in liquid ammonia to produce the triol 17 in 62% yield. Selective mono-tosylation of the primary hydroxyl group in 17, followed by treatment with mild base, afforded the spiro-epoxy compound 18 in 72%yield for the two steps. Epoxidation of the olefin in 18 with m-CPBA produced a 3:1 mixture of inseparable diastereomers 19 in 94% yield.

To measure the endothelial cell proliferation inhibitory activity of the novel cyclopentane analogue of fumagillol, chloroacetylcarbamoyl group, a side chain of TNP-470 (3), was introduced, and each diastereomer (20 and 20') was isolated by silica column chromatography. Anti-proliferating activities were evaluated against calf pulmonary artery endothelial (CPAE) cells, lymphoma EL-4 cells, and murine leukemia P388. IC₅₀



Figure 2.

 Table 1. In vitro endothelial cell proliferation inhibitory activities

 against CPAE, EL-4, and P-388 cell lines

Compound	IC ₅₀ (µg/mL)		
	CPAE	EL-4	P-388
20	8.8×10^{-2}	3.0×10^{-1}	≥10
20'	5.7×10^{-1}	1.1	≥10
Fumagillin (1)	3.0×10^{-3}	1.6×10^{-3}	≥10
TNP-470 (3)	2.0×10^{-4}	3.8×10^{-3}	≥10

values were colorimetrically measured by SRB (CPAE cell) or MTT (EL-4, P388) methods. The biological data are shown in Table 1. The prepared novel cyclopentane analogues of fumagillol showed endothelial cell proliferation inhibitory activity at μ M concentration. However, their activities turned out to be lower than those of fumagillin derivatives.

In summary, total synthesis of a novel cyclopentane analogue of fumagillol was carried out using glycolate Claisen rearrangement and intramolecular ester enolate alkylation as key steps. Studies on the structure-activity relationship of fumagillol analogues are underway.

Acknowledgments

This work was supported by Grants (01-PJ1-PG4-01PT01-0004 and 00-PJ2-PG-1-CD02-0018) from the Ministry of Health & Welfare, Republic of Korea.

References and notes

- 1. (a) Folkman, J. J. Natl. Cancer Inst. **1990**, 82, 4; (b) Hanahan, D.; Folkman, J. Cell **1996**, 86, 353.
- 2. Folkman, J. N. Engl. J. Med. 1971, 285, 1182.
- 3. Hanson, F. R.; Eble, E. J. Bacteriol. 1949, 58, 527.
- (a) Ingber, D. E.; Fujita, T.; Kishmoto, S.; Sudo, K.; Kanamaru, T.; Brem, H.; Folkman, J. *Nature* 1990, 345, 555; (b) Marui, S.; Itoh, F.; Kozai, Y.; Sudo, K.; Kishimoto, S. *Chem. Pharm. Bull* 1992, 40, 96; (c) Hong, C. I.; Kim, J. W.; Lee, S. J.; Ahn, S. K.; Choi, N. S.; Hong, R. K.; Chun, H. S.; Moon, S. K.; Han, C. K. W.O. Patent 9959986, 1999; (d) Han, C. K.; Ahn, S. K.; Choi, N. S.; Hong, R. K.; Moon, S. K.; Chun, H. S.; Lee, S. J.; Kim, J. W.; Hong, C. I.; Kim, D.; Yoon, J. H.; No, K. T. *Bioorg. Med. Chem. Lett.* 2000, 10, 39; (e) Fardis, M. Pyun, H.-J.; Tario, J.; Jin, H.; Kim, C. U.; Ruckman, J.; Lin, Y.; Green, L.; Hicke, B. *Bioorg. Med. Chem.* 2003, 11, 5051; (f) Pyun, H.-J.; Fardis, M.; Tario, J.; Yang, C. Y.; Ruckman, J.; Henninger, D.; Jin, H.; Kim, C. U. *Bioorg. Med. Chem. Lett.* 2004, 14, 91.
- (a) Sin, N.; Meng, L.; Wang, M. Q. W.; Wen, J. J.; Bornmann, W. G.; Crews, C. M. *Proc. Natl. Acad. Sci.* USA 1997, 94, 6099; (b) Liu, S.; Widom, J.; Kemp, C. W.; Crews, C. M.; Clardy, J. Science 1998, 282, 1324.
- (a) Kim, D.; Ahn, S. K.; Bae, H.; Choi, W. J.; Kim, H. S. *Tetrahedron Lett.* **1997**, *38*, 4437; (b) Kim, D.; Ahn, S. K.; Bae, H.; Kim, H. S. *Arch. Pharm. Res.* **2005**, *28*, 129.
- For total syntheses or approaches to fumagillin and analogues by other groups see: (a) Corey, E. J.; Snider, B. B. J. Am. Chem. Soc. 1972, 94, 2549; (b) Taber, D. F.; Christos, T. E.; Rheingold, A. L.; Guzei, I. A. J. Am. Chem. Soc. 1999, 121, 5589; (c) Vosburg, D. A.; Weiler, S.; Sorensen, E. J. Angew Chem. Int. Ed. 1999, 38, 971; (d) Picoul, W.; Urchegui, R.; Haudrechy, A.; Langlois, Y. Tetrahedron Lett. 1999, 40, 4797; (e) Moffat, D.; Simpkins, N. S. Synlett 2001, 661; (f) Boiteau, J.-G.; Van de Weghe, P.; Eustache, J. Org. Lett. 2001, 3, 2737; (g) Vosburg, D. A.; Weiler, S.; Sorensen, E. J. Chirality 2003, 15, 156; (h) Picoul, W.; Bedel, O.; Haudrechy, A.; Langlois, Y. Pure Appl. Chem. 2003, 75, 235; (i) Bedel, O.; Haudrechy, A.; Langlois, Y. Eur. J. Org. Chem. 2004, 3813.
- 8. All new compounds exhibited satisfactory spectroscopic and analytical data, and the ratio of stereoisomers was

determined by the ¹H NMR analysis and/or isolation of individual isomers. Selected data for the key intermediates 11 and 13: For 11 ¹H NMR (400 MHz, CHCl₃-d) δ 7.24-7.76 (m, 15H), 5.14 (t, 1H, J = 6.5 Hz), 4.48 (d, 1H, J = 11.3 Hz), 4.33 (d, 1H, J = 11.3 Hz), 4.21–4.26 (m, 2H), 4.00 (d, 1H, J = 10.2 Hz), 3.90 (dd, 1H, J = 10.8, 5.4 Hz), 3.82 (dd, 1H, J = 10.8, 5.4 Hz), 3.61 (s, 3H), 2.95 (dd, 1H, J = 0.2, 9.8 Hz), 1.88–1.92 (m, 2H), 1.58 (s, 3H), 1.42–1.51 (m, 1H), 1.37 (s, 3H), 1.29 (s, 3H), 1.08–1.11 (m, 2H), 1.06 (ii, 11), 1.57 (3, 511), 1.25 (3, 511), 1.06 1.11 (iii, 211), 1.06 (s, 9H), 0.85 (d, 6H, J = 6.6 Hz); ¹³C NMR (100 MHz, CHCl₃-d) δ 172.1, 137.3, 136.2, 136.1, 134.4, 134.2, 131.4, 130.7, 129.7, 128.8, 128.5, 128.3, 127.9, 127.8, 125.9, 107.2, 81.5, 79.9, 73.2, 64.6, 51.8, 50.8, 41.4, 38.9, 27.9, 27.8, 27.4, 25.9, 25.7, 22.9, 22.8, 19.7, 13.5; EI-MS (m/z) 673 (M⁺+1). For **13** ¹H NMR (400 MHz, CHCl₃-*d*) δ 7.26–7.38 (m, 5H), 5.28 (t, 1H, J = 7.2 Hz), 4.85 (d, 1H, J = 7.1 Hz), 4.64–4.69 (m, 1H), 4.57 (d, 1H, J = 11.5 Hz), 4.34 (d, 1H, J = 11.5 Hz), 3.74 (s, 3H), 2.98 (d, 1H, J = 7.1 Hz), 2.64 (dd, 1H, J = 14.3, 5.3 Hz), 2.42 (dd, 1H, J = 14.3, 5.3 Hz), 2.02-2.04 (m, 2H), 1.72 (s, 3H), 1.47-1.54 (m, 1H), 1.32 (s, 3H), 1.26 (s, 3H), 1.19–1.21 (m, 2H), 0.87 (d, 6H, J = 6.6 Hz); ¹³C NMR (100 MHz, CHCl₃-d) δ 172.3, 138.9, 131.1, 128.6, 127.9, 127.7, 113.7, 92.5, 82.5, 78.7, 68.1, 63.3, 60.8,

52.4, 39.1, 38.3, 28.0, 27.7, 26.3, 25.3, 23.0, 22.8, 21.4, 15.8, 14.6; EI-MS (*m*/*z*) 415 (M⁺-1).

- (a) Burke, S. D.; Fobare, W. F.; Pacofsky, G. J. J. Org. Chem. 1983, 48, 5221; (b) Sato, T.; Tajima, K.; Fujisawa, T. Tetrahedron Lett. 1983, 24, 729; (c) Kallmerten, J.; Gould, T. J. Tetrahedron Lett. 1983, 24, 5177.
- 10. Plausible transition states for the two transformations $(10 \rightarrow 11 \text{ and } 12 \rightarrow 13).$

