Bioorganic & Medicinal Chemistry Letters 20 (2010) 6978-6982

Contents lists available at ScienceDirect



Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Biarylimidazoles as inhibitors of microsomal prostaglandin E₂ synthase-1

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ARTICLE INFO

ABSTRACT

Article history: Received 24 August 2010 Revised 23 September 2010 Accepted 24 September 2010 Available online 19 October 2010

Keywords: mPGES-1 Inhibitor Biarylimidazole PGE2 Inflammation Microsomal prostaglandin E_2 synthase (mPGES-1) represents a potential target for novel analgesic and anti-inflammatory agents. High-throughput screening identified several leads of mPGES-1 inhibitors which were further optimized for potency and selectivity. A series of inhibitors bearing a biaryl imidazole scaffold exhibits excellent inhibition of PGE₂ production in enzymatic and cell-based assays. The synthesis of these molecules and their activities will be discussed.

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Inducible microsomal prostaglandin E_2 synthase-1 (mPGES-1) appears to be the predominant synthase involved in cyclooxygenase-2 (COX-2) mediated PGE₂ production.¹ Mice deficient in mPGES-1 show both a reduction in the production of inflammatory PGE₂ and a decrease in inflammatory responses in the collagen-induced arthritis model.^{2,3} Furthermore, mPGES-1 knock-out mice exhibited no thrombogenesis nor blood pressure changes.⁴ Thus, mPGES-1 inhibitors may achieve coxib-like efficacy in relieving pain and inflammation, while avoiding the adverse cardiovascular consequences associated with COX-2 mediated PGI₂ suppression.

An indole carboxylic acid series of mPGES-1 inhibitors derived from the FLAP (5-lipoxygenase activating protein) inhibitor, MK-886, has been previously described.⁵ However, this series suffered from high protein binding, which resulted in decreased potency in cells. Hence, high-throughput screenings were carried out to search for other molecular scaffolds that may address this issue. The assay involved a 30-s mPGES-1 enzymatic reaction followed by detection of PGE₂ using EIA (enzyme immune assay).⁶ Compound **2** was identified as a moderately potent mPGES-1 inhibitor (IC₅₀ = 660 nM; Table 1). In IL-1 β stimulated A549 epithelial lung carcinoma cells in presence of 2% and 50% FBS, compound **2** inhibited the production of PGE₂ (IC₅₀ = 3100 and 7000 nM, respectively) but not PGF_{2α} (data not shown). Given its low molecular weight and structural simplicity, structural–activity relationship (SAR) studies were initiated on this series of compound.

The biarylimidazole scaffold could be divided into four segments for SAR analysis: the 2-imidazole position, the central imidazole ring, the 4-imidazole position, and the 5-imidazole position. At the 2-imidazole position, three compounds have been prepared. Compound **1** which contains a mono-*ortho*-cyanophenyl ($IC_{50} = 1400 \text{ nM}$) and compound **3** with a bis-*ortho*-cyanofluoro phenyl ($IC_{50} = 1000 \text{ nM}$) showed less affinity for the mPGES-1 enzyme (Table 1). Hence, the inhibitors described hereafter all contain the bis-*ortho*-chlorofluoro substitution pattern at the 2-imidazole position as in compound **2** ($IC_{50} = 660 \text{ nM}$).

Moving the nitrogen on the central imidazole ring led to reduced activity against mPGES-1 (Table 2). Replacement of the central imidazole ring with an oxadiazole resulted in the complete loss of activity (compound **5**). A triazole derivative (compound **6**) showed superior inhibition in both the enzymatic (mPGES-1) and whole cell (A549) assays. These preliminary results suggest that the position of the H-bond donating NH is important for mPGES-1 activity.

For the 4-imidazole position, SAR studies show that *para*relationship between the imidazole and R^3 is better than *meta*and *ortho*- (data not shown). Hydrophobic groups are generally tolerated at the R^3 position (Table 3). Added flexibility of the linker resulted in lost of potency as can be seen with the alkyne linkage (compound **14**; IC₅₀ = 8 nM) which is superior over the alkene (compound **12**; IC₅₀ = 35 nM) and the alkane (compound **13**; IC₅₀ = 150 nM). However, any polar substitutions on the phenyl alkyne

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Table 1

SAR at the 2-imidazole position



^a Mean of two or more experiments.



Replacing the central imidazole ring



^a Mean of two or more experiments.

(e.g., carboxylic acid, amide, sulfone, sulfonamide, and tertiary alcohol, compounds **15a–e**) led to the complete loss of enzyme inhibition. Although polar groups are not tolerated at all in the R^3 region of the inhibitors, introduction of a nitrogen atom on the 4-phenyl ring boosted cellular activities of PGE₂ synthesis inhibition, as exemplified by comparing compound **16** (Table 4) with compound **14** (Table 3). Subsequent derivatization all contained a triple bond linker extending from the 4-pyridyl group. Non-polar substituents on the phenyl alkyne afforded good mPGES-1 inhibi-

tors, as did cyclohexyl (**19**) and cyclohexenyl (**20**) alkynes. The smaller cyclopropyl alkyne moiety (**21**) gave an inhibitor with slightly decreased potency, while the terminal alkyne analog (**22**) showed dramatically reduced potency. More polar heterocycle substituted alkynes were less potent inhibitors as well (compound **23**).

Addition of a substituent at the 5-imidazole position affected mPGES-1 activity (Table 5). In general, electron-withdrawing groups further increased the enzymatic potency (compounds **24**

Table 3

Entry

2

7

8

9

10

11

12

13

14

15а-е

SAR extending from the 4-phenyl group

R³

Cl~₅

mPGES-1

660

320

180

1400

110

570

35

150

8

>5000

 $IC_{50}^{a}(nM)$

A549 PGE₂ (2%/50% FBS)

 $IC_{50}^{a}(nM)$

3100/7000

2100/27,000

1700/27,000

>5000/>50,000

380/8700

1600/19,000

190/3300

920/13,000

280/3200

NA



SAR extending from the 4-pyridyl group



Entry	R ⁴	mPGES-1 IC_{50}^{a} (nM)	A549 PGE ₂ (2%/50% FBS) IC ₅₀ ^a (nM)
16	Contract of the second se	23	29/520
17	N	480	410/1800
18a-c	CI	ortho-9 meta-4 para-31	43/610 64/640 28/280
19	r Land	33	290/620
20		13	82/400
21		170	350/1800
22	- ser	2500	3300/4000
23	N	940	440/1400

^a Mean of two or more experiments.

Y = CO_2H , $CONH_2$, SO₂CH₃, SO₂NH₂, C(CH₃)₂OH

and **25**). Selected inhibitors were also tested in the human whole blood assay (HWBA) for inhibition of lipopolysaccharide-induced PGE₂ production.⁷ Compound **25**, containing a bromo substituent on the 5-imidazole position, showed an IC₅₀ of 1.6 μ M in the HWBA, which is only threefold less active than rofecoxib (IC₅₀ = 0.53 μ M) and 1.5-fold less active than etoricoxib (IC₅₀ = 1.1 μ M).^{8,9} Compound **25** also showed excellent bioavailability (127%) and a half-life of 4.8 h in rats (PO: 20 mpk in 60% PEG 200; IV: 5 mpk in 80% PEG 200). Inhibitors bearing a triazole (e.g., compound **28**) were not pursued further, since they generally exhibited lower potencies compared with the 5-bromo imidazoles.

The synthesis of compound **25** and similar biarylimidazole analogs is described in Scheme 1. Starting from commercially available arylmethylketones **29**, alpha bromination using bromine under acidic conditions gave alpha-bromoketones **30**. Intermediates

^a Mean of two or more experiments.

30 were then condensed with amidine **31** in refluxing aqueous KHCO₃/THF.¹⁰ The amidine was prepared in a separate reaction by addition of LHMDS to the corresponding benzonitrile, followed by cleavage of the silyl groups under aqueous acidic conditions.¹¹ The resulting imidazoles **32** were coupled with alkynes under standard Sonogashira coupling conditions. Compounds **33** were brominated through an electrophile substitution using *N*-bromosuccinimide, followed by aqueous workup, to give the desired 5-bromo-biarylimidazoles **34**.

In summary, the present study demonstrates that biarylimidazole can serve as an excellent scaffold to design potent and selective mPGES-1 inhibitors with good rat pharmacokinetic properties. Even though this series still display a large serum shift, probably due to their high lipophilicity, several biarylimidazole inhibitors are less shifted in presence of serum proteins when compared to the indole carboxylic acid series.⁵ In HWBA, the activity of compound **27** is approaching that of extensively-characterized COX-2 inhibitors rofecoxib and etoricoxib. It remains to be seen if inhibitors of mPGES-1 will demonstrate equivalent efficacy in inflammatory pain.

Table 5 SAR at the 5-imidazole position





^a Mean of two or more experiments.



Scheme 1. Reagents and conditions: (a) Br₂, HBr, AcOH, 0–40 °C, 1 h (54–74%); (b) KHCO₃ aq, THF, reflux, 3 h (50–70%); (c) Pd(PPh₃)₄, Cul, Et₃N, DMF, 65 °C, 18 h (50–80%); (d) NBS, CCl₄, rt, 2 h; 1% HBr aq (65-85%).

References and notes

- 1. Murakami, M.; Naraba, H.; Tanioka, T.; Semmyo, N.; Nakatani, Y.; Kojima, F.; Ikeda, T.; Fueki, M.; Ueno, A.; Oh, S.; Kudo, I. J. Biol. Chem. 2000, 275, 32783.
- Uematsu, S.; Matsumoto, M.; Takeda, K.; Akira, S. J. Immunol. 2002, 168, 5811.
 Trebino, C. E.; Stock, J. L.; Gibbons, C. P.; Naiman, B. M.; Wachtmann, T. S.; Umland, J. P.; Pandher, K.; Lapointe, J. M.; Saha, S.; Roach, M. L.; Carter, D.; Thomas, N. A.; Durtschi, B. A.; McNeish, J. D.; Hambor, J. E.; Jakobsson, P. J.;

Carty, T. J.; Perez, J. R.; Audoly, L. P. Proc. Natl. Acad. Sci. U.S.A. 2003, 100, 9044.

- Cheng, Y.; Wang, M.; Yu, Y.; Lawson, J.; Funk, C. D.; Fitzgerald, G. A. J. Clin. Invest. 2006, 116, 1391.
- Riendeau, D.; Aspiotis, R.; Ethier, D.; Gareau, Y.; Grimm, E.; Guay, J.; Guiral, S.; Juteau, H.; Mancini, J.; Méthot, N.; Rubin, J.; Friesen, R. W. *Bioorg. Med. Chem. Lett.* 2005, 15, 3352.
- Masse, F.; Guiral, S.; Fortin, L. J.; Cauchon, E.; Ethier, D.; Guay, J.; Brideau, C. J. Biomol. Screen. 2005, 10, 599.
- 7. Brideau, C.; Kargman, S.; Liu, S.; Dallob, A. L.; Ehrich, E. W.; Rodger, I. W.; Chan, C.-C. Inflamm. Res. **1996**, 45, 68.
- Chan, C.-C.; Boyce, S.; Brideau, C.; Charleson, S.; Cromlish, W.; Ethier, D.; Evans, J.; Ford-Hutchinson, A. W.; Forrest, M. J.; Gauthier, J. Y.; Gordon, R.; Gresser, M.; Guay, J.; Kargman, S.; Kennedy, B.; Leblanc, Y.; Leger, S.; Mancini, J.; O'Neill, G.

P.; Ouellet, M.; Patrick, D.; Percival, M. D.; Perrier, H.; Prasit, P.; Rodger, I.; Tagari, P.; Therien, M.; Vickers, P.; Visco, D.; Wang, Z.; Webb, J.; Wong, E.; Xu, L.-J.; Young, R. N.; Zamboni, R.; Riendeau, D. J. Pharmacol. Exp. Ther. **1999**, 290, 551.

- Riendeau, D.; Percival, M. D.; Brideau, C.; Charleson, D.; Dubé, D.; Ethier, D.; Falgueyret, J.-P.; Friesen, R. W.; Gordon, R.; Greig, G.; Guay, J.; Mancini, J.; Ouellet, M.; Wong, E.; Xu, L.; Boyce, S.; Visco, D.; Girard, Y.; Prasit, P.; Zamboni, R.; Rodger, I. W.; Gresser, M.; Ford-Hutchinson, A. W.; Young, R. N.; Chan, C.-C. J. Pharmacol. Exp. Ther. 2001, 296, 558.
- Li, B.; Chiu, C. K.-F.; Hank, R. F.; Murry, J.; Roth, J.; Tobiassen, H. Org. Process Res. Dev. 2002, 6, 682.
- 11. Thurkauf, A.; Hutchison, A.; Peterson, J.; Cornfield, L.; Meade, R.; Huston, K.; Harris, K.; Ross, P. C.; Gerber, K.; Ramabhadran, T. V. *J. Med. Chem.* **1995**, *38*, 2251.