PRODUCT STEREOSPECIFICITY IN THE MICROBIAL REDUCTION OF IMIDAZOL-1-YL METHYL ARYL KETONES

Randall Lis*, Walton B. Caldwell, Gregory I. Rudd, and William C. Lumma, Jr.

> Berlex Laboratories, Inc. 110 East Hanover Avenue Cedar Knolls, NJ, USA 07927

Georg Alexander Hoyer, Karl Petzoldt, Gerhard Cleve, and Gerhard Sauer

> Research Laboratories, Schering AG West Germany D-1000 Berlin (West) 65 Federal Republic of Germany

Abstract: Various microorganisms can be used to reduce ketoimidazole 3 to hydroxyimidazole 4. Only one enantiomer (i.e., $(\underline{R})-4$) is produced. In contrast, ketoimidazolium salt 2 is not reduced under identical conditions.

The importance of racemic imidazolium salt 1 as an active Class III antiarrhythmic agent¹ prompted us to investigate syntheses of its'



enantiomers. One possible route would be via the microbiological reduction of ketoimidazolium salt 2 to give $(\underline{R})-1$ and/or (S)-1 (eq. 1).



Attempted reduction of 2, using over 60 microbial organisms, was unsuccessful and afforded only recovered starting material. In contrast, reduction of ketoimidazole 3 with various microorganisms proved to be quite effective and afforded only one enantiomer² (i.e. (<u>R</u>)-4, <u>vide infra</u>) regardless of the microorganism used (eq. 2). High enantioselectivity has also been reported in the microbiological reductions of other aryl ketones.³



The results of the microbial reduction of 3 are summarized in Table I. The extent of reduction and chemical yield were determined by HPLC analysis of the crude lyophilized broths.⁴

| Reduction Medium | Time (hr) | Starting | Yield |
|--|--------------|-----------|-------|
| | | Remaining | |
| Saccharomyces carlsbergensis, immobilized | 232 | 15% | 16% |
| Kloeckera magna, immobilized | 232 | 2% | 21% |
| Hansenula subpelliculosa, distilled water resting cell, 3-fold | 332 | 3% | 27% |
| Hansenula subpelliculosa, distilled water resting cell, 5-fold | 188 | 1% | 36% |
| Kloeckera magna, distilled water resting cell, 3-fold | 236 | 11% | 69% |
| Kloeckera magna, distilled water resting cell, 5-fold | 332 | 24% | 69% |
| Kloeckera magna (Tris buffer), resting cell, 5-fold | 188 | 0% | 43% |
| Kloeckera jensenii (Tris buffer), resting cell, 3-fold | 332 | 11% | 45% |
| mol of alcohol | | | |

TABLE I: Result of the Microbial Reduction of 3

a = 100 x mol of starting ketone-mole of recovered ketone

The enantiomers of 4^5 have been prepared and their absolute configurations established by a single-crystal X-ray of the $(-)-\underline{1}$ -camphorsulfonic acid salt of $(\underline{S})-4$.¹ Derivatization of these enantiomers with $(\underline{S})-(-)-\alpha$ -methylbenzyl isocyanate (MBI) 5 gave the corresponding diastereomeric carbamates 6 and 7 (eq 3), which were also analyzed by HPLC.⁶ Treating the lyophilized broths⁷ in Table I with $(\underline{S})-(-)-\alpha$ -MBI afforded only carbamate 6 by HPLC analysis.



This indicates that only $(\underline{R})-4$ is produced from the microbial reduction of 3. This result is also consistent with Prelog's rule⁸ for predicting the stereochemistry of alcohols prepared by microbial reduction of their ketone precursors.

In conclusion, it is possible to prepare enantiomerically pure $(\underline{R})-4$ (and therefore $(\underline{R})-1)^2$ using various microbiological methods.

<u>General Procedure of Microbiological Reduction</u>: The yeast strains were cultivated in an aqueous nutrient medium containing per liter: glucose 50 g, and corn steep 20 g (pH 6.5). After inoculation, the flasks were agitated at 28°C for 70 h. Cells were harvested by centrifugation and resuspended for fermentation in distilled water or 0.1 M Tris/HCl buffer (pH 7.5). In addition, cells were entrapped by suspending in sodium alginate solution and dropping the mixture into CaCl₂ solution. The alginate beads were resuspended in aqueous transformation medium containing 10 g CaCl₂ per liter. After incubation on a rotary shaker at 28°C (substrate concentration 200 mg per liter) the cells and the alginate beads were filtered and the filtrate lyophilized.

REFERENCES AND NOTES

- R. Lis, T. K. Morgan, Jr., R. J. DeVita, D. D. Davey, W. C. Lumma, Jr., R. A. Wohl, J. Diamond, S. S. Wong, M. E. Sullivan, H. J. Reiser, J. Wiggins, and J. Hill, <u>The Pharmacologist</u> 1985, 27(3), 112, Abstr. 28; R. Lis, T. K. Morgan, Jr., R. J. DeVita, D. D. Davey, W. C. Lumma, Jr., R. A.Wohl, J. Diamond, S. S. Wong, and M. E. Sullivan, <u>J. Med. Chem</u>. in press.
- 2. Previous studies (ref 1) have shown that (R)-4 and (S)-4 can be converted into (R)-1 and (S)-1, respectively.
- O. Cervinka, L. Hub, Coll. Czech. Chem. Commun. 1966, 31, 2615;
 D. Ridley, M. Stralow, J. Chem. Soc., Chem. Commun. 1975, 400;
 K. Kabuto, M. Imuta, E. S. Kempner, H. Ziffer, J. Org. Chem. 1978, 43, 2357; M. Bucciarelli, A. Forni, I. Moretti, G. Torre, J. Chem. Soc.
 Commun. 1978, 456; Synthesis 1983, 897; and M. Imuta, K. I. Kawai, H. Ziffer, J. Org. Chem. 1980, 45, 3352.
- 4. HPLC Chromatographic Conditions: Column: Phenyl-4.6 mm X 250 mm, 5 micron, spherical (Chromegabond-Alkyl Phenyl, ES Industries, Marlton, NJ), Mobile Phase: 2/98 v/v 2-propanol/0.025 M ammonium acetate adjusted to pH 4.0 with H₂SO₄, Flow Rate: 1.5 mL/min, Temp.: 40°C, Detection: UV at 225 nm, Typical Retention Times: 3:t_r = 8.17 min; 4: t_r = 5.44 min.
- 5. Enantiomers $(\underline{R})-4$ and $(\underline{S})-4$ were prepared by the method described in reference 1.

 - (S)-4: CD (CH₃OH) $\lambda = 276$ nm, $\Delta \varepsilon = + 0.135$ $\lambda = 223$ nm, $\Delta \varepsilon = + 1.90$ $[\alpha]^{23} = +59.7^{\circ}$ (1N HCl)
- 6. HPLC Chromatographic Conditions: Column: C₆ 4.6 mm x 250 mm, 5 micron, spherical (Chromegabond-C₇, ES Industries, Marlton, NJ), Mobile Phase: 12/88 V/V 2-propanol/0.05[°] M ammonium acetate adjusted to pH 2.5 with H₂SO₄, Flow Rate: 1.5 mL/min., Temp: 50°C, Detection: U.V. at 254 nm, Typical Retention Times 6: $t_r = 7.12$ min.; 7: $t_r = 8.48$ min.
- 7. The crude lyophilized broths were purified by preparative TLC prior to derivatization. The carbamates were prepared by treating $(\underline{R})-4$ and $(\underline{S})-4$ with $(\underline{S})-(-)\alpha$ -methylbenzyl isocyanate (MBI) in 1-methyl-2-pyrrolidinone. Purification prior to derivatization minimizes interferences in the HPLC chromatogram.
- 8. V. Prelog, Pure Appl. Chem., 1964, 9, 119.

(Received in USA 23 October 1986)