

PII: S0960-894X(97)00242-4

The Synthesis of CD-ring modified 1α ,25-dihydroxy vitamin D analogues: Six-membered D-ring analogues I.

B. Linclau, P. De Clercq, M. Vandewalle*

University of Gent, Department of Organic Chemistry, Laboratory for Organic Synthesis, Krijgslaan, 281 (S.4), B-9000 GENT (Belgium)°

R. Bouillon and A. Verstuyf

Laboratorium voor Experimentele Geneeskunde en Endocrinologie, K.U. Leuven, Onderwijs en Navorsing Gasthuisberg, Herestraat, 49, B-3030 LEUVEN (Belgium)

Abstract : Vitamin D analogues, characterized by a cyclohexane D-ring and by the absence of a C-ring are described. @ 1997 Elsevier Science Ltd.

The observation that 1α ,25-dihydroxy-vitamin D₃ (1; calcitriol) is active in the regulation of cell proliferation and differentiation, next to the classical role in calcium-bone homeostasis, has led in recent years to the development of analogues capable of dissociating cell differentiating effects from calcemic effects.^{1,2} Among the three fragments of the vitamin D skeleton structural modifications of the side-chain and of the A-ring have been especially studied in the past.³



Scheme 1

Some years ago, we embarked on an extensive study of the structure-function relationship focussing on the least studied part of the molecule, i.e. the central CD-ring region.⁴ In this respect we decided stripping the

[°] Fax: (32-9) 264 49 98 - E-mail: pierre.declercq@rug.ac.be

molecule to its five-carbon backbone (C-8 to C-20 : i) and resubstituting it again in various ways. In the present paper we describe the synthesis of analogues 2 where the central part is replaced by only a cyclohexane D-ring (scheme 1). We decided to select a "D-ring" carrying a *gem*-dimethyl group at C-13 (steroid numbering) as these substituents mimic respectively the angular C-18 methyl group and C-12 in the parent steroid 1 and which are known to have an influence on restricting the side chain orientations.³ Also the C-20 configuration influences the side chain orientation. Indeed it has been shown⁵ that analogues of 1 with the unnatural 20-(S)-configuration can induce interesting differentiations between calcemic effects and new actions. We therefore decided to also introduce both C-20 configurations together with some other side chain modifications.

The synthetic strategy centers around the advanced intermediate 4 which can be obtained from (-)- α -pinene 5 according to a procedure described by Chapuis and Brauchli.⁶ The methoxycarbonyl substituent allows for the construction of the side chain while the keto function is a handle for the introduction of the C-8 formyl group in 3 needed for the Lythgoe coupling⁷ with the A-ring precursor 6.⁸



a) LDA, THF, 5-Br-1-pentene, HMPA, $-78^{\circ}C \rightarrow r.t.$, 5 h; b) LiAlH₄, Et₂O, r.t., 4 h; c) TsCl, Et₃N, DMAP, CH₂Cl₂, r.t., 20 h; d) LiAlH₄, Et₂O, Δ , 5 h; e) O₃, CH₂Cl₂ : 2.5 M NaOH, MeOH (4 : 1), $-78^{\circ}C$; f) PPTS, Me₂CO, H₂O, Δ , 3 h; g) Me₂S=CH₂, THF, r.t., 2 h; h) BF₃OEt₂, THF, r.t., 10 h; i) NaOMe, MeOH, r.t., 12 h; j) (*i*) 9-BBN, THF, r.t., 3.5 h; (*ii*) EtOH, NaOH, H₂O₂, 50°C, 1 h; k) SO₃.py, DMSO, Et₃N, CH₂Cl₂, $-8^{\circ}C$, 6 h; l) EtMgBr, THF, r.t., 3 h; m) PTSA, Me₂CO, H₂O, r.t., 30 h; n) Ph₃P=CH₂, THF, Δ , 3 h; o) TESCl, imid., DMF, r.t., 12 h; r) LiBF₄, 2% H₂O in MeCN, r.t., 3 h.

Scheme 2

For the synthesis of analogues **2a,b,c** with 20-*R* configuration (scheme 2) we decided to alkylate **4** with a 4-pentenyl group which would provide us with some flexibility for the construction of different side chains. Epimer **7** was obtained in 95% d.e. and separation was possible by preparative HPLC. This result is in accord

with Wicha's observation on steroids.⁹ The relative configuration¹⁰ was proven at the stage of 8 and 20-*epi*-8 (minor epimer) on the basis of MM2 conformational analysis¹¹ and n.O.e. experiments. In 20-*epi*-8 a 3.7% n.O.e. enhancement was observed between one of the 21-hydroxymethyl protons and the axial methyl group while no enhancement was found for 8; this is consistent with their calculated side chain orientations.

Reductive removal of the hydroxy group, *via* the tosylate, gave 9 which *via* ozonolysis¹² and acetal hydrolysis led to 10. Reaction with dimethylsulfonium methylide¹³, afforded an epimeric mixture of epoxides 11 (6:4). Lewis acid induced isomerization of 11 led to a separable mixture of aldehydes 12 and 13 (6:4 ratio). Base catalyzed equilibration gave 12 and 13 (ratio 6.5:1).¹⁴

The synthesis of analogue 2c, with a 24, 26, 27-trishomo side chain, now involved initial hydroboration of 9, followed by formation of methyl ester 15 which then led to 16. For the synthesis of the aldehyde 20 we decided to adopt a different route than described for 12, involving Wittig olefination and hydroboration. However again C-14 epimers 18 (circa 1:1) were obtained; separation was impossible as was the case for the corresponding aldehydes 20 and *epi-20*. We then found that HPLC separation was possible at the stage of acetals 19 and *epi-19*. Deprotection¹⁵ gave, without epimerization, the desired precursor 20,¹⁴



a) (*i*) LDA, THF, -30°C,; (*ii*) MeI, HMPA, -78°C, 3 h then r.t., 2 h; b) LiAlH4, Et₂O, r.t., 4 h; c) TsCl, NEt₃, DMAP, CH₂Cl₂, r.t., 20 h; d) (*i*) NaH, DMSO,65°C, 1.5 h; (*ii*) HC = C(OEE)Me₂ then **23**, DMSO, r.t., 2 h; e) PPTS, n-PrOH, r.t., 1 h; f) H₂ (1 atm), 5% Rh/Al₂O₃, EtOAc, r.t., 2 h; g) PPTS, acetone, H₂O, r.t., 30 h; h) TESCl, imidazole, DMF, r.t., 35 h; i) Ph₃P(Me)Br, t-BuOK, THF, Δ , 3 h; j) (*i*) BH₃.Me₂Sl, hexane, r.t., 3 h; (*ii*) H₂O₂, EtOH, NaOH, 65°C, 1 h; k) SO₃.py, DMSO, NEt₃, CH₂Cl₂, -8°C, 6 h.

Scheme 3

For the synthesis of the 20-*epi* analogues **2d**,**e** ester **4** was now methylated to **21** (scheme 3), as the sole isomer (compare **4** to **7**). The side chain was introduced *via* substitution of tosylate **22** with lithiated 3-(ethoxy-ethoxy)-3-methyl-but-1-yn. After hydrogenation and acetal hydrolysis, **24** was transformed into epimeric alcohols **25** (circa 1:1) as described for **16**. In contrast to **18**, epimers **25** could by separated by preparative HPLC. Finally oxidation afforded respectively the precursors **26** and **27**.¹⁴

Construction of the title compounds 2 involves the Lythgoe coupling⁷ of aldehydes 12, 13, 20, 26 and 27, with the A-ring phosphine oxide 6^8 and subsequent deprotection (TBAF). For the synthesis of 2a and 2b, the coupled products were reacted with MeMgBr prior to deprotection.

The affinity of the D-ring analogues 2 to the pig intestinal mucosa vitamin D receptor (VDR) was evaluated as described previously.¹⁶ The relative affinity of the analogues was calculated from their concentration needed

to displace 50% of $[^{3}H]1\alpha$,25(OH)₂D₃ from its receptor compared with the activity of 1α ,25(OH)₂D₃ (1 assigned a value of 100%).

The biological evaluation (see table) was determined in vitro on different cell lines (HL-60, MCF-7, MG-63, keratinocytes)³. The *in vivo* effect was tested in vitamin D-deplete normal NMRI mice by measuring calcium levels in serum. The values in the table are given in relation for those of the natural hormone 1 (value 100). BL 269 (2a) has a stronger affinity for the VDR compared to 1. This analogue shows an antiproliferative (MCF-7, keratinocytes) and prodifferentiating (MG-63) activity 3 times that of 1 and is 20 times less calcemic. It is noteworthy that the 20(S)-epimer 2d with unnatural configuration is devoid of all activity. This stands in sharp contrast to analogues with the natural ring where the 20(S) configuration mostly induces higher biological activities.⁵ Further details of the biological activities will be published elsewhere.

Table

Analogue	VDR	HL-60	MG-63	MCF-7	Keratino- cytes	Calcium Serum
2a (BL 269)	125	90	300	300	300	4
2b (BL 272)	1	2	3	1	30	< 0.1
2c (BL 473)	60	600	800	3000	1500	20
2d (BL 314)	0.9	1	1	0	8	< 0.1
2e (BL 315)	0.5	1	1	0	8	0.1

Acknowledgements. We thank the "NFWO", the 'Ministerie voor Wetenschapsbeleid" and THERAMEX S.A. for the financial support to the laboratory.

References and notes

- 1. Bouillon, R.; Van Baelen, H. Saudi Med. J. 1989, 10, 260.
- 2. DeLuca, H.F.; Burmester, J.; Darwish, H.; Krisinger, J. Comprehensive Medicinal Chemistry, Pergamon Press, New York 1990, vol. 3, 1129.
- 3. Bouillon, R.; Okamura, W.H.; Norman, A.W. Endocrine Reviews 1995, 16, 200.
- 4. (a) Bouillon, R.; De Clercq, P.; Pirson, P.; Vandewalle, M. Novel structural analogues of vitamin D; patent PCT/EP 93.202037.3; priority date 09-07-1993; (b) Sabbe, K.; D'Halleweyn, C.; De Clercq, P.; Vandewalle, M.; Bouillon, R.; Verstuyf, A. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 1697; (c) Gui-Dong, Zhu; Yongjun, Chen; Xiaoming, Zhou; Vandewalle, M.; De Clercq, P.; Bouillon, R.; Verstuyf, A. *Bioorg. Med.* Chem. Lett. 1996, 6, 1703;(d) For an analogous approach see Kutner, A.; Zhao, H.; Fitak, H.; Wilson, S.R. Bioorganic Chem. 1995,23, 22.
- 5. Binderup, L.; Latini, S.; Binderup, E.; Bretting, C.; Calverley, M.; Hansen, K. Biochem. Pharmacol. **1991**, *4*2, 1569.
- 6. Chapuis, C.; Brauchli, R. Helv. Chim. Acta 1992, 75, 1527.
- 7. Lythgoe, B.; Moran, T.A.; Nambudiry, M.E.N.; Tideswell, J.; Wright, P.W. J. Chem. Soc., Perkin Trans I 1978, 580; Kocienski, P.J.; Lythgoe, B. J. Chem. Soc., Perkin Trans. I 1978, 1290.
- 8. Baggiolini, E.G.; Iacobelli, J.A.; Hennessy, B.M.; Batcho, A.D.; Sereno, J.F.; Uskokovic, M.R. J.Org. Chem. 1996, 51, 3098.
 Wicha, J.; Bal, K. J. Chem. Soc., Perkins Trans, 1 1978, 1282.
- 10. Linclau, B.; Vandewalle, M.; Syntlett 1995, 10, 1063.
- 11. Still, W.C.; Mohamadi, F.; Richards, N.G.J.; Guida, W.C.; Lipton, M.; Liskang, R.; Chang, G.; Hendrickson, T., De Gunst, F.; Hasel, W.; MacroModel V3.0, Department of Chemistry, Colombia University, New York, NY 10027, USA.
- 12. Marshall, J.A.; Garofalo, A.W. J. Org. Chem. 1993, 58, 3675.
- 13. Corey, E.J.; Chaykovski, M. J. Am. Chem. Soc. 1965, 87, 1353.
- 14. ¹H-NMR (500 MHz) : J value for 14-H (after double irradiation of 8-H); 13, m (Δ J = 10 Hz); 14, 12.52 and 3.72 Hz; 21, 12.40 and 3.78 Hz; 27, 12.47 and 3.84 Hz; 28, m ($\Delta J = 10$ Hz).
- Lipshutz, B.H.; Harvey, D.F. Synth. Commun. 1982, 12, 267.
 Bouillon, R.; Allewaert, K.; Van Leeuwen, J.P.T.M.; Tan, B.K.; Xiang, D.Z.; De Clercq, P.; Vandewalle, M.; Pols, H.A.P.; Bos, M.P.; Van Baelen, H.; Birkenhäger, J.C. J. Biol. Chem. 1992, 267, 3044.