

A Journal of the Gesellschaft Deutscher Chemiker A DOCH International Edition Market Chemiker CDCh Chemiker Ch

Accepted Article

- Title: Total Synthesis of the Alleged Structure of Crenarchaeol enables Structure Revision
- Authors: Mira Holzheimer, Jaap S. Sinninghe Damsté, Stefan Schouten, Remco W. A. Havenith, Ana V. Cunha, and Adriaan J. Minnaard

This manuscript has been accepted after peer review and appears as an Accepted Article online prior to editing, proofing, and formal publication of the final Version of Record (VoR). This work is currently citable by using the Digital Object Identifier (DOI) given below. The VoR will be published online in Early View as soon as possible and may be different to this Accepted Article as a result of editing. Readers should obtain the VoR from the journal website shown below when it is published to ensure accuracy of information. The authors are responsible for the content of this Accepted Article.

To be cited as: Angew. Chem. Int. Ed. 10.1002/anie.202105384

Link to VoR: https://doi.org/10.1002/anie.202105384

WILEY-VCH

RESEARCH ARTICLE

Total Synthesis of the Alleged Structure of Crenarchaeol enables Structure Revision

Mira Holzheimer,^[a] Jaap S. Sinninghe Damsté,^{[e],[f]} Stefan Schouten,^{[e],[f]} Remco W. A. Havenith,^{[a],[b],[c]} Ana V. Cunha,^[d] and Adriaan J. Minnaard^{[a]*}

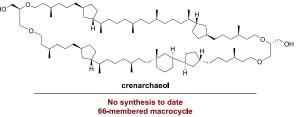
Dedication ((optional))

[a]	Dr. Mira Holzheimer, Prof. Dr. Remco W. A. Havenith, Prof. Dr. Stratingh Institute for Chemistry	Adriaan J. Minnaard	
	University of Groningen		
	Nijenborgh 7, 9747 AG, Groningen, The Netherlands		
[b]	Prof. Dr. Remco W. A. Havenith		
	Zernike Institute for Advanced Materials		
	University of Groningen		
	Nijenborgh 4, 9747 AG, Groningen, The Netherlands		
[c]	Prof. Dr. Remco W. A. Havenith		
	Ghent Quantum Chemistry Group, Department of Chemistry		
	Ghent University		
	Krijgslaan 281 (S3), B-9000 Gent, Belgium		
[d]	Dr. Ana V. Cunha		
	Eenheid Algemene Chemie (ALGC)		
	Vrije Universiteit Brussel (VUB)		
	Pleinlaan 2, 1050 Brussels, Belgium	4	
[e]	Prof. Dr. Jaap S. Sinninghe Damsté, Prof. Dr. Stefan Schouten		
	NIOZ Royal Netherlands Institute for Sea Research		
	Department of Marine Microbiology and Biogeochemistry		
	PO Box 59, 1790 AB Den Burg, The Netherlands		
[f]	Prof. Dr. Jaap S. Sinninghe Damsté, Prof. Dr. Stefan Schouten		
	Faculty of Geosciences, Department of Earth Sciences		
	Utrecht University		
	PO Box 80.021, 3508 TA Utrecht, The Netherlands		

Supporting information for this article is given via a link at the end of the document.

Abstract: Crenarchaeol is a glycerol dialkyl glycerol tetraether lipid produced exclusively in Archaea of the phylum Thaumarchaeota. This membrane-spanning lipid is undoubtedly the structurally most sophisticated of all known archaeal lipids and an iconic molecule in organic geochemistry. The 66-membered macrocycle possesses a unique chemical structure featuring 22 mostly remote stereocenters, and a cyclohexane ring connected by a single bond to a cyclopentane ring. Herein we report the first total synthesis of the proposed structure of crenarchaeol. Comparison with natural crenarchaeol allowed us to propose a revised structure of crenarchaeol, wherein one of the 22 stereocenters is inverted.

chains, contrary to the straight chain fatty acid glycerol ester lipids found in Bacteria and Eukarya.^[5] Apart from the difference in lipid linkage, the stereochemistry of the glycerol backbone in archaeal



66-membered macrocycle 22 stereocenters – four times 2 contiguous, 14 remote including 1 *all*-carbon quaternary Four 1,3-*trans*-substituted cyclopentane rings Unique 5-6 ring motif Highly flexible, aliphatic character



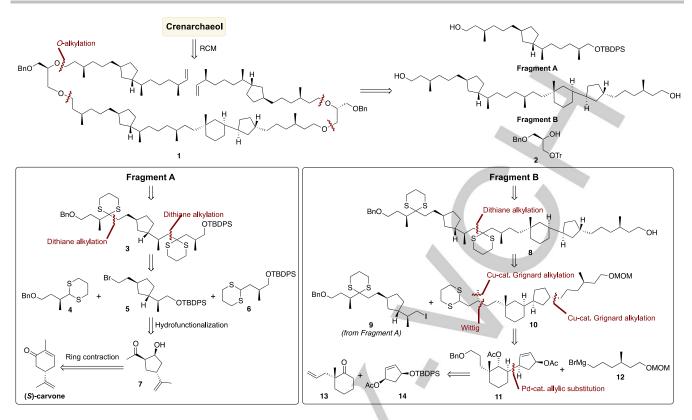
In 1990, Woese proposed to classify all living organisms in three domains of life: Archaea, Bacteria and Eukarya.^[1] Before that, 'archaeabacteria' were considered to belong to the Bacteria. Based on differences in their genome and lipidome, Archaea were ultimately recognized as separate, third domain.^[2] For a long time, Archaea were primarily associated with extreme habitats such as high temperature, extreme pH, and hypersaline environments.^[3] Growing interest over the years, however, led to the discovery of meso- and extremophilic Archaea in virtually any habitat on Earth.^[4] The cell membrane of Archaea is built up of diether or membrane-spanning tetraether lipids containing isoprenoid

Figure 1 The alleged structure of crenarchaeol.

isoprenoidal glycerol dialkyl glycerol tetraether lipids (GDGTs) is opposite to bacterial or eukaryotic glycerolipids, raising questions on the evolution of archaeal and bacterial/eukaryotic lipid membranes.^[6] The lipid composition of Archaea varies, depending on the species and environmental factors, and this is considered an adaptation to their habitat.^[7] The ether-linkages provide chemical stability against hydrolysis, and the presence of methyl-branches and cyclopentane moieties, which are formed by

Vanusc

RESEARCH ARTICLE



Scheme 1 Retrosynthetic analysis of crenarchaeol.

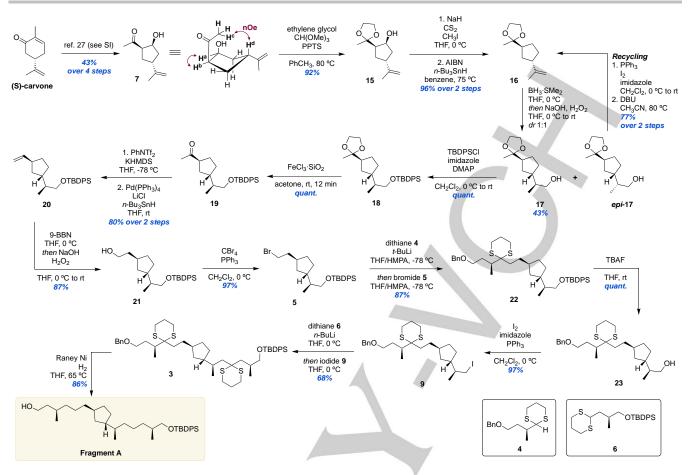
internal cyclization of the biphytanol chain,^[8] leads to decreased membrane permeability, allowing growth at extreme pH, salinity, and temperature.^[9] One archaeal GDGT - named crenarchaeol stands out from all other archaeal membrane lipids due to its unique chemical structure (Figure 1). Crenarchaeol is produced by a specific lineage of Archaea, the Thaumarchaeota, ^[10] and was first isolated from surface sediments of the Arabian Sea. After extensive GC-MS and NMR analysis, the structure and stereochemistry of this unique GDGT was proposed, a considerable achievement given the fact that the molecular complexity originates merely from its unusual hydrocarbon framework.^[11] It contains four 1,3-trans-substituted cyclopentane moieties. One of these is connected by a single bond to a cyclohexane ring, a structural feature rarely found in natural products.^[12] This feature of crenarchaeol is likely formed by further internal cyclization of the bicyclic biphytanyl moiety.^[5b] Crenarchaeol contains a total of 22 stereocenters, most of which are remote, including an all-carbon quaternary stereocenter. Recently, a parallel glycerol configuration of sedimentary crenarchaeol was inferred from chemical derivatization experiments.^[13] Montenegro et al. confirmed the structure of the bicyclic biphytanyl moiety in archaeal GDGTs by total synthesis,^[14] yet to date, there is no proof of structure of the tricyclic biphytanyl moiety of crenarchaeol and no total synthesis. The 5-6 ring motif of crenarchaeol is particularly interesting due to its complexity and uniqueness in nature. In order to ultimately confirm the structure and stereochemistry of crenarchaeol, we embarked on its total synthesis.

Results and Discussion

Our retrosynthetic analysis of crenarchaeol made use of the inherent symmetry of the bicyclic biphytanyl chain of the molecule (Scheme 1). It started with the disconnection of the central C-Cbond of the bicyclic biphytanyl moiety by intramolecular alkene metathesis and ether bond disconnection of 1. This led to two key intermediates, termed Fragment A and B, and protected glycerol building block 2. Fragment A can be further simplified via dithiane disconnections to arrive at building blocks 4 and 6, both carrying a methyl-branched stereocenter, and cyclopentane building block 5. In turn, 5 can be traced back to hydroxyketone 7, which is accessed from commercially available (S)-carvone via ring contraction. Syntheses of archaeal cis-[15] and trans-substituted[14] cyclopentane containing lipids have been previously reported. As we planned to build the macrocycle by alkylation of a suitably functionalized glycerol building block and ring-closing metathesis, we required differentially protected lipid chains containing the trans-substituted cyclopentane and the methyl-branches. Based on the stereochemical assessment of the bicyclic biphytane moiety in crenarchaeol¹⁷ and its subsequent confirmation provided by Helmchen et al.22, we planned the synthesis of the desired stereoisomer.

Retrosynthesis of Fragment B commenced with the C–C-bond disconnection of **8** arriving at dithiane **10** and iodide **9**, the latter originating from Fragment A. Further simplification of **10** by asymmetric Cu-catalyzed Grignard alkylations and a Wittig olefination delivered diacetate **11**. The 5-6 ring motif of **11** was disconnected at the C–C-bond joining the two carbocycles.^[16]

RESEARCH ARTICLE



Scheme 2 Synthesis of Fragment A.

We realized that for this challenging transformation an advanced intermolecular Pd-catalyzed asymmetric allylic alkylation could be instrumental, inspired by the work of Trost.^[17] By this, we arrived at building blocks **13** and **14**, readily accessible from pimelic acid and cyclopentadiene, respectively.

Synthesis of Fragment A

The synthesis of Fragment A was initiated by the preparation of known β -hydroxyketone 7 from (S)-carvone (Scheme 2). Via a four-step sequence involving a hydrolytic ring contraction,[18] 7 was obtained as single diastereomer, as confirmed by NOESY Notably, this sequence proved robust and scalable and allowed multigram synthesis of 7 (see SI). After acetal protection of 7, the hydroxyl group of 15 was removed by Barton-McCombie deoxygenation, providing 16 in excellent yield. Notably, acetal protection was necessary to avoid elimination of the β -hydroxyl group in the synthesis of the xanthate intermediate. Initially, we envisioned to stereoselectively install the methyl stereocenter adjacent to the 5-membered ring by means of Cu- or Co-catalyzed asymmetric hydroboration.^[19] No published method to perform the asymmetric hydroboration of the 1,1-disubstituted terminal alkene of 16 delivered 17 in acceptable yield and stereoselectivity, however. Thus, we resorted to non-stereoselective

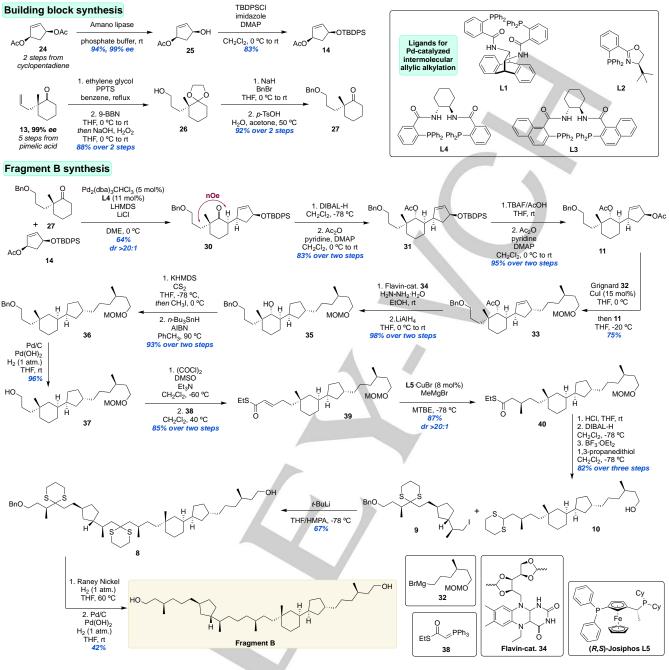
hydroboration-oxidation of **16** followed by diastereomer separation, giving **17** in 43% yield as single stereoisomer. The stereochemistry of the methyl-branched center in **17** was determined by amidation of its corresponding acid with phenylglycine methyl ester, followed by ¹H NMR analysis

Table 1. Optimization of the Pd-catalyzed allylic alkylation.
--

	AcO 14	OTBDPS	a) ₃ CHCl ₃ (5 mol%) and (11 mol%) 0 °C	0 H H 29	-OTBDPS
Entry ^[a]	Ligand	Base	Solvent ^[c]	Conversion ^[b] (yield) ^[c]	dr ^[d]
1	L1	LHMDS	THF	80%	25:75
2	L2	LHMDS	THF	40%	51:49
3	L3	LHMDS	THF	40%	81:19
4	L4	LHMDS	THF	40% (27%)	86:14
5	L4	LHMDS	PhCH₃	40%	85:15
6	L4	LHMDS	DME	42%	94:6
7	L4	NaHMDS	DME	10-15%	92:8
8	L4	LDA	DME	41%	92:8
9 ^[e]	L4	LHMDS	DME	full (53%)	93:7

[a] See SI for details. [b] Determined by $^1{\rm H}$ NMR. [c] Isolated yield. [d] Determined by $^{13}{\rm C}$ NMR of the crude product. [e] 1.6 eq. of LHMDS and 3 eq. LiCl were used.

RESEARCH ARTICLE



Scheme 3 Synthesis of Fragment B.

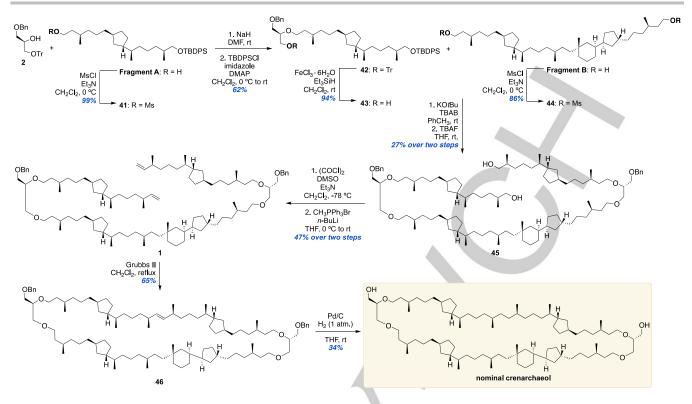
Synthesis of Fragment B

(see SI).^[20] In addition, the efficiency of the synthesis was further increased by 'recycling' of the undesired *epi-*17 by iodination and elimination, giving alkene **16** in 77% yield over two steps. After silyl protection of **17**, the acetal moiety of **18** was removed. Optimization of the reaction conditions, to minimize epimerization, resulted in treatment of **17** in acetone with FeCl₃ adsorbed to silica,^[21] giving **19** in quantitative yield with 3% epimerization. Ketone **19** was converted to the corresponding terminal alkene by enol-triflation and Pd-catalyzed triflate reduction, delivering **20** in 80% yield over two steps. Hydroboration-oxidation of **20** gave alcohol **21** in 87% yield, which was converted to the corresponding bromide **5** in excellent yield. With **5** in hand, the

stage was set for the first dithiane alkylation.^[22] After optimization of the lithiation conditions of **4** (prepared using known methods, see SI), the alkylation proceeded in high yield (87%) giving **22**. Desilylation followed by Appel iodination delivered iodide **9**, which serves as intermediate in the synthesis of both Fragment A and B. In turn, after identification of the optimal lithiation conditions, deprotonation of dithiane **6** with *n*-BuLi at 0 °C followed by addition of **9** produced bis-dithiane **3** in 68% yield. With the carbon skeleton of Fragment A constructed, the dithiane moieties and the benzyl ether of **3** were removed by Raney-nickel reduction in good yield, thus concluding the synthesis of Fragment A.

Next, the considerably more complex Fragment B was to be constructed. The synthesis started with the preparation of two

RESEARCH ARTICLE



Scheme 4 Completion of the synthesis of the proposed structure of crenarchaeol.

building blocks 14 and 27 (Scheme 3). The synthesis of cyclopentene 14 started from *meso*-diacetate 24, accessible in two steps from cyclopentadiene.^[23] Diacetate 24 was subjected to enzymatic desymmetrization^[24] in excellent yield and *ee*, followed by silyl protection giving 14. Cyclohexanone 27 was prepared according to the method developed by the Stoltz laboratory from allyl cyclohexanone 13,^[25] which was protected and subjected to hydroboration/oxidation to deliver 26. Omission of the protection of the ketone in 13 led to the formation of the corresponding hemiacetal. Benzylation and acetal hydrolysis provided the desired cyclohexanone 27 in 92% yield over two steps.

With acetate **14** in hand, we chose to investigate the key step – the intermolecular Pd-catalyzed Tsuji-Trost alkylation – with 2,2-dimethylcyclohexanone **28** as model substrate (Table 1).

We started by screening ligands L1-L4 (Scheme 3) in combination with Pd₂(dba)₃CHCl₃ in order to achieve good chiral induction. In presence of LHMDS as base and THF as solvent at 0 °C, (R,R)-ANDEN-Phenyl Trost L1 gave good conversion to the alkylation product 29, albeit with a dr of 75:25 favoring the undesired diastereomer (Table 1, entry 1). Under the same conditions, (R)-t-ButyIPHOX L2 failed to give chiral induction (Table 1, entry 2). When using DACH ligands L3 and L4, good diastereoselectivities of 81:19 and 86:14 were achieved (entries 3 and 4), yet with a low conversion of around 40% and in the case of L4 only 27% isolated yield. Since acceptable stereo-induction was achieved, we continued the optimization with L4. Changing the solvent to toluene or DME (Table 1, entry 5 and 6) did not result in higher conversion, but the latter gave the product with improved dr of 94:6. When using NaHMDS the conversion dropped significantly to around 10-15% (Table 1, entry 7), while LDA performed comparable to LHMDS (entry 8). Ultimately,

increasing the equivalents of LHMDS to 1.6 and using LiCl as additive resulted in full conversion (Table 1, entry 9). The product was isolated in 53% yield with an excellent dr of 93:7.

We decided to apply these conditions to acetate **14** and cyclohexanone **27**, and found this system to be superior to the model reaction. Product **30** was obtained in 67% yield with a *dr* >20:1, and no undesired diastereomer detected (Scheme 3). This variant of the intermolecular Pd-catalyzed asymmetric allylic alkylation further expands the toolbox of this type of reaction and we expect it to open up new avenues for future asymmetric construction of joint ring systems in a convergent manner.

Progressing the synthesis of Fragment B, the ketone moiety was reduced and acetylated, giving 31 as single diastereomer. Subsequent desilylation and acetylation delivered diacetate 11 in excellent yield. Notably, attempts to shorten this sequence by performing reduction, desilylation, and double acetylation led to significantly lower yields. This was due to the formation of a tricyclic product arising from S_N2' addition of the non-allylic hydroxy group to the double bond (see SI). With diacetate 11 in hand, a regioselective copper-catalyzed Grignard alkylation with 32 (prepared from (R)-citronellol, see SI) was performed providing a crude dr of 4:1 and, after separation of the isomers, alkylation product 33 in 75% yield as single stereoisomer. The double bond of 33 was reduced by a flavin-catalyzed diimide reduction^[26] followed by deacetylation providing 35 in 98% yield over two steps. The hydroxyl moiety of 35 was then removed by a Barton-McCombie deoxygenation reaction in excellent yield. After Pdcatalyzed hydrogenolysis of the benzyl ether in 36, alcohol 37 was oxidized to the corresponding aldehyde and subjected to a Wittig olefination delivering α,β -unsaturated thioester 39. The last methyl-branched stereocenter of Fragment B was then introduced in an excellent dr of 20:1 (see SI for details) by copper-catalyzed

RESEARCH ARTICLE

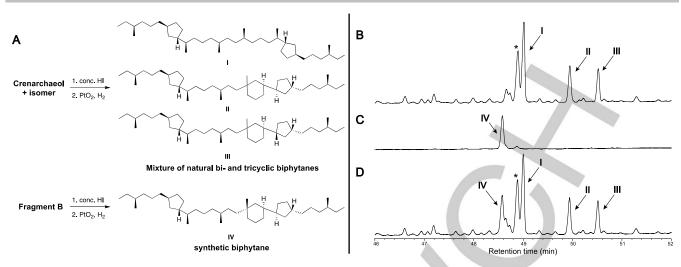


Figure 2 A: Conversion of natural crenarchaeol and Fragment B into biphytanes. B-D: Partial gas chromatograms of the formed biphytane(s). B: biphytanes I-III from the GDGTs in the Bligh Dyer extract of "*Ca. Nitrosotenuis uzonensis*".^[29] C: biphytane IV from Fragment B. D: Co-injection of IV with the biphytane mixture of "*Ca. N. uzonensis*". * Indicates an isomeric bicyclic biphytane, most likely originating from GDGT-4.

asymmetric conjugate addition of methylmagnesium bromide^[27] producing 40 in 87% yield. With the last stereocenter of the biphytane core of crenarchaeol set, the dithiane mojety of 10 was installed, after MOM deprotection of 40. through thioester reduction and treatment with 1.3propanedithiol in the presence of BF3:OEt2. Dithiane 10 was obtained in 82% over the three steps. Notably, dithiane synthesis in presence of the MOM ether resulted in a complex mixture of 10 and various trans-acetalization products. With 10 in hand, the last dithiane alkylation was performed, in presence of the free hydroxyl group. After optimization of the lithiation conditions, the reaction of lithiated 10 with iodide 9 smoothly provided the coupling product 8 in 67% yield, containing the entire carbon-skeleton of Fragment B. The synthesis of Fragment B was concluded by a two-step sequence, involving removal of the dithianes with Raney-nickel, followed by Pdcatalyzed hydrogenolysis of the remaining benzyl ether.

Endgame – Completion of the total synthesis of the proposed structure of crenarchaeol

After the successful stereoselective synthesis of both Fragment A and B, the macrocycle of crenarchaeol was assembled (Scheme 4). The endgame of the synthesis started with the O-alkylation of protected glycerol 2 with mesylate 41 prepared from Fragment A. During the reaction using sodium hydride in DMF, partial cleavage of the TBDPS ether was observed. Therefore, after O-alkylation, the silyl ether was reintroduced, giving alkylation product 42 in 62% yield. The trityl ether was removed delivering 43, the substrate for the next ether synthesis, in 94% yield. The double O-alkylation of 43 with bis-mesylate 44 came about after considerable experimentation, by reaction with KOtBu as the base in toluene in the presence of TBAB as phase-transfer catalyst. After desilylation of the crude double alkylation product, the desired diol 45 was obtained in a poor yield of 27% over the two steps. There are multiple factors complicating this reaction. It is a double O-alkylation of a bis-mesylate. The sheer size and flexibility of this electrophile plays a role in the reaction rate as we expect that the site of alkylation is not always exposed for reaction with the weak alkoxide nucleophile. In addition, small

amounts of elimination products were observed. Consequently, given the difficulty of this step, we continued with the synthesis. In order to perform the final ring closure of the macrocycle, **45** was converted to bis-alkene **1** by oxidation and Wittig reaction. The 66-membered macrocycle was closed by means of ringclosing metathesis with Grubbs 2nd generation catalyst,

a method often used for the construction of large rings.^[28] This provided **46** in 65% yield, given the size of the produced macrocycle a more than satisfactory result. In the final step, the double bond as well as the benzyl ethers were removed by hydrogenolysis with palladium on carbon in low yield of 34%, which could be partially attributed to the scale of the reaction. This concluded the synthesis of this structurally complex lipid and provided 1.2 mg of synthetic crenarchaeol. With both synthetic crenarchaeol and the tricyclic intermediate Fragment B in hand we sought to investigate the chemical structure of natural crenarchaeol. For this purpose, we re-isolated natural crenarchaeol in a laborious procedure (see SI) and made a comparison of their NMR spectra. Furthermore, we performed chemical derivatization in combination with GC-MS analysis.

Comparison of natural crenarchaeol and Fragment B by GC-MS

The Bligh Dyer extract of the thermophilic Thaumarchaeota "Ca. Nitrosotenuis uzonensis" (dominated by crenarchaeol and its cis-cyclopentyl isomer^[29], see Fig. 2A) has previously been treated with HI. This cleaves the ether bonds to produce a mixture of biphytane diiodides.[29] Reduction of the iodides with H_2/PtO_2 led to the corresponding hydrocarbons I-III, which were analyzed by GC-MS.^[29] This showed a ratio of bi- and tricyclic biphytanes of approximately 1:1 (Fig. 2B). As a direct comparison of the configuration of the tricyclic biphytane unit within synthetic and natural crenarchaeol was considered complicated, we subjected also Fragment B to this derivatization (Fig. 2A).^[29-30] This enabled a precise comparison by GC-MS. Treatment of fragment B with HI followed by reduction yielded biphytane IV which appeared, as expected, as a single peak in the gas chromatogram (Fig. 2C), but much to our surprise with a significantly different retention time than the supposedly identical II derived from natural

RESEARCH ARTICLE

crenarchaeol. The mismatch in chemical structure was confirmed by co-injection, showing retention time differences of **IV** and **II** or **III** of approximately 1.5 and 2 min, respectively (Fig. 2D).

Next, we turned our attention to the mass spectra of **II-IV** (see SI). The fragmentation patterns of natural **II** and **III** were equivalent to their previously reported mass spectra,^[29, 31] and featured the characteristic fragment m/z 262, originating from bond cleavage adjacent to the quaternary stereocenter. This fragment was also clearly visible in the mass spectrum of synthetic **IV**.

Furthermore, the remaining fragmentation patterns of **II/III** and **IV** are also virtually identical, providing strong evidence that the overall chemical connectivity of **II/III** and synthetic **IV** is identical. Thus, we concluded that **II** and **IV** are stereoisomers.

Comparison of Fragment B with isolated natural crenarchaeol by NMR

In order to elucidate the exact structural difference between synthetic Fragment B and the tricyclic biphytanyl moiety of natural crenarchaeol, we compared their NMR spectra. The ¹H and ¹³C signals of natural crenarchaeol^[11] and Fragment B were assigned by thorough 2D NMR analysis. In addition, the ¹³C signals of synthetic crenarchaeol were assigned based on the NMR analysis of Fragment B.

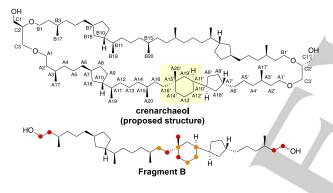


Figure 3 Alleged structure of crenarchaeol with carbon numbering and the synthetic Fragment B. Carbons with moderate ($\Delta\delta$ 0.25–1 ppm) and larger ¹³C chemical shift differences are marked in orange and red, respectively.

The comparison of selected ¹³C NMR signals of Fragment B and synthetic crenarchaeol with those of natural crenarchaeol is shown in Table 2 (see SI for a table with all signal assignments).

The carbon numbering is shown in Figure 3, and significant differences in ¹³C NMR shifts between Fragment B and natural crenarchaeol are marked in orange ($\Delta\delta$ 0.25–1 ppm) and red ($\Delta\delta$ >1 ppm). Upon comparison of the ¹³C NMR signals of Fragment B with those of natural crenarchaeol,^[11] it becomes clear that the majority of the chemical shifts of Fragment B are in very good agreement ($\Delta\delta$ < 0.25 ppm) with those of the tricyclic biphytane of

crenarchaeol. In particular, the ¹³C chemical shifts of the three cyclopentane rings (which are not connected to the cyclohexane ring) and their alkyl substituents are virtually identical (see SI).

Moderate chemical shift differences ($\Delta\delta$ 0.25–1 ppm) were ascribed to the cyclohexane ring carbons (A11', A12', A13' and A15') and the alkyl chain adjacent to the cyclohexyl ring (A16).

Large differences ($\Delta\delta$ >1 ppm) in chemical shift at the (sub)terminal carbons (A1, A1', A2 and A2') of Fragment B originate from the presence of primary hydroxyl moieties contrary to the ether linkages in crenarchaeol. More importantly, however, three carbon atoms around the allcarbon quaternary stereocenter elicit large differences in chemical shift at positions A14' ($\Delta\delta$ -5.96 ppm), A16' ($\Delta\delta$ -4.29 ppm) and A20' ($\Delta\delta$ +7.57 ppm), indicating a difference in structure around these positions. It is noteworthy that the ¹³C signals of the remaining stereocenters of the 5-6-ring system (A10' and A7') in Fragment B show no significant difference. In particular the good agreement of A10' is indicative for the ascribed stereochemistry of the single bond connecting the 5and 6-membered ring. It is expected that a difference in stereochemistry on A11' would translate to a significant ¹³C chemical shift difference in A10'. This indicates that, on these positions, the chemical structure of natural crenarchaeol matches that of Fragment B. The chemical shifts of synthetic nominal crenarchaeol (chemical shifts in brackets in Table 2) show the same pattern of chemical shift differences. It should be highlighted that there is no significant ¹³C chemical shift difference between Fragment B and the tricyclic biphytane of synthetic crenarchaeol (except for the terminal carbons A1/A1' and A2/A2') excluding an influence of the macrocyclic structure on the chemical shifts.

 Table 2. Comparison of ¹³C NMR values of natural crenarchaeol with

 Fragment B and synthetic nominal crenarchaeol.

	rragment D and synthetic nominal crenarchaeol.							
	Carbon number ^[a]	¹³ C shift natural crenarchaeol (ppm)	¹³ C shift Fragment (ppm) ^{b]}	В	$\Delta\delta$ (ppm) ^[c]			
	A1, A1'	70.23, 70.26	61.42 (70.28, 70.25)		-8.81, -8.84			
	A2, A2'	36.72, 36.75	40.14, 40.15 (36.74)		+3.42, +3.40			
	A11'	39.38	39.01 (39.10)		-0.37			
	A12'	32.27	31.80 (31.81)		-0.47			
	A13'	22.40	22.10 (22.12)		-0.30			
	A14'	44.13	38.17 (38.30)		-5.96			
	A15'	33.20	32.92 (32.93)		-0.28			
	A16	30.12	30.50 (30.51)		+0.38			
	A16'	37.80	33.51 (33.46)		-4.29			
_	A20'	22.55	30.12 (30.13)		+7.57			

[a] Assignments of ^{13}C NMR chemical shifts of crenarchaeol^{[11]} and Fragment B. Signals are reported relative to the solvent residual signal (CDCl₃ δ 77.16 ppm). [b] Corresponding signals of synthetic nominal crenarchaeol are shown in brackets.

Besides the good agreement of most of the ¹³C NMR chemical shifts of crenarchaeol and Fragment B, the ¹H NMR chemical shifts of A7', A10' and A11' correlate well (Table 3, see SI for all assignments). At position A19' (axial) and A20', only minor ¹H shift differences were observed. Only three positions show significant chemical shift differences: the equatorial proton of A14' ($\Delta\delta$ 0.27 ppm), A16' ($\Delta\delta$ 0.47 ppm) and the equatorial

RESEARCH ARTICLE

proton of A19' ($\Delta\delta$ 0.15 ppm). This provides further evidence that the difference in structure of natural and synthetic crenarchaeol is located around these positions.

Table 3. Comparison	of	¹Η	NMR	values	of	natural	crenarchaeol	and
Fragment B.								

Carbon number ^[a]	¹ H shift c (ppm)	renarchaeol	¹ H shift Fragment B (ppm) []]
A7'	1.79		1.78
A10'	1.47		1.46
A11'	1.17		1.12
A14'	1.15		1.42
A16'	1.31		1.78
A19'	ax.: 0.70; eq.: 1	1.39	ax.: 0.64; eq.: 1.52
A20'	0.84		0.79

[a] Assignments of ¹H NMR chemical shifts of crenarchaeol^[11] and Fragment B. Signals are reported relative to the solvent residual signal (CDCl₃ δ 77.16 ppm).

Since the relative and absolute stereochemistry of Fragment B is known, the methyl substituent A20' of Fragment B is assigned to be equatorial due to the 1,3-cis relationship of the methyl and cyclopentyl substituents on the cyclohexane ring. As a result of the deshielding γ -gauche effect, the ¹³C NMR chemical shift of axial substituents in cyclohexanes is more upfield relative to equatorial substituents.^[32] In Fragment B the ¹³C signal of methyl group A20' resonates at 30.12 ppm, while the methyl group A20' of natural crenarchaeol is shifted more upfield at 22.55 ppm. This strongly suggests that the methyl group A20' in natural crenarchaeol is in axial position in contrast to the initially proposed structure. To further support this, we considered the ¹³C chemical shifts of A16'. In Fragment B, the carbon atom A16' of the alkyl side-chain of the cyclohexyl ring is axially oriented. The ¹³C signal resonates at 33.51 ppm, whereas in crenarchaeol the ¹³C signal of A16' is shifted downfield to 37.80 ppm. Thus, the downfield shift of A16' in natural crenarchaeol strongly suggests equatorial

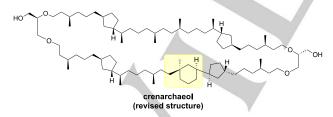


Figure 4 Revised chemical structure of natural crenarchaeol.

substitution of the alkyl chain substituent on the cyclohexyl ring. Further support comes from the computationally calculated ¹³C shift values for A16' and A20'. First, MD simulations in chloroform were carried out on fragment B and its isomer to determine the lowest energy conformations. Subsequently, the energies of the conformers from the MD trajectory were evaluated using DFT calculations and the chemical shifts calculated (See SI for the protocol and the calculated shifts). The DFT prediction is in good agreement with the upfield shift of methyl group A20' in natural crenarchaeol and the expected downfield shift of methylene A16'.

All in all this combined data provides overwhelming evidence for an inverted stereochemistry of crenarchaeol at A15' compared to Fragment B. On the basis of the evidence from chemical derivatization, NMR studies, and computation, we therefore propose a revised structure of crenarchaeol (Fig. 4), in which the stereochemistry of the *all*-carbon quaternary stereocenter is inverted compared to the original proposal.

Conclusion

The first total synthesis of the originally proposed structure of the thaumarchaeotal GDGT crenarchaeol has been achieved. The synthesis involved the stereoselective construction of a unique 5-6 ring motif as well as a late-stage 66-membered macrocyclization by means of RCM. The structure determination of crenarchaeol has a considerable history.[11] Due to the very complex structure, including 22 stereocenters, as well as the highly aliphatic character and its lack of rigidity, NMR-based structural studies have been heavily complicated. Furthermore, since this lipidic molecule does not have the tendency to crystallize, X-ray diffraction was not possible. The synthesis of the proposed structure of crenarchaeol and the key intermediate Fragment B enabled direct comparison with natural crenarchaeol by chemical derivatization and GC-MS analysis. This revealed a mismatch of the chemical structure of the tricyclic biphytane chain. Subsequently, detailed NMR analysis including computational simulation of ¹³C chemical shifts, of Fragment B and synthetic crenarchaeol, and comparison with natural crenarchaeol isolated from sea surface sediments was performed. Ultimately, from the spectroscopic data of fragment B, synthetic and natural crenarchaeol, we were able to revise the originally proposed structure beyond reasonable doubt. Through this extensive analysis we identified the inversion of just one out of the 22 stereocenters of crenarchaeol, namely the quaternary stereocenter embedded in the cyclohexane ring.

Total synthesis not only comprises the access to complex molecules, but serves also as a breeding ground for new synthetic methodology as well as probing current synthetic methods. Mistakes in the proposed structure of a natural product are by no means a rare occurrence.[33] The architectural and stereochemical complexity of a new unknown structure, in combination with very small amounts of isolated material often make assignments extremely difficult, in particular in a case such as crenarchaeol, which features almost no heteroatom functionalities and is highly flexible. By using the information gathered from the synthetic epimer of natural crenarchaeol, we were able to reassign the structure without the need to repeat the entire, very complex, synthesis. The correction of the structure of crenarchaeol has important implications for the study of its role in archaeal membranes. The current hypothesis is that the presence of crenarchaeol regulates membrane fluidity and packing, an important adaptation to temperature and pressure changes in the environment. As the stereochemistry of the guaternary center in crenarchaeol has a significant influence on its conformation,

RESEARCH ARTICLE

and thus membrane packing, we expect that an explanation (supported by for instance molecular dynamics simulations) for its role in membrane behavior is now within reach.

Acknowledgements

The authors would like to thank Dr. J. Buter and E. Jonkheim (University of Groningen) for their contribution to the isolation of crenarchaeol. P. van der Meulen and Dr. J. Kemmink (University of Groningen) are acknowledged for their assistance in NMR measurements. This work was sponsored by NWO Exact and Natural Sciences for the use of supercomputer facilities and RWAH and AV thank S. Dolas (SURF, NL) for allowing to perform experiments on the experimental AMD platform kleurplaat, maintained and operated by SURF Open Innovation Lab. This project received funding to JSSD from the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation program (grant agreement no. 694569, MICROLIPIDS).

Keywords: crenarchaeol • tetraether lipid • structure revision • archaea • total synthesis

- C. R. Woese, O. Kandler, M. L. Wheels, *Proc. Natl. Acad. Sci. USA* 1990, 87, 4576-4579.
- a) C. R. Woese, G. E. Fox, *Proc. Natl. Acad. Sci. USA* **1977**, *74*, 5088-5090; b) Y. Koga, H. Morii, *Microbiol. Mol. Biol. Rev.* **2007**, *71*, 97-120; c) R. Cavicchioli, *Nat. Rev. Microbiol.* **2011**, *9*, 51-61.
- [3] C. R. Woese, L. J. Magrum, G. E. Fox, J. Mol. Evol. 1978, 11, 245-252.
- [4] a) E. F. DeLong, *Curr. Op. Genet. Dev.* **1998**, *8*, 649-654; b) S.
 Schouten, E. C. Hopmans, R. D. Pancost, J. S. Sinninghe Damsté, *Proc. Natl. Acad. Sci. USA* **2000**, *97*, 14421-14426.
- [5] a) S. Jain, A. Caforio, A. J. Driessen, *Front. Microbiol.* 2014, 5, 641;
 b) L. Villanueva, J. S. Damste, S. Schouten, *Nat. Rev. Microbiol.* 2014, 12, 438-448.
- a) J. Lombard, P. Lopez-Garcia, D. Moreira, *Nat. Rev. Microbiol.* 2012, *10*, 507-515; b) Y. Koga, *J. Mol. Evol.* 2014, *78*, 234-242.
- [7] P. M. Oger, A. Cario, *Biophys. Chem.* 2013, 183, 42-56.
- [8] Z. Zeng, X. L. Liu, K. R. Farley, J. H. Wei, W. W. Metcalf, R. E. Summons, P. V. Welander, *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 22505-22511.
- [9] Y. Koga, Archaea **2012**, 2012, 789652.
- [10] a) J. S. Sinninghe Damsté, W. I. Rijpstra, E. C. Hopmans, M. Y. Jung, J. G. Kim, S. K. Rhee, M. Stieglmeier, C. Schleper, *Appl. Environ. Microbiol.* 2012, 78, 6866-6874; b) N. J. Bale, M. Palatinszky, W. I. C. Rijpstra, C. W. Herbold, M. Wagner, J. S. Sinnighe Damsté, *Appl. Environ. Microbiol.* 2019, *85*, e01332-01319.
- [11] J. S. Sinninghe Damsté, S. Schouten, E. C. Hopmans, A. C. T. van Duin, J. A. J. Geenevasen, *J. Lipid. Res.* 2002, 43, 1641-1651.
- [12] a) A. Matsuo, I. Terada, M. Nakayama, S. Hayashi, *Tet. Lett.* **1977**, 43, 3821-3824; b) A. V. Tkachev, M. M. Shakirov, V. A. Raldugin, *J. Nat. Prod.* **1991**, *54*, 849-853; c) F. Nagashima, M. Suzuki, S. Takaota, Y. Asakawa, *J. Nat. Prod.* **2001**, *64*, 1309-1317.
- [13] X.-L. Liu, D. A. Russell, C. Bonfio, R. E. Summons, Org. Geochem. 2019, 128, 57-62.
- [14] E. Montenegro, B. Gabler, G. Paradies, M. Seemann, G. Helmchen, Angew. Chem. Int. Ed. 2003, 42, 2419-2421.
- [15] a) G. Lecollinet, R. Auzély-Velty, M. Danel, T. Benvegnu, G. Mackenzie, J. W. Goodby, D. Plusquellec, J. Org. Chem. 1999, 64, 3139-3150; b) G. Lecollinet, A. Gulik, G. Mackenzie, J. W. Goodby, T. Benvegnu, D. Plusquellec, Chem. Eur. J. 2002, 8; c) M. Brard, W.

Richter, T. Benvegnu, D. Plusquellec, J. Am. Chem. Soc. 2004, 126, 10003-10012.

- [16] W. Kinouchi, R. Saeki, H. Kawashima, Y. Kobayashi, *Tet. Lett.* 2015, 56, 2265-2268.
- [17] B. M. Trost, D. L. Van Vranken, Chem. Rev. 1996, 96, 395-422.
- [18] W. B. Kover, J. Jones Jr., J. Braz. Chem. Soc. 1996, 7, 257-263.
- [19] a) J. Chen, T. Xi, X. Ren, B. Cheng, J. Guo, Z. Lu, Org. Chem. Front.
 2014, 1, 1306-1309; b) L. Zhang, Z. Zuo, X. Wan, Z. Huang, J. Am. Chem. Soc. 2014, 136, 15501-15504; c) W. J. Jang, S. M. Song, J. H. Moon, J. Y. Lee, J. Yun, J. Am. Chem. Soc. 2017, 139, 13660-13663.
- a) Y. Nagai, T. Kusumi, *Tet. Lett.* **1995**, *36*, 1853-1856; b) T.
 Yabuuchi, T. Ooi, T. Kusumi, *Chirality* **1997**, *9*, 550-555; c) T.
 Yabuuchi, T. Kusumi, *J. Org. Chem.* **2000**, *65*, 397-404.
- [21] K. S. Kim, Y. H. Song, B. H. Lee, C. S. Hahn, J. Org. Chem. 1986, 51, 404-407.
- [22] A. B. I. Smith, C. M. Adams, Acc. Chem. Res. 2004, 37, 365-377.
- [23] D. R. Deardorff, D. C. Myles, K. D. MacFerrin, *Tet. Lett.* 1985, 26, 5615-5618.
- [24] L. Tietze, C. Stadler, N. Böhnke, G. Brasche, A. Grube, Synlett 2007, 2007, 485-487.
- J. T. Mohr, M. R. Krout, B. M. Stoltz, *Org. Synth.* 2009, *86*, 194-211.
 C. Smit, M. W. Fraaije, A. J. Minnaard, *J. Org. Chem.* 2008, *73*, 9482-
- 9485.
- [27] R. Des Mazery, M. Pullez, F. López, S. R. Harutyunyan, A. J. Minnaard, B. L. Feringa, J. Am. Chem. Soc. 2005, 127, 9966-9967.
- [28] a) K. Arakawa, T. Eguchi, K. Kakinuma, J. Org. Chem. 1998, 63, 4741-4745; b) L. Yet, in Organic Reactions, Vol. 89 (Ed.: S. E. Denmark), John Wiley & Sons, Inc., 2016.
- [29] J. S. Sinninghe Damsté, W. I. C. Rijpstra, E. C. Hopmans, M. J. den Uijl, J. W. H. Weijers, S. Schouten, Org. Geochem. 2018, 124, 22-28.
- [30] a) O. Gräther, D. Arigoni, *J. Chem. Soc., Chem. Commun.* 1995, 405-406; b) M. Kaneko, F. Kitajima, H. Naraoka, *Org. Geochem.* 2011, *42*, 166-172; c) S. K. Lengger, Y. A. Lipsewers, H. de Haas, J. S. Sinninghe Damsté, S. Schouten, *Biogeosciences* 2014, *11*, 201-216.
- [31] S. Schouten, M. J. L. Hoefs, M. P. Koopmans, H.-J. Bosch, J. S. Sinnighe Damsté, Org. Geochem. 1998, 29, 1305-1319.
- [32] a) G. W. Buchanan, Can. J. Chem. 1982, 60, 2908-2913; b) E. Breitmaier, W. Voelter, Carbon-13 NMR Spectroscopy: High-Resolution Methods and Applications in Organic Chemistry and Biochemistry, 3 ed., VCH Verlagsgesellschaft mbH, Weinheim, Germany, 1987; c) M. E. Squillacote, J. M. Neth, Magn. Reson. Chem. 1987, 25, 53-56; d) J. Xiong, J. Wan, J. Ding, P.-P. Wang, G.-L. Ma, J. Li, J.-F. Hu, J. Nat. Prod. 2017, 80, 2874-2882.
- [33] K. C. Nicolaou, S. A. Snyder, Angew. Chem. 2005, 44, 1012-1044.

RESEARCH ARTICLE

Entry for the Table of Contents

revised structure of crenarch

Total synthesis of the proposed structure of crenarchaeol and comparison with the natural isolate has led to a revised structure of crenarchaeol, wherein one of the 22 stereocenters is inverted. The synthesis featured an asymmetric intermolecular Tsuji-Trost-type alkylation to access the unusual 5-6-ring motif as well as a late stage ring-closing metathesis to access the 66-membered macrocycle of crenarchaeol.

Institute and/or researcher Twitter usernames: @mira_holzheimer @AJMinnaard @StratinghInst @NIOZnews