## Synthesis of $\Delta^2$ -OPC-8:0 and OPC-6:0

Yuichi Kobayashi,\* Kaori Yagi, Takayuki Ainai

Department of Biomolecular Engineering, Tokyo Institute of Technology, 4259 Nagatsuta-cho, Midori-ku, Yokohama 226-8501, Japan Fax +81(45)9245789; E-mail: ykobayas@bio.titech.ac.jp

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**Abstract:**  $\Delta^2$ -OPC-8:0 (6), designed as a  $\beta$ -oxidation-insensitive analogue, was synthesized starting with 4-cyclopentene-1,3-diol monoacetate (5) in a stereocontrolled manner. The C(3)–C(8) moiety was first attached to the cyclopentene ring by using the Cucatalyzed S<sub>N</sub>2-type reaction with TBDPSO(CH<sub>2</sub>)<sub>6</sub>MgCl and was later converted into the full side chain by Wittig reaction. In addition, OPC-6:0 (3) was synthesized.

**Key words:** allylations, asymmetric synthesis, copper, cuprates, stereoselective synthesis

The linolenic acid cascade in plants leading to epi-jasmonic acid produces 12-oxophytodienoic acid (12-oxo-PDA) as the first metabolite possessing a five-membered ring, which is further transformed into 3-oxo-2-cis-(pent-2Z-enyl)cyclopentyloctanoic acid (OPC-8:0) by the reductase (Figure 1).<sup>1,2</sup> Subsequently,  $\beta$ -oxidation follows three times to produce sequentially OPC-6:0, OPC-4:0, and finally epi-jasmonic acid. It has become clear that epijasmonic acid is the key compound in the development, physiology, and defense of plants. These findings have suggested a biological function of the upper metabolites in the cascade. Indeed, tendril coiling response,<sup>3</sup> induction of secondary metabolites,<sup>4</sup> up-regulation of several genes,<sup>5</sup> and expression of the specific genes<sup>6</sup> have been reported for 12-oxo-PDA. However, a biological profile of OPCn:0 (n = 8, 6, 4) has not been studied, probably because of unavailability of these compounds in chemically pure form and in sufficient quantity.<sup>7,8</sup> A biological approach to these metabolites seems to suffer from low product selectivity and efficiency, because of the fairly fast  $\beta$ -oxidation of OPC-n:0 to its daughter OPC.9



Figure 1 Some metabolites of linolenic acid.

SYNLETT 2004, No. 14, pp 2582–2584 Advanced online publication: 20.10.2004 DOI: 10.1055/s-2004-834794; Art ID: U20004ST © Georg Thieme Verlag Stuttgart · New York The two side chains on the five-membered ring of these metabolites are oriented cis fashion, which restricts compatibility of reactions to be used for synthesis due to thermodynamic instability. Consequently, the synthetic methodologies developed for the prostaglandins which are based on the enolate chemistry would be inadequately applicable to synthesis of these metabolites.<sup>10</sup> Recently, our group developed copper-catalyzed installation of an alkyl chain onto 4-cyclopentene-1,3-diol monoacetate  $(5)^{11}$  and the reaction was successfully applied to synthesis of 12-oxo-PDA and OPC-8:0 in an enolate-free manner.<sup>12</sup> Herein, we report synthesis of  $\Delta^2$ -OPC-8:0 (6) as a  $\beta$ -oxidation-insensitive analogue by analogy of the  $\Delta^2$ prostaglandins<sup>13</sup> (Figure 2). The analogue would be useful for evaluation of OPC-8:0 (2).<sup>14</sup> In addition, we describe the first synthesis and characterization of OPC-6:0 (3).



Figure 2 Target and starting compounds of the present study.

Our approach to  $\Delta^2$ -OPC-8:0 (6) is summarized in Scheme 1. The key intermediate, diol 9, possessing the two carbon shorter side chain [C(3)–C(8)] was prepared from 5 through aldehyde 7 and lactone 8, and the full side chain was constructed by Horner–Emmons–Wittig reaction. The results of this investigation are delineated in Scheme 2.

According to our protocol to execute the S<sub>N</sub>2-type reaction, 3 equivalents of RMgCl was required with CuCN catalyst to complete the reaction with monoacetate 5. To reduce the quantity (3 equivalents) of TBDPSO(CH<sub>2</sub>)<sub>6</sub>MgCl, the hydroxyl group of 5 (>99% ee), prepared by the literature method,<sup>15</sup> was initially quenched with 1 equivalent of t-BuMgCl at 0 °C. Subsequently, 2 equivalents of TBDPSO(CH<sub>2</sub>)<sub>6</sub>MgCl was added at -18 °C to produce 10 in 81% yield with 92% regioselectivity after 4 hours. Mitsunobu inversion with HOAc, DIAD, and PPh<sub>3</sub> in toluene at -78 °C followed by hydrolysis afforded the alcohol (structure not shown) in 83% yield (2 steps). Claisen rearrangement of the alcohol with CH<sub>2</sub>=CHOEt at 180-190 °C for 3 days furnished aldehyde 11, which was converted into acid 12 in 86% yield. Lactonization with KI<sub>3</sub> followed by reaction with Bu<sub>3</sub>SnH under the radical conditions afforded lactone 13, which is equal to the key intermediate 8 proposed in Scheme 1.



**Scheme 1** Summary of  $\Delta^2$ -OPC-8:0 synthesis.

To complete the *cis*-pentenyl side chain, lactone **13** was hydrolyzed into hydroxyl acid, which upon esterification and silylation of the OH group afforded **14** in 80% yield. Reduction of ester **14** to aldehyde was followed by Wittig reaction with  $Ph_3P=CHEt$  to furnish *cis*-olefin **15**, which upon deprotection afforded the key diol **9** in quantitative yield.

In order to obtain aldehyde **21** for synthesis of  $\Delta^2$ -OPC-8:0 (6), direct oxidation of diol **9** with PCC was attempted (Scheme 3). However, the oxidation produced a mixture of **22** and **23**. To avoid this result, we then studied a transformation shown in Scheme 2. Silylation of diol **9** with TBSCl afforded **16**, which upon exposure to PPTS in EtOH and CH<sub>2</sub>Cl<sub>2</sub> (1:1) at 5–10 °C for 24 hours successfully afforded primary alcohol **17** in 89% yield.<sup>16</sup> PCC



Scheme 3 Attempted oxidation of diol 9,  $R = (Z)-CH_2CH=CHEt$ .

oxidation of **17** followed by Horner–Emmons–Wittig reaction under the Masamune conditions afforded the  $\alpha$ , $\beta$ unsaturated ester **18**. Desilylation of **18** followed by hydrolysis of the resulting alcohol **19** gave hydroxyl acid **20** in 79% yield from **18**. Finally, Jones oxidation furnished  $\Delta^2$ -OPC-8:0 (**6**) in 90% yield. Analytical data: IR (neat): 3196, 1738, 1699 cm<sup>-1</sup>. <sup>1</sup>H NMR:  $\delta = 0.96$  (t, J = 7.5 Hz, 3 H), 1.1–2.4 (m, 20 H), 5.36–5.46 (m, 2 H), 5.83 (d, J = 16 Hz, 1 H), 6.98–7.13 (dt, J = 16, 7 Hz, 1 H).<sup>17</sup>



Equation 1







**Scheme 2** Synthesis of  $\Delta^2$ -OPC-8:0.

Next, the key diol 9 used above was subjected to Jones oxidation to afford OPC-6:0 (3) in 68% yield (Equation 1):  $[\alpha]_D^{23}$  +43 (c 0.274, CHCl<sub>3</sub>). IR (neat): 3080, 1738, 1709  $cm^{-1}$ . <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta = 0.95$  (t, J = 7.5 Hz, 3 H), 1.18–1.48 (m, 8 H), 1.56–1.72 (m, 2 H), 1.76–2.42 (m, 10 H), 5.27-5.49 (m, 2 H). <sup>13</sup>C NMR (75 MHz,  $CDCl_3$ ):  $\delta = 14.2, 20.7, 22.6, 24.7, 24.8, 27.4, 28.0, 29.3,$ 33.9, 35.4, 38.7, 53.7, 126.2, 133.1, 179.2, 220.3. In addition, 3 was exposed to aqueous LiOH for epimerization at C(11) (Equation 2). The 11-epimer of 3 (i.e., 24) thus synthesized in 81% yield was identical with the racemic 11epimer reported in the literature<sup>8c</sup> by <sup>1</sup>H NMR, <sup>13</sup>C NMR, and IR spectroscopy. Purity of OPC-6:0 (3) was >95% by calculation of the peak heights at  $\delta = 38.7$  and 53.7 ppm for **3** and  $\delta = 41.2$  and 55.1 ppm for **24**, respectively, in the  $^{13}$ C NMR spectrum of **3**.<sup>18</sup>

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- (17) Spectral data ( $^{1}$ H NMR at 300 MHz and  $^{13}$ C NMR at 75 MHz in CDCl<sub>3</sub>) and specific rotations of the intermediates. Lactone 13:  $[\alpha]_D^{24}$  –5.7 (*c* 0.53, CHCl<sub>3</sub>). IR (neat): 1771, 1111 cm<sup>-1</sup>. <sup>1</sup>H NMR:  $\delta$  = 0.97 (s, 9 H), 1.10–1.36 (m, 10 H), 1.42–2.20 (m, 5 H), 2.35 (dd, *J* = 19, 6 Hz, 1 H), 2.42 (dd, *J* = 19, 10 Hz, 1 H), 2.78–2.92 (m, 1 H), 3.58 (t, *J* = 6 Hz, 2 H), 4.95 (t, *J* = 6 Hz, 1 H), 7.33–7.43 (m, 6 H), 7.63–7.69 (m, 4 H). <sup>13</sup>C NMR:  $\delta$  = 19.3, 25.7, 26.9, 28.5, 28.7, 28.9, 29.5, 30.6, 32.5, 33.1, 40.5, 42.8, 63.9, 86.2, 127.6, 129.6, 134.2, 135.6, 178.1. Ester 14:  $[\alpha]_D^{23}$  +0.10 (*c* 1.98, CHCl<sub>3</sub>). IR (neat): 1741, 1112 cm<sup>-1</sup>. <sup>1</sup>H NMR:  $\delta = 0.55$  (q, J = 8 Hz, 6 H), 0.93 (t, J = 8 Hz, 9 H), 1.04 (s, 9 H), 1.08–1.93 (m, 15 H), 2.20 (dd, J = 15, 5.5 Hz, 1 H), 2.32–2.44 (m, 1 H), 2.46 (dd, J = 15, 7.5 Hz, 1 H), 3.64 (t, J = 6.5 Hz, 2 H), 3.65 (s, 1)3 H), 4.17–4.24 (m, 1 H), 7.33–7.43 (m, 6 H), 7.64–7.70 (m, 4 H). <sup>13</sup>C NMR:  $\delta$  = 4.9, 6.9, 19.3, 25.9, 26.9, 28.1, 28.4, 29.5, 29.7, 31.7, 32.7, 32.8, 39.6, 43.8, 51.4, 64.1, 75.0, 127.6, 129.6, 134.3, 135.7, 174.9. Olefin **15**:  $[\alpha]_D^{25}$  +2.2 (*c* 0.368, CHCl<sub>3</sub>). IR (neat): 1428, 1112, 701 cm<sup>-1</sup>. <sup>1</sup>H NMR: δ = 0.56 (q, J = 8 Hz, 6 H), 0.95 (t, J = 8 Hz, 9 H), 0.96 (t, J = 7.5 Hz, 3 H), 1.04 (s, 9 H), 1.15–1.88 (m, 16 H), 1.98– 2.24 (m, 4 H), 3.65 (t, J = 6.5 Hz, 2 H), 4.06–4.19 (m, 1 H), 5.24-5.48 (m, 2 H), 7.34-7.43 (m, 6 H), 7.64-7.70 (m, 4 H). <sup>13</sup>C NMR:  $\delta$  = 5.0, 7.0, 14.4, 19.3, 20.8, 22.4, 25.9, 26.9, 28.5, 28.8, 29.8, 31.8, 32.7, 33.4, 39.9, 48.7, 64.1, 75.5, 127.6, 129.5, 129.9, 131.1, 134.3, 135.7. Diol **9**: [α]<sub>D</sub><sup>24</sup> +12  $(c \ 0.388, \text{CHCl}_3)$ . IR (neat): 3350, 1056 cm<sup>-1</sup>. <sup>1</sup>H NMR:  $\delta =$ 0.98 (t, J = 7.5 Hz, 3 H), 1.19-1.95 (m, 17 H), 2.20-2.50 (m, 17 H), 2.20-2.505 H), 3.64 (t, J = 6.5 Hz, 2 H), 4.14–4.26 (m, 1 H), 5.34–5.49 (m, 2 H). <sup>13</sup>C NMR:  $\delta = 14.5, 20.9, 22.8, 25.9, 28.9, 29.1,$ 29.8, 31.8, 32.9, 33.2, 40.1, 47.9, 63.1, 75.5, 128.7, 132.1. Hydroxyl acid **20**: IR (neat): 3410, 1699, 1652 cm<sup>-1</sup>. <sup>1</sup>H NMR:  $\delta = 0.96$  (t, J = 7.5 Hz, 3 H), 1.13–2.44 (m, 21 H), 4.15–4.25 (m, 1 H), 5.22–5.51 (m, 2 H), 5.82 (d, J = 16 Hz, 1 H), 6.98–7.13 (dt, J = 16, 7 Hz, 1 H). <sup>13</sup>C NMR:  $\delta = 14.4$ , 20.8, 22.7, 28.0, 28.6, 29.0, 29.5, 31.7, 32.4, 33.1, 40.0, 47.8, 75.5, 120.8, 128.7, 132.3, 152.3, 171.8.
- (18) Both **3** and **24** appeared on TLC with the same  $R_f$  value.