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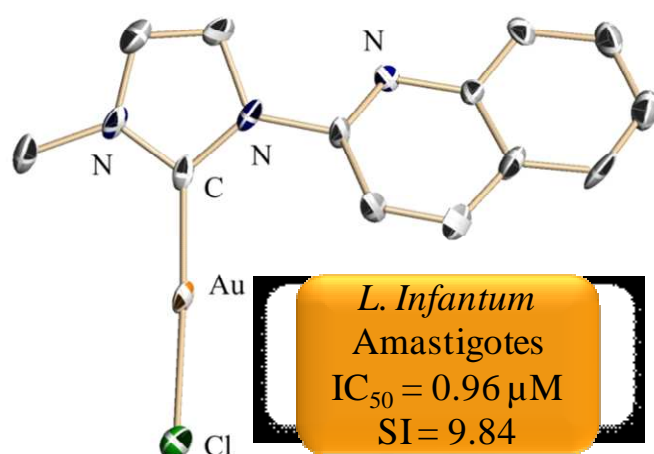
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Synthesis, characterization, and antileishmanial activities of gold(I) complexes involving quinoline functionalized *N*-heterocyclic carbenes

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Abstract

A series of new mononuclear cationic or neutral gold(I) complexes containing quinoline functionalized *N*-heterocyclic carbene(s) (NHC(s)) were synthesized and fully characterized by spectroscopic methods. The X-ray structures of two key compounds are presented. Proligands and their corresponding gold(I) complexes together with previously described silver(I) and gold(I) bis(NHC-quinoline) and gold(I) bis(NHC-methylbipyridine) complexes were evaluated *in vitro* towards *Leishmania infantum*. In parallel, the *in vitro* cytotoxicity of these molecules was assessed on the murine macrophages J774A.1. All gold(I) compounds show potent antileishmanial activity against *L. infantum* promastigotes and three of them are also efficient against *L. infantum* intracellular amastigotes. Structure-activity and toxicity relationships enables to evidence a lead-compound (**6**) displaying both a high activity and a good selectivity index.

Keywords: gold, *N*-heterocyclic carbene, quinoline, *Leishmania infantum*, J774.1A, antileishmanial activity.

1. Introduction

The *Leishmania* parasite is transmitted in their promastigote form to the mammalian host by the bite of infected sandflies (Phlebotominae) during their blood meal. Then promastigotes target cells of the immune system, macrophages and dendritic cells, where they differentiate into amastigotes forms, living inside the digestive vacuole of these host cells, responsible for leishmaniasis. Among tropical diseases, leishmaniasis remains one of the most neglected in terms of available controls and affects more and more countries. In 2007 this parasitic disease was endemic in 88 countries, in 2012 in 98 countries (mainly in tropical and sub-tropical regions, but also in southern Europe, especially around the Mediterranean area), and the number of cases continues to increase still certainly underestimated because reporting of this disease is mandatory in only 33 of the 98 countries involved. Factors and/or events such as mass migration of non-immune populations, deteriorating of socio-economic conditions and increase of immune-deficient people also lead to a resurgence of leishmaniasis worldwide. Actually this disease affects every year an estimated 1.3 million people and is the second most lethal parasitic infection worldwide with about 50.000 annual deaths. Leishmaniasis symptomatology may range from visceral disease affecting the haemopoietic organs, to

cutaneous and muco-cutaneous lesions that may cause disfiguring destruction of mucous membranes [1,2]. Until now, there is no available vaccine for humans whereas dogs can be preventively treated by the CaniLeish[®] (Europe), Leishmune[®] or Leish-Tec[®] (Brazil) vaccines [3,4]. In human medicine, very few drugs are available: pentavalent antimonials, liposomal amphotericin B, pentamidine, paromomycin, miltefosine. Sitamaquine and Fexinidazole are the only new ones actually in phase II clinical trial [5,6]. These drugs present main limitations including non-oral routes of administration (except for miltefosine), very expensive cost of liposomal amphotericin B (making this last drug unaffordable for most of affected patients), teratogenicity of miltefosine (prohibiting its use in many women) and finally, severe adverse effects of both pentamidine and pentavalent antimonials, leading to patients withdrawing from treatments and emergence of resistant strains. In this context, novel, effective, and safe drugs having reduced side effects in their treatment regimens against *Leishmania* is a major priority for health researchers [1].

Numerous synthetic organic compounds, including quinoline derivatives, that showed antileishmanial activities equal to or better than those of the standard drugs have been recently reviewed [7-11]. On the other side, advancements in the rational design of metal-based therapeutic agents have increased after the important discovery of cisplatin, and its derivatives as anticancer drugs [12,13], which has been the main impetus for the expansion of metal complexes in cancer and other pathologies. The peculiar biological properties of some organometallic compounds have helped in the development of new drugs based on metal complexes for major medical human problems including mainly cancer along with bacterial, viral and parasitic diseases [14]. In particular, the similarity between metabolic pathway of *Leishmania* parasites and tumor cells [15] and the fact that many antiprotozoal drugs bind to DNA, has led to the use of transition metal complexes as metallointercalators. Iridium, ruthenium, gold, copper, platinum and rhodium complexes with organic drugs or metallointercalating ligands are examples that have been explored for their antileishmanial activity [16-18]. However, there are just few gold based compounds explored against leishmaniasis [19].

Very recently, we focused our research on heteroditopic *N*-heterocyclic carbene (NHC) gold(I) complexes for their biomedical potential in anticancer and parasitic diseases such as malaria [20,21]. In particular, we have previously shown that mononuclear cationic gold(I) complexes containing aromatic amino-functionalized NHC exhibit *in vitro* potent and non toxic antiplasmodial activity against the chloroquino-resistant strain FcM29-Cameroon with

IC₅₀ values up to 330 nM [20]. In this work, we report the preparation and characterization of three mononuclear gold(I) complexes involving quinoline-NHC ligands. A family of molecules, based on aromatic nitrogen containing NHCs and including prolignands and mononuclear group 11 complexes, was evaluated both for their *in vitro* antiprotozoal activity toward *L. infantum* promastigotes and their *in vitro* cytotoxicity on J774.1 murine macrophages. Moreover, the most selective ones were tested for their *in vitro* activity toward *Leishmania* intracellular amastigotes.

2. Results and discussion

2.1. Chemistry

2.1.1. Synthesis of quinoline functionalized imidazolium salts

The imidazolium salt **1** was prepared from commercially available reagents, by heating an excess of 1-methylimidazole and 2-chloroquinoline in toluene (Scheme 1). 1-(4-Methylthiophenyl)-1*H*-imidazole was efficiently synthesized by a copper-catalyzed *N*-arylation of imidazole with 4-bromothioanisole in the presence of K₂CO₃ at 205 °C without solvent. The prolignands **2** and **3** were simply obtained after a quaternization step of 1-arylimidazole (aryl = mesityl for **2** and methylthiophenyl for **3**) with one equivalent of 2-chloroquinoline at 170 °C without solvent (Scheme 2). The most notable features in the ¹H and ¹³C NMR spectra of the imidazolium salts are the resonances for imidazolium protons (*H*₂) located at 10.89, 12.17 and 11.02 ppm and the corresponding imidazolium carbons (*C*₂) in the range of 135.2-137.8 ppm, for **1**, **2** and **3** respectively. The mass (FAB) or high resolution mass (ES) spectra of all compounds exhibit the classical peak corresponding to [M – Cl]⁺ cations.

Scheme 1. Synthesis of imidazolium salts **1** and **1'** and silver(I) bis(NHC) (**4**), gold(I) bis(NHC) (**5**) and gold(I) NHC (**6**) complexes.

Scheme 2. Synthesis of imidazolium salts **2** and **3** and gold(I) bis(NHC) complexes **7** and **8**.

2.1.2. Synthesis and characterization of mononuclear gold(I) complexes

The synthesis of gold(I) as well as silver(I) bis(NHC) complexes were previously described [20]. We have chosen the convenient transmetalation route to prepare the neutral gold(I) complex **6**. Firstly, the silver precursor complex was prepared by deprotonation of the

imidazolium salt **3**, with an excess of the mild base Ag₂O in acetonitrile at 40 °C. The carbene transfer reaction was then carried out *in situ*, by adding one equivalent of Au(SMe₂)Cl, with respect to the ligand. The cationic mononuclear gold(I) complexes **7** and **8** were prepared according a classical way, from the *N*-functionalized imidazolium salts **2** or **3** and one half equivalent of Au(SMe₂)Cl with sodium acetate as a mild base in hot *N,N*-dimethylformamide (120 °C). Gold(I) complexes **6-8** were isolated as white or beige powders with good yields (74-85%); all the synthesized complexes are highly stable towards air and moisture and soluble in CH₃CN, MeOH, H₂O and DMSO. NMR spectroscopy unequivocally demonstrates the formation of the gold(I) complexes; the ¹³C spectra show the resonance for the carbene carbon atoms at 170.1, 181.4 and 180.8 ppm, for **6**, **7** and **8**, respectively. These values are in the range of reported values for Au^I-NHC complexes having C-Au-X (X = halide) or C-Au-C motifs [22]. The elemental analysis of the gold(I) complexes correspond to the general formula AuLCl for **6** and [AuL₂][Cl] for **7** and **8** and the high resolution mass spectra (ES) exhibit the classical peaks corresponding to the cationic fragments [M + H]⁺ for the neutral complex **6** and [M – X]⁺ for the cationic compounds **7** and **8**.

2.1.3. Crystal structures of complexes **6** and **7**

Crystals of **6** suitable for X-ray diffraction analysis were obtained by slow evaporation from an acetonitrile solution of this compound (see Figure 1).

Figure 1: Crystal structure of **6** depicted at 50% level. Hydrogen atoms have been omitted for clarity. Selected bond lengths [Å] and angles [°]: Au-C 1.96(2), Au-Cl 2.28(1), C-Au-Cl 177.3(6).

In the case of **7**, slow evaporation from a methanolic solution led to crystals suitable for X-ray diffraction analysis (see Figure 2).

Figure 2: Crystal structure of the cationic part of **7** depicted at 30% level. Non-coordinating anions and hydrogen atoms have been omitted for clarity. Selected bond lengths [Å] and angles [°]: Au-C1 1.997(6), Au-C22 2.016(6), C1-Au-C22 178.9(2).

In both structures the gold atom is coordinated in a classical linear manner for gold(I) compounds. The gold carbon distances are in the normal ranges for NHC-Au-Cl and NHC-Au-NHC compounds. It is notable that in both cases the nitrogen atoms of the quinoline systems are localized away from the gold atoms and that the quinoline rings systems are not coplanar with the NHC ring systems ($C-N-C-N = 148^\circ$ for **6** and $127 - 137^\circ$ for **7**).

2.2. Biological activity

Figure 3. Compounds **9** and **10** also used in this study.⁹

All molecules **1-10** were first screened *in vitro* on the promastigote stage of *L. infantum* by determining their inhibitory concentrations 50% (IC_{50}) and compared to different antileishmanial reference-drugs: amphotericin B, miltefosine and pentamidine. In order to assess their selectivity of action, all molecules were also evaluated *in vitro* as regards of their cytotoxicity, measured by the cytotoxic concentrations 50% (CC_{50}) on the murine J774A.1 macrophages cell line (positive control = doxorubicin), giving access to the corresponding selectivity indexes ($SI = CC_{50} / IC_{50}$). The most selective compounds were then screened on the intracellular amastigote stage of *L. infantum*, the pathological relevant form of *Leishmania*.

Except proligands **1**, **1'** and **9**, all tested compounds including imidazolium salts, mononuclear silver(I) and gold(I) complexes presented an antileishmanial IC_{50} lower than 15 μM against promastigotes of *L. infantum*. The carbene precursors **2** and **3** exhibited relatively good antileishmanial IC_{50} values of 9.81 and 11.96 μM but they can only be considered as globally aspecific toxic molecules because of their very low CC_{50} values. First informations concerning the organic cationic proligands are emerging from these results. All imidazolium salts comprise an aromatic nitrogen containing arm, namely a quinoline moiety (**1**, **1'**, **2** and **3**) or a methyl-bipyridine entity (**9**), but differ in their non functionalized groups, methyl or aryl (mesityl for **2** and methylthiophenyl for **3**). The highest lipophilicity of **2** and **3** correlates with the higher antiproliferative activity against promastigotes, when compared to methyl-substituted imidazoliums (**1**, **1'** and **9**), a trend that we have already demonstrated in anticancer investigation of gold(I) aliphatic amino-NHCs complexes [21]. The silver(I) bis(NHC) complex (**4**) showed a potent but poorly specific antiprotozoal activity with respect

to the starting proligand **1'**, with an IC_{50} value of $9.37\ \mu M$ and a selectivity index of 1.61. The antileishmanial activity of the corresponding gold(I) complexes **5-8** and **10** was highly improved when compared to the corresponding proligands, with IC_{50} in the micromolar and submicromolar range, from 0.39 to $1.86\ \mu M$. The best values on *Leishmania* promastigotes were obtained for the three cationic Au(I) bis(NHC-quinoline) species with IC_{50} of 0.39, 0.42 and $0.43\ \mu M$ for **5**, **7** and **8**, respectively. Moreover as regards the toxicity, while the complex **7** appears to be non selective ($CC_{50} = 0.61$) although it is less toxic than the starting imidazolium salt **2** ($CC_{50} = 0.09$), compounds **5**, **6** and **8** exhibit interesting selectivity index comprised between 3.02 and 6.19. From these initial results, the later three complexes were selected for additive assays on the intracellular amastigote stage of *L. infantum*, in order to estimate the real potential of these organometallic molecules as efficient and selective antileishmanial compounds. They all display good antileishmanial activity on intracellular amastigotes with IC_{50} values below $1\ \mu M$, the best value being $0.24\ \mu M$ for **8**. More interestingly, the neutral complex **6** stands out with a selectivity index near to 10 (9.84). This compound display an anti promastigotes activity 38 times and 3 times higher than the one of amphotericin B and pentamidine respectively, but 5.8 times lower than the one of miltefosine, and an anti amastigotes activity 4.3 times lower than the one of miltefosine also. Overall, structure-activity relationships can be brought out for this family of gold complexes. First, this set of results evidenced the key role of the gold cation in antiprotozoal activity against *L. infantum* promastigotes and intracellular amastigotes. In this case, it seems that the carbenic ligand acts mainly as a carrier for the metal center towards its target(s). Second, even if the efficiency of compounds **5-8** and **10** are in the same order, the charge of the complex constitutes an important factor for the selectivity and the neutral gold complex **6** can be considered as the hit molecule in this study. To the best of our knowledge, the herein tested complexes represent the first examples of gold(I) functionalized NHC with efficacious antileishmanial activity.

In general, with the exception of antimonials, the use of metallodrugs and especially gold compounds as antileishmanial agents was scarcely explored and warrants deeper insight to elucidate molecular targets and modes of action. From the literature, Colotti and co-workers evaluated the antiarthritic gold(I) drug auranofin and $[AuCl(PEt_3)]$ (PEt: triethylphosphine) as antileishmanial agents [23]. Tests *in vitro* showed activity on the promastigote stage of *L. infantum* with IC_{50} of 9.68 and $16.59\ \mu M$, respectively. Because gold(I) compounds are known to be highly thiophilic, the authors have performed enzymatic assays on trypanothione

reductase (TR), which contains a dithiol motif in its active site; they have demonstrated that both auranofin and $[\text{AuCl}(\text{PEt}_3)]$ cause a pronounced inhibition of TR, a key enzyme of *L. infantum* polyamine-dependent redox metabolism and a validated target for antileishmanial drugs. Navarro *et al.* reported a gold(III) complex, containing a DNA intercalating dppz ligand (dppz: dipyrindo[3,2-a:2',3'-c]-phenazine) able to induce dose-dependent antiproliferative activity against promastigotes of *L. mexicana* with a minimal inhibitory concentration (MIC) of 3.4 nM and a lethal dose (LD_{26}) of 17 nM for 48 h. DNA interaction studies showed that this gold(III) complex may interact with calf thymus DNA and plasmid pUC 119, so the authors proposed that the potent leishmanicidal activity may be associated to the interaction of this gold-dppz compound with the parasite DNA by intercalation mode [24]. The same authors reported also a proposed dinuclear gold(I) triphenylphosphine complex with the bioactive coligand pyridine-2-thiol *N*-oxide (mpo). This compound showed *in vitro* potent activity against promastigotes of *L. mexicana* and *L. braziliensis* and *Trypanosoma. cruzi* epimastigotes and low unspecific cytotoxicity on mammalian macrophages. The coordination of gold to mpo induced a potentiation of the antiproliferative effect against both *Leishmania* species and *T. cruzi*, which could be associated to the inhibition of NADH fumarate reductase absent in mammalian cells [25]. The antiprotozoal activity and proposed or supposed targets of gold complexes confirms the strong interest to develop and study new gold-based molecules as antileishmanial compounds. Within this framework, the next step of our work will be the pharmacomodulation of our lead compound **6** by fine tuning either the non-functionalized group (alkyl or aryl) or/and the quinoline entity, in order to improve both activity and selectivity in particular toward intracellular amastigotes forms of *Leishmania*, and afterwards to understand the mechanism(s) of action of these new promising gold(I) functionalized NHC metallodrugs and to study their *in vivo* activity on murine model.

3. Conclusion

A new family of compounds, namely five imidazoliums salts, one silver(I) and five gold(I) complexes based on aromatic nitrogen functionalized NHCs and including six new synthesized and fully characterized ones, was screened *in vitro* for its antileishmanial activity against both the promastigote and the intramacrophagic amastigote forms of *L. infantum*. Moreover, the cytotoxicity was assessed on the murine J774A.1 macrophages, in order to determine their selectivity of action. From this studies, gold(I) NHC-quinoline and in particular a neutral complex proved to be promising metallodrugs with potent and selective

action against the pathological relevant form of *Leishmania*. By taking into account that 1) quinoline derivatives are one of the most studied families of organic compounds for leishmanicidal activity evaluation (due to their easily modulable scaffold) and 2) gold(I) complexes are good candidates for parasitic pathogens such as *Leishmania*, we aim to develop further synergetic metal-drugs based on Au(I)-NHC and quinolines or aminoquinolines derivatives, easy to set up and with high pharmacological activity and low toxicity.

4. Experimental section

4.1. General Information

Unless otherwise stated, all reactions were performed in air. All reagents were used as received from commercial suppliers. 1-Mesitylimidazole [26], 3-Methyl-1-(quinolin-2-yl)-1*H*-imidazol-3-ium hexafluorophosphate (**1'**), 3-Methyl-1-([6-(pyridin-2-yl)pyridin-3-yl]methyl)-1*H*-imidazol-3-ium bromide (**9**) and complexes **4**, **5** and **10** [20] were prepared according literature procedure. Reactions involving silver compounds were performed with the exclusion of light. ¹H (300 or 400 MHz) and ¹³C NMR spectra (75 or 101 MHz) were recorded at 298 K on Bruker AV300 or Bruker AV400 spectrometers in CDCl₃, CD₃OD, CD₃CN as solvents. Elemental analyses were carried out by the “Service de Microanalyse du Laboratoire de Chimie de Coordination (Toulouse). Mass spectrometry analysis were performed on a Nermag R1010 (FAB⁺/meta-nitrobenzylalcohol (MNBA)) spectrometer and HRMS were performed with a Thermo Finnigan MAT 95 XL spectrometer using electrospray ionization (ESI) by the “Service de Spectrométrie de Masse de Chimie UPS-CNRS” (Toulouse).

4.1.1. Preparation of imidazolium salts

Scheme 3. Numbering of H (¹H NMR) and C (¹³C NMR). These notations are used in the following experimental section.

4.1.1.1 3-Methyl-1-(quinolin-2-yl)-1*H*-imidazol-3-ium chloride (**1**)

1-Methylimidazole (3.2 mL, 40.22 mmol) and 2-chloroquinoline (1.88 g, 11.49 mmol) were stirred in toluene (6 mL) in a closed pressure tube at 130 °C for 24 h. After cooling to room temperature, the solvent was evaporated and the resulting brown paste was triturated with acetone and diethyl ether successively. The beige solid obtained was filtered and dried under

vacuum (1.455 g, 52% yield). Anal. Calcd. For $C_{13}H_{12}N_3Cl$: C, 63.55; H, 4.92; N, 17.10. Found: C, 63.38; H, 5.02; N, 17.03. 1H NMR (400 MHz, CD_3CN) δ 10.89 (s, 1H, H_2), 8.64 (d, $J = 8.8$ Hz, 1H, H_8), 8.44 (t, $J = 1.9$ Hz, 1H, H_5), 7.31 (d, $J = 8.8$, 1H, H_7), 8.11 – 8.03 (m, 2H, H_{10} , H_{13}), 7.90 (ddd, $J = 8.4$, 6.9, 1.5 Hz, 1H, H_{12}), 7.73 (ddd, $J = 8.2$, 6.9, 1.2 Hz, 1H, H_{11}), 7.67 (t, $J = 1.9$ Hz, 1H, H_4), 4.10 (s, 3H, CH_3). ^{13}C NMR (101 MHz, CD_3OD) δ 146.0 (1C, C_6), 144.8 (1C, C_{14}), 141.1 (1C, C_8), 135.5 (1C, C_2), 131.3 (1C, C_{12}), 128.3 (1C, C_{13}), 128.0 (1C, C_{10}), 127.8 (1C, C_9), 127.8 (1C, C_{11}), 124.6 (1C, C_7), 119.1 (1C, C_5), 111.4 (1C, C_4), 35.9 (1C, CH_3). MS (FAB $^+$): m/z 210 $[M - Cl]^+$.

4.1.1.2. 3-Mesityl-1-(quinolin-2-yl)-1H-imidazol-3-ium chloride (2)

1-Mesitylimidazole (211 mg, 1.132 mmol) and 2-chloroquinoline (196 mg, 1.198 mmol) were stirred in a closed pressure tube at 170 °C for 12 h. After cooling to room temperature, the crude product was dissolved in CH_2Cl_2 and precipitated with diethyl ether to afford a white solid, which was filtered and dried under vacuum (131 mg, 33% yield). Anal. Calcd. For $C_{21}H_{20}N_3Cl$: C, 72.09; H, 5.76; N, 12.01. Found: C, 72.22; H, 5.82; N, 12.08. 1H NMR (300 MHz, $CDCl_3$) δ 12.17 (t, $J = 1.5$ Hz, 1H, H_2), 9.45 (d, $J = 8.9$ Hz, 1H, H_8), 9.07 (t, $J = 1.7$ Hz, 1H, H_5), 8.60 (d, $J = 8.9$ Hz, 1H, H_7), 8.08 (dd, $J = 8.6$, 0.8 Hz, 1H, H_{13}), 7.98 (dd, $J = 8.4$, 0.8 Hz, 1H, H_{10}), 7.85 (ddd, $J = 8.6$, 6.9, 1.5 Hz, 1H, H_{12}), 7.68 (ddd, $J = 8.4$, 6.9, 1.2 Hz, 1H, H_{11}), 7.37 (t, $J = 1.7$ Hz, 1H, H_4), 7.08 (d, $J = 1.2$ Hz, 2H, H_{Mes}), 2.38 (s, 3H, CH_3), 2.25 (s, 6H, CH_3). ^{13}C NMR (101 MHz, $CDCl_3$) δ 146.0 (1C, C_6), 144.9 (1C, C_{14}), 141.8 (1C, C_8), 141.5 (1C, C_{Mes}), 137.8 (1C, C_2), 134.0 (2C, C_{Mes}), 131.3 (1C, C_{12}), 130.7 (1C, C_{Mes}), 130.0 (2C, C_{Mes}), 128.7 (1C, C_{13}), 128.5 (1C, C_9), 128.2 (1C, C_{10}), 128.1 (1C, C_{11}), 123.9 (1C, C_7), 119.9 (1C, C_5), 114.1 (1C, C_4), 21.1 (1C, C_{Mes}), 17.9 (2C, C_{Mes}). HRMS (ES $^+$): $m/z = 314.1657$ $[M - Cl]^+$.

4.1.1.3. 3-(4-Methylthiophenyl)-1-(quinolin-2-yl)-1H-imidazol-3-ium chloride (3)

1-(4-Methylthiophenyl)-1H-imidazole was synthesized according to a modified literature procedure. Imidazole (3.24 g, 47.57 mmol), 4-bromothiophenyl (4.83 g, 23.78 mmol), K_2CO_3 (3.29 g, 23.78 mmol) and a catalytic amount of $CuSO_4$ were stirred in a closed pressure tube at 205 °C for 10 h. After cooling to room temperature, the crude product was extracted with MeOH, filtered and evaporated to dryness. The resulting residue was purified by column chromatography on silica gel with a mixture of CH_2Cl_2 and methanol (90:10) as eluent to give 1-(4-methylthiophenyl)-1H-imidazole as a yellow solid (3.99 g, 88% yield). 1H NMR (300 MHz, $CDCl_3$) δ 7.93 (s, 1H), 7.40 – 7.31 (m, 4H), 7.28 (s, 1H), 7.24 (s, 1H), 2.54 (s, 3H) [25]. 1-(4-Methylthiophenyl)-1H-imidazole (500 mg, 26.28 mmol) and 2-chloroquinoline (456 mg,

27.86 mmol) were stirred in a closed pressure tube at 170 °C for 12 h. After cooling to room temperature, the white solid was triturated with CH₂Cl₂, filtered, and dried under vacuum. (729 mg, 86% yield). Anal. Calcd. For C₁₉H₁₆N₃ClS: C, 64.49; H, 4.56; N, 11.87. Found: C, 64.30; H, 4.62; N, 11.88. ¹H NMR (300 MHz, DMSO-*d*₆) δ 11.02 (s, 1H, *H*₂), 8.95 – 8.82 (m, 2H, *H*₅, *H*₈), 8.65 (t, *J* = 2.0 Hz, 1H, *H*₄), 8.49 (d, *J* = 8.9 Hz, 1H, *H*₇), 8.23 – 8.07 (m, 2H, *H*₁₀, *H*₁₃), 8.06 – 7.90 (m, 3H, *H*_{SMePh}, *H*₁₂), 7.78 (ddd, *J* = 8.2, 6.9, 1.2 Hz, 1H, *H*₁₁), 7.56 (d, 2H, *J* = 8.7 Hz, 2H, *H*_{SMePh}), 2.59 (s, 3H, *H*_{SMePh}). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 145.9 (1C, *C*₆), 145.7 (1C, *C*₁₄), 141.8 (1C, *C*_{SMePh}), 141.7 (1C, *C*₈), 135.2 (1C, *C*₂), 132.2 (1C, *C*₁₂), 131.8 (1C, *C*_{SMePh}), 128.8 (1C, *C*₁₃), 128.8 (1C, *C*₁₀), 128.5 (1C, *C*₉), 128.3 (1C, *C*₁₁), 127.1 (2C, *C*_{SMePh}), 123.0 (2C, *C*_{SMePh}), 122.6 (1C, *C*₇), 120.4 (1C, *C*₅), 113.6 (1C, *C*₄), 15.0 (1C, *C*_{SMePh}). HRMS (ES⁺): *m/z* = 318.1060 [*M* - Cl]⁺.

4.1.2. Preparation of complexes

4.1.2.1. Complex 6

Under a nitrogen atmosphere and protection of the light, a Schlenk tube was charged with **1** (76.7 mg, 0.312 mmol) in 6 ml of acetonitrile and Ag₂O (72.3 mg, 0.312 mmol). The reaction mixture was stirred at 40 °C for 24 h. After that Au(SMe₂)Cl (110 mg, 0.374 mmol) was added and the mixture was stirred for 4 h. The solution was filtered through a pad of celite and the solvent removed under reduced pressure to give a pale yellow solid (0.117 g, 85% yield). Crystals suitable for X-ray diffraction analysis were obtained by slow evaporation of an acetonitrile solution of **6**. Anal. Calcd. For C₁₃H₁₁N₃ClAu: C, 35.35; H, 2.51; N, 9.51. Found: C, 35.22; H, 2.72; N, 9.48. ¹H NMR (400 MHz, CD₃CN) δ 8.55 (d, *J* = 8.7 Hz, 1H, *H*₇), 8.47 (d, *J* = 8.7 Hz, 1H, *H*₈), 8.14 – 8.03 (m, 2H, *H*₁₀, *H*₁₃), 7.93 (d, *J* = 2.0 Hz, 1H, *H*₅), 7.88 (ddd, *J* = 8.5, 6.9, 1.5 Hz, 1H, *H*₁₂), 7.71 (ddd, *J* = 8.1, 6.9, 1.2 Hz, 1H, *H*₁₁), 7.39 (d, *J* = 2.0 Hz, 1H, *H*₄), 3.97 (s, 3H, CH₃). ¹³C NMR (101 MHz, CD₃CN) δ 170.1 (1C, *C*₂), 149.55 (1C, *C*₆), 146.41 (1C, *C*₁₄), 139.43 (1C, *C*₈), 130.99 (1C, *C*₁₂), 128.60 (1C, *C*₁₃), 128.05 (1C, *C*₁₀), 127.80 (1C, *C*₉), 127.45 (1C, *C*₁₁), 123.18 (1C, *C*₇), 120.56 (1C, *C*₅), 116.10 (1C, *C*₄), 38.77 (1C, CH₃). HRMS (ES⁺): *m/z* = 442.0385 [*M* + H]⁺.

4.1.2.2. Complex 7

Under a nitrogen atmosphere, sodium acetate (31 mg, 0.377 mmol) was added to a mixture of **2** (110 mg, 0.314 mmol) and Au(SMe₂)Cl (46.3 mg, 0.157 mmol) in dry DMF (3 mL) at 100 °C. The mixture was then heated to 120 °C and this temperature was maintained for 2 h. After cooling to room temperature, CH₂Cl₂ (1 mL) and Et₂O (50 mL) were added and the solution was kept at -4 °C for 2 h to afford a white solid. After filtration, complex **7** was dried

under vacuum (100 mg, 74% yield). Crystals suitable for X-ray diffraction analysis were obtained by slow evaporation of a methanolic solution of **7**. Anal. Calcd. For $C_{42}H_{38}N_6ClAu$: C, 58.71; H, 4.46; N, 9.78. Found: C, 58.62; H, 4.42; N, 9.63. 1H NMR (300 MHz, CD_3OD) δ 8.29 (d, $J = 2.0$ Hz, 1H, H_5), 8.25 (d, $J = 8.6$ Hz, 1H, H_8), 8.04 – 7.88 (m, 3H, H_7 , H_{10} , H_{13}), 7.83 – 7.74 (m, 1H, H_{12}), 7.66 (ddd, $J = 8.2$, 6.9, 1.2 Hz, 1H, H_{11}), 7.52 (d, $J = 2.0$ Hz, 1H, H_4), 6.91 (d, $J = 1.2$ Hz, 2H, H_{Mes}), 2.25 (s, 3H, H_{Mes}), 1.96 (s, 6H, H_{Mes}). ^{13}C NMR (101 MHz, CD_3OD) δ 181.4 (1C, C_2), 147.4 (1C, C_6), 144.7 (1C, C_{14}), 138.1 (1C, C_8), 138.0 (1C, C_{Mes}), 133.5 (1C, C_{Mes}), 132.9 (2C, C_{Mes}), 129.4 (1C, C_{12}), 127.4 (2C, C_{Mes}), 126.7 (1C, C_{13}), 126.3 (1C, C_9), 126.2 (1C, C_{10}), 125.9 (1C, C_{11}), 122.4 (1C, C_7), 119.8 (1C, C_5), 113.5 (1C, C_4), 18.3 (1C, C_{Mes}), 14.9 (2C, C_{Mes}). HRMS (ES^+): $m/z = 831.1658 [M - H]^+$.

4.1.2.3. Complex **8**

Under a nitrogen atmosphere, sodium acetate (32.83 mg, 0.400 mmol) was added to a mixture of **3** (118 mg, 0.334 mmol) and $Au(SMe_2)Cl$ (49 mg, 0.167 mmol) in dry DMF (3 mL) at 100 °C. The mixture was then heated to 120 °C and this temperature was maintained for 2 h. After cooling to room temperature, CH_2Cl_2 (1 mL) and Et_2O (10 mL) were added and the solution was kept at -4 °C for 2 h to afford a beige solid. After filtration, complex **8** was dried under vacuum (99 mg, 74% yield). Anal. Calcd. For $C_{38}H_{30}N_6ClS_2Au$: C, 52.63; H, 3.49; N, 9.69. Found: C, 52.72; H, 3.62; N, 9.74. 1H NMR (400 MHz, CD_3OD) δ 8.22 (d, $J = 2.0$ Hz, 1H, H_5), 8.14 – 8.04 (m, 2H, H_8 , H_{13}), 7.90 (d, $J = 8.5$ Hz, 1H, H_7), 7.86 – 7.80 (m, 2H, H_{10} , H_4), 7.80 – 7.72 (m, 1H, H_{12}), 7.66 – 7.56 (m, 3H, H_{SMePh} , H_{11}), 7.13 (d, $J = 8.6$ Hz, 2H, H_{SMePh}), 2.46 (s, 1H, H_{SMePh}). ^{13}C NMR (101 MHz, CD_3OD) δ 180.8 (1C, C_2), 149.2 (1C, C_6), 146.2 (1C, C_{14}), 141.0 (1C, C_{SMePh}), 139.6 (1C, C_8), 136.0 (1C, C_{SMePh}), 131.2 (1C, C_{12}), 128.4 (1C, C_{13}), 128.0 (1C, C_{10}), 127.8 (1C, C_9), 127.7 (1C, C_{11}), 126.0 (2C, C_{SMePh}), 125.4 (2C, C_{SMePh}), 123.8 (1C, C_7), 122.0 (1C, C_5), 115.9 (1C, C_4), 14.1 (1C, C_{SMePh}). HRMS (ES^+): $m/z = 823.2840 [M - H]^+$.

4.1.3. Crystallographic data for **4-6**

All data were collected at low temperature using oil-coated shock-cooled crystals on a Bruker-AXS APEX II diffractometer with $MoK\alpha$ radiation ($\lambda = 0.71073$ Å). The structures were solved by direct methods [28] and all non hydrogen atoms were refined anisotropically using the least-squares method on F^2 [29].

6: $C_{13}H_{11}AuClN_3$, Mr = 441.66, crystal size = 0.40 x 0.30 x 0.30 mm³, monoclinic, space group $P2_1/n$, $a = 3.983(2)$ Å, $b = 12.613(4)$ Å, $c = 24.744(8)$ Å, $\beta = 91.35(2)^\circ$, $V = 1242.8(7)$

\AA^3 , $Z = 4$, $T = 173(2)$ K, 9449 reflections collected, 1784 unique reflections ($R_{\text{int}} = 0.1715$), $R1 = 0.0659$, $wR2 = 0.1471$ [$I > 2\sigma(I)$], $R1 = 0.1213$, $wR2 = 0.1732$ (all data), residual electron density = 3.189 e \AA^{-3} .

7: $\text{C}_{42}\text{H}_{39}\text{AuCl}_2\text{N}_6$, $M_r = 895.66$, crystal size = $0.20 \times 0.05 \times 0.05 \text{ mm}^3$, orthorhombic, space group $P2_12_12_1$, $a = 15.1793(4) \text{ \AA}$, $b = 17.4742(5) \text{ \AA}$, $c = 29.2211(8) \text{ \AA}$, $V = 7750.8(4) \text{ \AA}^3$, $Z = 8$, $T = 173(2)$ K, 76886 reflections collected, 14933 unique reflections ($R_{\text{int}} = 0.0439$), $R1 = 0.0302$, $wR2 = 0.0533$ [$I > 2\sigma(I)$], $R1 = 0.0466$, $wR2 = 0.0573$ (all data), residual electron density = 1.516 e \AA^{-3} .

CCDC-1040256 (**6**) and CCDC-1040257 (**7**) contain the supplementary crystallographic data. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

4.1.4. Antileishmanial evaluation

Leishmania species used in this study were *Leishmania infantum* MHOM/MA/67/ITMAP-263 (CNR *Leishmania*, Montpellier, France) expressing luciferase activity.

4.1.4.1. Antileishmanial activity on promastigotes

The effects of the tested compounds on the growth of *Leishmania infantum* promastigotes were assessed by Luciferase Assay. Briefly, promastigotes in log-phase in RPMI 1640 medium supplemented with 10% fetal calf serum (FCS), 2 mM L-glutamine and antibiotics (100U/mL penicillin, 100 $\mu\text{g/mL}$ streptomycin and 50 $\mu\text{g/mL}$ geneticin), were incubated at an average density of 10^6 parasites/mL in sterile 96-well plates with various concentrations of compounds dissolved in DMSO (final concentration less than 0.5% v/v), in duplicate. Appropriate controls treated by DMSO, amphotericin B, miltefosine and pentamidine (reference drugs purchased from Sigma Aldrich) were added to each set of experiments. After a 72 h incubation period at 24 °C, each plate-well was then microscope-examined for detecting possible precipitate formation. To estimate the luciferase activity of promastigotes, 80 μl of each well are transferred in white 96-well plates, Steady Glow reagent (Promega) was added according to manufacturer's instructions, and plates were incubated for 2 min.. The luminescence was measured in Microbeta Luminescence Counter (PerkinElmer). Inhibitory concentration 50% (IC_{50}) was defined as the concentration of drug required to inhibit by 50% the metabolic activity of *Leishmania infantum* promastigotes compared to the control. IC_{50} were calculated by non-linear regression analysis processed on dose-response curves, using

TableCurve 2D V5 software. IC₅₀ values represent the mean value calculated from three independent experiments.

4.1.4.2. Antileishmanial activity on intracellular amastigotes

The effects of the tested compounds on the growth of *L. infantum* intracellular amastigotes were assessed in the following way. 100 µL of J774A.1 cells were seeded 96-well plates at an average density of 1.10^5 cells/mL and incubated for 24 h at 37 °C and 5% CO₂. *L. Infantum* promastigotes were centrifuged at 900 g for 10 min and the supernatant replaced by the same volume of RPMI 1640 complete medium at pH 5.4 and incubated for 24 h at 27 °C. Then, half of the macrophages medium was removed, J774A.1 cells were then infected by 100µl of acidified promastigotes at an average density of 1.10^6 cells/mL (10:1 ratio) and plates incubated for 24 h at 37 °C and 5% CO₂. Half of the infected macrophages medium was removed and medium containing various concentrations of tested-compounds was added (final DMSO concentration being inferior to 0.5% v/v) in duplicate. Appropriate controls treated with or without solvent (DMSO), and various concentrations of amphotericin B, miltefosine and pentamidine were added to each set of experiments. After 120 h incubation at 37 °C and 5% CO₂, 100 µl of well supernatant was removed and Steady Glow reagent (Promega) was added according to manufacturer's instructions. Plates were incubated for 3 min. 100 µL of each well were transferred in white 96-well plates and the luminescence was measured in Microbeta Luminescence Counter (PerkinElmer). Inhibitory concentration 50% (IC₅₀) was defined as the concentration of drug required to inhibit by 50% the metabolic activity of *L. Infantum* intracellular amastigotes compared to the control. IC₅₀ were calculated by non-linear regression analysis processed on dose-response curves, using TableCurve 2D V5 software. IC₅₀ values represent the mean value calculated from three independent experiments.

4.1.5 Cytotoxicity evaluation

The evaluation of the tested molecules cytotoxicity by MTT assay on the J774A.1 cell line (mouse macrophage cell line, Sigma-Aldrich) was done according to Mosmann with slight modifications [30]. Briefly, cells (5.10^4 cells/mL) in 100 µL of complete medium, [RPMI 1640 supplemented with 10% fetal calf serum (FCS), 2 mM L-glutamine and antibiotics (100U/mL penicillin and 100 µg/mL streptomycin)] were seeded into each well of 96-well plates and incubated at 37 °C and 5% CO₂. After a 24 h incubation, 100 µL of medium with various product concentrations and appropriate controls were added and the plates were incubated for 72 h at 37

°C and 5% CO₂. Each plate-well was then microscope-examined for detecting possible precipitate formation before the medium was aspirated from the wells. 100 µL of MTT solution (0.5 mg/mL in RPMI) were then added to each well. Cells were incubated for 2 h at 37 °C and 5% CO₂. After this time, the MTT solution was removed and DMSO (100 µL) was added to dissolve the resulting formazan crystals. Plates were shaken vigorously (300 rpm) for 5 min. The absorbance was measured at 570 nm with a microplate spectrophotometer (Eon BioTek). DMSO was used as blank and doxorubicin (purchased from Sigma Aldrich) as positive control. CC₅₀ were calculated by non-linear regression analysis processed on dose–response curves, using TableCurve 2D V5 software. CC₅₀ values represent the mean value calculated from three independent experiments.

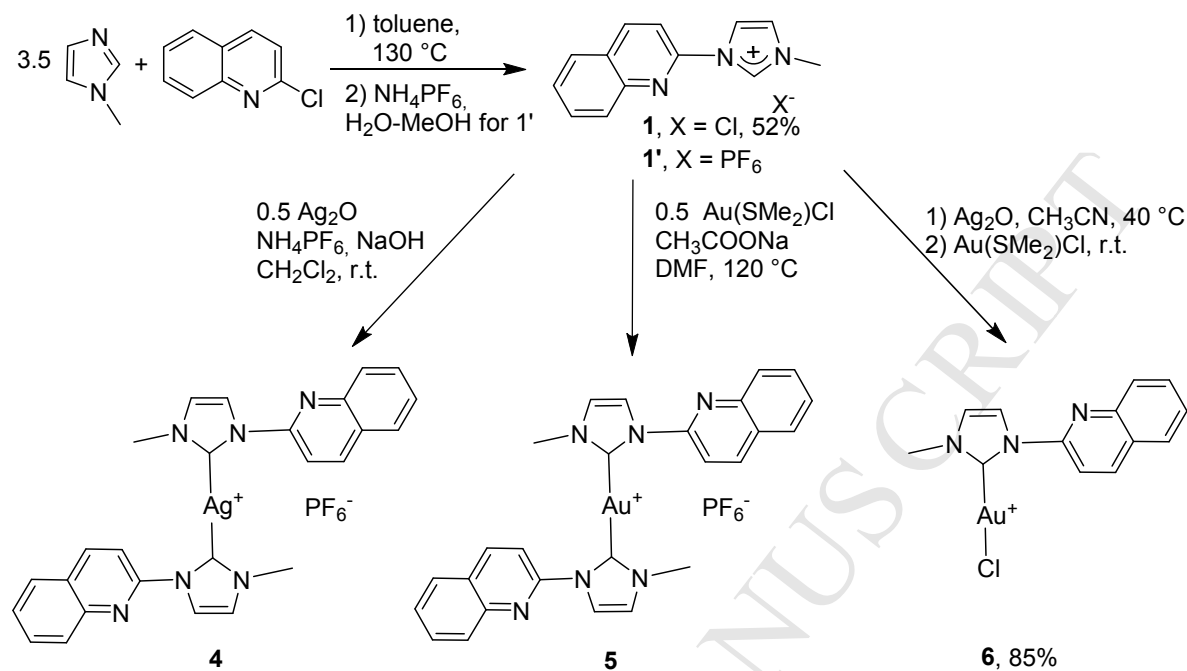
Table 1. *In vitro* antileishmanial activity and cytotoxicity of compounds 1-9.

References

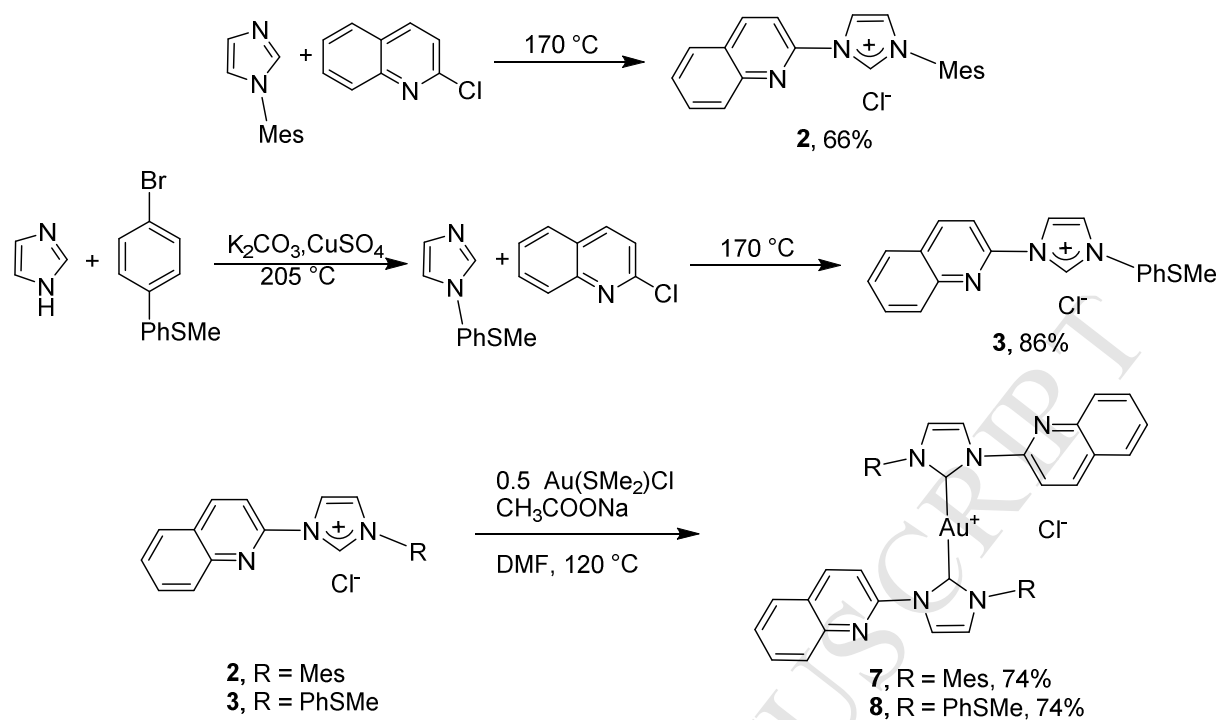
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List of Schemes, Figures and Table.



Scheme 1. Synthesis of imidazolium salts **1** and **1'** and silver(I) bis(NHC) (**4**), gold(I) bis(NHC) (**5**) and gold(I) NHC (**6**) complexes.



Scheme 2. Synthesis of imidazoliums salts **2** and **3** and gold(I) bis (NHC) complexes **7** and **8**.

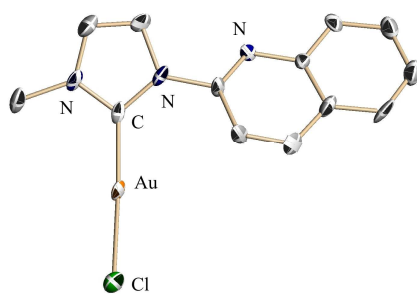


Figure 1: Crystal structure of **6** depicted at 50% level. Hydrogen atoms have been omitted for clarity. Selected bond lengths [Å] and angles [°]: Au-C 1.96(2), Au-Cl 2.28(1), C-Au-Cl 177.3(6).

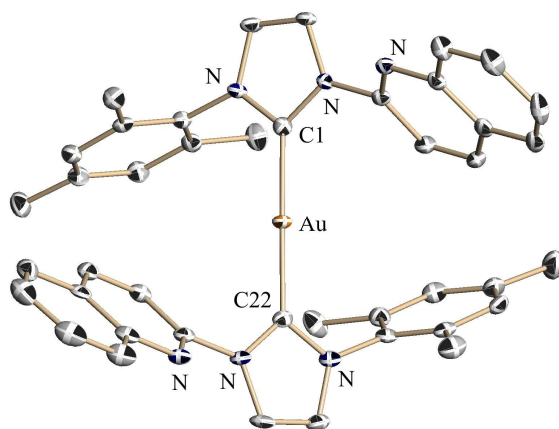


Figure 2: Crystal structure of the cationic part of **7** depicted at 30% level. Non-coordinating anions and hydrogen atoms have been omitted for clarity. Selected bond lengths [Å] and angles [°]: Au-C1 1.997(6), Au-C22 2.016(6), C1-Au-C22 178.9(2).

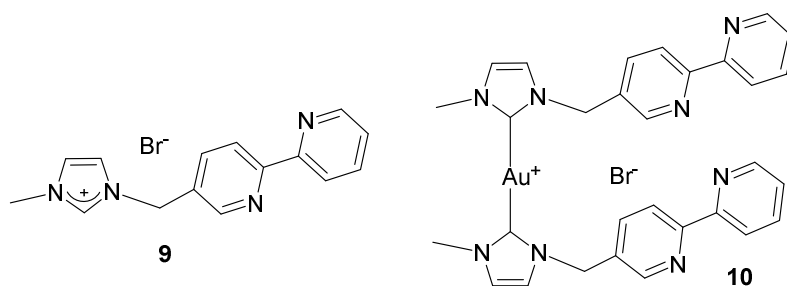
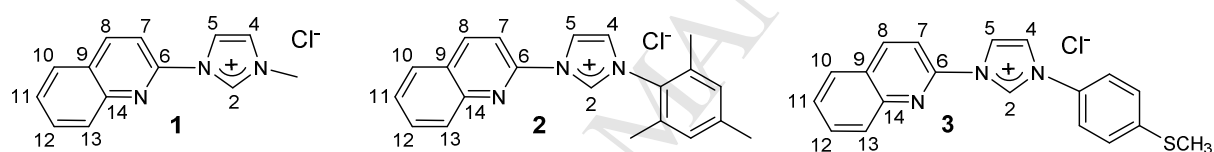


Figure 3. Compounds **9** and **10** also used in this study.



Scheme 3. Numbering of H (^1H NMR) and C (^{13}C NMR). These notations are used in the following experimental section.

Table 1. *In vitro* antileishmanial activity and cytotoxicity of compounds **1-9**.

Compound	<i>L. infantum</i> promastigotes	J774A.1	Selectivity index Pro	<i>L. infantum</i> intracellular amastigotes	Selectivity index Ama
	IC ₅₀ (μM) ^a	CC ₅₀ (μM) ^a	CC ₅₀ / IC ₅₀ (Pro)	IC ₅₀ (μM) ^a	CC ₅₀ / IC ₅₀ (Ama)
1	82.82 ± 18.46	48.36 ± 16.72	0.58	-	-
1'	> 100	-	-	-	-
2	9.81 ± 1.54	0.92 ± 0.08	0.09	-	-
3	11.96 ± 4.22	5.17 ± 0.86	0.43	-	-
4	9.37 ± 0.69	15.13 ± 2.37	1.61	-	-
5	0.39 ± 0.14	2.06 ± 0.16	5.33	0.40 ± 0.23	5.19
6	1.53 ± 0.62	9.45 ± 0.42	6.19	0.96 ± 0.55	9.84
7	0.42 ± 0.13	0.26 ± 0.08	0.61	-	-
8	0.43 ± 0.26	1.29 ± 0.45	3.02	0.24 ± 0.06	5.45
9	> 100	-	-	-	-
10	1.86 ± 0.22	2.89 ± 0.83	1.56	-	-
Amphotericin B^b	0.04 ± 0.03	3.28 ± 1.41	79.92	0.07 ± 0.03	49.86
Miltefosine^b	8.83 ± 2.77	155.63 ± 15.25	17.63	4.17 ± 2.58	37.32
Pentamidine^b	0.51 ± 0.28	0.53 ± 0.57	1.04	-	-
Doxorubicin^a	-	0.08 ± 0.03	-	-	-

^a Mean of three independent experiments^b Amphotericin B, Miltefosine and Pentamidine were used as antileishmanial compounds of reference.^c Doxorubicin was used as positive control of cytotoxicity.

Highlights

Gold(I) *N*-heterocyclic carbene-quinoline complexes as anti-leishmanial agents.

Potent activity *in vitro* against promastigote and amastigote stages of *L. infantum*.

IC₅₀ values in the micro- and submicromolar range.

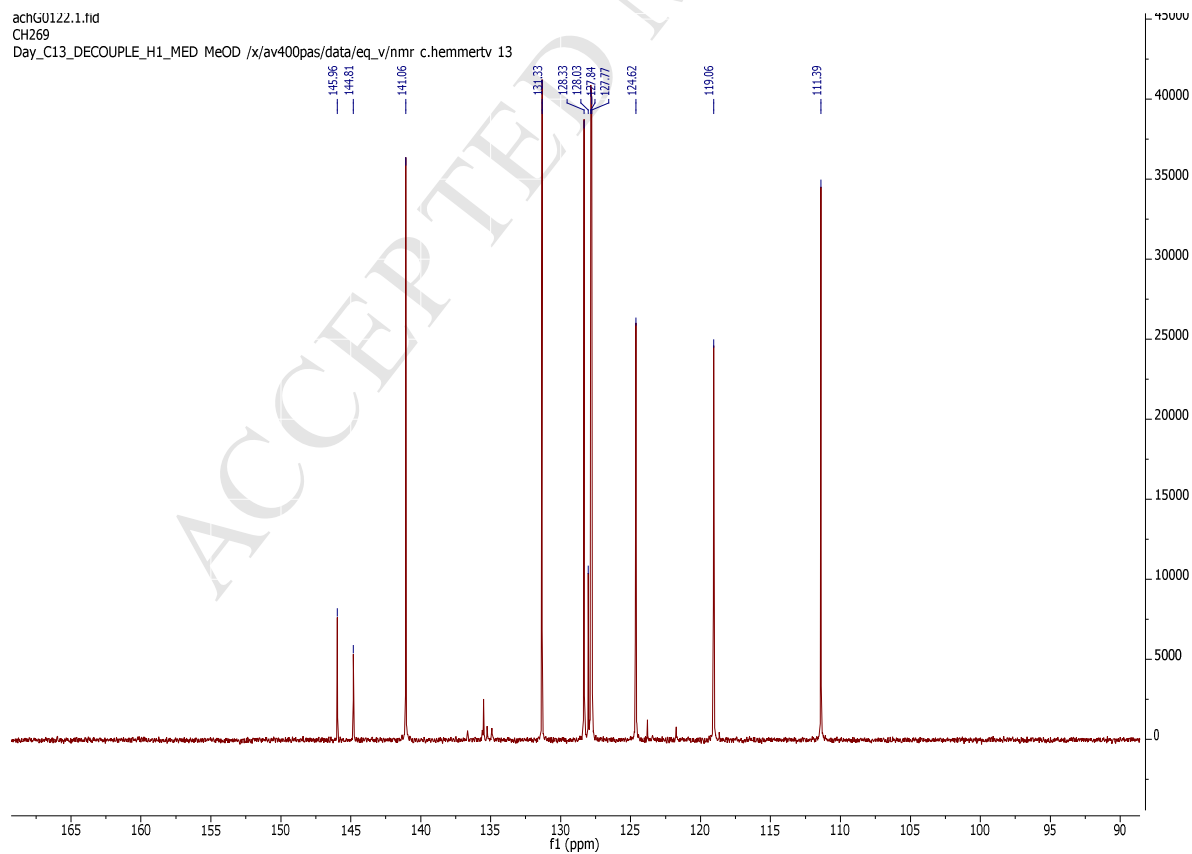
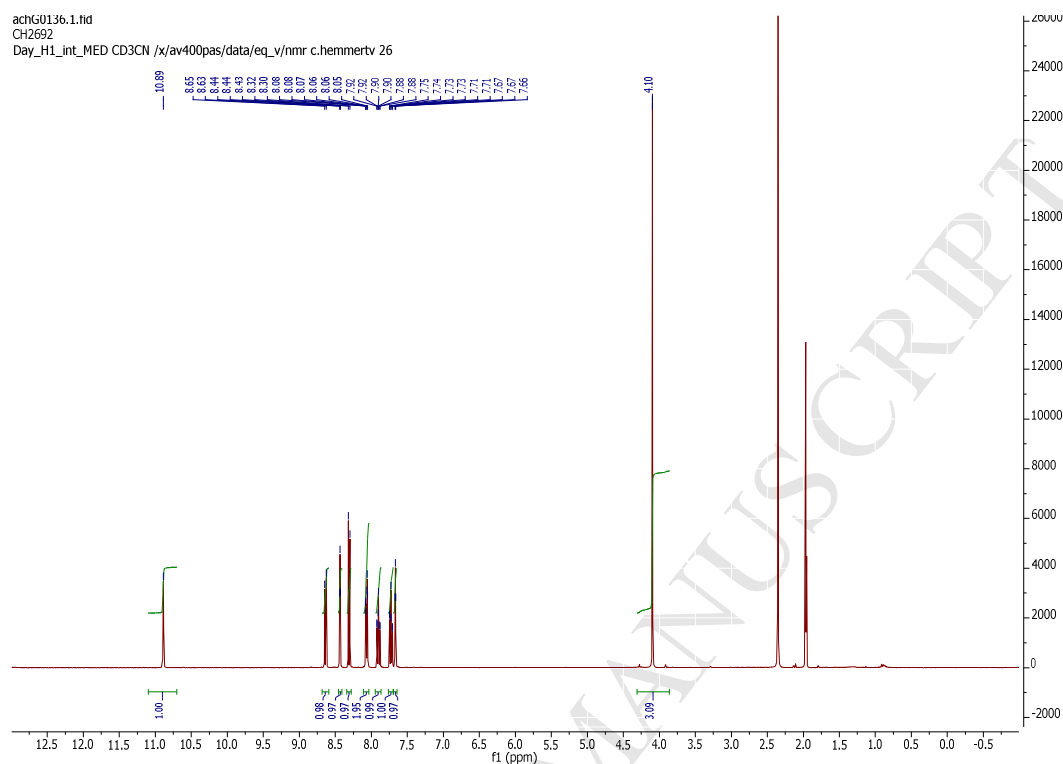
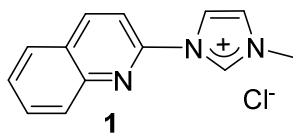
The most promising complex **6** shows an IC₅₀ = 0.96 μM (amastigotes) with SI = 9.84.

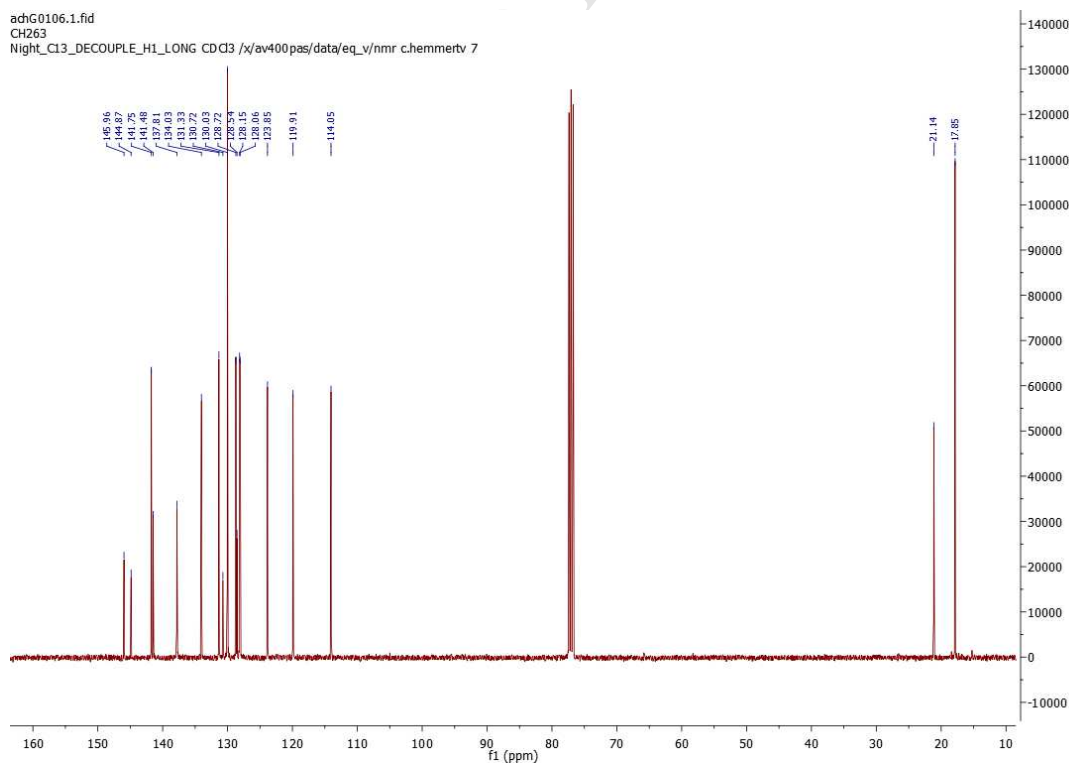
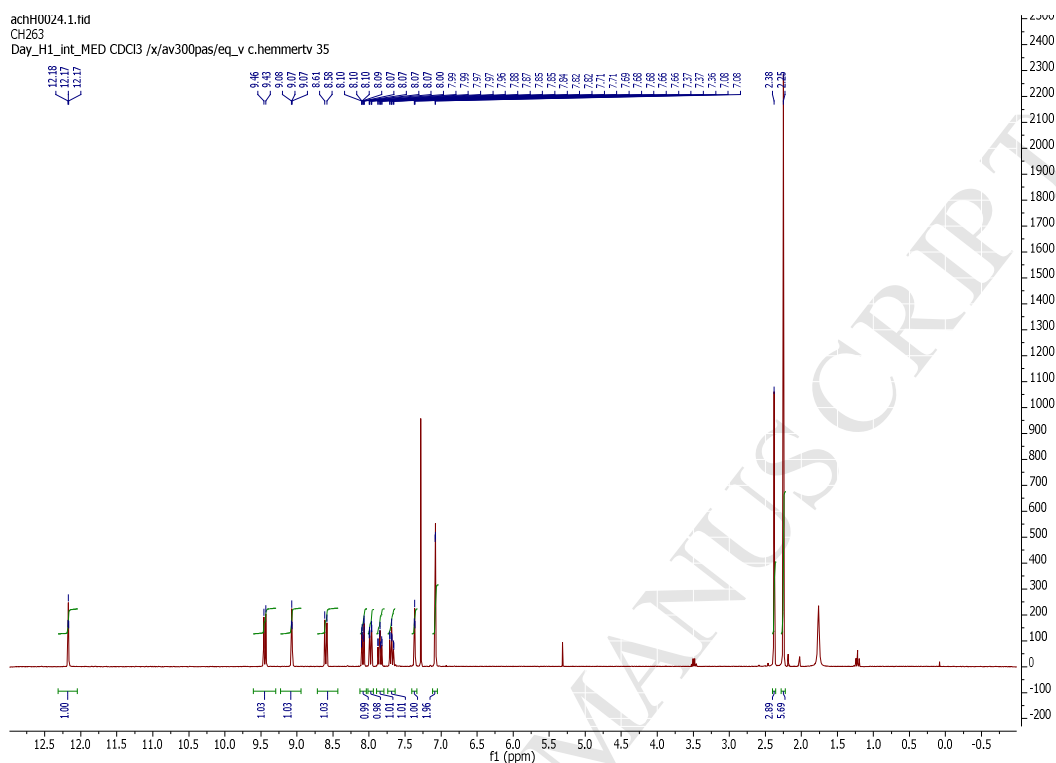
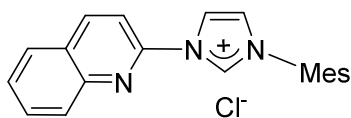
**Synthesis, characterization, and antileishmanial activities of gold(I)
complexes involving quinoline functionalized *N*-heterocyclic carbenes**

Lucie Paloque, Catherine Hemmert, Alexis Valentin, Heinz Gornitzka

¹H and ¹³C NMR spectra of proligands **1-3** and gold(I) complexes **6-8**

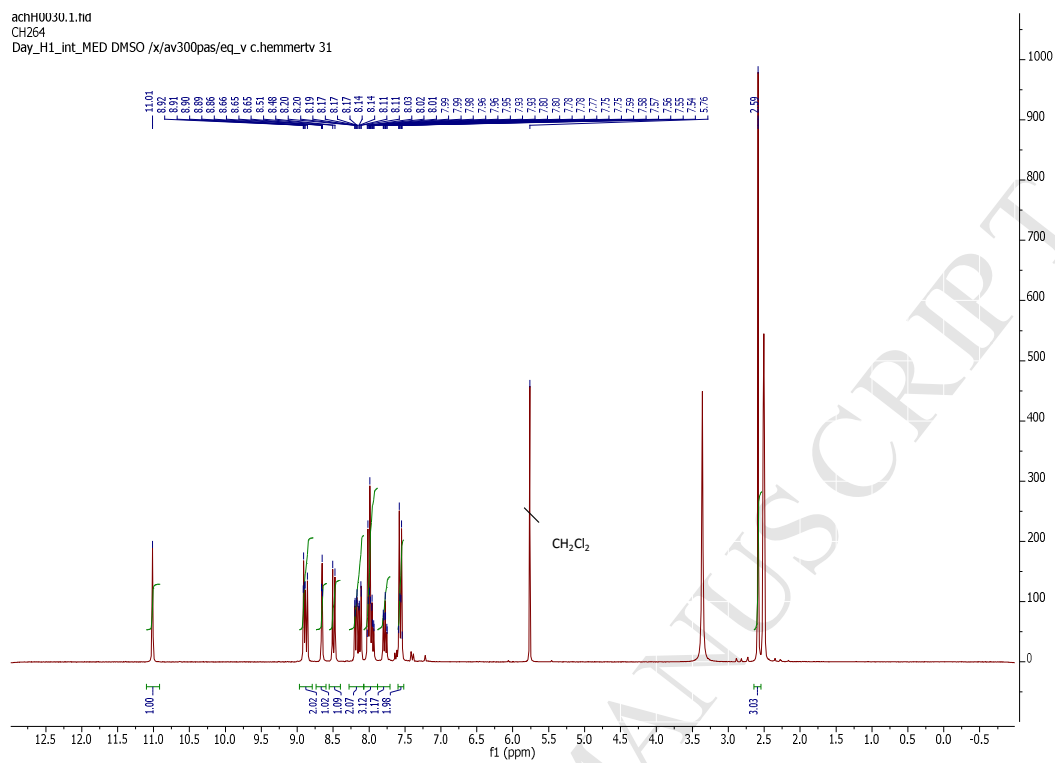
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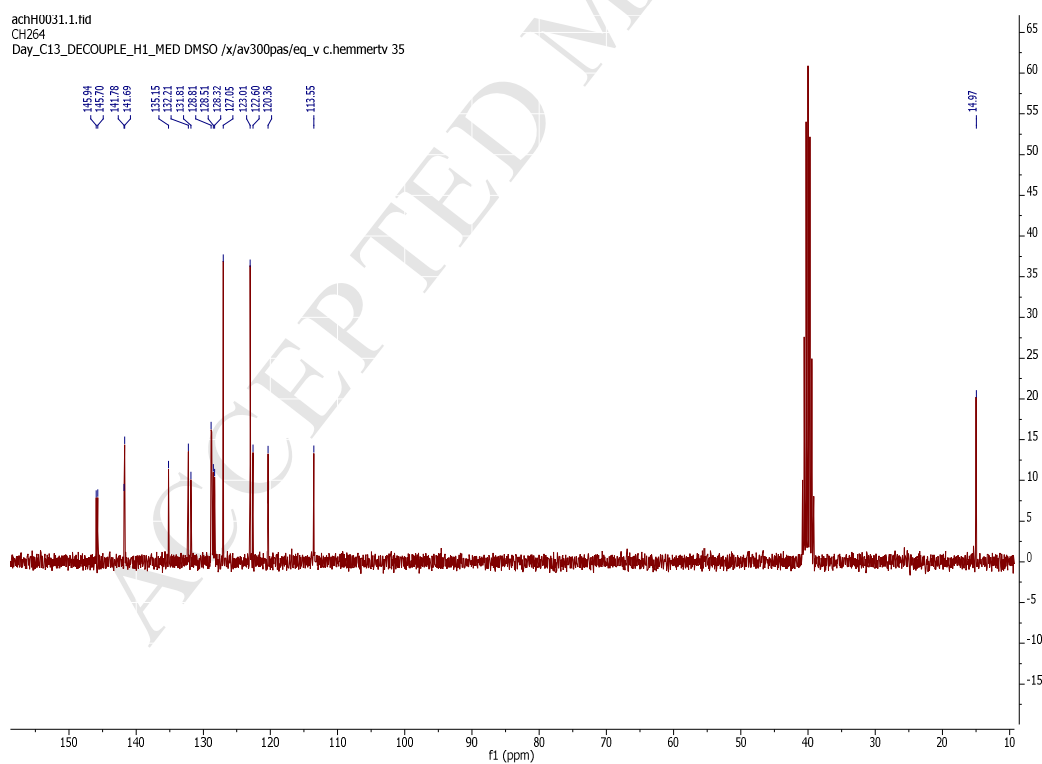


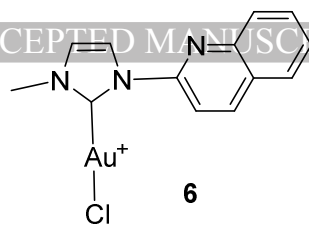
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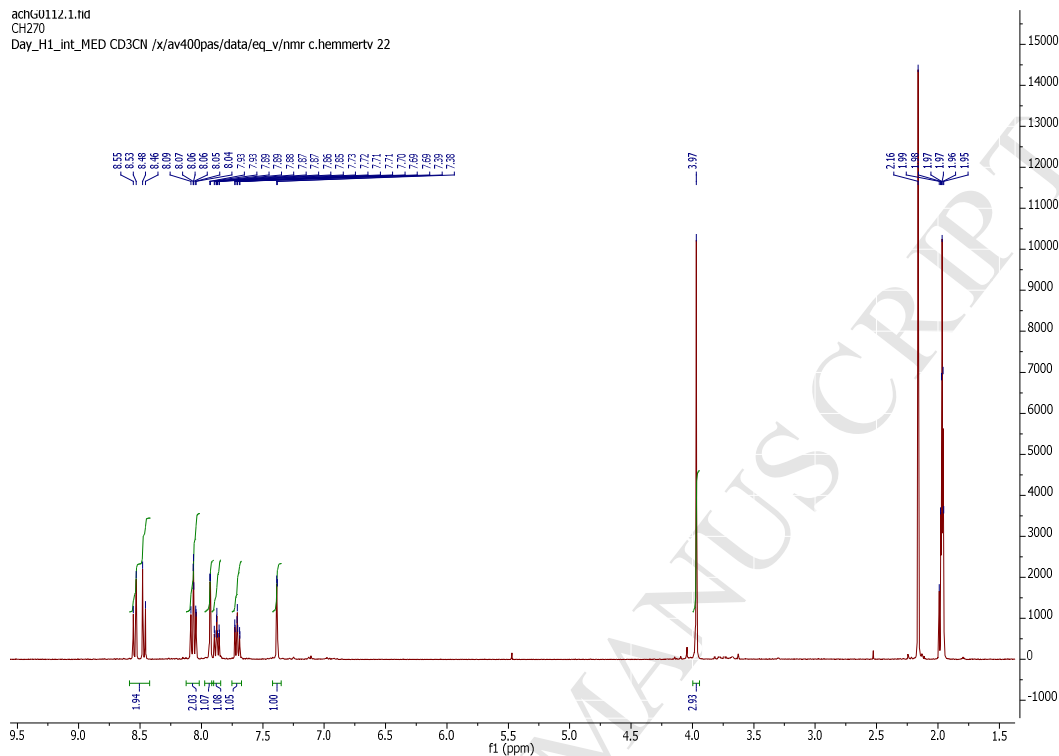
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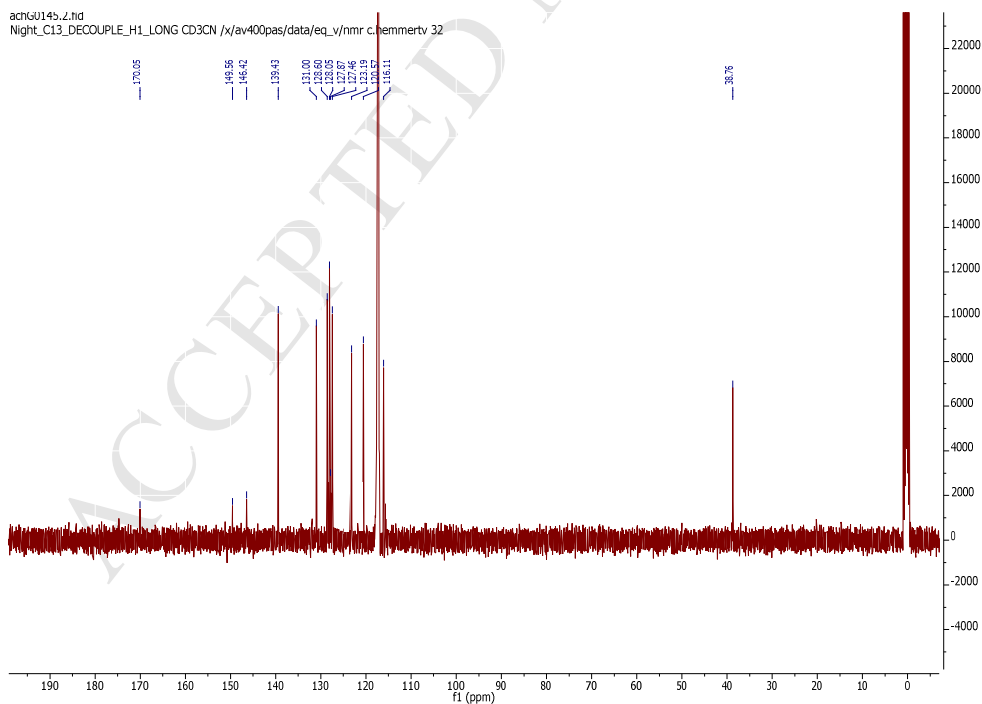
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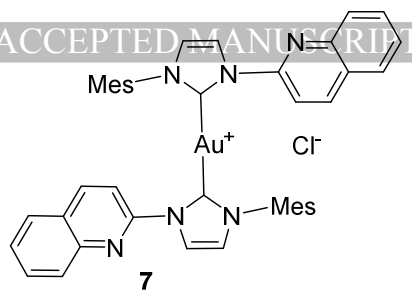
Day_H1_int_MED CD3CN /x/av400pas/data/eq_v/nmr c.hemmertv 22



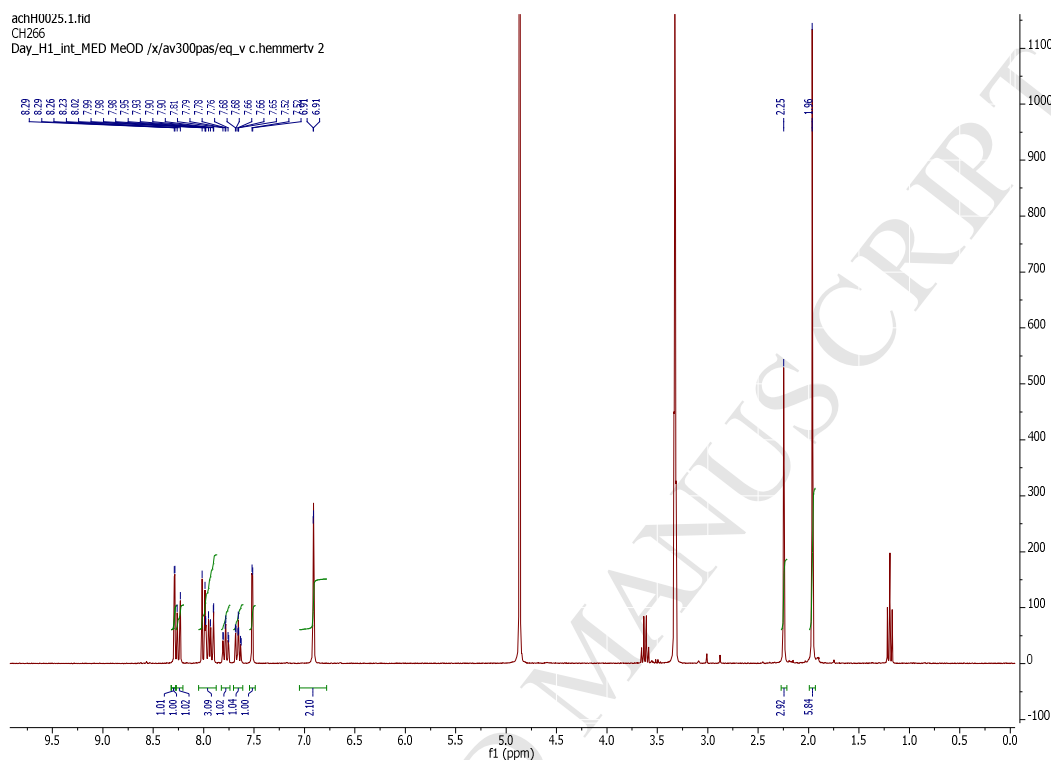
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Night_C13_DECOUPLE_H1_LONG CD3CN /x/av400pas/data/eq_v/nmr c.hemmertv 32





achH0025.1.tid
CH266
Day_H1_int_MED MeOD /x/av300pas/eq_v c.hemmerv 2



achG0118.1.tid
CH266
Day_C13_DECOUPLE_H1_MED MeOD /x/av400pas/data/eq_v/nmr c.hemmerv 7

