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Structure-activity relationships of 2-substituted phenyl-*N*-phenyl-2-oxoacetohydrazonoyl cyanides as novel antagonists of exchange proteins directly activated by cAMP (EPACs)

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ABSTRACT

Exchange proteins directly activated by cAMP (EPACs) are critical cAMP-dependent signaling pathway mediators that play important roles in cancer, diabetes, heart failure, inflammations, infections, neurological disorders and other human diseases. EPAC specific modulators are urgently needed to explore EPAC's physiological function, mechanism of action and therapeutic applications. On the basis of a previously identified EPAC specific inhibitor hit ESI-09, herein we have designed and synthesized a novel series of 2-substituted phenyl-*N*-phenyl-2-oxoacetohydrazonoyl cyanides as potent EPAC inhibitors. Compound **31** (ZL0524) has been discovered as the most potent EPAC inhibitor with IC₅₀ values of 3.6 μM and 1.2 μM against EPAC1 and EPAC2, respectively. Molecular docking of **31** onto an active EPAC2 structure predicts that **31** occupies the hydrophobic pocket in cAMP binding domain (CBD) and also opens up new space leading to the solvent region. These findings provide inspirations for discovering next generation of EPAC inhibitors.

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Cyclic adenosine monophosphate (cAMP, **1**), generated from adenosine triphosphate (ATP) by adenyl cyclase, is a second messenger for intracellular signal transduction in many different organisms. cAMP-mediated signaling events were considered to be transduced largely by protein kinase A (PKA) until the discovery of exchange proteins directly activated by cAMP (EPACs)/cAMP regulated guanine nucleotide exchange factor (cAMP-GEF).¹⁻⁴ These two intracellular receptor families mediated the major physiological effects of cAMP in mammalian cells through the cAMP binding domain (CBD) which acts as a molecular switch for controlling various cellular activities.^{5,6} The identification of EPACs opens new avenues for cAMP signaling research.⁷ Unlike PKA, EPAC proteins do not have kinase activity and activate the Ras superfamily small GTPases Rap1 and Rap2 in response to the generation of intracellular cAMP. There are two isoforms of mammalian EPACs, EPAC1 which is more ubiquitously expressed and EPAC2 which is

mainly found in CNS, pancreatic islets and adrenal gland.² EPAC1 and EPAC2 are structurally homologous but functionally nonredundant. A number of studies have revealed that EPAC proteins are critically involved in a variety of human diseases such as cancer, inflammation, bacterial and viral infections, central nervous system disorders, energy homeostasis and obesity, and cardiac functions.⁸⁻¹⁴

Given the physiological and pathophysiological significance of EPAC proteins, the development of pharmacological EPAC modulators is needed.¹⁵ Most EPAC agonists are derivatives of cAMP,¹⁶ while non-cyclic nucleotide ligands usually display EPAC inhibitory activities, except a very recently reported small molecule partial agonist with modest potency.¹⁷ Cheng, et al developed a sensitive and robust fluorescence-based high throughput (HTS) assay,¹⁸ which led to the discovery of a series of non-cyclic nucleotide EPAC inhibitors.¹⁹ After extensive modifications by our team, dihydropyrimidine **2** (HJC0198, Fig. 1) was obtained with an IC₅₀ value of 4.0 μM against EPAC2. It selectively blocks cAMP-induced EPAC activation without

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affecting cAMP-mediated PKA activation at the concentration of 25 μM .²⁰ Diphenylamine **3** (MAY0132, Fig. 1), also developed by our team, exhibited potent and selective EPAC2 inhibitory activity with an IC_{50} value of 0.4 μM and inhibited cAMP-mediated EPAC2 GEF activity with IC_{50} of 1 μM , while displaying no significant inhibition against EPAC1 at 100 μM .^{21,22} Subtype selective compound will not only help understanding diverse functions of EPACs but also may contribute to therapeutic safety in the future, thus further modifications on compound **3**, a good EPAC2 selective inhibitor, are undergoing in our lab. Compound **4** ((*R*)-CE3F4, Fig. 1), identified by Courileau and coworkers via HTS screening,²³ displayed EPAC1 inhibitory activity with an IC_{50} value of 4.2 μM with 10-fold selectivity over EPAC2.²⁴ In our assays, its inhibitory activities against EPAC1 and EPAC2 are 5.5 and 17 μM , respectively. Compound **5** (ESI-09, Fig. 1) is another promising hit discovered by our team, displaying micromolar inhibitory activities against both EPAC1 and EPAC2.²⁵⁻²⁷ Herein, we describe the discovery of a series of novel EPAC inhibitors such as compound **31**, which displays enhanced potency against EPAC proteins in comparison with chemical lead **5**.

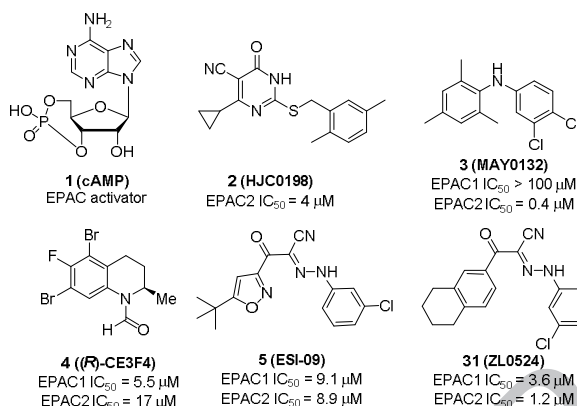


Fig. 1. The structures of cAMP and representative EPAC antagonists including newly identified **31** (ZL0524) reported in this paper.

While previously reported optimizations on compound **5** focused on the phenyl ring A (Fig. 2) and substituents on isoxazole ring,²⁸ other alternatives to replace isoxazole ring were never explored. Herein, we propose to replace the isoxazole with its bioisosteric substituted phenyl ring B (Fig. 2) to explore new chemical space and improve EPAC inhibitory potency.²⁹ Docking studies of compound **5** into the cAMP binding domain of active EPAC2 suggested that 3-chloro phenyl ring A and *tert*-butylisoxazole occupy two hydrophobic pockets and play important roles for their interaction.¹⁵ Compound **6** was synthesized and gave an IC_{50} value of 13.3 μM , showing the phenyl bioisosteric replacement was a viable route forward.

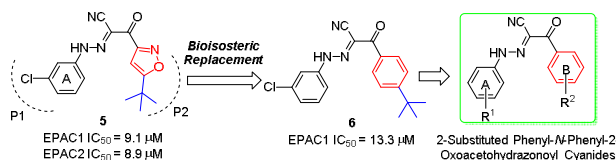
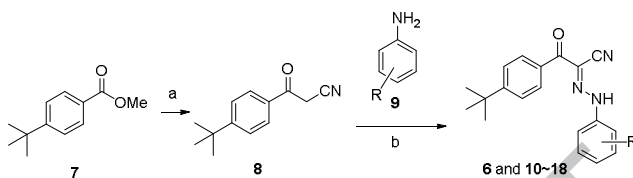


Fig. 2. Design strategy of 2-substituted phenyl-*N*-phenyl-2-oxoacetohydrazonoyl cyanides using a bioisosteric replacement approach.

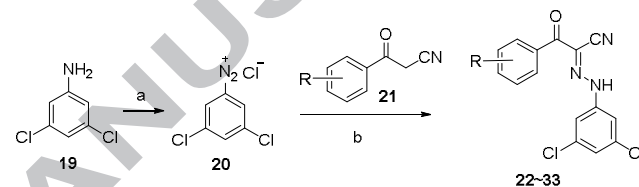
The first series of compounds were designed to keep the *tert*-butyl phenyl group B intact and the synthetic route is depicted in Scheme 1. Starting material methyl 4-(*tert*-butyl)benzoate (**7**) was reacted with CH_3CN in the presence of CH_3Li to give 3-(4-(*tert*-butyl)phenyl)-3-oxopropanenitrile (**8**).³⁰ Anilines **9** were

transformed into corresponding diazo salts and coupled with **8** in the presence of NaOAc to produce compounds **6** and **10–18** in good yields.



Scheme 1. Synthetic route to access compounds **6** and **10–18**. Reagents and conditions: (a) CH_3CN , CH_3Li , -78°C , 1h, 64%; (b) i): 10% HCl, NaNO_2 , H_2O , rt.; ii): NaOAc, EtOH, rt., 67% to quant. for two steps.

The second series of compounds were designed to replace *tert*-butyl group on phenyl ring B, and the synthetic protocol is depicted in Scheme 2. 3,5-Dichloroaniline **19** was transformed into its corresponding diazo salt and coupled with different 3-oxo-3-phenylpropanenitriles **21** in the presence of NaOAc to give compounds **22–33** in moderate to good yields. All the structures and purity of synthesized compounds were confirmed by ^1H NMR, ^{13}C NMR and HR-MS.³¹



Scheme 2. Synthetic route of compounds **22–33**. Reagents and conditions: (a) 10% HCl, NaNO_2 , H_2O , rt; (b) NaOAc, EtOH, rt., 37% to quant.

Compounds were then evaluated for their abilities to inhibit EPAC1 and EPAC2-mediated Rap1b-bGDP exchange activity using purified recombinant full-length EPAC1 and EPAC2 proteins.³² For promising compounds, the IC_{50} values against both EPAC1 and EPAC2 were determined (Table 1 and Table 2). Compound **5** was used as the reference compound for comparison with IC_{50} values of 10.8 μM and 4.4 μM against EPAC1 and EPAC2, respectively. Based on our previous SAR study, 3-Cl or 3- CF_3 on phenyl ring A which resides in a hydrophobic pocket P1 is favorable for EPAC binding. Thus, for most newly designed compounds, we chose to retain these substituents. Compared to compound **6**, compound **10** with an additional 5-Cl substituent has 2.5-fold improvement (IC_{50} = 5.4 μM) on EPAC1 inhibitory activity and 11-fold improvement (IC_{50} = 2.5 μM) on EPAC2 inhibitory activity. In addition, substituent at 5-position is more favorable than that with a same group at 4-position on this phenyl ring A (compound **11** vs **12**, and compound **16** vs **17**). Triple substituents at 3,4,5-position displayed slightly decreased EPAC inhibition (compounds **13** and **18**), but compound **18** has a 4.6-fold EPAC2/EPAC1 selectivity. 3,5-Di CF_3 substituted compound **14** showed similar EPAC inhibitory activities to those of compound **10**.

Table 1. IC_{50} values of substituted 2-(4-(*tert*-butyl)phenyl)-*N*-phenyl-2-oxoacetohydrazonoyl cyanides for inhibiting EPAC1/2 GEF activity.

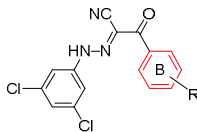
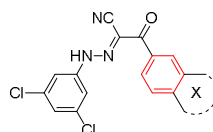
Entry	R	Rap1b-bGDP	
		EPAC1 IC_{50} (μM) ^a	EPAC2 IC_{50} (μM)




5	-	10.8 ± 1.6	4.4 ± 0.5
6	3-Cl	13.3 ± 3.0	28.4 ± 5.6
10	3,5-di Cl	5.4 ± 0.7	2.5 ± 0.6
11	3-Cl, 4-F	12.4 ± 0.9	ND ^b
12	3-Cl, 5-F	6.4 ± 1.4	3.4 ± 0.6
13	3,4,5-tri F	24.4 ± 5.7	15.8 ± 3.8
14	3,5-di CF ₃	5.5 ± 0.7	6.3 ± 0.9
15	3-CF ₃ , 4-Cl	6.1 ± 1.0	12.4 ± 3.9
16	3-Cl, 5-CF ₃	5.6 ± 0.9	5.4 ± 1.1
17	3-Cl, 4-CF ₃	50.8 ± 14.6	ND
18	3,4,5-tri Cl	13.9 ± 3.6	3.0 ± 0.4

^aThe values are the mean ± SE, n = 3. ^bND: not determined.

The first round of optimization illustrated that all compounds obtained via bioisosteric replacement of *tert*-butyl isoxazole ring with *tert*-butyl phenyl group retained both EPAC1 and EPAC2 inhibitory activities, while nearly half of them displayed more potent inhibitory activities than reference compound **5**. 3,5-di Cl substituents on phenyl ring A are favored and compound **10** exhibited the most potent EPAC1 and EPAC2 inhibitory activities. Thus, we kept this fragment intact and further modified substituents on phenyl ring B which occupies the other hydrophobic pocket P2 (Table 2). We investigated 3-Cl (**22**) and 4-Cl (**23**) first, but both are detrimental to EPAC inhibition. It appears that the bulk size of substituents on phenyl ring B is critical. We then explored electron donating group 4-OCF₃ (**24**) and electron withdrawing group 4-CO₂Me (**25**). Interestingly, neither of them showed increased inhibition against EPAC1 compared to compound **10**. Introduction of 4-Ph and 3,4-di OMe substituents on phenyl ring B led to compounds **26** and **27** which have a complete loss of EPAC inhibitory activities. The various rings including piperidine (**28**), morpholine (**29**) and cyclohexane (**30**) were also investigated, but none of them displayed more potent EPAC inhibition than compound **10**. However, compounds **31**–**33** with fused rings (e.g. 1,2,3,4-tetrahydronaphthalene, naphthalene and quinoline) all displayed improved EPAC inhibitory activities when compared to aforementioned compounds that have different substituents on phenyl ring B. Among them, compound **31** exhibited the most potent EPAC inhibitory activities with IC₅₀ values of 3.6 μM and 1.2 μM against EPAC1 and EPAC2, respectively. In Fig. 3, examples of dose-response curves are shown for compounds **5**, **6**, **14** and **31**.

Table 2. IC₅₀ values of substituted 2-phenyl-*N*-(3,5-dichlorophenyl)-2-oxoacetohydrazonoyl cyanides for inhibiting EPAC1/2 GEF activity.

Entry	R or X	Rap1b-bGDP EPAC1	Rap1b-bGDP EPAC2
			
			

		IC ₅₀ (μM) ^a	IC ₅₀ (μM)
10	4- <i>tert</i> butyl	5.4 ± 0.7	2.5 ± 0.6
22	4-Cl	>300	ND ^b
23	3-Cl	>300	ND
24	4-OCF ₃	40.9 ± 6.8	ND
25	4-CO ₂ Me	29.3 ± 6.2	ND
26	4-Ph	>300	ND
27	3,4-di OMe	>300	ND
28	4-Piperidin-1-ylmethyl	39.4 ± 4.7	ND
29	4-Morpholinomethyl	14.6 ± 5.7	ND
30	4-Cyclohexane	32.4 ± 9.5	ND
31		3.6 ± 0.3	1.2 ± 0.1
32		11.3 ± 1.4	2.5 ± 0.5
33		19.6 ± 6.7	ND

^aThe values are the mean ± SE. ^bND: not determined.

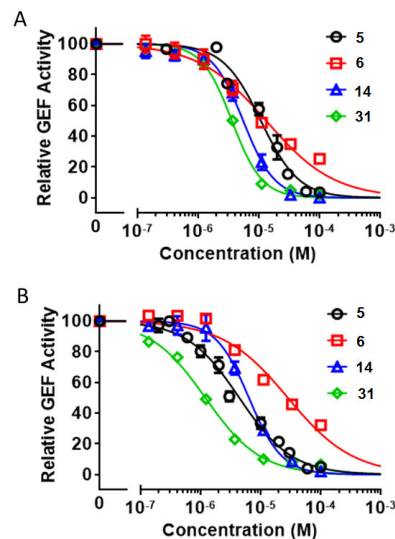


Fig. 3. A) Relative inhibitory activity for EPAC1-mediated Rap1b-bGDP exchange. B) Relative inhibitory activity for EPAC2-mediated Rap1b-bGDP exchange. Dose-dependent inhibition of EPAC1/2 GEF activity by compounds **5**, **6**, **14**, and **31**, in the presence of 20 μM cAMP. Relative GEF activity were presented as normalized reaction rate constant (means ± SEM, n = 3) described in the method.

Molecular docking study of compound **31** was performed using the Schrödinger Small-Molecule Drug Discovery Suite taking advantage of known X-ray crystal structures of activated EPAC2 protein.³³ The model shows that compound **31** occupies the CBD of EPAC2 and forms hydrogen bonds with Arg448 and the cyano-hydrazine substituent (Fig. 4A). The fused tetrahydronaphthalene ring B lays in the same pocket as *tert*-butyl isoxazole ring of ESI-09 does and is surrounded by

hydrophobic Ala416, Leu406, Val447, Val386, etc. Interestingly, ring A of compound **31** extends to the opposite direction (the solvent region) compared to ESI-09 (Fig. 4B). This docking mode suggests that there exists a large space deep pocket remaining to be explored in the CBD domain of EPAC protein for the discovery of next generation ligands.

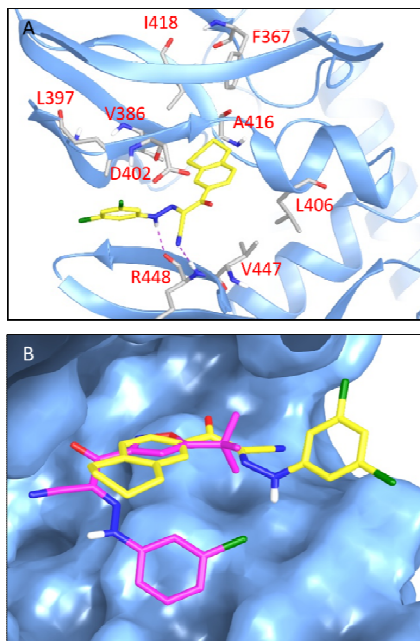


Fig. 4. Docking studies of compound **31**. A) Predicted binding pose of antagonist **31** docking at the cAMP binding domain B (CBD) of EPAC2 (PDB Code 3CF6). Compound **31** is shown in yellow. Key residues are displayed in sticks representation. Hydrogen bond is shown in purple dotted line; B) Overlay of molecular docking pose of **31** and **5** binding at the CBD of EPAC2. Compound **31** is shown in yellow, and **5** in magenta.

We have designed and synthesized a new series of 2-substituted phenyl-*N*-phenyl-2-oxoacetohydrazonoyl cyanides as EPAC inhibitors via simple chemistry with inexpensive starting material and synthetic ease suitable for scale up. Among those new molecules, compound **31** (ZL0524) was the most potent EPAC inhibitory activities with IC_{50} values of 3.6 μ M and 1.2 μ M against EPAC1 and EPAC2, respectively. Docking studies of ZL0524 with activated EPAC2 reveal that it occupies the CBD2 hydrophobic pocket, forms hydrogen bonds with Arg448 and extended to the solvent region. The findings provide us inspirations to rationally design larger molecules to reach a deeper binding pocket as next generation EPAC inhibitors. *In vivo* efficacy studies of **31** in infectious disease models (e.g. rickettsiosis) are under way, and the results will be reported in due course.

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31. Spectra data of the representative compounds: (*E*)-2-(4-(*tert*-Butyl)phenyl)-*N*-(3,5-dichlorophenyl)-2-oxoacetohydrazonoyl cyanide (**10**). Yellow solid, 69%. ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ 12.43 (s, 1H), 7.81 (d, J = 8.0 Hz, 2H), 7.56 (d, J = 8.0 Hz, 2H), 7.31 (s, 2H), 1.33 (s, 9H). ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$) δ 187.29, 166.62, 156.14, 145.04, 135.30, 133.76, 130.36, 125.25, 123.96, 115.50, 115.30, 111.49, 35.28, 31.29. HRMS (ESI): m/z ($M + \text{Na}$) $^+$ calcd for $\text{C}_{19}\text{H}_{17}\text{Cl}_2\text{N}_3\text{NaO}$: 396.0646, found: 396.0670. (*E*)-2-(4-(*tert*-Butyl)phenyl)-*N*-(3-chloro-5-fluorophenyl)-2-oxoacetohydrazonoyl cyanide (**12**). Yellow solid, quant. ^1H NMR (300 MHz, MeOD) δ 7.88 (d, J = 8.4 Hz, 2H), 7.59 (d, J = 8.4 Hz, 2H), 7.21 (s, 1H), 7.04 (d, J = 10.2 Hz, 1H), 6.94 (d, J = 8.4 Hz, 1H), 1.39 (s, 9H). ^{13}C NMR (75 MHz, MeOD) δ 187.07, 164.99, 161.71, 156.55, 144.61, 135.93, 135.77, 133.19, 129.84, 124.80, 112.00, 111.96, 111.60, 111.26, 109.70, 101.86, 101.50, 34.61, 30.09. HRMS (ESI): m/z ($M + \text{H}$) $^+$ calcd for $\text{C}_{19}\text{H}_{18}\text{ClFN}_3\text{O}$: 358.1122, found: 358.1123. (*E*)-*N*-(3,5-Bis(trifluoromethyl)phenyl)-2-(4-(*tert*-butyl)phenyl)-2-

oxoacetohydrazonoyl cyanide (**14**). Yellow solid, 67%. ^1H NMR (300 MHz, MeOD) δ 7.88 (m, 4H), 7.69 (s, 1H), 7.62 – 7.56 (m, 2H), 1.40 (s, 9H). ^{13}C NMR (75 MHz, MeOD) δ 187.29, 156.50, 144.04, 133.30, 133.18, 132.86, 132.41, 131.97, 129.82, 128.53, 124.92, 124.70, 121.32, 117.71, 116.96, 116.90, 116.85, 116.80, 116.75, 116.23, 115.80, 115.76, 109.47, 34.57, 30.05. HRMS (ESI): m/z ($M + H$)⁺ calcd for $\text{C}_{21}\text{H}_{18}\text{F}_6\text{N}_3\text{O}$: 442.1354, found: 442.1355. (*E*)-2-(4-(*tert*-Butyl)phenyl)-*N*-(3-chloro-5-(trifluoromethyl)phenyl)-2-oxoacetohydrazonoyl cyanide (**16**). Yellow solid, 76%. ^1H NMR (300 MHz, DMSO- d_6) δ 12.58 (s, 1H), 7.81 (d, $J = 7.7$ Hz, 2H), 7.56 (m, 5H), 1.32 (s, 9H). ^{13}C NMR (75 MHz, DMSO- d_6) δ 187.37, 156.13, 144.79, 135.46, 133.68, 132.39, 131.95, 130.36, 125.16, 120.80, 120.00, 115.71, 111.78, 111.33, 35.23, 31.22. HRMS (ESI): m/z ($M + \text{Na}$)⁺ calcd for $\text{C}_{20}\text{H}_{17}\text{F}_3\text{N}_3\text{ClNaO}$: 430.0910, found: 430.0956. (*E*)-*N*-(3,5-Dichlorophenyl)-2-oxo-2-(5,6,7,8-tetrahydronaphthalen-2-yl)acetohydrazonoyl cyanide (**31**). Yellow solid, 59%. ^1H NMR (300 MHz, DMSO- d_6) δ 12.42 (s, 1H), 7.68 (s, 1H), 7.58 (d, $J = 8.0$ Hz, 1H), 7.34 (s, 3H), 7.21 (d, $J = 8.0$ Hz, 1H), 2.81 (s, 4H), 1.78 (s, 4H). ^{13}C NMR (75 MHz, DMSO) δ 186.93, 144.98, 142.79, 136.83, 135.34, 133.44, 131.65, 129.19, 127.36, 123.94, 115.57, 115.23, 111.44, 29.39, 29.28, 22.95, 22.84. HRMS (ESI): m/z ($M + H$)⁺ calcd for $\text{C}_{19}\text{H}_{16}\text{N}_3\text{Cl}_2\text{O}$: 372.0670, found: 372.0667.

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33. Molecular docking studies: The docking study was performed with Schrödinger Small-Molecule Drug Discovery Suite. The crystal structure of EPAC2 (PDB code: 3CF6) was downloaded from RCSB PDB Bank and prepared with Protein Prepared Wizard. During this step, hydrogens were added, crystal waters were removed, and partial charges were assigned using the OPLS-2005 force field. The 3D structures of ESI-09 and ZL0524 were created with Schrödinger Maestro, and the initial lowest energy conformations were calculated with LigPrep. For all dockings, the grid center was chosen on the centroid of included ligand of PDB structure CBD-B site and a $24 \times 24 \times 24$ Å grid box size was used. All dockings were employed with Glide using the XP protocol. Docking poses were incorporated into Schrödinger Maestro for a visualization of ligand-receptor interactions and overlay analysis.

Graphical Abstract

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Structure-Activity Relationships of 2-Substituted Phenyl-*N*-Phenyl-2-Oxoacetohydrazonoyl Cyanides as Novel Antagonists of Exchange Proteins Directly Activated by cAMP (EPACs)

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