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2-(Anilinomethyl)imidazolines as α_{1A} Adrenergic Receptor Agonists: 2'-Heteroaryl and 2'-Oxime Ether Series

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Abstract—A series of 2'-heteroaryl and 2'-oxime anilinomethylimidazolines was prepared and evaluated in in vitro functional assays for cloned human α_{1A} , α_{1B} , and α_{1D} receptor subtypes. Potent and selective α_{1A} agonists have been identified in these series. © 2002 Elsevier Science Ltd. All rights reserved.

Alpha₁ adrenergic receptors have an important physiological role in the cardiovascular and urogenital systems.^{1–3} Three α_1 adrenergic receptor subtypes (i.e., α_{1A} , α_{1B} , and α_{1D}) have been confirmed via cloning techniques.^{4–9} The expression levels of these subtypes varies in different tissues; therefore, development of subtype specific ligands may result in drugs with improved therapeutic profiles.¹⁰ For example, the selective α_{1A} antagonist tamsulosin has been shown to have fewer side effects in the treatment of benign prostatic hyperplasia than the non-selective α_1 antagonist terazosin.^{11,12} Similarly, the selective α_{1A} agonist NS-49 may be effective in the treatment of stress urinary incontinence with a lower potential for systemic blood pressure effects.^{13,14} The improved therapeutic profile of NS-49 is due to the predominance of the α_{1A} subtype in urethal smooth muscle. We are interested in evaluating selective α_{1A} agonists as potential therapeutic agents for the treatment of stress urinary incontinence.

2'-Carboxylic acid and methyl ester 2-(anilinomethyl)imidazolines were reported to be potent pressor agents and believed to be acting through α_1 adrenergic activation.¹⁵ Our evaluation of a 2'-methyl ester (Fig. 1) at the three cloned human α_1 adrenergic receptors¹⁶ revealed that it was a potent full agonist at all three subtypes: α_{1A} (pEC₅₀=9.1); α_{1B} (pEC₅₀=9.4); α_{1D} (pEC₅₀=9.3).

Efforts to develop a selective α_{1A} agonist focused on the replacement of the methyl ester. Heterocycles^{17–23} and oxime ethers^{24–27} have been used as ester bioisosteres. Isosteric replacement of the methyl ester with a variety of heterocycles and oxime ethers resulted in the discovery of α_{1A} agonists that are both potent and selective. The preparation and biological activity of a series of 2'-heteroaryl and 2'-oxime ethers of 2-(anilinomethyl)-imidazolines are described herein.

The 2'-substituted 2-(anilinomethyl)imidazolines were prepared according to the general synthetic scheme illustrated in Scheme 1.

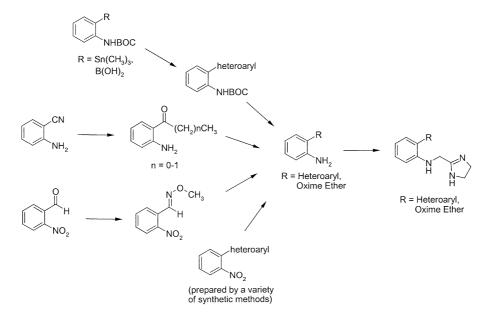
The requisite nitro intermediates were prepared by a variety of synthetic methods. 2-(2-Nitrophenyl)oxazole,^{28,29} 3-methyl-5-(2-nitrophenyl)isoxazole,³⁰ 5-(2-nitrophenyl)isoxazole,³¹ and 4-(2-nitrophenyl)isoxazole³¹ were prepared



Figure 1.

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Scheme 1.

according to literature procedures. The nitro intermediate for compound **21** was prepared by a method analogous to that described for 5-(2-nitrophenyl)isoxazole in which guanidine hydrochloride was used in the presence of catalytic NaOH to form the 2-aminopyrimidine ring. The nitro intermediates for compounds 12-13, 17, and 19 were prepared under Ullmann cross-coupling conditions using 2-bromonitrobenzene and the corresponding bromo- or iodo-substituted heterocycles.³² 2-(2-Nitrophenyl)furan was prepared by Stille³³ coupling of 2nitrobromobenzene and 2-tributylstannylfuran in DMF with tetrakis(triphenylphosphine)palladium as a catalyst. The analogous thiophene intermediate was obtained in a similar manner using 2-nitroiodobenzene, 2-tributylstannylthiophene, and dichlorobis(triphenylphosphine)palladium (II) as a catalyst. The nitro intermediates for oxadiazoles 14 and 15 were prepared according to the general procedures described by Orlek and co-workers.¹⁷ 4-Methyl-5-(2-nitrophenyl)oxazole was prepared from 2-nitrobenzaldehyde and 1-(toluene-4-sulfonyl)ethyl isocyanide³⁴ using the general procedure described by Possel et al.³⁵ The nitro intermediate for oxazole 6 was prepared from 2-bromo-2'-nitroacetophenone using potassium acetate and acetic acid followed by ammonium acetate and acetic acid under refluxing conditions. 2-Methyl-4-(2-nitrophenyl)thiazole was prepared from 2-bromo-2'-nitroacetophenone and thioacetamide. Treatment of 2-bromo-2'-nitroacetophenone with tetrabutylammonium azide in acetonitrile between 0 and 25°C followed by treatment of the resultant azide with polymer-bound triphenylphosphine and acetyl chloride in toluene at room temperature gave the nitro intermediate for methyloxazole 3. The reaction of 2-nitrobenzaldehyde with methoxylamine hydrochloride in refluxing EtOH gave the desired nitro intermediate for oxime 26.

Reduction of the previously described nitro intermediates as well as commercially available 5-(2-nitrophenyl)oxazole

provided the majority of desired anilines. The nitro groups of the isoxazole intermediates were reduced with stannous chloride and HCl.³¹ 2-(2-Nitrophenyl)furan and 2-(2-nitrophenyl)pyridine were reduced with 35% hydrazine in EtOH in the presence of 10% Pd/C. The nitro intermediates for oxadiazoles 14 and 15 and oxime 26 were reduced with sodium sulfide using the general conditions described by Lin and Lang.³⁶ Commercially available 5-(2-nitrophenyl)furfural was reduced by the general method described by Kano and co-workers using titanium (IV) chloride and sodium borohydride to give the desired aniline intermediate for compound 16.³⁷ 4-(2-Nitrophenyl)-2-methylthiazole was reduced by catalytic hydrogenation with platinum oxide in ethanol. Catalytic hydrogenation of 2-(2-nitrophenyl)thiophene was carried out in ethanol and acetic acid using 10% palladium on carbon to give the desired aniline. The remaining nitro intermediates for compounds 1, 3, 6–7, 10, 12-13, 19, 21, and 26 were reduced by catalytic hydrogenation using 10% Pd/C in EtOH.

The remaining aniline intermediates were prepared by a variety of synthetic methods as described below. Stille coupling of 2-chloropyrazine and (2-trimethylstannylphenyl-carbamic acid tert-butyl ester³⁸ with tetrakis(triphenylphosphine)palladium (0) in the presence of catalytic copper (I) bromide followed by deprotection with trifluoroacetic acid provided 2-(2-aminophenyl)pyrazine en route to 18. Suzuki coupling of [2-(N-t-bocamino)phenyl]boronic acid with 2-bromo-3-methylpyridine and 2-bromo-3-trifluoromethylpyridine according to the general procedure described by Snieckus³⁹ followed by N-deprotection with trifluoroacetic acid provided the aniline intermediates for compounds 20 and 22, respectively. Finally, the aniline intermediates for oximes 23–25 and 27 were prepared by treatment of the corresponding anilino ketones with the O-alkyl hydroxylamine hydrochlorides in EtOH under refluxing conditions. The requisite anilino ketones for compounds 25 and **27** were obtained via Grignard reaction between anthranilonitrile and ethylmagnesium chloride followed by hydrolysis with sulfuric acid. The anilino ketones for compounds **23** and **24** were commercially available.

The anilines described above were coupled to 2-(chloromethyl)-2-imidazoline hydrochloride⁴⁰ as described in the literature.^{41,42} All compounds except **3–4**, **14–15**, **18**, and **23–24** were converted to either their hydrogen chloride or fumarate salts. ¹H NMR and mass spectroscopic data were consistent for all final products. Representative ¹H, ¹³C NMR, and mass spectroscopic data are provided for compound **20**.⁴³

The 2'-heteroaryl substituted 2-anilinomethylimidazolines listed in Table 1 range from weak partial agonists to potent full agonists at the α_{1A} receptor. The α_{1A} agonists oxymetazoline, naphazoline, and cirazoline are included for comparison.

Potency and efficacy at the α_{1A} receptor as well as selectivity versus the α_{1B} and α_{1D} subtypes are influenced by the heteroatoms of the 2'-heteroaromatic ring. The relative positions of the heteroatoms can significantly impact potency and selectivity. For example, isoxazole **8** is a potent full agonist at all three receptor subtypes

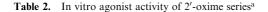
 Table 1. In vitro agonist activity of 2'-heteroaryl series^a

whereas isoxazole 11 is a less potent but selective α_{1A} agonist. A similar comparison can be made for oxazoles 10 and 1, respectively. Heteroatom substitution may also influence activity at the three receptor subtypes. For example, oxazole 1 was a moderately potent and selective α_{1A} agonist that was inactive at the α_{1B} and α_{1D} subtypes. However, replacement of the oxazole nitrogen with a carbon resulted in a compound (i.e., furan 2) that was significantly more potent at all three receptor subtypes and fully efficacious at α_{1B} and α_{1D} . While pyridine 17 and pyrazine 18 are both potent full agonists at the α_{1A} receptor, the presence of the second nitrogen in the pyrazine ring resulted in a significant loss of potency and efficacy at the α_{1B} receptor. Finally, a comparison between thiophene 12 and furan 16 reveals that substitution of oxygen for sulfur resulted in an increase in α_{1A} potency and efficacy. These observations suggest that the heteroatoms of the 2'-aromatic ring contribute significantly to the ligand-receptor interactions at all three receptor subtypes. These heteroatoms (i.e., oxygen and nitrogen) may act as hydrogen-bond acceptors, which in turn could either facilitate or inhibit receptor activation.

Substitution of the 2'-heteroaryl ring may also have a significant effect on subtype potency, efficacy, and selectivity. The α_{1B} and α_{1D} receptors appear to be

			1–16	17-22			
					pEC ₅₀ (% Max. of phenylephrine) ^b		
Compd	W	Х	Υ	Z	α_{1A}	α_{1B}	α_{1D}
Oxymetazoline					7.7 (74)	< 5.3	< 5.3
Naphazoline					7.2 (72)	< 5.3	< 5.3
Cirazoline					7.9 (93)	7.2 (72)	6.9 (31)
1	С	Ν	С	0	7.76 (107)	< 5.30	< 5.30
2	С	С	С	0	9.33 (101)	8.2 (95)	7.79 (90)
3	С	Ν	C–Me	0	6.96 (126)	< 5.30	< 5.30
4	Ν	C–Me	S	С	8.35 (98)	7.16 (21)	< 5.30
5	С	С	С	S	8.65 (99)	8.24 (103)	7.74 (59)
6	С	0	C–Me	Ν	6.49 (64)	< 5.30	< 5.30
7	0	С	Ν	C–Me	7.82 (95)	< 4.00	< 4.00
8	С	С	Ν	0	9.10 (134)	8.43 (97)	8.54 (114)
9	0	Ν	C–Me	С	7.41 (86)	7.02 (27)	< 4.00
10	0	С	С	Ν	9.18 (121)	8.79 (123)	8.3 (103)
11	С	Ν	0	С	7.34 (99)	<4.00	< 4.00
12	С	С	C–Me	S	7.44 (49)	< 4.00	< 4.00
13	S	С	С	C–Me	9.07 (104)	6.95 (78)	7.31 (83)
14	0	Ν	C–Me	Ν	6.18 (92)	< 5.30	< 5.30
15	Ν	0	C–Me	Ν	6.14 (57)	6.17 (43)	< 5.30
16	С	С	C–Me	0	8.57 (98)	7.33 (48)	< 5.70
17	Ν	C C	С	С	8.17 (103)	7.52 (101)	7.38 (35)
18	Ν	C	N	Ċ	8.34 (94)	6.47 (48)	6.74 (38)
19	N	Č	C	Ň	8.00 (105)	7.11 (102)	7.09 (60)
20	C–Me	Č	Č	N	7.61 (104)	<4.00	< 4.00
21	C	Ň	C-NH ₂	N	7.37 (97)	< 5.30	7.54 (44)
22	C-CF3	С	C	Ν	6.17 (78)	< 4.00	< 4.00

^aSee ref 16 for description of the assay. Each entry represents the mean of at least two experiments, with an average SEM of ± 0.12 . ^b% of phenylephrine response (40 μ M).





Compd	Isomer	R	R′	pEC ₅₀ (% Max. of phenylephrine) ^b		
				α_{1A}	α_{1B}	α_{1D}
23	Ε	CH ₃	CH ₃	7.68 (92)	< 5.30	< 5.30
24	E	CH ₂ CH ₃	CH ₃	7.28 (102)	< 5.30	< 5.30
25	Ε	CH ₃	CH ₂ CH ₃	8.48 (112)	< 4.00	< 4.00
26	Ε	CH ₃	Ĥ	7.29 (94)	< 5.30	< 5.30
27	Ζ	CH ₃	CH ₂ CH ₃	7.06 (141)	< 5.30	< 5.30

^aSee ref 16 for description of the assay. Each entry represents the mean of at least two experiments, with an average SEM of ± 0.12 . ^{b%} of phenylephrine response (40 μ M).

Table 3. Comparison of in vitro functional and binding data^a

	Funct	tional assay	In vitro binding (pIC ₅₀)			
Compd	α_{1A}	α_{1B}	$\alpha_{1\mathrm{D}}$	$\alpha_{1\mathrm{A}}$	α_{1B}	α_{1D}
1 7 8 10 17 19	7.76 (107) 7.82 (95) 9.10 (134) 9.18 (121) 8.17 (103) 8.00 (105)	<4.00 8.43 (97)) 7.38 (35)	6.34 6.72 7.45 7.34 7.09 6.72	6.64 6.49 6.88 6.35 6.23 5.66	6.53 6.46 7.42 7.39 6.93 6.38
26	7.29 (94)	< 5.30	< 5.30	6.55	6.79	7.37

^aSee refs 16 and 46 for descriptions of the assays. Each entry represents the mean of at least two experiments, with an average SEM of ± 0.12 for the functional assay and ± 0.11 for the binding assay. ^{b%} of phenylephrine response (40 μ M).

less able to accommodate substitution of the 2'-heteroaromatic ring. For example, methyl substitutions of thiophene 5, furan 2, isoxazole 8, and pyridine 17 to give compounds 12, 16, 9, and 20, respectively, result in a greater loss of potency and/or efficacy at the α_{1B} and α_{1D} receptors relative to α_{1A} . In fact, one may possibly exploit this steric constraint of the α_{1B} and α_{1D} receptors to design α_{1A} agonists with greater selectivity.

Finally, replacement of the methyl group of pyridine **20** with a trifluoromethyl group (i.e., **22**) resulted in a loss of potency at the α_{1A} receptor, suggesting that electron density of the heteroaromatic ring may be important for interaction with this receptor subtype.

The in vitro α_1 functional profiles of the 2'-oxime ethers are listed in Table 2. All of the compounds listed in this series are selective, full agonists at the α_{1A} receptor subtype. An increase in steric bulk of the (*E*)-*O*-methyl oxime ethers resulted in an increase in potency at the α_{1A} receptor. Thus, as size increased from the formyl (**26**) to the acetophenone (**23**) and finally the propiophenone derivative (**25**), potency at the α_{1A} receptor increased more than an order of magnitude. In contrast, an increase in size of the *O*-alkyl group resulted in a slight decrease in potency at the α_{1A} receptor (i.e., *O*-methyl oxime ether **23** vs *O*-ethyl oxime ether **24**). The effect of configuration about the oxime double bond on α_{1A} agonist activity was significant. While both compounds were fully efficacious at the α_{1A} receptor, the (*E*)-*O*-methyl oxime ether (**25**) was more than 25 times as potent as the corresponding *Z*-isomer (**27**).

While a number of compounds in the 2'-heteroaryl and 2'-oxime ether series were potent and selective for the α_{1A} receptor, none were selective for the α_{1B} and α_{1D} subtypes. This is consistent with the fact that most imidazoline-type α_1 adrenergic agonists exhibit some degree of selectivity for the α_{1A} subtype.^{44,45} Furthermore, the functional activity observed at the three receptor subtypes did not generally correlate with their binding affinities.⁴⁶ For example, oxazoles 1 and 7 were at least 100-fold more potent at the α_{1A} receptor relative to α_{1B} and α_{1D} ; however, the compounds bound equally well to all three subtypes (see Table 3). The lack of correlation between the α_1 subtype selectivity in the functional and binding assays has been attributed to a difference in intrinsic activity at the three receptor subtypes.45

2'-Heteroaryl and 2'-oxime anilinomethylimidazolines have been identified that were potent, selective, and fully efficacious at the α_{1A} receptor. Functional activity at the α_{1A} , α_{1B} , and α_{1D} subtypes did not generally correlate with receptor binding affinity. In the 2'-heteroaryl series, the heteroatoms appeared to play a significant role in ligand-receptor interactions. Also, the α_{1B} and α_{1D} receptor subtypes have greater steric constraints in the vicinity of the heteroaryl ring than does the α_{1A} receptor. All of the oxime ethers were fully efficacious at the α_{1A} receptor progressively increased as steric bulk at the oxime carbon was increased. In the case of the *O*methyl propiophenone derivative, the *E*-isomer was preferred.

References and Notes

1. Schwinn, D. A.; Dwatra, M. M. Adv. Pharmacol. 1998, 42, 390.

- 2. Michel, M. C.; Taguchi, K.; Schafers, R. S.; Williams, T. J.;
- Clarke, D. E.; Ford, A. P. D. W. *Adv. Pharmacol.* **1998**, *42*, 394. 3. Ruffolo, R. R., Jr.; Hieble, J. P. *Eur. Urol.* **1999**, *36*, 17.
- J. Kullolo, K. K., JI., Hicole, J. F. Lur. Orol. 1999, 30, 17.
- 4. Cotecchia, S.; Schwinn, D. A.; Randall, R. R.; Lefkowitz, R. J.; Caron, M. G.; Kobilka, B. K. *Proc. Natl. Acad. Sci. U.S.A.* **1988**, *85*, 7159.
- 5. Lomasney, J. W.; Cotecchia, S.; Lorenz, W.; Leung, W.-Y.;
- Schwinn, D. A.; Yang-Feng, T. L.; Brownstein, M.; Lefkowitz, R. J.; Caron, M. G. J. Biol. Chem. 1991, 266, 6365.
- 6. Perez, D. M.; Piascik, M. T.; Graham, R. M. Mol. Pharmacol. 1991, 40, 876.
- 7. Schwinn, D. A.; Lomasney, J. W.; Loreng, W.; Szklut, P. J.;
- Fremeau, R. T.; Yang-Feng, T. L.; Caron, M. G.; Lefkowitz, R. J.; Cotecchia, S. J. Biol. Chem. **1990**, 265, 8183.
- 8. Hirasawa, A.; Horie, K.; Tanaka, T.; Takagaki, K.; Murai, M.; Yano, J.; Tsujimoto, G. *Biochem. Biophys. Res. Commun.* **1993**, *195*, 902.
- 9. Esbenshade, T. A.; Hirasawa, A.; Tsujimoto, G.; Tanaka,
- T.; Yano, J.; Minneman, K. P.; Murphy, T. J. Mol. Pharmacol. 1995, 44, 977.
- 10. Hieble, J. P.; Ruffolo, R. R., Jr. Drugs Pharm. Sci. 1998, 89, 231.
- 11. Na, Y. J.; Guo, Y. L.; Gu, F. J. Med. 1998, 29, 289.
- 12. Lee, E.; Lee, C. Br. J. Urol. 1997, 80, 606.
- 13. Taniguchi, N.; Hamada, K.; Ogasawara, T.; Ukai, Y.;
- Yoshikuni, Y.; Kimura, K. Eur. J. Pharmacol. 1996, 318, 1289.
- 14. Craig, D. A.; Forray, C. C.; Gluchowski, C.; Branchek, T. A. US Patent 5,610,174, 1997. *Chem. Abstr.* **1997**, *126*, 117873.
- 15. Brown, R. E. US Patent 3,754,002, 1973. Chem. Abstr. 1973, 79, 92219.
- 16. Human α_{1A} (clone #137–12), α_{1B} (clone # 37–11) and α_{1D} (clone # 16–7) adrenoceptors were expressed in Rat-1 fibroblast cells. Receptor activation was determined via calcium mobilization through the Gq coupled PLC pathway using calcium-sensitive fluorescent dyes (Calcium Green-Molecular Probes C 3011) measured by a Fluorescent Light Imaging Plate Reader (FLIPR). Eleven-point concentration–response curves were calculated as a percent of the 40 µM phenylephrine response in which the highest sample concentration was typically 5 µM.
- 17. Orlek, B. S.; Blaney, F. E.; Brown, F.; Clark, M. S. G.; Hadley, M. S.; Hatcher, H.; Riley, G. J.; Rosenberg, H. E.; Wadsworth, H. J.; Wyman, P. *J. Med. Chem.* **1991**, *34*, 2726. 18. Jenkins, S. M.; Wadsworth, H. J.; Bromidge, S.; Orlek, B. S.; Wyman, P. A.; Riley, G. J.; Hawkins, J. *J. Med. Chem.* **1992**, *35*, 2392.
- 19. Street, L. J.; Baker, R.; Book, T.; Reeve, A. J.; Saunders, H.; Willson, T.; Marwood, R. S.; Patel, S.; Freedman, S. B. *J. Med. Chem.* **1992**, *35*, 295.
- 20. Saunders, J.; MacLeod, A. M.; Merchant, K.; Showell, G. A.; Snow, J.; Street, L. J.; Baker, R. *J. Chem. Soc., Chem. Commun.* **1988**, 1618.
- 21. Saunders, J.; Cassidy, M.; Freedman, S. B.; Harley, E. A.; Iversen, L. L.; Kneen, C.; MacLeod, A. M.; Merchant, K. J.; Snow, R. J.; Baker, R. J. *Med. Chem.* **1990**, *33*, 1128.
- 22. MacLeod, A. M.; Baker, R.; Freedman, S. B.; Patel, S.; Merchant, K. J.; Roe, M.; Saunders, J. J. Med. Chem. **1990**, 2052.

- 23. Wadsworth, H. J.; Jenkins, S. M.; Orlek, B. S.; Cassidy, F.; Clark, M. S. G.; Brown, F.; Riley, G. J.; Graves, D.; Hawkins, J.; Nahlor, C. B. *J. Med. Chem.* **1992**, *35*, 1280.
- 24. Toja, E.; Bonetti, C.; Butti, A.; Hunt, P.; Fortin, M.; Barzaghi, F.; Formento, M. L.; Maggioni, A.; Nencioni, A.; Galliani, G. *Eur. J. Med. Chem.* **1991**, *26*, 853.
- 25. Bromidge, S. M.; Brown, F.; Cassidy, F.; Clark, M. S. G.; Dabbs, S.; Hadley, M. S.; Hawkins, J.; Loudon, J. M.; Naylor,
- C. B.; Orlek, B. S.; Riley, G. J. J. Med. Chem. **1997**, 40, 4265. 26. Bromidge, S. M.; Brown, F.; Cassidy, F.; Clark, M. S. G.;
- 20. Bronndge, S. M., Brown, F., Cassidy, F., Clark, M. S. G., Dabbs, S.; Hadley, M. S.; Loudon, J. M.; Orlek, B. S.; Riley, J. G. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 787.
- 27. Bromidge, S. M.; Brown, F.; Cassidy, F.; Clark, M. S. G.; Dabbs, S.; Hawkins, J.; Loudon, J. M.; Orlek, B. S.; Riley,
- J. G. Bioorg. Med. Chem. Lett. 1992, 2, 791.
- 28. Cass, W. E. J. Am. Chem. Soc. 1942, 64, 785.
- 29. Jung, M. E.; Dansereau, S. M. K. Heterocycles 1994, 39, 767.
- 30. Sakamoto, T.; Kondo, Y.; Uchiyama, D.; Yamanaka, H. *Tetrahedron* **1991**, *47*, 5111.
- 31. Wolf, A. D.; Rorer, M. P. European Patent 0 083 975, 1987. Chem. Abstr. 1983, 99, 175812.
- 32. Shimizu, N.; Kitamura, T.; Watanabe, K.; Yamaguchi, T.; Shigyo, H.; Ohta, T. *Tetrahedron Lett.* **1993**, *34*, 3421.
- 33. Stille, J. K. Angew. Chem., Int. Ed. Engl. 1986, 25, 508.
- 34. Van Leusen, A. M.; Bouma, R. J.; Possel, O. Tetrahedron Lett. 1975, 40, 3487.
- 35. Possel, O.; van Leusen, A. M. Heterocycles 1977, 7, 77.
- Lin, Y.; Lang, S. A., Jr. J. Heterocyc. Chem. 1980, 17, 1273.
 Kano, S.; Tanaka, Y.; Sugino, E.; Hibino, S. Synthesis 1980, 9, 695.
- 38. Navas, F., III; Tang, F. L. M.; Schaller, L. T.; Norman, M. H. *Bioorg. Med. Chem.* **1998**, *6*, 811.
- 39. Guillier, F.; Nivoliers, F.; Godard, A.; Marsais, F.; Queguiner, G.; Siddiqui, M. A.; Snieckus, V. J. Org. Chem. **1995**, 60, 292.
- 40. Prepared from chloroacetonitrile via ester imidate formation and treatment with ethylenediamine; see: Klarer, W.; Urech, E. *Helv. Chim. Acta* **1944**, *27*, 1762.
- 41. Copp, F. C.; Roberts, P. T.; Frenkel, A. D.; Collard, D. German Patent DE 2756638, 1978; *Chem. Abstr.* **1978**, *89*, 109498.

42. Saari, W. S.; Halczenko, W.; Randall, W. C.; Lotti, V. J. J. Med. Chem. 1983, 26, 1769.

- 43. *N*-(4,5-Dihydro-1*H*-imidazol-2-ylmethyl)-2-(3-methyl-2pyridinyl)aniline fumarate: ¹H NMR (400 MHz; DMSO- d_6) δ 2.12 (s, 3H), 3.67 (s, 4H), 4.12 (d, *J*=5.8 Hz, 2H), 5.53 (br d, *J*=6.0 Hz, 1H), 6.38 (s, 2H), 6.66 (d, *J*=8.2 Hz, 1H), 6.72 (t, *J*=7.4 Hz, 1H), 7.00 (d, *J*=6.8 Hz, 1H), 7.21 (t, *J*=7.6 Hz, 1H), 7.25 (m, 1H), 7.69 (d, *J*=7.7 Hz, 1H), 8.42 (d, *J*=4.3 Hz, 1H); ¹³C NMR (100 MHz; DMSO- d_6) δ 19.76, 40.65, 45.84, 11.82, 117.79, 123.10, 126.46, 129.81, 130.68, 132.78, 135.84, 139.48, 145.36, 147.30, 157.24, 168.55, 170.61. HRESI-MS: [M+H]⁺ 267.1610, found 267.1599.
- 44. Minneman, K. P.; Theroux, T. L.; Hollinger, S.; Han, C.; Esbenshade, T. A. *Mol. Pharmacol.* **1994**, *46*, 929.
- 45. Zhong, H.; Minneman, K. P. Eur. J. Pharmacol. 1999, 375, 261.
- 46. Affinity of compounds at α_1 adrenoceptor subtypes was determined by radioligand binding techniques using membranes prepared from Rat-1 fibroblasts expressing human α_{1A} , α_{1B} , or α_{1D} adrenoceptors as previously described: Gobel, J.; Saussy, D. L.; Goetz, A. S. *J. Pharmacol. Toxicol.* **1999**, *42*, 237.