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Imidazole moiety replacements in the 3-(1*H*-benzo[d]imidazol-2-yl)pyridin-2(1*H*)-one inhibitors of insulin-like growth factor receptor-1 (IGF-1R) to improve cytochrome P450 profile

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Abstract—A series of 3-(1*H*-benzo[d]imidazol-2-yl)pyridin-2(1*H*)-one inhibitors of insulin-like growth factor receptor-1 (IGF-1R) were examined in which the pendant imidazole moiety was replaced to improve selectivity for IGF-1R inhibition over cytochrome P450 (CYP). Synthesis and SAR of these compounds is presented. © 2007 Elsevier Ltd. All rights reserved.

Insulin-like growth factor receptor-1 (IGF-1R) plays an important role in normal cell physiology, and is also implicated in the establishment and maintenance of the malignant phenotype.¹ IGF-1R is a tyrosine kinase receptor that is activated by the binding of its ligands, IGF-I or IGF-II, to the extracellular domain. Ligand binding causes a conformational change resulting in autophosphorylation of tyrosine residues in the intracellular kinase domain resulting in activation of both the mitogenic (Ras/Raf/MAPK) and survival pathways (PI3K/Akt/mTor).^{1,2} In addition, recent studies have shown that IGF-1R regulates cell adhesion and motility.3 Overexpression of IGF-1R and IGF-I has been demonstrated in a variety of tumors,⁴ including glioma, lung, ovary, breast, carcinomas, sarcomas, and melanoma. Therefore, IGF-1R seems to be a very promising target for cancer therapy.⁵ As a consequence, the identification of potent inhibitors of IGF-1R has recently

attracted considerable attention and several classes of inhibitors have been described. 6

We have recently reported the discovery and biological activity associated with the benzimidazole-pyridone derivatives stemming from the identification of 1 as a prototype of a new structural class of IGF-1R kinase inhibitors.⁷ Further optimization of **1** by installing basic amine side chains at the C-4-position of pyridone led to the identification of 2, 3, and 4 with IC_{50} 's of 390, 530, and 180 nM, respectively.⁸ Additionally, compound 4 reduced autophosphorylation of IGF-1R in CD8-IGF1R mouse xenografts when dosed in vivo. However, compounds 1-4 exhibited potent cytochrome P450 (CYP) inhibition profiles across 3A4, 2C9, 2C19, 2D6, and 1A2 isozymes (Table 1). Such potent inhibition of drug-metabolizing CYP enzymes may cause unfavorable drug-drug interactions in the clinic.⁹ Many lead candidate molecules in pharmaceutical development fail due to potent inhibition of one or more CYP enzyme isoforms. Consequently, we sought to improve the CYP inhibition of this chemotype while maintaining or improving the IGF-1R enzyme potency. By examining the structural elements within the chemotype 1, it was reasoned that the pendant imidazole moiety could be responsible for potent CYP inhibition, as some

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Table 1. IGF1R and CYP activities¹¹ for compounds **2**, **3**, and **4**

IC ₅₀ (µM)	2	3	4
IGF-1R	0.39	0.53	0.18
CYP1A2	< 0.03	< 0.04	< 0.04
CYP2C9	< 0.03	< 0.04	< 0.04
CYP2C19	0.1	< 0.04	< 0.04
CYP2D6	5.7	0.56	ND
CYP3A4-BFC	0.08	0.06	< 0.046

imidazole derivatives such as ketoconazole and clotrimazole are known to show high affinity for the heme iron of major drug-metabolizing CYP enzymes Figure 1.^{9b,10}

The crystal structure of **1** bound to a truncated IGF-1R revealed that the pendant imidazole ring is solvent exposed, and is not involved in any favorable hydrogen bonding interactions with the receptor.^{7a} Therefore, we explored replacing of the imidazole ring with other groups in an effort to improve selectivity for IGF-1R inhibition over cytochrome P450 inhibition.

At the outset, we elected to replace the imidazole moietv with both the bromo and cvano groups as these groups are considered versatile chemical handles for analog synthesis. Accordingly, the syntheses of bromo derivative 10 and cyano derivative 24 were accomplished as outlined in Schemes 1 and 2. Commercially available 4-bromo-2-methyl-6-nitro aniline (5) was reduced to furnish diamine, which was immediately treated with 2-methoxy-4-iodo-pyridine-3-carboxalde $hvde^{12}$ (7) to afford benzimidazole 8. The methoxy group in 8 was cleaved, and the resultant iodopyridone was heated with (S)-phenyl alaninol to provide 10. Suzuki coupling with aryl boronic acids was rather sluggish presumably for solubility reasons, and therefore the primary hydroxyl was protected as its trityl ether 11 which underwent Suzuki couplings smoothly with several aryl boronic acids. Finally, the trityl group was deprotected with trifluoroacetic acid to afford biaryls (12, 13, and 14).



Figure 1. Representative benzimidazole IGF-1R inhibitors.



Scheme 1. Reagents and conditions: (a) $SnCl_2 2H_2O$ EtOH, reflux, 59%; (b) 2-methoxy-4-iodo pyridine-3-carboxaldehyde (7), MeOH, rt, 50%; (c) 4 N HCl in dioxane, H₂O, rt, 100%; (d) phenyl alaninol, Et₃N, DMF, 90 °C, 89%; (e) triphenylmethyl chloride, Et₃N, DMAP, CH₂Cl₂, 72%; (f) aryl boronic acids, Pd(PPh₃)₄, K₃PO₄, THF, reflux, 60–67%; (g) TFA, CH₂Cl₂, 100%.

Similarly, the cyano derivative 24, which is useful to synthesize other heterocycles, was synthesized starting from 3-methyl-4-nitro-benzonitrile (15). The nitro group was reduced and the resultant aniline 16 was trifluoroacetylated. The amide 17 was subjected to nitration by treating with potassium nitrate in sulfuric acid, and then the trifluoroacetyl group was cleaved. The nitroaniline 19 was reduced and the resultant phenylene diamine 20 was immediately treated with the 2-methoxy-4-iodo-pyridine-3-carboxaldehyde¹² (7) to furnish the benzimidazole 21. The methoxy group was cleaved to give the iodo pyridone 22, which was heated with meta-chloro phenethylamine 23 to afford 24. The nitrile group was hydrolyzed to the corresponding acid 25, which was used to make amides 27-32. Additionally, the cyano compound 24 was also converted to imidate ester 33 by exposure to anhydrous HCl gas in ethanol. Heating 33 with several 1,3 or 1,4 diamines furnished imidazolines (34, 35, and 36) or tetrahydropyrimidine 37, respectively. The carbon attached imidazole 39 was built from the imidate ester 33 using a two-step procedure.¹³

As outlined in Table 2, biaryls **12–14** represented the first examples with a reduced cytochrome P450 inhibition profile in this series of IGF-1R inhibitors. These results suggest that the pendant imidazole was primarily responsible for potent CYP inhibition as anticipated. Interestingly, the bromo compound **10** displayed reduced CYP3A4 inhibition, while still retaining significant affinity for other CYP isozymes.



Scheme 2. Reagents and conditions: (a) Fe, HOAc, 74%; (b) trifluoroacetic anhydride, Et₃N, CH₂Cl₂, 100%; (c) KNO₃, concd sulfuric acid, 52%; (d) 2 M NH₃ in MeOH, 95%; (e) SnCl₂·2H₂O, EtOH reflux, 90%; (f) 2-methoxy-4-iodo-pyridine-3-carboxaldehyde (7), I₂, MeOH, 46%; (g) 4 N HCl in dioxane, H₂O, 100%; (h) 2-amino-1-(3-chlorophenyl)-ethanol (23), DMF, Et₃N, 80 °C, 94%; (i) 6 N HCl, 80 °C, 54%; (j) DPPA, Et₃N, DMF, amines, 40–60%; (k) anhydrous HCl, EtOH, 90%; (l) *N*-methyl 1,3-propane diamine, EtOH, reflux, 100%; (m) NH₂Me, heat, 100%.

However, the biaryls (12-14) were found to be less potent against IGF-1R relative to the imidazole containing compounds (2, 3, and 4). With these promising initial results in improving the CYP profile our efforts shifted to cyano compound 24 and derivatives thereof Table 3.

Table 2. IGF1R and CYP activities¹¹ for imidazole replacements

			1	
$IC_{50} \ (\mu M)$	10	12	13	14
IGF-1R ^a	1.4	3.1	5.1	4.0
CYP1A2	0.052	0.58	3.2	12.0
CYP2C9	0.72	2.3	4.0	11.0
CYP2C19	0.32	0.53	1.0	2.2
CYP2D6 ^b	ND	ND	ND	ND
CYP3A4	3.1	3.6	63	6.8

 a IC₅₀ values represent the average of two determinations and standard deviations ranged from 0.16 to 0.84.

^b Overlap with reference, IC₅₀ could not be determined.

The compound that lacks imidazole, 26, displayed a fourfold reduction in IGF-1R enzyme potency with an improved CYP profile. Examination of various functional groups at the imidazole location led to the identification of amides that retained IGF-1R enzyme potency while improving the CYP profile. The primary and secondary amides (27 and 28) displayed equipotency of 180 nM while the tertiary amide (29) was less active (550 nM). It was found that increasing bulk on the secondary amide led to a decrease in enzyme potency as represented by compounds 28 and 30 through 32. The enhanced IGF-1R potency and acceptable CYP profile (3.7 µM for 3A4) of secondary amide 28 prompted us to make the corresponding methyl amidine 38, which displayed similar IGF-1R potencies of 170 nM with an improved CYP profile across all isozymes. In order to fine-tune the potency of amidine containing compounds, cyclic amidines such as imidazolines were synthesized. We were delighted to observe a significant improvement in IGF-1R activity with these imidazolines (34, 35, and 36) and tetrahydropyrimidine 37 with a reduction in affinity for CYP enzymes. For instance, N-methyl imidazoline 34 displayed an impressive 93 nM IGF-1R activity and even further reduction in cytochrome P450 inhibition (>17 µM against all isozymes). Alpha methyl substitutions on N-methyl imidazoline as represented by 35 and 36 further enhanced selectivity of IGF1R versus CYP inhibition with 46 nM for compound 36 while its CYP3A4 affinity was 13 µM.

The corresponding six-membered tetrahydro pyrimidine 37 also maintains excellent IGF-1R inhibition (68 nM) with less affinity for CYP enzymes (>8.5 μ M). Interestingly, the unsaturated version of imidazoline, the carbon attached imidazole 39, regained some of its CYP inhibition. Taken together, these findings demonstrate that saturated heteroaryls such as imidazolines were optimal for greater separation of IGF-1R activity versus CYP inhibition. The reduced CYP inhibition of these imidazolines could be in part due to enhanced basicity. Further investigation of these imidazolines for selectivity over other tyrosine kinases, cell-based activities, and optimization for desirable ADME properties are ongoing.

In summary, the results described herein have demonstrated that the pendant imidazole was primarily responsible for potent cytochrome P450 inhibition. By replacing this moiety with other groups such as imidazolines, greater separation of IGF-1R activity versus CYP inhibition was observed. Future studies will expand on

Table 3. IGF1R and CYP activities¹¹ for imidazole replacements



Compound	R ₁	IC ₅₀ ^a -IGF1R (µM)	CYP1A2	CYP2C9	CYP2C19	CYP3A4
4	Imidazole	0.18	< 0.046	< 0.04	< 0.04	< 0.046
26	Н	0.77	0.38	1.5	0.26	0.71
27	CONH ₂	0.18	ND	ND	ND	ND
28	CONHMe	0.18	21.0	2.9	2.1	3.7
29	CONMe ₂	0.55	15	2.8	1.7	2.2
30	CONH-cyclopropyl	0.27	93	8.1	3.5	10
31	CONH-cyclopentyl	1.3	9.8	3.5	0.48	1.6
32	CONH-cyclohexyl	2.5	59	6.6	1.3	11
34	N	0.093	53	91	17	35
35	N H	0.054	10	67	4.7	4.3
36	N R R R R R R R R R R R R R R R R R R R	0.046	11	47	5.1	13
37	N N H K	0.068	20	100	8.5	16
38	NH N H r ⁵⁵	0.17	25	76	3.9	15
39	N N H	0.14	5.8	7.8	2.0	0.36

 a IC₅₀ values represent the average of two determinations. Standard deviations ranged from 0.03 to 0.26.

these results to identify compounds with superior ADME properties for further in vivo evaluation.

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References and notes

- (a) Baserga, R. Expert Opin. Ther. Targets 2005, 9, 753;
 (b) Garcia-Echeverria, C. IDrugs 2006, 9, 415;
 (c) Baserga, R. Oncogene 2000, 183, 1;
 (d) Valentinis, B.; Baserga, R. J. Clin. Pathol. Mol. Pathol. 2001, 54, 133.
- (a) Pollak, M. N.; Schernhammer, E. S.; Hankinson, S. E. Nat. Rev. Cancer 2004, 4, 505; (b) Yee, D. Br. J. Cancer 2006, 94, 465; (c) Hofmann, F.; Garcia-Echeverria, C. Drug Discov. Today 2005, 10, 1041; (d) Bahr, C.; Groner,

B. Growth Horm. IGF Res. 2004, 14, 287; (e) Zhang, H.; Yee, D. Expert Opin. Investig. Drugs 2004, 13, 1569.

- (a) Reiss, K.; Wang, J. Y.; Romano, G.; Furnari, F. B.; Cavenee, W. K.; Morrione, A.; Tu, X.; Baserga, R. *Oncogene* 2000, 19, 2687; (b) Reiss, K.; Wang, J. Y.; Romano, G.; Tu, X.; Peruzzi, F.; Baserga, R. *Oncogene* 2001, 20, 490.
- 4. Yu, H.; Rohan, T. J. Natl. Cancer Inst. 2000, 92, 1472.
- 5. Baserga, R. Cancer Res. 1995, 55, 249.
- (a) Garcia-Echeverria, C.; Pearson, M. A.; Marti, A.; Meyer, T.; Mestan, J.; Zimmermann, J.; Gao, J.; Brueggen, J.; Capraro, H. G.; Cozens, R.; Evans, D. B.; Fabbro, D.; Furet, P.; Porta, D. G.; Liebetanz, J.; Martiny-Baron, G.; Ruetz, S.; Hoffman, F. *Cancer Cell* 2004, *5*, 231; (b) Mitsiades, C. S.; Mitsiades, N. S.; McMullan, C. J.; Poulaki, V.; Shringarpure, R.; Akiyama, M.; Hideshima, T.; Chauhan, D.; Joseph, M.; Libermann, T. A.; Garcia-Echeverria, C.; Pearson, M. A.; Hoffmann, F.; Anderson, K. C.; Kung, A. L. *Cancer Cell* 2004, *5*, 221; (c) Bell, I. M.; Stirdivant, S. M.; Ahern, J.; Cullberson, C.; Darke, P. L.; Dinsmore, C. J.; Drakas, R. A.; Gallicchio, S. N.; Graham, S. L.; Heimbrook, D. C.; Hall, D. L.; Hua, J.;

Kett, N. R.; Kim, A. S.; Kornienko, M.; Kuo, L. C.; Munshi, S. K.; Quigley, A. G.; Reid, J. C.; Trotter, W.; Waxman, L. H.; Williams, T. M.; Zartman, C. B. Biochemistry 2005, 44, 9430; (d) Girnita, A.; Girnita, L.; del Prete, F.; Bartolazzi, A.; Larsson, O.; Axelson, M. Cancer Res. 2004, 64, 236; (e) Vasilcanu, D.; Girnita, A.; Girnita, L.; Vasilcanu, R.; Axelson, M.; Larsson, O. Oncogene 2005, 23, 7854; (f) Blum, G.; Gazit, A.; Levitzki, A. J. Biol. Chem. 2003, 273, 40442; (g) Girnita, A.; Girnita, L.; del Prete, F.; Bartolazzi, A.; Larsson, O.; Axelson, M. Cancer Res. 2004, 64, 236; (h) Sarma, P. K. S.; Tandon, R.; Gupta, P.; Dastidar, S. G.; Ray, A.; Das, B.; Cliffe, I. A. Expert Opin. Ther. Patents 2007, 17, 25; (i) Mulvihill, M. J.; Ji, Q.-S.; Werner, D.; Beck, P.; Cesario, C.; Cooke, A.; Cox, M.; Crew, A.; Dong, H.; Feng, L.; Foreman, K. W.; Mak, G.; Nigro, A.; O'Connor, M.; Saroglou, L.; Stolz, K. M.; Sujka, I.; Volk, B.; Weng, Q.; Wilkes, R. Bioorg. Med. Chem. Lett. 2007, 17, 1091.

- 7. (a) Wittman, M.; Carboni, J.; Attar, R.; Balasubramanian, B.; Balimane, P.; Brassil, P.; Beaulieu, F.; Chang, C.; Clarke, W.; Dell, J.; Eummer, J.; Frennesson, D.; Gottardis, M.; Greer, A.; Hansel, S.; Hurlburt, W.; Jacobson, B.; Krishnanathan, S.; Lee, F. Y.; Li, A.; Lin, T.-A.; Liu, P.; Ouellet, C.; Sang, X.; Saulnier, M.; Stoffan, K.; Sun, Y.; Velaparthi, U.; Wong, H.; Yang, Z.; Zimmermann, K.; Zoeckler, M.; Vyas, D. J. Med. Chem. 2005, 48, 5639; (b) Carboni, J. M.; Lee, A. V.; Hadsell, D. L.; Rowley, B. R.; Lee, F. Y.; Bol, D. K.; Camuso, A. E.; Gottardis, M.; Greer, A. F.; Ho, C. P.; Hurlburt, W.; Li, A.; Saulnier, M.; Velaparthi, U.; Wang, C.; Wen, M.-L.; Westhouse, R. A.; Wittman, M.; Zimmermann, K.; Rupnow, B. A.; Wong, T. W. Cancer Res. 2005, 65, 3781; (c) Haluska, P.; Carboni, J. M.; Loegering, D. A.; Lee, F. Y.; Wittman, M.; Saulnier, M. G.; Frennesson, D. B.; Kalli, K. R.; Conover, C. A.; Attar, R. M.; Kaufmann, S. H.; Gottardis, M.; Erlichman, C. Cancer Res. 2006, 66, 362.
- (a) Velaparthi, U.; Wittman, M.; Liu, P.; Stoffan, K.; Zimmermann, K.; Sang, X.; Carboni, J.; Li, A.; Greer, A.; Chiehying, C.; Jacobsen, B.; Sun, Y.; Jacobson, B.; Langley, D.; Balasubramanian, B.; Vyas, D. *Bioorg. Med. Chem. Lett.* 2007, *17*, 2317; (b) Wittman, D. M.; Balasubramanian, B.; Stoffan, K.; Velaparthi, U.; Krishnanathan, S.; Carboni, J.; Li, A.; Greer, A.; Chieying, C.; Jacobsen, B.; Sun, Y.; Hansel, S.; Zoeckler, M.; Vyas, D. *Bioorg. Med. Chem. Lett.* 2007, *17*, 974.
- (a) Dresser, G. K.; Spence, D.; Bailey, D. Clin. Pharmacokinet. 2000, 28, 41; (b) Tang, C.; Chiba, M.; Nishime, J.;

Hochman, J. H.; Chen, I.-W.; Williams, T. M.; Lin, J. H. Drug Metab. Dispos. 2000, 28, 680.

- 10. Yamaguchi, H. N. Yakuzaishikai Zasshi **1997**, 49, 1829.
- 11. (a) IGF-1R In Vitro Assay. The primary screen is an in vitro kinase assay that utilized the fusion of glutathione-S-transferase (GST) to the IGF-1R receptor cytoplasmic domain that contains the catalytic activity. The cytoplasmic sequence of the human IGF-1R was expressed as a recombinant protein using baculovirus insect cells and was purified by affinity chromatography on glutathione-Sepharose. The IGF-1 receptor tyrosine kinase was assayed using the synthetic polymer poly(Glu/Tyr) (Sigma Chemicals) as a phosphoacceptor substrate. Each reaction mixture was in a total volume of 50 µl and contained 125 ng of enzyme, poly(Glu/Tyr) at 50 µg/ml (2.5 ng/well final), and 1–25 μ M ATP, and 0.1 μ Ci [γ -³³P]ATP. The mixtures contained also 20 mM MOPS, pH 7.0, 5 mM MnCl₂, 0.5 mM dithiothreitol, and bovine serum albumin at 0.1 mg/ml. The reaction mixtures were incubated at 27 °C for 1 h and kinase activity was determined by quantitation of the amount of radioactive phosphate transferred to the poly(Glu/Tyr) substrate. Incorporation was measured by acid precipitation of the proteins and scintillation counting. Compounds were dissolved in dimethylsulfoxide to a concentration of 10 mM and were added to the kinase assays such that the final concentration of dimethylsulfoxide was no more than 1%, which has been shown to have no effect on kinase activity;

(b) *CYP Enzyme Assay*. The ability of test compounds to inhibit CYP microsomes (supersomes) was measured using 3-cyano-7-ethoxycoumarin (CYP1A2 and CYP2C19), 7-benzyloxy-4-trifluoromethyl coumarin (CYP3A4), 7-methoxy-4-trifluoromethyl coumarin (CYP2C9), and 3-[2-(*N*,*N*-diethyl-*N*-thylamino)ethyl]-7-methoxy-4-methyl coumarin (CYP2D6) as substrates. Assays were performed in 384-well microplates in a total volume of 30 µl. Incubations were performed with microsomes derived from baculovirus-infected cells, which were obtained from BD Gentest (Woburn, MA).

- 12. Fang, F. G.; Xie, S.; Lowery, M. W. J. Org. Chem. 1994, 59, 6142.
- The imidate ester was heated with aminoacetaldehyde diethyl acetal and the resultant amidine was subjected to acid mediated cyclization. For example, see: Walsh, J. S.; Wang, R.; Bagan, E.; Wang, C. C.; Wislocki, P.; Miwa, G. T. J. Med. Chem. 1987, 30, 150.