

Design, Synthesis and Anti-itch Activity Evaluation of Aromatic Amino Acid Derivatives as Gastrin-Releasing Peptide Receptor Antagonists

Ri-Sheng Yao*, Ting-Ting Li, Jun Xu, Lai-En Jiang and Ban-Feng Ruan

School of Medical Engineering, Hefei University of Technology, Hefei 230009, People's Republic of China

Abstract: Eight aromatic amino acid derivatives (**9a-9h**) as gastrin-releasing peptide receptor (GRPR) antagonists were designed and synthesized. For the design, the tertiary structure of GRPR was predicted by blast searching in the premise of 1u19 protein as the template. Eight target compounds were docked into the binding pocket to investigate their possible binding interactions. Their anti-itch activities were tested by intrathecal injection using Kunming mice as experimental animals and chloroquine phosphate as a modeling medium. Compounds **9e** and **9f** significantly inhibited scratching behaviors. The anti-itch activities of these compounds decreased with the following sequence: **9e** > **9f** = **9d** > **9b** > **9g** > **9a** > **9h** > **9c** predicted by computer-aided drug design (CADD) and **9e** > **9f** > **9b** > **9h** > **9g** > **9d** > **9a** > **9c** evaluated by preliminary test. They were broadly in line with activity order pretested by CADD. It showed that the predicted tertiary structure of GRPR could be used for antipruritic drug design.

Key Words: Anti-itch activity, Aromatic amino acid derivatives, Computer-aided drug design, Gastrin-releasing peptide receptor, 2-(1*H*-imidazol-1-yl) ethanol, Molecular modeling, Pruritus, Tertiary structure.

1. INTRODUCTION

Itch is an unpleasant sensation that causes the desire or reflex to scratch. It usually caused by skin disease, biliary disease, thyroid disease, kidney disease, cancer, AIDS and so on [1-3]. Chronic itch is difficult to treat [4]. However, gastrin-releasing peptide receptor, situated in the central nervous system, is the first gene directly related to the itch which confirmed by Yan-Gang Sun and Zhou-Feng Chen in 2007 [5, 6]. So GRPR may be a central target for therapeutic treatment of chronic and intractable pruritus [5].

Gastrin-releasing peptide receptor belongs to the family of the bombesin-like peptides (BLP) [7]. The human GRPR has 384 amino acids. Hydrophobic analysis of the predicted GRPR structure revealed seven regions of hydrophobic amino acids consistent with a seven-transmembrane structure typical for G protein-coupled receptors [8]. However, the tertiary structure of GRPR had not been reported.

In humans, GRPR is highly expressed in the pancreas and is also expressed in the stomach, adrenal cortex and brain [9]. GRPR activation has been proposed to be important in the mediation of a number of human disorders including disorders of lung development, various pulmonary diseases, CNS disorders, and the growth/differentiation of human cancers [10].

There have been a large number of different compounds reported to function as GRPR antagonists. They can be divided into six general classes. These six classes include substituted substance P analogs (class 1), [D-Phe¹²] bombesin analogs (class 2), modified position 13-14 bombesin or posi-

tion 26-27 GRP analogs (class 3), desMet¹⁴ or GRP²⁷ analogs (class 4), peptoids (class 5), and finally the nonpeptide analogs, kuwanon G and kuwanon H (class 6) [10-12]. Hereinto, PD 176252 is a peptoid antagonist that has nanomolar affinity for GRPR ($K_i = 1$ nM). Subsequent studies demonstrated that PD 176252 inhibited the growth of lung cancer cells, potentiated the growth inhibitory effects of histone deacetylase inhibitors; inhibited GRP/Bnstimulated signaling in lung cancer cells (Ca^{2+} and tyrosine phosphorylation of p125^{F_{AK}}) and the stimulation of increases in c-fos mRNA and growth, and in rats had an anxiolytic effect *in vivo* [13]. All classes are peptides or peptoid antagonists, except for class 6, which are flavone derivatives, isolated from extracts of the mulberry tree *Morus bombycis* [14].

So far, few small molecule compounds with anti-itch activity as GRPR antagonists have been reported. The GRPR antagonists are mainly peptides, and they are not suitable as molecular design reference for the high molecular weights and complex structures. The non-peptides, kuwanon G and kuwanon H (Fig. 1), have too many rotatable bonds and are difficult to synthesize by chemical methods. So they also don't fit as molecular design reference. The peptoids, especially PD 176252 with high affinity, can be used as reference. To discover some innovative drugs for anti-itch, the tertiary structure of GRPR was predicted with a homologous protein, then with the reported GRPR antagonists, especially PD 176252, eight compounds (Fig. 2) were designed, synthesized and preliminarily tested the anti-itch activity.

2. RESULTS AND DISCUSSION

2.1. Chemistry

Compounds **9a-9h** were composed of three fragments, see Fig. (2). They were synthesized of L-phenylalanine or L-tryptophan (fragment 2) as the basic skeleton by esterifica-

*Address correspondence to this author at the School of Medical Engineering, Hefei University of Technology, Hefei 230009, People's Republic of China; Tel: +86-0551-2901771; Fax: +86-0551-2904675; E-mail: rishengyao@163.com

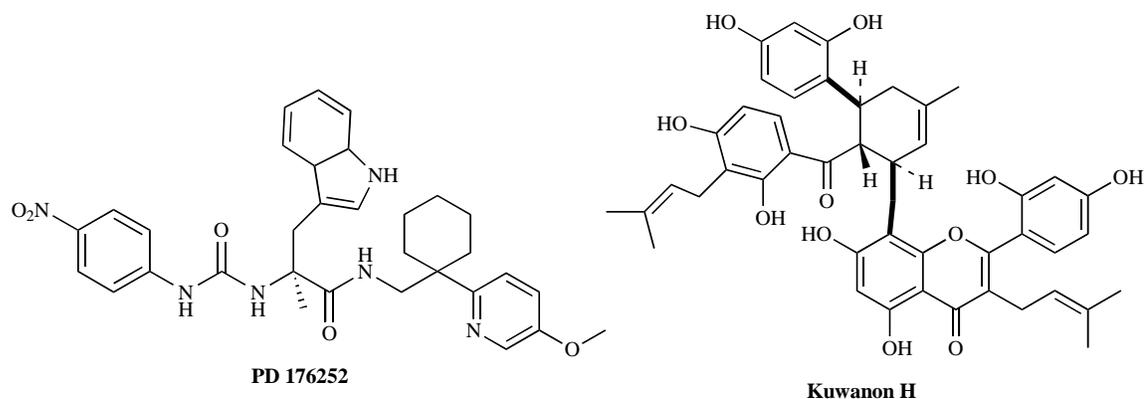
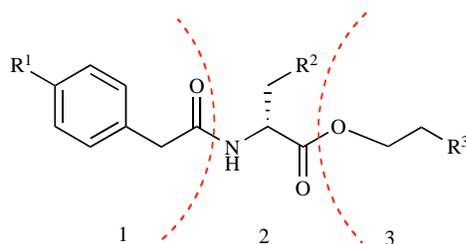


Fig. (1). The Structures of PD 176252 and Kuwanon H.



Comp. No	R ¹	R ²	R ³
9a	NO ₂	phenyl	1-imidazole
9b	F	phenyl	1-imidazole
9c	CH ₃	phenyl	1-imidazole
9d	NO ₂	3-indole	1-imidazole
9e	F	3-indole	1-imidazole
9f	CH ₃	3-indole	1-imidazole
9g	F	3-indole	4-morpholine
9h	F	3-indole	4-(1-methylpiperazine)

Fig. (2). The Structures of Compounds **9a-9h**.

tion with the methanol, amidation with alphetoluic acid derivatives (fragment 1), then deprotection and esterification with ethanol derivatives (fragment 3).

In Scheme 1, ethyl 2-(1*H*-imidazol-1-yl) acetate (**1**) was prepared in 79.0% yield by refluxing imidazole in THF with ethyl 2-chloroacetate for 10 h [15, 16]. NaBH₄ was slowly added to a solution of **1** in MeOH at 0 °C. Then heating to 50 °C, 2-(1*H*-imidazol-1-yl) ethanol (**2**) was obtained for 8 h in a good yield [17]. While 2-morpholinoethanol and 2-(4-methylpiperazin-1-yl) ethanol were synthesized with the total yields of 76.6% and 72.2% applying the same preparation protocols followed in Scheme 1. The only difference was the use of morpholine and methylpiperazine instead of imidazole used in Scheme 1.

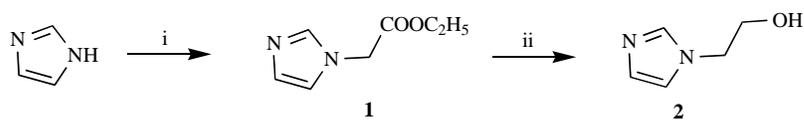
As shown in Scheme 2, compounds **9a-9f** were synthesized using L-Phenylalanine or L-Tryptophan as starting material. Firstly, SOCl₂ was added to MeOH at 0 °C for 1 h. After the addition of **3** and refluxing for 5 h, **4** was obtained [18]. Secondly, **7** was carried out by the dropwise addition of

the new prepared acetyl chloride derivative **6** to a solution of **4**. Then, the hydrolysis of **7** was conducted in a mixed solvent of MeOH and water in a (4:1, V/V) ratio and in the presence of Na₂CO₃ [19]. Finally, compounds **9a-9f** were prepared by the reaction of DCC condensation. Compounds **9g** and **9h** were synthesized applying the same preparation protocols followed in Scheme 2. The only difference was the use of 2-morpholinoethanol and 2-(4-methylpiperazin-1-yl) ethanol instead of 2-(1*H*-imidazol-1-yl) ethanol.

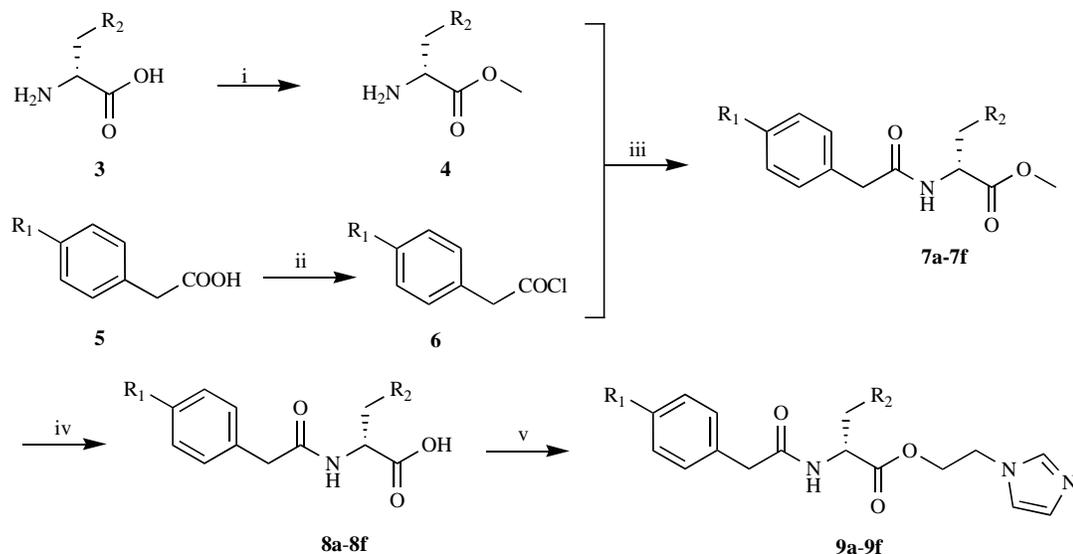
The structures of intermediates and target compounds were confirmed by FT-IR and ¹H NMR.

2.2. Molecular Modeling

In order to get a better insight into the binding affinity, molecular modeling studies were performed using Molecular Operating Environment (MOE). 2008. 10 software. Since no crystal structure of GRPR is yet available, the tertiary structure of GRPR was predicted using the crystal structure given in PDB code 1u19 (Fig. 3).



Scheme 1. Reagents and Conditions: (i) Ethyl 2-chloroacetate, NEt_3 , THF, Reflux, 10 h, 79%; (ii) NaBH_4 , MeOH, 0°C to 50°C , 8 h, 90%.



Scheme 2. Reagents and Conditions: (i) SOCl_2 , MeOH, 0°C to Reflux, 5 h; (ii) $(\text{COCl})_2$, NEt_3 , Cat. DMF, DCM, N_2 , 0°C to rt, 2 h; (iii) NEt_3 , DCM, N_2 , 0°C to rt, 5 h; (iv) Na_2CO_3 , MeOH/ H_2O (4:1), rt, 6 h; (v) 2-(1H-imidazol-1-yl)ethanol, DCC, cat. DMAP, THF, rt, 15 h.

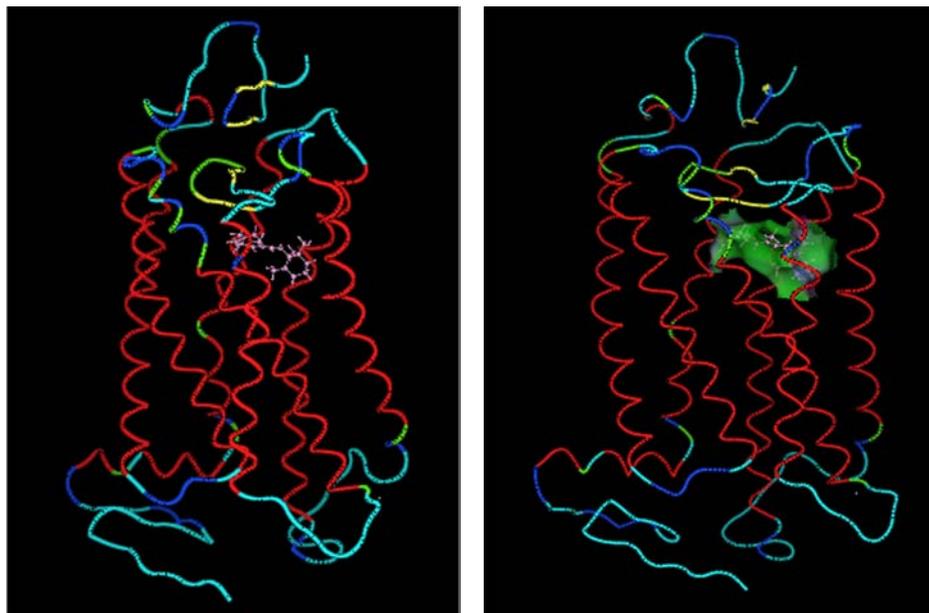


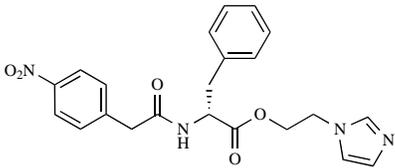
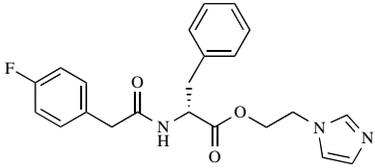
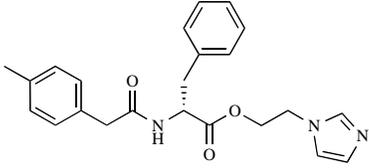
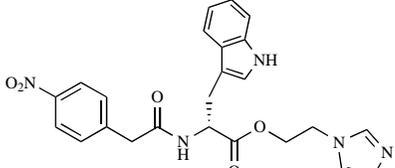
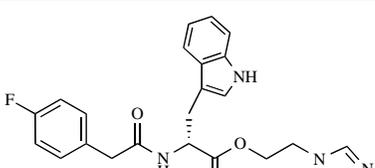
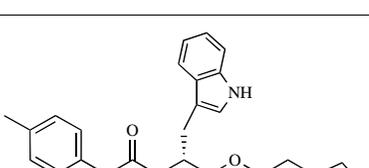
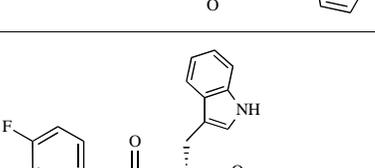
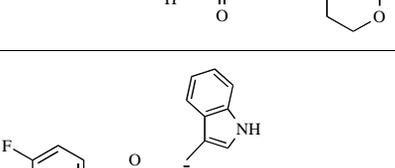
Fig. (3). The Tertiary Structure of GRPR.

As well as GRPR, 1u19 protein belongs to the family of G protein coupled receptor. By blast searching, the two proteins have a high degree of similarity in the structure and the identities and positives are 22% and 42%, respectively. So 1u19 protein was chosen as the template of homology modeling.

Compounds **9a-9h** were docked over the ligand atoms in the binding site. They occupy nearly the same space in the binding pocket, and are overlaid well over the skeleton of the ligand.

In compound **9a** (Fig. 4), the benzene ring of L-phenylalanine establishes an arene-cation interaction with Lys-188. The left C=O forms a hydrogen bonding interaction with the backbone NH of Lys-265 and the right C=O forms a hydrogen bonding interaction with Val-187 through H_2O . The 4-nitro-benzene shows an arene-cation interaction with His-186. The binding interactions of compounds **9b** and **9c** (Table 1) are the same with **9a**. Compounds **9d** and **9f** both have one more interaction with His-207 than **9a**. Compound **9e** as well as **9a**, interacts with Lys-265, Val-187, and His-

Table 1. The Binding Interactions of Compounds 9a-9h by CADD

Comp. No	Structure	Binding Interactions
9a		Lys-188, Lys-265, Val-187, His-186
9b		Lys-188, Lys-265, Val-187, His-186
9c		Lys-188, Lys-265, Val-187, His-186
9d		Lys-188, Lys-265, Val-187, His-186, His-207
9e		Lys-265, Val-187, His-186, His-207
9f		Lys-188, Lys-265, Val-187, His-186, His-207
9g		Lys-265, Val-187, His-186, His-207
9h		Lys-265, Val-187, His-186, His-207

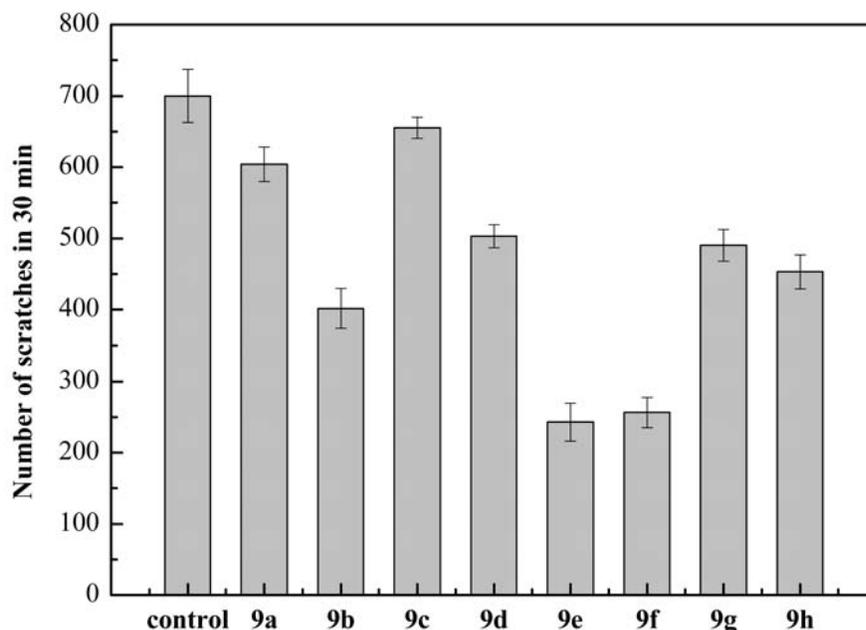


Fig. (5). The Anti-itch Activity Results of Compounds **9a-9h**. The First line was the Number of Scratching Behaviors of Control Animals and the others were the Numbers of Scratching Behaviors of Treatment Animals. All the Treatments Resulted in a Significant Decrease in Scratching Numbers Compared to the Control ($p < 0.05$).

3. CONCLUSION

Eight compounds were designed and synthesized. Compounds **9e** and **9f**, which had prominence activities, were screened out as GRPR activity antagonists. The optimum structure of fragment 1 was 2-(4-fluorophenyl) acetic acid, fragment 2 was L-tryptophan and fragment 3 was 2-(1H-imidazol-1-yl) ethanol. The anti-itch activity of them decreased with the following sequence: **9e** > **9f** = **9d** > **9b** > **9g** > **9a** > **9h** > **9c** by CADD and **9e** > **9f** > **9b** > **9h** > **9g** > **9d** > **9a** > **9c** by preliminary test. On the whole, they were broadly in line with activity order pretested by CADD. It showed that the predicted GRPR tertiary structure was of a certain reference value and it can be used for antipruritic drug design.

4. EXPERIMENTAL SECTION

4.1. General

The ^1H NMR spectra were recorded on a Bruker 300 MHz spectrometer. The chemical shifts were expressed in ppm using residual CDCl_3 and $(\text{CD}_3)_2\text{SO}$ as reference. The FT-IR spectra were recorded in solid-state KBr dispersion on a Perkins-Elmer FT-IR spectrophotometer. The melting points were taken on a Buchi apparatus. All reactions were monitored by TLC with HuanghaiGF 254 silica gel-coated plates. Commercially obtained reagents were used without further purification. Compounds **1**, **2** and **4** were prepared according to the literature methods as described [15-18].

4.2. Synthesis

4.2.1. General Procedure for the Synthesis of Compounds **7a-7h**

The mixture of alphaltoluic acid derivatives (**5**) (18 mmol), DCM (30 mL), TEA (2.6 mL, 18 mmol) and DMF (0.1 mL) was stirred at 0°C under the protection of N_2 gas.

$(\text{COCl})_2$ (1.84 mL, 21.6 mmol) in DCM (10 mL) was slowly added to the mixture. After the dropwise addition, the mixture was stirred at room temperature for 2 h. Solvent and excess $(\text{COCl})_2$ was removed under vacuum. The brown solid was obtained, which was directly used in the next step.

The suspension of new prepared acetyl chloride derivative **6** in DCM (15 mL) was added to the mixture of **4** (18 mmol), TEA (5.2 mL, 36 mmol) and DCM (20 mL) at 0°C under the protection of N_2 gas. After the dropwise addition, the mixture was stirred at room temperature for 5 h. Compounds **7a-7h** were obtained by column chromatography (silica gel, 200-300 mesh, ethyl acetate/petroleum ether, 1:2, V/V).

4.2.1.1. (R)-methyl-2-(2-(4-nitrophenyl)acetamido)-3-phenylpropanoate (**7a**)

The title compound was separated by column chromatography. Yield (5.62 g, 91.2%); yellow solid; ^1H NMR (CDCl_3) δ : 6.94-8.17 (m, 9H), 4.88 (m, 1H), 3.72 (s, 3H), 3.66 (s, 2H), 3.11 (d, 2H); IR ν/cm^{-1} : 3284, 2925, 1743, 1665, 1542, 1345, 1280, 1172, 825, 701.

4.2.1.2. (R)-methyl-2-(2-(4-fluorophenyl)acetamido)-3-phenylpropanoate (**7b**)

The title compound was separated by column chromatography. Yield (5.23 g, 92.3%); cream white solid; ^1H NMR (CDCl_3) δ : 6.89-7.26 (m, 9H), 4.85 (m, 1H), 3.71 (s, 3H), 3.50 (s, 2H), 3.06 (dd, 2H); IR ν/cm^{-1} : 3285, 2953, 1755, 1669, 1546, 1512, 1364, 1222, 705.

4.2.1.3. (R)-methyl-3-phenyl-2-(2-p-tolylacetamido) propanoate (**7c**)

The title compound was separated by column chromatography. Yield (4.89 g, 87.3%); white solid; ^1H NMR (CDCl_3) δ : 6.87-7.24 (m, 9H), 4.84 (m, 1H), 3.69 (s, 3H), 3.48 (s,

2H), 3.02 (d, 2H), 2.34 (s, 3H); IR ν/cm^{-1} : 3319, 2925, 1757, 1646, 1533, 1438, 1373, 1275, 1171, 782, 699.

4.2.1.4. (R)-methyl-3-(1H-indol-3-yl)-2-(2-(4-nitrophenyl)acetamido)propanoate (7d)

The title compound was separated by column chromatography. Yield (6.03 g, 87.8%); brown solid; $^1\text{H NMR}$ (CDCl_3) δ : 7.03-8.04 (m, 9H), 4.91 (m, 1H), 3.72 (s, 3H), 3.54 (s, 2H), 3.29 (d, 2H); IR ν/cm^{-1} : 3381, 2925, 1734, 1655, 1520, 1438, 1346, 1106, 741.

4.2.1.5. (R)-methyl-2-(2-(4-fluorophenyl)acetamido)-3-(1H-indol-3-yl)propanoate (7e)

The title compound was separated by column chromatography. Yield (5.82 g, 91.2%); cream white solid; $^1\text{H NMR}$ (CDCl_3) δ : 8.07 (s, 1H), 6.74-7.44 (m, 9H), 4.90 (m, 1H), 3.68 (s, 3H), 3.45 (s, 2H), 3.25 (d, 2H); IR ν/cm^{-1} : 3407, 3291, 2923, 1733, 1662, 1547, 1513, 1451, 1361, 1220, 830, 746.

Compounds **7g** and **7h** are the same as **7e**.

4.2.1.6. (R)-methyl-3-(1H-indol-3-yl)-2-(2-p-tolylacetamido)propanoate (7f)

The title compound was separated by column chromatography. Yield (5.74 g, 91.1%); white solid; $^1\text{H NMR}$ (CDCl_3) δ : 8.02 (s, 1H), 6.72-7.42 (m, 9H), 4.90 (m, 1H), 3.66 (s, 3H), 3.48 (s, 2H), 3.24 (d, 2H), 2.32 (s, 3H); IR ν/cm^{-1} : 3331, 2924, 2855, 1751, 1652, 1517, 1461, 1377, 1198, 753, 659.

4.2.2. General Procedure for the Synthesis of Compounds 8a-8h

Compound **7** (5 mmol) was added to a solution of MeOH (32 mL), H_2O (8 mL) and Na_2CO_3 (0.79 g, 7.5 mmol) [19]. The mixture was stirred at room temperature for 6 h. MeOH was removed under vacuum, and H_2O (20 mL) was added to the residue. pH was adjusted to 5 to 6 using 3 N HCl. The mixture was extracted with ethyl acetate (50 mL \times 3). The organic layer was separated, dried over anhydrous Na_2SO_4 and then evaporated under vacuum.

4.2.2.1. (R)-2-(2-(4-nitrophenyl)acetamido)-3-phenylpropanoic acid (8a)

The title compound was evaporated under vacuum. Yield (1.53 g, 93.2%); yellow solid; mp 165~168 $^\circ\text{C}$; $^1\text{H NMR}$ ($\text{CD}_3\text{SO}-d_6$) δ : 12.73 (s, 1H), 7.19-8.53 (m, 9H), 4.45 (m, 1H), 3.57 (s, 2H), 3.01 (d, 2H); IR ν/cm^{-1} : 3319, 2894, 1711, 1621, 1520, 1347, 1246, 1050, 831, 700.

4.2.2.2. (R)-2-(2-(4-fluorophenyl)acetamido)-3-phenylpropanoic acid (8b)

The title compound was evaporated under vacuum. Yield (1.41 g, 94.0%); cream white solid; mp 152~154 $^\circ\text{C}$; $^1\text{H NMR}$ (CDCl_3) δ : 6.94-7.26 (m, 9H), 4.81 (m, 1H), 3.51 (s, 2H), 3.08 (d, 2H); IR ν/cm^{-1} : 3295, 2928, 1709, 1613, 1559, 1355, 1251, 1225, 1158, 825, 700.

4.2.2.3. (R)-3-phenyl-2-(2-p-tolylacetamido)propanoic acid (8c)

The title compound was evaporated under vacuum. Yield (1.34 g, 90.4%); white solid; mp 124~125 $^\circ\text{C}$; $^1\text{H NMR}$ (CDCl_3) δ : 6.91-7.25 (m, 9H), 4.80 (m, 1H), 3.49 (s, 2H),

3.06 (d, 2H), 2.35 (s, 3H); IR ν/cm^{-1} : 3358, 2924, 1770, 1743, 1611, 1534, 1396, 1209, 1170, 784, 700.

4.2.2.4. (R)-3-(1H-indol-3-yl)-2-(2-(4-nitrophenyl)acetamido)propanoic acid (8d)

The title compound was evaporated under vacuum. Yield (1.68 g, 91.4%); brown solid; mp 211~214 $^\circ\text{C}$; $^1\text{H NMR}$ ($\text{CD}_3\text{SO}-d_6$) δ : 10.84 (s, 1H), 6.94-8.50 (m, 9H), 4.51 (m, 1H), 3.59 (s, 2H), 3.15 (d, 2H); IR ν/cm^{-1} : 3402, 3305, 2926, 1711, 1618, 1515, 1348, 1243, 747.

4.2.2.5. (R)-2-(2-(4-fluorophenyl)acetamido)-3-(1H-indol-3-yl)propanoic acid (8e)

The title compound was evaporated under vacuum. Yield (1.62 g, 95.2%); cream white solid; mp 196~197 $^\circ\text{C}$; $^1\text{H NMR}$ ($\text{CD}_3\text{SO}-d_6$) δ : 10.83 (s, 1H), 8.32 (s, 1H), 6.94-7.52 (m, 9H), 4.47 (m, 1H), 3.42 (s, 2H), 3.14 (dd, 2H); IR ν/cm^{-1} : 3408, 3309, 2925, 1709, 1615, 1508, 1460, 1218, 1170, 793, 746.

Compounds **8g** and **8h** are the same as **8e**.

4.2.2.6. (R)-3-(1H-indol-3-yl)-2-(2-p-tolylacetamido)propanoic acid (8f)

The title compound was evaporated under vacuum. Yield (1.63 g, 96.9%); gray oily substance; $^1\text{H NMR}$ ($\text{CD}_3\text{SO}-d_6$) δ : 10.83 (s, 1H), 8.23 (s, 1H), 6.94-7.53 (m, 9H), 4.48 (m, 1H), 3.38 (s, 2H), 3.06 (d, 2H), 2.25 (s, 3H); IR ν/cm^{-1} : 3403, 2924, 1728, 1624, 1515, 1344, 1232, 1100, 743.

4.2.3. General Procedure for the Synthesis of Compounds 9a-9f

A mixture of compound **8** (4 mmol), THF (10 mL), DCC (0.99 g, 4.8 mmol) and DMAP (10 mg) was stirred at 0 $^\circ\text{C}$ for 1 h. Compound **2** in THF (5 mL) was added to the solution. The mixture was stirred at room temperature for 15 h. Target compound was obtained by column chromatography (silica gel, 200-300 mesh, DCM/MeOH, 70:1, V/V).

4.2.3.1. (R)-2-(1H-imidazol-1-yl)ethyl-2-(2-(4-nitrophenyl)acetamido)-3-phenylpropanoate (9a)

The title compound was separated by column chromatography. Yield (1.52 g, 90.1%); brown oily substance; $^1\text{H NMR}$ (CDCl_3) δ : 6.92-8.15 (m, 12H), 4.78 (m, 1H), 4.36 (t, 2H), 4.15 (t, 2H), 3.61 (s, 2H), 3.01 (d, 2H); IR ν/cm^{-1} : 3206, 2941, 1746, 1641, 1532, 1350, 1283, 1178, 820, 698.

4.2.3.2. (R)-2-(1H-imidazol-1-yl)ethyl-2-(2-(4-fluorophenyl)acetamido)-3-phenylpropanoate (9b)

The title compound was separated by column chromatography. Yield (1.43 g, 90.5%); yellow oily substance; $^1\text{H NMR}$ (CDCl_3) δ : 8.02 (s, 1H), 6.72-7.42 (m, 9H), 4.90 (m, 1H), 3.66 (s, 3H), 3.48 (s, 2H), 3.24 (d, 2H), 2.32 (s, 3H); IR ν/cm^{-1} : 3201, 2932, 1747, 1658, 1543, 1509, 1355, 1222, 1159, 825, 702.

4.2.3.3. (R)-2-(1H-imidazol-1-yl)ethyl-3-phenyl-2-(2-p-tolylacetamido)propanoate (9c)

The title compound was separated by column chromatography. Yield (1.41 g, 90.0%); white oily substance; $^1\text{H NMR}$ (CDCl_3) δ : 6.81-7.35 (m, 12H), 4.75 (m, 1H), 4.21 (t, 2H),

4.08 (t, 2H), 3.49 (s, 2H), 2.95 (d, 2H), 2.34 (s, 3H); IR ν/cm^{-1} : 3206, 2941, 1746, 1641, 1532, 1350, 1283, 1178, 820, 698.

4.2.3.4. (R)-2-(1H-imidazol-1-yl)ethyl-3-(1H-indol-3-yl)-2-(2-(4-nitrophenyl)acetamido)propanoate (9d)

The title compound was separated by column chromatography. Yield (1.72 g, 93.2%); brown oily substance; ^1H NMR (CDCl_3) δ : 8.78 (s, 1H), 6.65-8.02 (m, 12H), 4.81 (m, 1H), 4.26 (t, 2H), 3.96 (t, 2H), 3.55 (s, 2H), 3.17 (dd, 2H); IR ν/cm^{-1} : 2924, 1744, 1662, 1518, 1346, 1184, 822, 745.

4.2.3.5. (R)-2-(1H-imidazol-1-yl)ethyl-2-(2-(4-fluorophenyl)acetamido)-3-(1H-indol-3-yl)propanoate (9e)

The title compound was separated by column chromatography. Yield (1.63 g, 93.8%); yellow oily substance; ^1H NMR (CDCl_3) δ : 8.67 (s, 1H), 6.58-7.38 (m, 12H), 4.81 (m, 1H), 4.22 (t, 2H), 3.89 (t, 2H), 3.46 (s, 2H), 3.17 (dd, 2H); IR ν/cm^{-1} : 3274, 1744, 1657, 1509, 1356, 1223, 825, 745.

4.2.3.6. (R)-2-(1H-imidazol-1-yl)ethyl-3-(1H-indol-3-yl)-2-(2-p-tolylacetamido)propanoate (9f)

The title compound was separated by column chromatography. Yield (1.62 g, 94.1%); white oily substance; ^1H NMR (CDCl_3) δ : 8.36 (s, 1H), 6.56-7.38 (m, 12H), 4.81 (m, 1H), 4.20 (t, 2H), 3.86 (t, 2H), 3.48 (s, 2H), 3.15 (dd, 2H), 2.32 (s, 3H); IR ν/cm^{-1} : 3265, 2924, 1744, 1656, 1514, 1438, 1355, 1233, 1187, 745.

4.2.3.7. (R)-2-morpholinoethyl-2-(2-(4-fluorophenyl)acetamido)-3-(1H-indol-3-yl)propanoate (9g)

The title compound was separated by column chromatography. Yield (1.59 g, 87.6%); yellow oily substance; ^1H NMR (CDCl_3) δ : 8.13 (s, 1H), 6.82-7.46 (m, 9H), 4.90 (m, 1H), 4.20 (t, 2H), 3.65 (t, 4H), 3.48 (s, 2H), 3.26 (d, 2H), 2.53 (t, 2H), 2.38 (t, 4H); IR ν/cm^{-1} : 3293, 2926, 1740, 1657, 1509, 1457, 1354, 1222, 1115, 744.

4.2.3.8. (R)-2-(4-methylpiperazin-1-yl)ethyl-2-(2-(4-fluorophenyl)acetamido)-3-(1H-indol-3-yl)propanoate (9h)

The title compound was separated by column chromatography. Yield (1.68 g, 90.0%); yellow oily substance; ^1H NMR (CDCl_3) δ : 8.22 (s, 1H), 6.85-7.45 (m, 9H), 4.89 (m, 1H), 4.16 (t, 2H), 3.65 (t, 4H), 3.44 (s, 2H), 3.26 (d, 2H), 2.53 (t, 2H), 2.42 (s, 8H), 2.27 (s, 3H); IR ν/cm^{-1} : 3278, 2925, 1742, 1658, 1511, 1450, 1353, 1226, 1125, 742.

4.3. Biological Evaluation

4.3.1. Animals

In all biological experiments, Kunming mice, three of either sex, 6 to 12 weeks of age, and 18 to 22 g of weight, were used as experimental animals. The animals were housed for a week before this experiment in an air-conditioned room with controlled temperature (22 ± 2 °C) and illumination time (08: 00 to 20: 00). Food and water were provided ad libitum [20, 21].

4.3.2. Solution Preparation

4.3.2.1. The Preparation of Chloroquine Phosphate Solution

With saline as a solvent, different concentrations (200 $\mu\text{g}/100$ μL , 400 $\mu\text{g}/100$ μL , 600 $\mu\text{g}/100$ μL , 645 $\mu\text{g}/100$ μL ,

800 $\mu\text{g}/100$ μL) of chloroquine phosphate solutions were prepared, filtered by 0.22 μm water film, and loaded 1 mL in each sample device in the sterile console.

4.3.2.2. The Preparation of Eight Target Compounds Solutions

With saline as a solvent, and 2 mL Tween 80 and 2.5 mL glycerol as solubilizers, 10 nmol/2.5 μL eight target compounds solutions were prepared, filtered by 0.22 μm water film, and loaded 1 mL in each sample device in the sterile console.

4.3.3. Statistical Analysis

Data were expressed as mean \pm standard deviation and they were analyzed by Student's t-test. Values of $p < 0.05$ were considered significant.

CONFLICT OF INTEREST

The author(s) confirm that this article content has no conflicts of interest.

ACKNOWLEDGMENTS

This work was supported by Key Program of Natural Science Research by Education Department of Anhui Province (2011AJZR0919) and Condition Construction Cost of Hefei University of Technology (407-037022).

REFERENCES

- [1] Oliveira, F.A.; Lima, R.C.; Cordeiro, W.M.; Vieira, G.M.; Chaves, M.H.; Almeida, F.R.; Silva, R.M.; Santos, F.A.; Rao, V.S. Pentacyclic triterpenoids, α , β -amyriins, suppress the scratching behavior in a mouse model of pruritus. *Pharmacol. Biochem. Behav.*, **2004**, *78*, 719-725.
- [2] Ständer, S.; Weishaar, E.; Metang, T.; Szepletowski, J.C.; Carstens, E.; Ikoma, A.; Bergasa, N.V.; Gieler, U.; Misery, L.; Wallengren, J.; Darsow, U.; Streit, M.; Metz, D.; Luger, T.A.; Greaves, M.W.; Schmelz, M.; Yosipovitch, G.; Bernhard, J.D. Clinical classification of itch: a position paper of the international forum for the study of itch. *Acta. Derm. Venereol.*, **2007**, *87*, 291-294.
- [3] Twycross, R.; Greave, M.W.; Handwerker, H.; Jones, E.A.; Libretto, S.E.; Szepletowski, J.C.; Zylicz, Z. Itch: scratching more than the surface. *Q. J. Med.*, **2003**, *96*, 7-26.
- [4] Lee, S.; Xie, J.; Chen, X.Y. Peptides and peptide hormones for molecular imaging and disease diagnosis. *Chem. Rev.*, **2010**, *110*, 3087-3111.
- [5] Sun, Y.G.; Chen, Z.F. A gastrin-releasing peptide receptor mediates the itch sensation in the spinal cord. *Nature*, **2007**, *448*, 700-704.
- [6] Swain, M.G. Gastrin-releasing peptide and pruritus: More than just scratching the surface. *Hepatology*, **2008**, *48*, 681-683.
- [7] Baldwin, G.S.; Patel, O.; Shulkes, A. Phylogenetic analysis of the sequences of gastrin-releasing peptide and its receptors: biological implications. *Regul. Pept.*, **2007**, *143*, 1-14.
- [8] Corjay, M.H.; Dobrzanski, D.J.; Way, J.M.; Viallet, J.; Shapira, H.; Worland, P.; Sausville, E.A.; Battey, J.F. Two distinct bombesin receptor subtypes are expressed and functional in human lung carcinoma cells. *J. Biol. Chem.*, **1991**, *266*, 18771-9.
- [9] Xiao, D.; Wang, J.; Hampton, L.L.; Weber, H.C. The human gastrin-releasing peptide receptor gene structure, its tissue expression and promoter. *Gene*, **2001**, *264*, 95-103.
- [10] Jensen, R.T.; Battey, J.F.; Spindel, E.R.; Benya, R.V. International union of pharmacology. LXVIII. Mammalian bombesin receptors: nomenclature, distribution, pharmacology, signaling, and functions in normal and disease states. *Pharmacol. Rev.*, **2008**, *60*, 1-42.

- [11] Jensen, R.T.; Coy, D.H. Progress in the development of potent bombesin receptor antagonists. *Trends. Pharmacol. Sci.*, **1991**, *12*, 13-19.
- [12] Tokita, K.; Katsuno, T.; Hocart, S.J.; Coy, D.H.; Llinares, M.; Martinez, J.; Jensen, R.T. Molecular basis for selectivity of high affinity peptide antagonists for the gastrin-releasing peptide receptor. *J. Biol. Chem.*, **2001**, *276*, 36652-36663.
- [13] Ashwood, V.; Brownhill, V.; Higginbottom, M.; Horwell, D.C.; Hughes, J.; Lewthwaite, R.A.; McKnight, A.T.; Pinnock, R.D.; Pritchard, M.C.; Suman-Chauhan, N.; Webb, C.; Williams, S.C. PD 176252-The first high affinity non-peptide gastrin-releasing peptide (BB₂) receptor antagonist. *Bioorg. Med. Chem. Lett.*, **1998**, *8*, 2589-2594.
- [14] Mihara, S.; Hara, M.; Nakamura, M.; Sakurawi, K.; Tokura, K.; Fujimoto, M.; Fukai, T.; Ncmura, T. Non-peptide bombesin receptor antagonists, kuwanon G and H, isolated from mulberry. *Biochem. Biophys. Res. Commun.*, **1995**, *213*, 594-599.
- [15] Zaderenko, P.; Gil, M.S.; Ballesteros, P. Synthesis and regioselective hydrolysis of 2-Imidazol-1-ylsuccinic esters. *J. Org. Chem.*, **1994**, *59*, 6268-6273.
- [16] Martin, R.M.; Ortega, E.; Rojas, M.L.; Vicente, M.A.; Banares, M.A. Sonocatalysis and basic clays. Michael addition between imidazole and ethyl acrylate. *Catal. Lett.*, **2002**, *84*, 201-204.
- [17] Wu, Y.M.; Ge, Y.H.; Xue, Z.J. A New Synthesis method of indole-4-carboxaldehyde. *Chin. J. Syn. Chem.*, **2004**, *12*, 333-335.
- [18] Brenner, M.; Huber, W. Herstellung von α -Aminosäureestern durch Alkoholyse der Methylester. *Helv. Chim. Acta.*, **1953**, *36*, 1109-1115.
- [19] Lakshminarayana, N.; Prasad, Y.R.; Gharat, L.; Thomas, A.; Ravikumar, P.; Narayanan, S.; Srinivasan, C.V.; Gopalan, B. Synthesis and evaluation of some novel isochroman carboxylic acid derivatives as potential anti-diabetic agents. *Eur. J. Med. Chem.*, **2009**, *44*, 3147-57.
- [20] Fan, Y.M.; Zang, H.T.; Yin, G.F.; Lin, C.Y.; Hu, D.S.; Ru, L.Q. Primary afferent tract tracing of itch signal in rat itch modle. *Chin. J. Derm. Venereol.*, **2006**, *20*, 595-596.
- [21] Nojima, H.; Carstens, E. Quantitative assessment of directed hind limb scratching behavior as a rodent itch model. *J. Neurosci Meth.*, **2003**, *126*, 137-43.
- [22] El-Deeb, I.M.; Lee, S.H. Design and synthesis of new anticancer pyrimidines with multiple-kinase inhibitory effect. *Bioorg. Med. Chem.*, **2010**, *18*, 3860-3874.

Received: July 18, 2011

Revised: April 25, 2012

Accepted: April 26, 2012