Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Aryloxy cyclohexyl imidazoles: A novel class of antileishmanial agents st

Nagarapu Srinivas^a, Shraddha Palne^b, Nishi^b, Suman Gupta^b, Kalpana Bhandari^{a,*}

^a Medicinal and Process Chemistry Division, Central Drug Research Institute, M.G. Marg, Lucknow, U.P. 226001, India
^b Division of Parasitology, Central Drug Research Institute, Lucknow 226001, India

ARTICLE INFO

ABSTRACT

Article history: Received 27 August 2008 Revised 19 November 2008 Accepted 24 November 2008 Available online 27 November 2008

Keywords: Leishmania Azoles Imidazoles Aryloxy cyclohexane Thirteen novel aryloxy cyclohexane-based mono and bis imidazoles were synthesized and evaluated in vitro as antileishmanials against *Leishmania donovani* and cytotoxicity assessed. These compounds were better than the existing drugs, sodium stibogluconate and pentamidine in respect to IC_{50} and SI values. Promising compounds were tested further in vivo. Among all, the bis methylimidazole with 2-fluoro, 4-nitro aryloxy group (**9**) exhibited significant in vivo inhibition of 77.9%, thus providing new structural lead for antileishmanials.

© 2008 Elsevier Ltd. All rights reserved.

Leishmaniasis is a group of parasitic diseases of global distribution transmitted by the bite of the infected female phlebotomine sandfly¹ and manifest with visceral, cutaneous, and mucocutaneous forms. This disease is currently prevalent in four continents, being endemic in 88 countries, threatening 350 millions worldwide.² Chemotherapy for these parasitic diseases is generally ineffective mainly due to the emergence of drug-resistant strains, significant toxicity, variable efficacy, lack of oral bioavailability and high cost of the therapeutic agents.^{3,4} The pentavalent antimonials are widely used as primary therapy whereas alternative drugs include amphotericin B, pentamidine, paromomycin, miltefosine and azoles,^{5,6} all suffer from one or more of the above deficiencies. Drug resistance, high toxicity and high treatment costs necessitate the need for novel therapeutics.⁷

Among potential orally active drugs for the treatment of these complex diseases, sterol biosynthesis inhibitors offer an attractive possibility, as *Leishmania* parasites synthesize specific sterols which seem to be essential for cell proliferation and viability.^{8,9} Azole antifungal agents, have been used as antileishmanial agents since 1980s,^{10–12} inhibit the growth of *Leishmania* amastigotes in culture systems by inhibiting the cytochrome P-450-mediated 14a-demethylation of lanosterol, blocking ergosterol synthesis, and causing accumulation of 14a-methyl sterols.^{8,10} Metronidazole and *N*-substituted azoles (ketoconazole, miconazole, fluconazole and itraconazole) are well-tolerated drugs^{13,14} that are potentially active against *Leishmania*, but their use in the treatment of cutaneous and visceral leishmaniasis has produced conflicting results.^{15,16}

Based on above report, we recently prepared a series of novel aryloxy benzocycloalkyl azoles and found that they were highly active in vitro against *Leishmania donovani* and also exhibited significant in vivo activity in *L. donovani*/Hamster model. In view of above and our continuation of studies on chemotherapy of *Leishmania*, we decided to synthesize an expanded series of aryloxy cycloalkyl azoles, and investigated their biological effects against the *Leishmania* parasites and the results are reported in this communication.

The compounds used in the present study were prepared from cyclohexanone. The synthetic route for the preparation of 2,6-bisimidazolyl-methyl-1-aryloxycyclohexane (**5–9**) is outlined in Scheme 1. Mannich reaction on cyclohexanone with 2 moles of pyrrolidine gave 2,6-bis-pyrrolidin-1-ylmethyl-cyclohexanone (**2**) which on reaction with imidazole in presence of ethanol/water resulted in the replacement of pyrrolidine moiety with imidazole (**3**). The keto intermediate **3** was then transformed to corresponding hydroxy derivative **4** in a single diasteriomeric form, which on reaction with proper aryl halides furnished the desired 2,6-bis-imidazolylmethyl-cyclohexyl aryl ethers (**5–9**) (Scheme 1).

For the synthesis of 1-(2-aryloxy-cyclohexylmethyl)-1*H*-imidazoles **13–17**, cyclohexanone was reacted with pyrrolidine under Mannich conditions in the presence of L-proline¹⁷ to give 2-pyrrolidin-1-ylmethyl-cyclohexanone (**10**). Subsequent replacement of the pyrrolidine with imidazole followed by reduction led to the *cis/trans* mixture of 2-imidazol-1-ylmethyl-cyclohexanol **12a**/ **12b**.¹⁸ The *trans* isomer **12b** (major product) was condensed with substituted aryl halides to obtain the corresponding ethers **13–17** (Scheme 1). For SAR studies we also synthesized the directly connected imidazole derivatives viz: 1-aryloxy-2, 6-bisimidazolyl cyclohexane **21–23** (Scheme 2). Bromination of cyclohexanone¹⁹

 $^{^{\,\}pm}\,$ CDRI communication number 7527.

^{*} Corresponding author. Tel.: +91 522 2612411x18; fax: +91 522 2623405. *E-mail address*: bhandarikalpana@rediffmail.com (K. Bhandari).

⁰⁹⁶⁰⁻⁸⁹⁴X/\$ - see front matter \odot 2008 Elsevier Ltd. All rights reserved. doi:10.1016/j.bmcl.2008.11.094



Scheme 1. Reagents and conditions: (a) $(HCHO)_3$, pyrrolidine hydrochloride (2.0 equiv), isopropanol; (b) Imidazole, ethanol: water (2:3); (c) NaBH₄; (d) K(*t*-OBu), DMSO, substituted aryl halides (a-c); (e) $(HCHO)_3$, pyrrolidine (1.0 equiv), *L*-proline (0.3 equiv), DMSO.

gave the 2,6-dibromo derivative **18** which on reaction with imidazole followed by reduction with NaBH₄ gave the 2,6-di-imidazol-1yl cyclohexanol **20**. Condensation of this hydroxy compound with substituted aryl halides gave the required aryloxy bisimidazolyl cyclohexanes **21–23**.

All the compounds shown in Schemes 1 and 2 were obtained as racemic mixtures. Their structures, including the relative configurations depicted in Scheme 1. The conformations were established through the complete analysis of their 1D and 2D NMR, NOESY and NOE enhancement studies, MS and IR spectra. In representative cases the unambiguous assignment of all significant ¹H and ¹³C NMR signals was performed. Careful examination of the ¹H, ¹³C NMR of bisimidazol-1-yl-methyl-cyclohexanol 4 suggested it as symmetrical conformation. The 2D NMR (Dept-90, 135, COSY, HMBC and HSOC) also supported the same. Inharmoniously the C-1 proton appeared as singlet at δ 3.36 (instead of a multiplet or triplet). After etherification (5-9) the same singlet shifted down field, δ 4.41–4.54 (no change in symmetry). The ¹H–¹H COSY spectrum of **4** also revealed that there are no characteristic correlations with adjacent protons (H-2, H-6). However, the conformation of cyclohexyl ethers 5-9 and 21-23 was determined from the analysis of the splitting patterns and shifting of the ¹H NMR signals together with a 2D NOESY and NOE enhancement studies. Previously Claudon and his co workers have established the thermodynamically stable conformation of 2,6 di-benzyl substituted cyclohexanols.²⁰ They affirmed that both 2_{eq} , 6_{eq} , 1_{ax} and 2_{eq} , 6_{eq} , 1_{eq} conformers were 100% stable and the reactivity varies



Figure 1. Characteristic and key NOESY correlations for 5.

(~40% more for equatorial OH). The significant correlations between H-1 and its α -protons in the NOESY spectra of **5** supported the fact that the aryloxy function is at axial position. Hence the plane of symmetry (σ) is possible in chair as well as in boat conformations. Further, NOE correlations from H-1 to H-2_{ax}, H-7a, H-7b and also with H-8, H-10 and H-11 (considering symmetrical) (Fig. 1) were observed but no interaction was found between H-1 and H-4 (these 1, 4 flag pole interactions possible in boat conformation). An examination of Drieding models also revealed that these correlations were possible only if the cyclohexyl ring occurs in a chair conformation with two imidazole groups in equatorial and aryloxy at axial position (Fig. 1).

The ¹H NMR of 2, 6-Di-imidazol-1-yl-cyclohexanol **20** was quite different from bisimidazol-1-yl-methyl-cyclohexanol **4**. The H-1 proton appearing as double doublet (J = 8.82, 3.66 Hz) at δ 4.48–4.52 and H-2, H-6 protons were found as two individual multiplets at δ 2.5, δ 2.9, respectively. After etherification (**21–23**), the same pattern of ¹H signals multiplicity was maintained (with little down field shifting of H-1, H-2 and H-6). In ¹³C all the carbons showed individual signals (no σ is found). The H-7 and H-8 (imidazole protons) appeared at δ 8.01–8.09 and 7.65–7.66 rather than its regular position, that is, δ 7.48–7.51 and 7.05–7.09, respectively).²¹ These can interact (di-pole interaction) with phenoxy oxygen only if the cyclohexyl ring is in skew boat conformation (by Drieding models). This conformation is highly energetic and the key NOE enhancement was found between H-1 and H-2 (Fig. 2) (**21**).

The compounds selected for study were evaluated in vitro against transgenic *L. donovani* promastigotes²² and intracellular amastigotes²² at various concentrations taking sodium stibogluconate and pentamidine as a control and cytotoxicity responses²³ were assessed using mouse macrophage cell line (J-774-A-1). All the compounds killed promastigotes and amastigotes in concentration dependent manner and showed 100% inhibition of parasites at a maximum concentration of 10 µg/ml. IC₅₀ of antileishmanial activity was calculated by Probit analysis.²⁴ Based on IC₅₀ and SI values six compounds were further evaluated for in vivo activity



Scheme 2. Reagents and conditions: (a) Br₂, CCl₄; (b) Imidazole, DMF; (c) NaBH₄; (d) NaH, DMF, substituted aryl halides.



Figure 2. NOE enhancements for 21.

Table	1
-------	---

In vitro and in vivo antileishmanial activity of synthesized imidazoles

Sl.No.	Compound	und In vitro assessment [@]				
		Anti promastigote activity IC ₅₀ (μg/ml)	Anti amastigote activity. [#] IC ₅₀ (µg/ml) (C.I)	^a Cytotoxicity CC ₅₀ (μg/ml)	Selective index (SI)	In vivo activity (dose-50 mg/ kg \times 10 i.p.) percent inhibition
1	4	9.4	10.77	>100	9.28	
2	5	3.284	1.13	94.47	*83.60	51.00
3	6	5.524	1.97	73.88	*37.51	52.00
4	7	9.57	2.437	69.39	28.47	
5	8	6.725	0.588	14.44	24.557	
6	9	3.57	1.17	39.37	*33.64	77.90
7	13	0.81	3.83	35.59	9.292	
8	14	0.82	4.57	44.59	9.757	
9	15	4.23	6.72	63.11	9.39	
10	16	2.37	2.13	13.84	6.49	
11	17	1.08	0.71	>100	*140.84	55.35
12	21	2.06	3.0	96.43	*32.143	36.8
13	22	4.67	8.76	>100	11.4	
14	23	6.13	3.02	90.6	*30	56.69
15	Sodium					
Stibogluconate	946.52	46.54	297.38	6.38	84.10	
					(20 mg/Kg)	
16	Pentamidine	0.643	12.11	31.31	2.58	92 (40 mg/Kg)

[®] All the compounds showed concentration dependent response against extra cellular promastigotes and intracellular amastigotes.

[#] $IC_{50} = >15 \ \mu g/ml = Inactive; \ IC_{50} = 5-15 \ \mu g/ml = moderately active; \ IC_{50} = <5 \ \mu g/ml = highly active compounds.$

* Compounds having $IC_{50} = <5 \mu g/ml$ (in vitro antiamastigote activity) and SI = >30 were picked up for in vivo evaluation.

^a CC₅₀ (cytotoxic concentration for 50% inhibition in cell viability) was evaluated against J-774A-1 cell line.

intraperitoneally at 50 mg/kg \times 10 i.p dose against L.donovani/ Hamster model. 25

The in vitro biological activities of bis and mono imidazoles have shown encouraging results. Table 1 displays IC₅₀ and SI values of synthesized bis and mono imidazoles against promastigotes and intracellular amastigotes. The IC₅₀ and SI values for amastigotes of the test derivatives indicate that all compounds exhibited high activity against L. donavani (IC₅₀ $0.58-8.76 \mu g/ml$), better than the reference drugs sodium stibogluconate (IC₅₀ = $46.54 \mu g/ml$) and Pentamidine (IC₅₀ = 12.11 μ g/ml). Among the bisimidazolyl methyl series (5-9) all the compounds appeared highly active exerting a strong inhibitory effect on the amastigote form of parasite with IC₅₀ in the range of 0.588 to 2.437 μ g/ml, while three compounds (5, 6 and 9) produced an interesting selective antiamastigote activity (SI > 30). Concerning monoimidazolyl methyl analogues **13–17**, though the tested derivatives displayed a strong inhibitory activity on the intracellular amastigote IC₅₀ ranging from 0.71 to 4.57 μ g/ml, but the selective index of all the compounds was below 10 except 17 which showed a high SI value of 140.84. Among the corresponding bisimidazolyl derivatives (21-23), two compounds 21 and 23 expressed interesting antiamastigote activity (IC₅₀ of 3.00 and 3.02 μ g/ml), with SI > 30.

The hydroxyl intermediate (4), showed an IC₅₀ 10.77 and SI 9.28. It is apparent from activity results (Table 1) that on introduction of aryl moiety the activity increased several folds $(0.588-2.437 \,\mu g/ml)$. Further the compounds of bisimidazolyl series 5-9 with one carbon spacer were found more potent than the corresponding bisimidazolyl derivatives 21-23 as well as monoimidazolyl analogues 13-16 (except 17), revealing the presence of a carbon spacer between the cyclohexane and imidazole rings for better activity profile. The overall activity profile of compounds (5-9, 21-23, 13-17) demonstrated that there is a small difference in their IC₅₀ values. Thus, the biological activity was slightly influenced by the type of substituent attachment at the 2 and 4-position of the aryloxy nucleus. However, it is interesting to note that while the NO₂ group at position 2 (7, 15) renders the molecule moderately active, the same group at position 4 enhances the activity (6, 14). Moreover, the presence of a fluorine atom at 2 position together with 4-NO₂ further confers increased selectivity 9, 17 and 23.

Six compounds (**5**, **6**, **9**, **17**, **21** and **23**) of SI above 30 were tested further for in vivo leishmanicidal activity and the results are presented in Table 1. Compound **9** with bis methylimidazolyl moiety and with a 2-fluoro, 4-nitro aryloxy group exhibited significant in vivo activity with 77.9% inhibition of parasite growth. Compound **17** and **23** displayed medium activity with 55.35% and 56.69% inhibition, respectively, while other three showed moderate activity. It is interesting to note that in all the three series (**5–9**, **13–17** and **21–23**) the highest activity (in vitro as well as in vivo) was shown by the compounds with a 2-fluoro and 4-NO₂ aryloxy moiety. This finding indicates that aryloxy moiety with a 2-fluoro and 4-NO₂ substituent should be investigated for the development of highly selective antileishmanial compounds.

In conclusion, this study has identified aryloxy cyclohexyl imidazoles as an entirely new structural class of azoles with antileishmanial activity both in vitro and in vivo. The potent activity and simple synthesis of these imidazoles suggest that they are potential candidates for the development of new antileishmanial drugs. Further studies on these compounds and optimization of its structure leading to novel analogues with superior biological properties are on going in our laboratories.

Acknowledgments

N.S. and S.P. thank the UGC and CSIR (India), respectively, for the award of SRF, Nishi to MES (India) for financial support. The authors are thankful to Anoop Srivastava for providing technical assistance and Dr. Neena Goyal for providing luciferase transfected parasites.

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2008.11.094.

References and notes

 Guerin, P. J.; Olliaro, P.; Sundar, S.; Boelaert, M.; Croft, S. L.; Desjeux, P.; Wasunna, M. K.; Bryceson, A. D. Lancet Infect. Dis. 2002, 2, 494.

- Garnier, T.; Mantyla, A.; Jarvinen, T.; Lawrence, J.; Brown, M.; Croft, S. J. Antimicrob. Chemother. 2007, 60, 802–810.
- 3. Croft, S. L. Trends Pharmacol. Sci. 1988, 9, 376.
- 4. Berman, J. D. Rev. Infect. Dis. 1988, 10, 560.
- 5. Jha, T. K. Trans. R. Soc. Trop. Med. Hyg. 1983, 77, 167.
- 6. Sampaio, S. A.; Castro, R. M.; Dillon, N. L. Int. J. Dermatol. 1971, 10, 178.
- 7. Maertens, J. A. Clin. Microbiol. Infect. 2004, 10, 1.
- 8. Beach, D. H.; Goad, L. J.; Holz, G. G. Mol. Biochem. Parasitol. 1988, 31, 149.
- Hart, D. T.; Lauwers, W. J.; Willemsens, G.; Vanden Bossche, H.; Opperdoes, F. R. Mol. Biochem. Parasitol. 1989, 33, 123.
- 10. Chance, M. L. Ann. Trop. Med. Parasitol. 1995, 89, S37.
- (a) Croft, S. L.; Yardley, V. Curr. Pharmaceut. Design 2002, 8, 319; (b) Rangel, H.; Dagger, F.; Hernandez, A.; Liendo, A.; Urbina, J. A. Antimicrob. Agents Chemother. 1996, 40, 2785.
- Abdely, H. M.; Graybill, J. R.; Loebenberg, D.; Melby, P. C. Antimicrob. Agents Chemother. 1999, 43, 2910.
- (a) Borelli, D. A. Rev. Infect. Dis. 1987, 9, 57; (b) Dogra, J.; Saxena, V. N. Int. J. Parasitol. 1996, 26, 1413.

- 14. Haughan, P. A.; Chance, M. L.; Goad, L. J. Biochem. Pharmacol. **1992**, 4411, 2199. 1.
- 15. Gangneux, J. P.; Dullin, M.; Sulahian, A.; Garin, Y. J. F.; Derouin, F. Antimicrob. Agents Chemother. **1999**, 43, 172.
- 16. Alkhawajah, A. Ann. Saudi Med. **1998**, 18, 412.
- 17. Cordova, A. Acc. Chem. Res. 2004, 372, 102.
- Epameinondas, X.; Zarevucka, M.; Saman, D.; Wimmerova, M.; Fragiskos, N.; Kolisis, ; Zdenek, W. *Tetrahedron: Asymmetry* 2006, *17*, 2987.
- 19. Org. Synthesis, Coll. Vol. VI, 711-713.
- Claudon, M. M.; Delpnech, J. J.; Lapicqne, A.; Nicole, D. Tetrahedron 1978, 34, 95.
- Bhandari, K.; Srinivas, N.; Shiva Keshava, G. B.; Shukla, P. K. Eur. J. Med. Chem. 2008. doi:10.1016/j.ejmech.2008.01.006.
- Ashutosh; Gupta, S.; Ramesh; Sundar, S.; Goyal, N. Antimicrob. Agents Chemother. 2005, 49, 3776.
- 23. Heuber, W.; Koella, J. C. Acta tropica **1993**, 55, 257.
- 24. Finney, D. J. Probit analysis, 3rd ed.; Cambridge University Press, 1971.
- 25. Gupta, S.; Tiwari, S.; Bhaduri, A. P.; Jain, G. K. Acta Tropica 2002, 84, 165.