

Article



# Synthesis, X-ray Single Crystal Structure, Molecular Docking and DFT Computations on *N*-[(1*E*)-1-(2*H*-1,3-Benzodioxol-5-yl)-3-(1*H*-imidazol-1-yl) propylidene]-hydroxylamine: A New Potential Antifungal Agent Precursor

# Reem I. Al-Wabli<sup>1</sup>, Alwah R. Al-Ghamdi<sup>1</sup>, Hazem A. Ghabbour<sup>1,2</sup>, Mohamed H. Al-Agamy<sup>3,4</sup>, James Clemy Monicka<sup>5</sup>, Issac Hubert Joe<sup>6</sup> and Mohamed I. Attia<sup>1,7,\*</sup>

- <sup>1</sup> Department of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, P.O. Box 2457, Riyadh 11451, Saudi Arabia; ralwabli@KSU.EDU.SA (R.I.A.-W.); toto24se@hotmail.com (A.R.A.-G.); ghabbourh@yahoo.com (H.A.G.)
- <sup>2</sup> Department of Medicinal Chemistry, Faculty of Pharmacy, Mansoura University, Mansoura 35516, Egypt
- <sup>3</sup> Department of Pharmaceutics, College of Pharmacy, King Saud University, P.O. Box 2457, Riyadh 11451, Saudi Arabia; malagamy@ksu.edu.sa
- <sup>4</sup> Microbiology and Immunology Department, Faculty of Pharmacy, Al-Azhar University, Cairo 11884, Egypt
- <sup>5</sup> Muslim Arts College, Thiruvithancode 629174, Tamil Nadu, India; clemymonicka@gmail.com
- <sup>6</sup> Centre for Molecular and Biophysics Research, Mar Ivanios College, Thiruvananthapuram 695015, Kerala, India; hubertjoe@gmail.com
- <sup>7</sup> Medicinal and Pharmaceutical Chemistry Department, Pharmaceutical and Drug Industries Research Division, National Research Centre (ID: 60014618), El Bohooth Street, Dokki, Giza 12622, Egypt
- \* Correspondence: mattia@ksu.edu.sa; Tel.: +966-11-4677-337

Academic Editor: Diego Muñoz-Torrero

Received: 4 January 2017; Accepted: 21 February 2017; Published: 28 February 2017

**Abstract:** Mycoses are serious health problem, especially in immunocompromised individuals. A new imidazole-bearing compound containing an oxime functionality was synthesized and characterized with different spectroscopic techniques to be used for the preparation of new antifungal agents. The stereochemistry of the oxime double bond was unequivocally determined via the single crystal X-ray technique. The title compound 4,  $C_{13}H_{13}N_3O_3\cdot C_3H_8O$ , crystallizes in the monoclinic space group  $P2_1$  with a = 9.0963(3) Å, b = 14.7244(6) Å, c = 10.7035(4) Å,  $\beta = 94.298(3)^\circ$ , V = 1429.57(9) Å<sup>3</sup>, Z = 2. The molecules were packed in the crystal structure by eight intermolecular hydrogen bond interactions. A comprehensive spectral analysis of the title molecule **4** has been performed based on the scaled quantum mechanical (SQM) force field obtained by density-functional theory (DFT) calculations. A molecular docking study illustrated the binding mode of the title compound **4** into its target protein. The preliminary antifungal activity of the title compound **4** was determined using a broth microdilution assay.

Keywords: crystal structure; imidazole; benzodioxole; oxime; DFT

# 1. Introduction

The incidence of systemic fungal infections (mycoses) has increased drastically in recent years, mainly in immunosuppressed or immunocompromised individuals with AIDS, cancer or undergoing organ transplantation [1,2]. Failure of the available antifungal agents to treat fungal infections is primarily due to dramatic increase in resistance to the conventional antifungal drugs leading to morbidity and mortality in patients facing life-threatening fungal infections. In order to overcome

this serious problem, the development of new alternative antifungal drug therapies with improved efficacy, broader activity and favorable safety profile has attracted a great deal of interest [3,4].

Azole-based compounds having either an imidazole or triazole pharmacophore moiety in their structure, constitute the mainstay of the antifungal chemotherapeutic agents used in the clinic [5,6]. Azoles competitively inhibit cytochrome P450-dependent lanosterol  $14\alpha$ -demethylase (CYP51) resulting in depletion of ergosterol in fungi making them unable to grow in a normal way [7,8]. A screening the literature revealed that most of the available imidazole-bearing antifungal agents have two carbon spacers between the imidazole moiety and an aromatic residue, while few antifungals have a three carbon spacer between the pharmacophore and the aromatic part [9–11].

On the other hand, the benzodioxole moiety is found in a sizable number of biologically active compounds with a wide range of activities [12–16]. The title molecule features both the 1,3-benzodioxole moiety and the imidazole nucleus connecting to each other through a three carbon bridge. Therefore, the current investigation deals with the synthesis, molecular characterization and single crystal X-ray structure of a new oxime derivative, namely *N*-[(1*E*)-1-(2*H*-1,3-benzodioxol-5-yl)-3-(1*H*-imidazol-1-yl)propylidene]hydroxylamine (4) to be utilized as a potential precursor for imidazole-bearing antifungal agents. The stereochemistry of the imine functionality in the title molecule 4 was certainly determined via the single crystal X-ray crystallography technique. In addition, density-functional theory (DFT) computations were also performed as a useful tool to investigate the electronic structure and molecular geometry of the title molecule 4. Molecular docking studies were conducted in order to predict the biological activity of compound 4.

#### 2. Results and Discussion

#### 2.1. Chemistry

Scheme 1 illustrates the synthetic pathway which was adopted to synthesize the target compound **4**. The synthesis commenced with a Mannich reaction under acidic conditions using the commercially available 1-(2*H*-1,3-benzodioxol-5-yl)ethanone (**1**). Subsequently, the formed Mannich base hydrochloride **2** was smoothly transformed in aqueous solution into the ketone **3** via a nucleophilic substitution reaction using imidazole. The target oxime **4** was ultimately obtained using the standard procedure for oxime formation with hydroxylamine hydrochloride in ethanol in the presence of potassium hydroxide [17].



**Scheme 1.** Synthesis of the target oxime **4**. *Reagents and conditions*: (i)  $HN(CH_3)_2$ .HCl,  $(CH_2O)_n$ , conc. HCl, ethanol, reflux, 2 h; (ii) Imidazole, water, reflux, 5 h; (iii) H<sub>2</sub>NOH.HCl, KOH, ethanol, reflux, 18h.

#### 2.2. Crystal Structure of the Title Compound 4

The configuration of the target compound 4 was confirmed via X-ray crystallography. A suitable single colorless crystal of dimensions,  $0.40 \times 0.23 \times 0.11$  mm, was selected for X-ray diffraction analysis. The labeled displacement ellipsoid plot of this molecule is shown in Figure 1. Figure 2 depicts the packing of the molecules in the crystal structure.



**Figure 1.** ORTEP diagram of the title compound drawn at 40% ellipsoids for non-hydrogen atoms showing two molecules and one isopropanol molecule as a solvent.



**Figure 2.** Crystal packing showing intermolecular hydrogen bonds as dashed lines along the *b* and *c* axes.

The single crystal X-ray molecular structure therefore conformed the assigned (*E*)-configuration of the imine group in the target compound. The crystal structure contains two independent molecules with one isopropanol molecule as a solvent in the asymmetric unit. The benzodioxole ring (C1/C2/O1/C3/O2/C4–C7) forms dihedral angles of 44.04(3)° and 20.04(2)° with the imidazole ring (N2/C11/N3/C12/C13) for molecules A and B, respectively. The crystal structure is stabilized by eight hydrogen bonds along the *b* and *c*-axis (Table 1).

$D-H\cdots A$	D-H	$\mathbf{H} \cdots \mathbf{A}$	$D \cdots A$	D–H···A
O3A-H3OA…N3A <sup>i</sup>	1.00(5)	1.73(5)	2.722(5)	176(6)
O3B–H3OB…N3B <sup>ii</sup>	0.98(6)	1.70(6)	2.672(5)	175(8)
O4-H1O4…N1A	0.91(7)	2.38(7)	3.127(6)	139(6)
C3A–H3AB…O3A <sup>iii</sup>	0.9900	2.4900	3.128(6)	122.00
C3B–H3BB····O3B <sup>iv</sup>	0.9900	2.3600	3.080(6)	129.00
C11A–H11A…O4 <sup>ii</sup>	0.9500	2.2700	3.179(7)	159.00
C13A–H13A…O2A <sup>iv</sup>	0.9500	2.5600	3.459(6)	158.00
C13B-H13B····O2B <sup>iii</sup>	0.9500	2.5200	3.424(6)	160.00
C9B-H9BB····N1A <sup>v</sup>	0.9900	2.5800	3.502(6)	154.00

**Table 1.** Hydrogen-bond geometry (Å,  $^{\circ}$ ) of the title molecule 4.

Symmetry codes: (i) x - 1, y, z; (ii) x + 1, y, z; (iii) x, y, z - 1; (iv) x, y, z + 1; (v) -x + 1, y + 1/2, -z + 1.

# 2.3. Structural Geometry Analysis

Equilibrium structural geometry of the title molecule 4 has been evaluated by a potential energy surface (PES) scan study. The flexible dihedral angles of C<sub>11</sub>-C<sub>12</sub>-N<sub>13</sub>-C<sub>14</sub>, C<sub>11</sub>-C<sub>10</sub>-C<sub>5</sub>-C<sub>4</sub> and  $C_{11}$ - $C_{10}$ - $N_{18}$ - $O_{19}$  were scanned from 0° to 360° and their optimum energy was studied to identify stable conformations of the title compound 4. The global minimum energy is -893.45 and -893.49 Hartree for the conformers of compound 4 in the gas and solution phases, respectively (various conformers of compound 4 are shown in Figure S1). From this PES analysis, we have identified the minimum energy conformer of this molecule which was chosen for the subsequent studies. The optimized structure of the studied compound 4 with atoms numbering is depicted in Figure 3. Optimized bond lengths, bond angles and dihedral angles have been presented in Table 2. The formation of intramolecular hydrogen bonding is exposed by the intramolecular contacts to H<sub>22</sub>…N<sub>18</sub> occur with H…O distance of 2.455 Å, which is shorter than the van der Waals separation between the O and H atoms (2.75 Å) [18]. Shortening of the C–N bond lengths  $N_{13}$ – $C_{14}$ ,  $N_{13}$ – $C_{17}$ ,  $N_{16}-C_{17}$  and  $N_{16}-C_{15}$  is typical for double bonds and it is due to resonance interactions. The linear fitting graphs (Figure S2) are drawn to study the correlation between the average experimental values and computed results. Statistical analysis revealed that the results of the computed solvation model are in a good agreement with the average experimental XRD values. Therefore, this method was considered to compute spectral vibrations, natural bond orbital and Frontier orbital energy analyses for the title molecule 4.



Figure 3. Optimized molecular structure of compound 4.

Bond Length (Å) Bond Angle (°)						Dihedral A	angle (°)				
_	Ca	lculated		_	Calculated		F		Calculated		
Parameters	Gas Phase	Solution Phase	Exp.	Parameters	Gas Phase	Solution Phase	Exp.	Parameters	Gas Phase	Solution Phase	Exp.
C1-C2	1.3947	1.3945	1.379	C1C2C3	121.40	121.44	121.60	C1-C2-C3-C4	-0.32	-0.29	1.20
C2-C3	1.3753	1.3764	1.362	C1C2O9	109.69	109.67	110.05	C1C2C3H20	179.6	179.72	-179.12
C3-C4	1.4049	1.405	1.395	C1C6C5	117.55	117.55	117.05	C1C2O9C8	10.37	10.84	-4.10
C4–C5	1.4001	1.4005	1.394	C1-C6-H22	121.74	121.53	121.62	C1O7C8O9	16.91	17.41	-4.55
C5-C6	1.4181	1.4183	1.411	C107C8	105.45	105.50	105.41	C1O7C8H23	135.54	135.89	115.78
C6-C1	1.3707	1.3719	1.370	C2C1C6	122.44	122.37	122.65	C2-C1-O7-C8	-10.64	-10.95	1.95
C1-07	1.3743	1.374	1.376	C2C1O7	109.26	109.35	109.85	C2-C3-C4-C5	0.39	0.51	-1.00
C2-O9	1.3695	1.3695	1.382	C2-C3-C4	116.93	116.98	117.25	C2-C3-C4-H21	-179.8	-179.96	178.51
C3-H20	1.0821	1.0822	0.950	C2O9C8	105.31	105.44	105.57	C2-C1-C6-C5	0.21	0.09	1.80
C4-H21	1.0814	1.0815	0.950	C3-C2-O9	128.89	128.87	128.35	C2C1C6H22	-179.59	179.94	-179.81
C5-C10	1.4846	1.4847	1.482	C2-C3-H20	121.55	121.72	121.41	C3-C4-C5-C6	-0.17	-0.44	1.05
C6-H22	1.0805	1.0808	0.950	C4-C3-H20	121.52	121.30	121.44	C3-C2-C1-O7	-178.62	-178.62	179.60
C8–O7	1.4317	1.4372	1.424	C3-C4-C5	122.28	122.18	121.95	C3-C4-C5-C10	178.73	178.73	177.75
C8–O9	1.4360	1.4401	1.418	C3-C4-H21	117.57	117.55	119.03	C3-C2-O9-C8	-170.62	-170.62	177.8
C8-H23	1.0892	1.0877	0.990	C5-C4-H21	120.15	120.27	119.03	C4-C5-C6-C1	0.13	0.13	2.30
C8-H24	1.0966	1.0943	0.990	C4-C5-C6	119.38	119.48	119.62	C4-C5-C6-H22	-179.72	-179.72	179.78
C10-C11	1.5127	1.5133	1.519	C4-C5-C10	121.01	120.74	121.35	C4-C5-C10-C11	16.37	16.37	19.05
C10-N18	1.2861	1.2868	1.288	C5-C10-C11	121.63	121.26	119.86	C4-C5-C10-N18	-163.31	-163.31	164.15
C11-C12	1.5435	1.5427	1.523	C5-C10-N18	116.24	116.28	117.56	C6-C1-C2-C3	0.002	0.004	-1.60
C11-H25	1.0897	1.0894	0.990	C5-C6-H22	120.71	120.91	121.63	C6C1C2O9	178.67	178.67	178.55
C11-H26	1.0912	1.0915	0.990	C6-C5-C10	119.60	119.78	118.95	C6-C1-O7-C8	170.55	170.55	-178.00
C12-N13	1.4574	1.4620	1.459	C6-C1-O7	128.27	128.27	127.61	C6-C5-C4-H21	-179.7	-179.96	-179.51
C12–H27	1.0902	1.0894	0.990	O7–C8–O9	107.29	106.84	109.45	C6-C5-C10-C11	-164.46	-164.46	159.60
C12-H28	1.0924	1.0911	0.990	O7-C8-H23	109.53	109.56	109.83	C6-C5-C10-N18	15.85	15.85	17.25
N13-C14	1.3815	1.3802	1.370	O7-C8-H24	109.50	109.44	109.81	O7-C1-C2-O9	0.06	0.13	1.40
N13-C17	1.3678	1.3629	1.349	O9-C8-H23	109.38	109.46	109.80	O7-C1-C6-C5	178.42	178.54	179.65
C14-C15	1.3716	1.3713	1.360	O9-C8-H24	109.17	109.18	109.81	O7-C1-C6-H22	-1.73	-1.64	0.59
C14–H29	1.0778	1.0776	0.950	C10-C11-C12	111.71	111.99	111.31	O9-C2-C3-C4	-178.68	-178.61	178.55
C15–N16	1.3751	1.3795	1.377	C10-C11-H25	110.75	110.32	109.42	O9-C2-C3-H20	1.32	1.52	-1.53
C15-H30	1.0790	1.0792	0.950	C10-C11-H26	108.46	108.41	109.42	C10-C5-C6-C1	-179.05	-179.29	-178.45
N16-C17	1.3142	1.3197	1.323	C11-C10-N18	122.12	122.46	122.83	C10-C5-C4-H21	-0.79	-1.78	-1.11
C17-H31	1.0802	1.0798	0.950	C11-C12-N13	112.49	112.05	111.10	C10-C5-C6-H22	1.09	0.90	1.52
N18019	1.4067	1.4061	1.409	C12-C11-H25	109.89	109.89	109.42	C5-C4-C3-H20	-179.53	-179.49	178.63
O19-H32	0.9631	0.9647	0.985	C12-C11-H26	108.47	108.56	109.43	C12-N13-C14-C15	176.72	177.67	176.55

 Table 2. Optimized structural geometry parameters for compound 4.

The used numbering of atoms is as shown in Figure 3.

#### 2.4. Natural Bond Orbital Analysis

Natural bond orbital (NBO) analysis describes the (hyper)conjugative interactions between donor-acceptor orbitals in order to understand intramolecular charge-transfer phenomenon of the molecular system [19]. Selective donor-acceptor interactions of the oxime 4 are listed along with their occupancy and stabilization energy values in Table 3. The hyperconjugative interactions of  $\pi(C_1-C_6) \rightarrow \pi^*(C_2-C_3)$ ,  $\pi(C_1-C_6) \rightarrow \pi^*(C_4-C_5)$ ,  $\pi(C_2-C_3) \rightarrow \pi^*(C_1-C_6)$ ,  $\pi(C_2-C_3) \rightarrow \pi^*(C_4-C_5)$ ,  $\pi(C_4-C_5) \rightarrow \pi^*(C_1-C_6)$ , and  $\pi(C_4-C_5) \rightarrow \pi^*(C_2-C_3)$  were 20.27, 17.86, 18.94, 19.07, 17.00 and 17.47 kcal/mol, respectively. This could be attributed to the charge delocalization leading to ring resonance effect. The lone pair conjugative interactions of LP(2)O<sub>7</sub> $\rightarrow \pi^*(C_1-C_6)$ , LP(2)O<sub>9</sub> $\rightarrow \pi^*(C_2-C_3)$ , LP1(N<sub>13</sub>) $\rightarrow \pi^*(C_{14}-C_{15})$ , and LP1(N<sub>13</sub>) $\rightarrow \pi^*(N_{16}-C_{17})$  have stabilization energy of 25.80, 27.07, 30.67, and 46.28 kcal/mol, respectively. The large E(2) values revealed the occurrence of strong electron delocalization over the ring moieties. These interactions were observed as an increase in the electron density (ED) of the C–C antibonding orbital, which weakens the respective bonds. The  $C_{10}=N_{18}$  bond length (1.2861 A) is significantly shorter than the other –CN bonds and the electron density (ED) of this antibonding orbital was decreased to 0.18707, which is an evidence for the rehybridization [20]. More conjugative and hyperconjugative interactions were formed in the lone-pair, C–C and C–C bond orbital overlap which confirms the intramolecular charge-transfer (ICT) causing the stabilization of the molecular structure of the title oxime 4.

**Table 3.** Second-order perturbation theory analysis of Fock matrix in natural bond orbital basis forcompound 4.

Donor (i)	Occupancy(e)	Acceptor (j)	Occupancy(e)	E(2) <sup>a</sup> kcal/mol	E(j) – E(i) <sup>b</sup> (a.u)	F( <i>i,j</i> ) <sup>c</sup> (a.u)
$\pi(C_1 - C_6)$	1.70639	$\pi^*(C_2 - C_3)$	0.36424	20.27	0.29	0.070
$\pi(C_1 - C_6)$	1.70639	$\pi^*(C_4 - C_5)$	0.37847	17.86	0.30	0.067
$\pi(C_2 - C_3)$	1.69340	$\pi^*(C_1 - C_6)$	0.33036	18.94	0.30	0.068
$\pi(C_2 - C_3)$	1.69340	$\pi^*(C_4 - C_5)$	0.37847	19.07	0.30	0.069
$\pi(C_4 - C_5)$	1.69274	$\pi^*(C_1 - C_6)$	0.33036	17.00	0.29	0.063
$\pi(C_4 - C_5)$	1.69274	$\pi^*(C_2 - C_3)$	0.36424	17.47	0.28	0.063
$\pi(C_4 - C_5)$	1.69274	$\pi^*(C_{10}-N_{18})$	0.18707	17.62	0.28	0.064
$\pi(C_{14}-C_{15})$	1.85899	$\pi^*(N_{16}-C_{17})$	0.37926	14.88	0.28	0.061
$LP2(O_7)$	1.85977	$\pi^*(C_1 - C_6)$	0.33036	25.80	0.36	0.090
$LP2(O_9)$	1.85270	$\pi^*(C_2 - C_3)$	0.36424	27.07	0.35	0.093
$LP1(N_{13})$	1.55644	$\pi^*(C_{14}-C_{15})$	0.30606	30.67	0.29	0.087
$LP1(N_{13})$	1.55644	$\pi^*(N_{16}-C_{17})$	0.37926	46.28	0.28	0.103
$LP1(N_{18})$	1.95533	$\sigma^*(C_{10}-C_{11})$	0.03261	8.70	0.83	0.076
LP1(O <sub>19</sub> )	1.99160	$\sigma^*(C_{11}-H_{25})$	0.01285	0.67	1.11	0.024
LP2(O <sub>19</sub> )	1.90883	$\pi^*(C_{10}-N_{18})$	0.18707	15.97	0.35	0.068

The used numbering of atoms is as shown in Figure 3. <sup>a</sup>: E(2) means energy of stabilization interactions; <sup>b</sup>: Energy difference between donor-to-acceptor, *i* and *j* NBO orbitals; <sup>c</sup>: F(i,j) is the Fock matrix element between *i* and *j* NBO orbitals.

#### 2.5. Vibrational Analysis

The title molecule 4 consists of 32 atoms and their characteristic vibrations are described by 90 normal modes. FT-Raman and FT-IR spectra of the title compound 4 are shown in Figures 4 and 5, respectively. Theoretical and experimental spectral data of the target oxime 4 are presented in Table 4, including calculated and fundamental wavenumbers, FT-IR- and FT-Raman intensities along with the tentative vibrational assignment. The calculated vibrational wavenumbers were corrected by scaled quantum mechanical force-field (SQMFF) method [21] by selective scaling factor approach [22]. Using this force-field method, the calculated wavenumbers are reproducing the experimental values with a mean deviation of 15 cm<sup>-1</sup>. Internal valence coordinates and scaling factors information of the title compound 4 modes are given in Tables S1 and S2, respectively.

808

799

752

743

814

-

-

809

798

785

756

21.360

13.387

32.319

2.956

7.71

28.60

4.73 6.19

Wavenur	nber		Intensity				
Expt.		Calc.	тра	Barman b	- Assignment with PFD % (>10%)		
ν <sub>IR</sub> (cm <sup>-1</sup> )	ν <sub>Raman</sub> (cm <sup>−1</sup> )	ν <sub>Scal</sub> (cm <sup>-1</sup> )	- IK" (km∙mol <sup>-1</sup> )	$(m^2 \cdot sr^{-1})$			
		3830	131.118	7.45	ν (O <sub>19</sub> -H <sub>32</sub> ) (100)		
3144	3145	3128	1.497	8.37	$\nu_{\text{ring II}}$ (C–H) (99)		
3121	3120	3118	2.054	7.851	$\nu_{\text{ring II}}$ (C–H) (99)		
3114	3118	3103	4.324	10.20	$v_{\text{ring II}}$ (C–H) (99)		
-	3073	3096	3.035	6.57	$\nu_{\text{ring II}}$ (C–H) (99)		
3018	-	3056	1.286	5.39	$v_{\rm ring I}$ (C–H) (99)		
3007	3001	3026	2.149	10.90	$v_{\text{ring I}} (C-H) (99)$		
2993	-	3017	3.215	11.70	$v_{\text{ring I}}$ (C–H) (99)		
-	2970	2977	9.230	2.62	$v_{as}$ (CH <sub>2</sub> ) Me III(93)		
2943	2940	2944	29.934	17.00	$v_{as}(CH_2)$ We $I(75) + v_s(CH_2)$ We $I(25)$		
-	-	2934	25.071 4 531	7.86	$v_{\rm s}$ (CH <sub>2</sub> ) Me III (98)		
_	2908	2889	6.873	10.60	$v_{as}$ (CH <sub>2</sub> ) Me II (98)		
-	2845	2836	118.557	27.20	$v_{s}$ (CH <sub>2</sub> ) Me I (74) + $v_{2s}$ (CH <sub>2</sub> ) Me I (26)		
_	1629	1629	14.078	73.10	$v_{\rm s}(C_{10}-N_{18})$ (71)		
1595	1606	1591	24.766	100.00	$v_{ring I}$ (CC) (53) + $\beta$ (CH) Ring I (16)		
-	-	1581	3.985	20.60	$v_{\text{ring I}}$ (CC) (58) + $\beta$ (CH) Ring I (14)		
-	-	1560	0.336	14.40	<i>Sci</i> (HC) Me I (89)		
1503	1516	1504	44.673	2.85	β (CH) Ring II (41) + $\nu$ (C <sub>17</sub> –N <sub>16</sub> ) (28) + $\nu$ (NC) (12)		
-	-	1492	179.197	6.28	$\beta$ (CH) Ring I (50) + $\nu_{ring I}$ (CC) (32)		
1493	1494	1490	17.736	6.84	$\nu$ (C <sub>14</sub> –C <sub>15</sub> ) (35) + $\beta$ (CH) Ring II (29) + $\nu$ (NC) (11)		
1450	-	1457	14.740	4.35	$Sci$ (CH <sub>2</sub> ) Me II (37) + $\tau$ (CN) (31) + $Sci$ (CH <sub>2</sub> ) Me III (16)		
-	-	1444	3.326	21.40	ω (CH <sub>2</sub> ) Me I (88)		
1439	1440	1442	25.303	19.50	$Sci (CH_2) Me II + \tau (CN) (25) + Sci (CH_2) Me III (14)$		
1428	-	1424	55.376	5.77	$v_{\text{ring I}}$ (CC) (44) + $\beta$ (CH) Ring I (34)		
1401	1403	1390	17.021	13.50	$\tau$ (CN) (47) + $\omega$ (CH <sub>2</sub> ) Me III (30)		
1364	-	1369	2.376	5.68	$v$ (NC) (28) + $\tau$ (CN) (17) + $\omega$ (CH <sub>2</sub> ) Me III (12) + $\rho$ (CH <sub>2</sub> ) Me III (12) + twi (CH <sub>2</sub> ) Me III (12)		
1350	1346	1341	2.477	15.10	$\beta$ (CH) Ring II (39) + $\nu$ (C <sub>17</sub> –N <sub>16</sub> ) (31) + $\nu$ (NC) (15)		
1313	1314	1324	38.626	10.90	$β$ (CH) Ring I (40) + ν (CC) (37) + $δ_a$ (Ring I) (10)		
-	-	1301	90.153	46.50	$v_{\text{ring I}}$ (CC) (32) + $\beta$ (CH) Ring I (15)		
1280	1291	1293	9.747	28.50	$\beta$ (CH) Ring II (55) + $\nu$ (C <sub>17</sub> -N <sub>16</sub> ) (16)		
-	1271	1276	18.733	10.70	$\omega$ (CH <sub>2</sub> ) Me II (62)		
1255	1254	1251	4.921	3.83	$Twi (CH_2)$ Me II (34) + $\tau$ (CN) (24) + $\rho$ (CH <sub>2</sub> ) Me III (10) + $Twi (CH_2)$ Me III (10)		
-	-	1225	26.089	28.90	ω (CH <sub>2</sub> ) Me I (37)		
1225	1232	1224	199.125	31.10	$\beta$ (NOH) (23) + CO (13) + CC (12)		
-	-	1222	78.688	30.70	$\nu$ (NC) (16) + CN (13) + $\beta$ (CH) Ring II (13) + Twi (CH <sub>2</sub> ) Me I (11)		
-	-	1208	103.083	19.30	$\beta$ (NOH) (23) + $\nu_{\text{ring I}}$ (CC) (16) + $\nu$ (CO) (10) + $\beta$ (CH) Ring I (10)		
-	-	1202	120.699	19.90	$v_{\text{ring I}}$ (CC) (30) + $v$ (C5–C10) (10) + $\beta$ (NOH) (10)		
1173	1174	1151	4.028	2.22	$v_{\text{ring I}}$ (CC) (25) + $\beta$ (CH) King I (25)		
1147	1141	1136	6.792 E (82	2.61	$\gamma$ (NC) (28) + $IWI$ (CH <sub>2</sub> ) Me I (14) + $\gamma_{ring I}$ (CC) (14) + $\beta$ (CH) King I (12)		
-	- 1107	1124	5.66Z	2.44	$\rho(CH_2)$ Ne I (50) + $\tau(CO)$ (10) + $v_{ring I}$ (CC) (10)		
1085	107	1097	12.714 54 779	15.00	$\delta$ (Ring I) (20) + $\gamma$ (CO) (16) + $\gamma$ (NC) (14)		
-	1050	1079	51 301	13.80	$\beta$ (CH) Ring II (43) + $\gamma$ (NC) (32) + $\gamma$ (C14–C15)(15)		
1035	1030	1032	12.857	19.20	$\tau$ (CN) (25) + $\rho$ (CH <sub>2</sub> ) Me III (21) + <i>Twi</i> (CH <sub>2</sub> ) Me III (21)		
1016	-	1019	121.778	4.74	$\gamma$ (OC) (36) + $\tau$ (CN) (11)		
957	956	1000	50.386	9.32	$v$ (CC) (26) + $v_{ring I}$ (CC) (14)		
-	-	972	6.193	14.00	ν (CC)(66)		
936	-	966	16.460	14.90	δ (Ring I) (34) + $ν$ (NC) (27) + $ν$ (CN) (13)		
-	-	935	3.579	3.29	ω (CH) Ring I (86) γ (N <sub>12</sub> –O <sub>10</sub> ) (34) + τ (CN) (14) +ρ (CH <sub>2</sub> ) Me III (10) + <i>Twi</i> (CH <sub>2</sub> )		
924	922	921	100.257	9.39	$Me III (10) = (D_1 + V(14)) + (C_1 + V(14)) $		
895	893	896	54.208	6.21	$\gamma$ (CO) (70) + $\delta$ (King I) (14)		
8/8 854	-	883 951	61.051 24.424	8.07 2.27	$\nu$ (CU) (24) + $\nu$ (N <sub>18</sub> –O <sub>19</sub> ) (16) + $\delta$ (King I) (13)		
004	000	801 841	24.434 2.112	2.27	ω (CH) Ring I (77)		
-	-	818	7.104	4.86	v (Ring II) (89)		
		010		1.00	· · · · · · · · · · · · · · · · · · ·		

 $\begin{aligned} & \omega \ (CH) \ Ring \ II \ (84) \\ & \nu_{ring \ I} \ (CC) \ (33) + \nu \ (CO) \ (18) + \nu \ (CO) \ (13) \\ & \omega \ (CH) \ Ring \ II \ (77) + ORO \ (11) \\ & \tau \ (CN) \ (40) + \rho \ (CH_2) \ Me \ III \ (17) + twi \ (CH_2) \ Me \ III \ (17) \end{aligned}$ 

Table 4. Calculated vibrational wavenumbers, observed FT-IR and FT-Raman frequencies, FT-IR and FT-Raman intensities and their assignments with PED % for compound 4.

Wavenum	ıber		Intensity		
Expt.		Calc.	тра	Barman b	Assignment with PED % (>10%)
$\nu_{IR}$ (cm <sup>-1</sup> )	ν <sub>Raman</sub> (cm <sup>−1</sup> )	ν <sub>Scal</sub> (cm <sup>-1</sup> )	(km·mol <sup>-1</sup> )	$(m^2 \cdot sr^{-1})$	
-	-	743	2.742	5.72	$\tau$ (CN) (26) + $\rho$ (CH <sub>2</sub> ) Me III (11) + <i>twi</i> (CH <sub>2</sub> ) Me III (11)
-	-	733	9.478	12.10	$\tau$ (CN) (37) + $\rho$ (CH <sub>2</sub> ) Me III (15) + <i>twi</i> (CH <sub>2</sub> ) Me III (15)
725	721	706	33.336	14.00	$\omega$ (CH) Ring II (82) + $\tau$ (CN) (14)
716	-	704	3.019	15.40	$\gamma$ (Ring I) (18) + $\delta$ (Ring I) (11) + $\gamma$ (Ring I) (11)
-	-	690	5.722	5.12	<i>puc</i> (Ring I) (61) + $\omega$ (CC) (13) + $\tau_a$ (Ring I) (10)
-	692	650	5.478	2.91	$\omega$ (CC) (15) + $\tau_a$ (Ring I) (12) + $\tau$ (Ring I) (10)
658	658	629	17.050	1.48	$\tau_a (\text{Ring II}) (69) + \tau (CN) (17)$
621	626	614	18.672	4.91	$\delta$ (Ring II) (17) + $\nu$ (C <sub>12</sub> -N <sub>13</sub> ) (15) + $\nu$ (CC) (11)
578	573	592	11.347	7.42	$\tau_a$ (Ring II) (62) + $\omega$ (CC) (11)
556	-	582	14.299	9.94	δ (Ring I) (30)
492	490	530	5.947	2.11	$\tau$ (CN) (20) + $\omega$ (CC) (14) + puc (14)
-	-	447	5.385	7.04	$\gamma$ (Ring I) (20) + OC (19) + $\delta$ (Ring I) (15)
433	449	440	19.054	7.96	$\beta$ (CNO) (21) + $\delta$ (Ring I) (15) + $\nu$ (CC) (10)
425	-	424	86.609	9.89	τ (NO) (72)
417	-	405	13.261	4.70	$\tau_a$ (Ring I) (36) + <i>butt</i> (35) + $\tau$ (Ring I) (14)
-	387	361	1.486	7.82	$\tau$ (CN) (26) + $\beta$ (CN) (20) + $\tau$ (CN) (10)
-	-	341	1.034	10.40	$\tau$ (CN) (30) + Sci (13) + OC (12)
-	330	333	0.753	9.09	$\tau$ (CN) (67) + $\beta$ (CN) (11)
-	303	303	4.676	15.80	$\tau$ (CN) (38) + $\tau$ (CN) (15) + <i>butt</i> (11)
-	277	260	1.201	20.60	$\tau$ (CN) (69)
-	230	222	0.661	34.50	$\tau$ (CN) (51) + $\rho$ (OC) (10)
-	-	215	0.930	29.90	$\tau_a (\text{Ring II}) + (24) + \tau (\text{Ring I}) (24)$
-	-	180	0.384	13.60	$\tau$ (CN) (73) + $\beta$ (CC) (10)
-	-	138	0.183	22.60	$\tau$ (CN) (37) + <i>Sci</i> (20)
-	-	126	7.961	18.10	$\tau$ (Ring I) (40) + $\tau$ (CN)(18) + $\tau$ (CN) (18) + $\tau$ (CN) (14)
-	-	91	5.187	119.00	$\tau$ (CN) (27) + $\tau$ (Ring I) (17) + $\omega$ (CC) (12) + $\tau$ (CN)(10)
-	-	72	1.075	156.00	τ (CN) (99)
-	-	64	0.692	162.00	$\tau$ (CN) (78)
-	-	29	1.601	104.00	τ (CN) (98)
-	-	20	1.176	312.00	τ (CN) (99)
-	-	16	0.169	328.00	$\tau$ (CN) (86)

Table 4. Cont.

<sup>a</sup>: IR intensity; <sup>b</sup>: Raman intensity; Ring I: benzodioxole ring; Ring II: imidazole ring; v: stretching; Me: methylene;  $v_s$ : symmetric stretching;  $v_{as}$ : asymmetric stretching;  $\beta$ : bending;  $\tau$ : torsion; puc: puckering;  $\omega$ : wagging; *twi*: twisting;  $\rho$ : rocking; *Sci*: scissoring;  $\gamma$ : out-of-plane bending;  $\delta$ : in-plane bending; *butt*: butterfly mode;  $\tau_a$ : out-of-plane torsion.



Figure 4. Experimental (upper) and simulated (lower) FT-Raman spectra of compound 4.



Figure 5. Experimental (upper) and simulated (lower) FT-IR spectra of compound 4.

#### 2.5.1. Imidazole Ring Vibrations

The calculated IR and Raman imidazole C–H stretching vibrations at 3118 and 3145 cm<sup>-1</sup> are in close agreement with the experimental wavenumbers observed at 3144, 3125, 3120 cm<sup>-1</sup> [23,24]. The FT-IR bands observed at 1503, 1350, 1280 and 1108 cm<sup>-1</sup> and the FT-Raman ones at 1516, 1346, 1291, 1107 and 1050 cm<sup>-1</sup> are definitely attributed to imidazole CH in-plane bend modes which are coupled with several other modes. The FT-IR bands at 752 and 725 cm<sup>-1</sup> and the weak FT-Raman bands at 721 cm<sup>-1</sup>, can be assigned to the out-of-plane C–H wagging modes, as supported by theliterature data [25]. The C<sub>14</sub>=C<sub>15</sub> stretching mode is the most characteristic vibration of the heterocyclic imidazole ring which was observed as medium intensity bands at 1493 (FT-IR) and at 1494 (FT-Raman) cm<sup>-1</sup>. The FT-IR and FT-Raman vibrations of the imidazole ring out-of-plane torsion modes appeared at 658, 578 (FT-IR) and 573 (FT-Raman) cm<sup>-1</sup>.

# 2.5.2. Methylene Group Vibrations

The neighboring rings  $\pi$ -system and nitrogen lone pair affects the spectral behavior of  $sp^3$  hybridized methylene moiety. Methylene asymmetric and symmetric C–H stretching vibrations usually appear in the region 2960–2840 cm<sup>-1</sup> [26]. The asymmetric (2970 and 2946 cm<sup>-1</sup>) and symmetric (2908 and 2845 cm<sup>-1</sup>) stretching contributing bands appeared as a medium intensity band in the FT-Raman spectrum of the title compound **4**. Lowering of symmetric stretching wavenumbers could be attributed to the hyperconjugative interaction between the nitrogen lone pair and  $\sigma^*$ (C–H) bond. The rocking, wagging and twisting vibrational modes of compound **4** appeared in the region of 1400–900 cm<sup>-1</sup> [27]. The CH<sub>2</sub> scissoring mode appeared as a characteristic band near 1450 and 1439 cm<sup>-1</sup> in the FT-IR spectrum and at 1440 cm<sup>-1</sup> in the FT-Raman spectrum of the title oxime **4**. The normal coordinate analysis (NCA) supports the FT-Raman bands at 1254 and 1271 cm<sup>-1</sup> and the FT-IR bands at 1255 cm<sup>-1</sup> for the unambiguous assignment of CH<sub>2</sub> twisting and wagging modes.

# 2.5.3. Benzodioxole Ring Vibrations

Aromatic C–H stretching mode appears in the region of 3000–3100 cm<sup>-1</sup>. Weak FT-Raman bands identified at 3073 and 3001 cm<sup>-1</sup> have been assigned to the C–H stretching mode. Usually the bands due to ring C–H in-plane and out-of-plane bending vibrations are observed in the region of 1000–1300 and 750–1000 cm<sup>-1</sup>, respectively. In the title compound **4**, the C–H in-plane bending vibration has been observed as a medium intensity FT-IR band at 1313 cm<sup>-1</sup> and as a weak band at 1314 cm<sup>-1</sup> in the FT-Raman spectrum. The medium to weak intensity bands observed at 854 (FT-IR), 850 (FT-Raman), 814 (FT-Raman) and 808 (FT-IR) cm<sup>-1</sup> have been assigned to C–H out-of-plane bending mode. Ring

C–C stretching modes have been identified in the FT-IR spectrum of the target compound 4 at 1595, 1428 and 1173 cm<sup>-1</sup> while they appeared at 1606 and 1174 cm<sup>-1</sup> in the FT-Raman spectrum. The ring asymmetric deformation and torsion modes have been identified at 716, 556, 492 and 417 cm<sup>-1</sup> in the FT-IR and at 490 cm<sup>-1</sup> in the FT-Raman spectra of the title molecule 4.

# 2.6. Frontier Molecular Orbital Analysis

A detailed knowledge of molecular electron density distribution and electron motion are necessary to understand molecular recognition and chemical reactivity of the molecule. The LUMO and HOMO orbital energy analysis of the title compound **4** has been computed using DFT method in the solution phase (isopropanol). The electron charge cloud is located at the piperonal ring in highest occupied molecular orbital (HOMO) while it is located mainly at the phenyl ring in the lowest unoccupied molecular orbital (LUMO). The energy of the HOMO is -6.23 eV and LUMO is -1.60 eV giving rise to a HOMO-LUMO energy gap of 4.63 eV. The HOMO-LUMO energy gap value supports the intramolecular charge-transfer interactions within the title molecule. The HOMO and LUMO orbital diagram is shown in Figure 6.



Figure 6. (a) HOMO (b) LUMO orbital of compound 4.

# 2.7. NMR Chemical Shift Analysis

The carbon and hydrogen chemical shift values of the title oxime 4 were calculated based on Gauge-independent atomic orbital (GIAO) method at B3LYP/6-311++G(d,p) level of theory [28]. The computed values are presented in Table 5. The correlation graphs were plotted over the observed and predicted chemical shift values of the title molecule 4 and the linear fitting plots are shown in Figure S3.

Ca	rbon Atoms ( <sup>13</sup>	C)	Hydrogen Atoms ( <sup>1</sup> H)			
Atoms —	Value	(ppm)	• -	Value (ppm)		
	Calc.	Exp.	Atoms	Calc.	Exp.	
C1	127.25	148.0	H20	5.81	6.84	
C2	127.81	148.3	H21	7.01	7.06	
C3	89.62	108.5	H22	5.89	7.13	
C4	107.84	120.5	H23	5.40	6.04	
C5	107.85	130.3	H24	5.20	6.04	
C6	89.80	106.1	H25	1.92	3.14	
C8	87.78	101.7	H26	2.23	3.14	
C10	132.24	153.7	H27	3.40	4.17	
C11	29.66	28.2	H28	3.64	4.17	
C12	38.54	43.3	H29	6.05	6.89	
C14	100.78	119.9	H30	5.94	7.19	
C15	109.30	128.3	H31	6.38	7.66	
C17	115.68	137.5				

Table 5. Calculated and experimental chemical shift values for compound 4.

The used numbering of atoms is as shown in Figure 3.

The phenyl carbons signals usually appear in the region of 120–140 ppm. The C1 and C2 in the title molecule **4** were observed at 148.0 and 148.3 ppm, respectively. This downfield chemical shift values revealed that, these carbon atoms bounded with the electronegative oxygen atoms. In general, imidazole ring protons signals occur in the region of 6–8 ppm. The <sup>1</sup>H-NMR spectrum of the target oxime **4** showed signals at 6.89, 7.19 and 7.66 ppm which correspond to the protons of the imidazole ring. The aromatic piperonal ring protons were observed at 6.84 (as doublet), 7.06 (as doublet) and 7.13 (as singlet) ppm. There is a good agreement between the calculated and observed chemical shift values of the title molecule **4** with a correlation coefficient ( $R^2$ ) values = 0.994 and 0.953 for <sup>13</sup>C and <sup>1</sup>H, respectively.

#### 2.8. Molecular Docking Study

Molecular docking is an important technique to predict the biological activity of chemical compounds [29]. The target compound 4 was energy minimized using DFT method with the help of Gaussian program [30]. The target protein (cytochrome P450-dependent (CYP51) lanosterol 14 $\alpha$ -demethylase enzyme) for antifungal azoles has been identified based on a multilevel neighborhoods of atoms' (MNAs) algorithm model by the PASS online server [31]. This target protein (PDB code: 1EA1) was downloaded from the research collaboratory structural bioinformatics (RCSB) protein data bank [32]. The target compound 4 was docked using AutoDock Tools 1.5.4 (The Scripps Research Institute, La Jolla, CA, USA) interfaced with the AutoDock 4.2 program in the rigid docking methodology [33,34]. The binding free energy and inhibition constant of the proper conformation of the title compound 4 was predicted to be -5.06 kcal/mol and 195.75  $\mu$ M, respectively. The hydrogen bond interaction of protein-ligand complex is shown in Figure 7. The amino acid residues LEU321 and PRO386 of the target protein are bounded with the title compound 4 by an intermolecular hydrogen bonding interaction. The docking study illustrated the affinity of compound 4 toward its target protein with a good binding energy value (-5.06 kcal/mol) and hence its suitability as a potential precursor to prepare new antifungal agents.



Figure 7. Binding pose of compound 4 with its target protein.

# 2.9. Antifungal Activity of the Title Compound 4

Table 6 presents the preliminary antifungal activity of the tested oxime 4 as well as the reference standard drug, ketoconazole. Compound 4 exhibited a MIC value of 987.43 µmol/L against *C. albicans, C. tropicalis, C. parapsilosis* and *Aspergillus niger* in the broth microdilution assay. While inactive itself, taken together the body of evidence suggests that compound 4 could be used as a starting material to prepare new antifungal agents with better antifungal profile.

Common d No	MIC (µmol/L)						
Compound No	C. albicans	C. tropicalis	C.parapsilosis	Aspergillus niger			
4	987.43	987.43	987.43	987.43			
Ketoconazole	7.53	15.05	15.05	15.05			

**Table 6.** Antifungal activity of compound 4 and ketoconazole against different *Candida* species and *Aspergillus niger*.

# 3. Experimental

#### 3.1. General

Melting points were measured using a Gallenkamp melting point device, and are uncorrected. Infrared (IR) spectra (as KBr disks) were recorded on FT-IR Spectrum BX device (Perkin Elmer, Ayer Rajah Crescent, Singapore). The NMR samples were dissolved in either CDCl<sub>3</sub> or DMSO- $d_6$  and the NMR spectra were recorded using a Bruker NMR spectrometer (Bruker, Reinstetten, Germany), at 500 MHz for <sup>1</sup>H and 125.76 MHz for <sup>13</sup>C at the Research Center, College of Pharmacy, King Saud University, Saudi Arabia. Chemical shifts are expressed in  $\delta$ -values (ppm) relative to TMS as an internal standard. Mass spectra were recorded using a Quadrupole 6120 LC/MS equipped with an electrospray ionization (ESI) source (Agilent Technologies, Palo Alto, CA, USA). Silica gel TLC (thin layer chromatography) plates (silica gel precoated aluminium cards with 254 nm fluorescent indicator) from Merck (Darmstadt, Germany) were used for thin layer chromatography. Visualization was performed by illumination with a UV light source (254 nm).

#### 3.2. Synthesis

#### 3.2.1. Synthesis of 1-(2H-1,3-Benzodioxol-5-yl)-3-(1H-imidazol-1-yl)propan-1-one (3)

A catalytic amount of concentrated hydrochloric acid (0.5 mL) was added to a mixture containing paraformaldehyde (0.81 g, 9.0 mmol), dimethylamine hydrochloride (2.20 g, 27 mmol) and1-(2*H*-1,3-benzodioxol-5-yl)ethanone (**1**, 3.28 g, 20 mmol) in absolute ethanol (15 mL). The reaction mixture was refluxed for two hours, cooled and acetone (30 mL) was added to precipitate the Mannich base hydrochloride (**2**) which was collected by filtration and dried. Imidazole (1.36 g, 20 mmol) was added to a solution of compound **2** (2.58 g, 10 mmol) in water (10 mL) and the solution was refluxed for five hours, cooled and the precipitated solid was filtered off to afford compound **3**. Compound **3** was re-crystallized from ethanol to give 1.15 g (47%) of the pure pivotal ketone **3** as a white solid m.p. 150–152 °C. IR (KBr): v (cm<sup>-1</sup>) 3115, 2968, 1757 (C=O), 1637 (C=N), 1600, 1494, 1255, 750; <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$  (ppm) 3.37 (t, *J* = 6.5 Hz, 2H, –CH<sub>2</sub>–CH<sub>2</sub>–N), 4.42 (t, *J* = 6.5 Hz, 2H, –CH<sub>2</sub>–CH<sub>2</sub>–N), 6.05 (s, 2H, –O–CH<sub>2</sub>–O–), 6.84 (d, *J* = 8.0 Hz, 1H, Ar–H), 6.99 (s, 1H, –N–CH=CH–N=), 7.04 (s, 1H, –N–CH=CH–N=), 7.39 (d, *J* = 1.5 Hz, 1H, Ar–H), 7.50 (dd, *J* = 1.5, 8.0 Hz, 1H, Ar–H), 7.64 (s, 1H, –N–CH=N–); <sup>13</sup>C-NMR (CDCl<sub>3</sub>):  $\delta$  (ppm) 39.6 (–CH<sub>2</sub>–CH<sub>2</sub>–N), 41.7 (–CH<sub>2</sub>–CH<sub>2</sub>–N), 102.0 (–O–CH<sub>2</sub>–O–), 107.7 (Ar–CH), 108.0 (Ar–CH), 119.2 (–N–CH=CH–N=), 124.4, 129.1 (Ar–CH, –N–CH=CH–N=), 131.0, (Ar–C), 137.4 (–N–CH=N–), 148.4, 152.3 (Ar–C), 194.6 (C=O); MS *m*/z (ESI): 245.0 [M + H]<sup>+</sup>.

#### 3.2.2. Synthesis of (1E)-1-(2H-1,3-Benzodioxol-5-yl)-N-hydroxy-3-(1H-imidazol-1-yl)propan-1-imine (4)

Potassium hydroxide (1.12 g, 20 mmol) was added to a mixture containing hydroxylamine hydrochloride (1.39 g, 20 mmol), ketone **3** (2.44 g, 10 mmol) in ethanol (10 mL). The reaction mixture was refluxed under stirring for 18 hours, cooled to room temperature and the insoluble matter was collected by filtration. The filtrate was concentrated under reduced pressure and the residue was poured onto ice-cold water (15 mL). The obtained solid was filtered off and dried to yield 1.7 g (64%) of the target oxime **4** as a white solid m.p. 142–144 °C. Re-crystallization of the oxime **4** from isopropanol gave colorless single crystals which were suitable for X-ray analysis. <sup>1</sup>H-NMR (DMSO-*d*<sub>6</sub>):  $\delta$ (ppm) 3.14 (t, *J* = 7.0 Hz, 2H, –CH<sub>2</sub>–CH<sub>2</sub>–N), 4.17 (t, *J* = 7.0 Hz, 2H, –CH<sub>2</sub>–N), 6.04 (s, 2H, –O–CH<sub>2</sub>–O–),

6.84 (d, *J* = 7.5 Hz, 1H, Ar–H), 6.89 (s, 1H, –N–CH=CH–N=), 7.06 (dd, *J* = 1.5, 8.0 Hz, 1H, Ar–H), 7.13 (d, *J* = 1.0 Hz, 1H, Ar–H), 7.19 (s, 1H, –N–CH=CH–N=), 7.66 (s, 1H, –N–CH=N–), 11.41 (s, 1H, OH); <sup>13</sup>C-NMR (DMSO–*d*<sub>6</sub>):  $\delta$ (ppm) 28.2 (–CH<sub>2</sub>–CH<sub>2</sub>–N), 43.3 (–CH<sub>2</sub>–CH<sub>2</sub>–N), 101.7 (–O–CH<sub>2</sub>–O–), 106.1, 108.5 (Ar–CH), 119.9 (–N–CH=CH–N=), 120.5, 128.3 (Ar–CH, –N–CH=CH–N=), 130.3 (Ar–C), 137.5 (–N–CH=N–), 148.0, 148.3 (Ar–C), 153.7 (C=N–OH); MS *m*/*z* (ESI): 260.1 [M + H]<sup>+</sup>.

#### 3.3. Crystal Structure Determination

Slow evaporation of the alcoholic (isopropanol) solution of the title compound 4 furnished its colourless block single crystals. The X-ray diffraction measurement of the target oxime 4 was conducted on a SMART APEXII CCD diffractometer (Bruker, Karlsruhe, Germany) equipped with graphite monochromatic CuKådiation ( $\lambda = 1.54178$  Å) at 296 (2) K. Cell refinement and data reduction were done by Bruker SAINT [35]. SHELXS-97 [36] was used to solve and refine the title structure. The final refinement of the crystal structure of the title oxime 4 was performed by full- matrix least-squares techniques with anisotropic thermal data for non hydrogen atoms on  $F^2$ . All the hydrogen atoms were placed in the calculated positions and constrained to ride on their parent atoms. Multi-scan absorption correction was applied by the use of SADABS software [35]. The crystallographic data and refinement information are summarized in Table 7. Crystallographic data of compound 4 have been deposited with the Cambridge Crystallographic Data Center (supplementary publication number CCDC-1508986). Copies of the data may be obtained free of charge from the Director, CCDC, 12 Union Road, Cambridge, CB2 1EZ, UK (deposit@ccdc.cam.ac.uk).

Chemical Formula	2(C <sub>13</sub> H <sub>13</sub> N <sub>3</sub> O <sub>3</sub> )·C <sub>3</sub> H <sub>8</sub> O					
Molecular weight	578.62					
Crystal system, space group	Monoclinic, P2 <sub>1</sub>					
Temperature (K)	296					
a, b, c (Å)	9.0963(3), 14.7244(6), 10.7035(4)					
β(°)	94.298(3)					
$V(Å^3)$	1429.57(9)					
Z	2					
Radiation type	Cu Kα					
$\mu$ (mm <sup>-1</sup> )	0.81					
Crystal size (mm)	0.40 imes 0.23 imes 0.11					
Data collection						
Diffractometer	Bruker APEX-II CCD diffractometer					
Absorption correction	Multi-scan, SADABS Bruker 2014					
T <sub>min</sub> , T <sub>max</sub>	0.740, 0.916					
No. of measured, independent and observed $[I > 2\sigma(I)]$ reflections	6654, 3851, 2788					
R <sub>int</sub>	0.039					
Re	efinement					
$R[F^2 > 2\sigma(F^2)]^a$ , $wR(F^2)^b$ , S	0.059, 0.144, 1.03					
No. of reflections	3851					
No. of parameters	393					
No. of restraints	1					
H-atom treatment	H atoms treated by a mixture of independent and constrained refinement					
$\Delta \rho_{max}$ , $\Delta \rho_{min}$ (e·Å <sup>-3</sup> )	0.26, -0.23					

**Table 7.** The crystallographic data and refinement information.

<sup>a</sup>: R is the residual factor for the reflections; <sup>b</sup>: wR is the weighted residual factor for all the reflections.

#### 3.4. FT-IR and FT-Raman Measurements

The FT-Raman spectrum of the oxime 4 was recorded in the spectral range of 3500–50 cm<sup>-1</sup> using a Bruker RFS-27 FT-Raman spectrophotometer (Bruker, Billerica, MA, USA). The 1064 nm line of Nd:YAG laser operating at 100 mW power was used for excitation. The FT-IR spectrum of the oxime 4 was recorded with a spectral resolution of 2 cm<sup>-1</sup> in the 4000–400 cm<sup>-1</sup> range. Solid sample in KBr pellets was used in the FT-IR measurements.

#### 3.5. Quantum Chemical Calculations

Optimized structural geometry and harmonic vibrational wavenumbers have been calculated at DFT/B3LYP/6-311++G(d,p) level of basis set in the gas phase. The polarizable continuum model (PCM) using the integral equation formalism (IEF) variant is the self-consistent reaction field (SCRF) to predict the structural parameters and vibrational wavenumbers of the title molecule 4 and isopropanol has defined as the solvent in implicit solvation model. This calculation has been performed in the presence of isopropanol by placing the title molecule 4 in a cavity within the solvent reaction field. Normal coordinate analysis (NCA) has been performed to obtain detailed explanation of the molecular motion relating to the normal modes using the MOLVIB program version 7.0 written by Sundius [37,38]. According to the scaled quantum mechanical force-field (SQMFF) procedure [21], selective scaling has been performed in the natural internal coordinate representation [22]. The simulated IR and Raman spectra of the title compound 4 have been plotted using pure Lorentizian band shapes with a bandwidth of full width height maximum of 10 cm<sup>-1</sup>. Second order interactions between the filled orbital of one subsystem and vacant orbitals of another subsystem have been understood with the aid of natural bonding orbitals (NBO) analysis [19] using NBO 3.1 program [39] as implemented in the Gaussian '09 package [30] at the DFT/B3LYP level. Molecular docking analysis using AutoDock 4.2 program [33] predicted the antifungal activity of the title compound 4.

# 3.6. Antifungal Activity

# 3.6.1. Materials

The reference standard antifungal drug, ketoconazole, was purchased from Sigma-Aldrich Co. (St. Louis, MO, USA). Liquid RPMI 1640 medium supplemented with L-glutamine was obtained from Gibco-BRL, Life Technologies (Paisley, Scotland). Sabouraud Dextrose Agar (SDA) was obtained from Merck Co. (Darmstadt, Germany). Dimethyl sulfoxide (100%) was used to dissolve ketoconazole, and/or the tested compound **4** to give an initial concentration of 2048 mg/L.

# 3.6.2. Organisms

The used fungal strains are *Candida albicans* (ATCC 90028), *Candida tropicalis* (ATCC 66029), *Candida parapsilosis* (ATCC 22019) and *Aspergillus niger* (ATCC 16404).

# 3.6.3. Preparation of Fungal Inocula

The inocula of the standard mold *Aspergillus niger* strain have been prepared by removing the sporulated *A. niger* from the Sabouraud Dextrose agar slant with a microbiological loop and the spores have been suspended in 10 mL of sterile water. The suspension has been filtered through sterile gauze to remove hyphae. The resulting suspension of conidia has been vigorously mixed using a vortex. The suspension has been adjusted to  $1 \times 10^5$  CFU/mL using spectrophotometer. This fungal suspension has been diluted 1:5 with RPMI medium to obtain suspensions having 2× of the required final concentration. This conidial suspension had a final concentration of  $1 \times 10^4$  CFU/mL when mixed with the tested solution of compound **4**. On the other hand, the inocula of the standard yeast strains of *C. albicans, C. tropicalis* and *C. parapsilosis* have been prepared by suspending five representative colonies, obtained from 24 to 48 h culture on Sabauraud Dextrose agar medium, in sterile distilled water. The final inoculum concentration must be between  $0.5 \times 10^5$  and  $2.5 \times 10^5$  CFU/mL.

#### 3.6.4. Preparation of the Tested Compound Solution

Briefly, a twofold dilution series of the tested compound **4** has been prepared in a double strength RPMI 1640 culture medium. Ten serial dilutions were prepared to give concentrations ranged from 1024 mg/L to 2 mg/L.

#### 3.6.5. Antifungal Susceptibility Studies

Minimum Inhibitory Concentrations (MICs) have been determined by broth microdilution testing as described previously by EUCAST [40]. The experiment was carried out in duplicate. Briefly, one mL of RPMI 1640 medium from each of the bottle containing the corresponding concentration of the tested compound **4** has been transferred into sterile 7 mL Sterilin tubes (Thermo Fisher Scientific, Waltham, MA, USA). The RPMI 1640 medium containing 1024 mg/L of the tested compound **4** has been dispensed to tube 1, the medium containing 512 mg/L has been dispensed to tube 2, the medium containing 256 mg/L has been dispensed to tube 3 and so on to tube 10 for the medium containing 2 mg/L of the tested compound **4**. One mL of the medium has been dispensed in tubes 11 (positive control) and 12 (negative control). One mL of the diluted inoculum suspension has transferred to each tube except tube 12 to bring the tested compound **4** dilutions to the required final test concentrations. The tubes were incubated at 35 °C for 72 h. The MIC of the tested compound **4** was determined visually by recording the degree of growth inhibition in each tube.

# 4. Conclusions

The synthesis and spectroscopic characterization of *N*-[(1*E*)-1-(2*H*-1,3-benzodioxol-5-yl)-3-(1*H*-imidazol-1-yl)propylidene]hydroxylamine (**4**) as new antifungal precursor has been reported. Computational studies on the target oxime **4** revealed that the theoretical wavenumbers are in a fair agreement with the observed wavenumbers except those associated with H-bonding. CH<sub>2</sub> symmetric stretching wavenumber is red-shifted due to the hyperconjugative interaction between the nitrogen lone pair and  $\sigma^*(C-H)$  bond. Single crystal X-ray analysis of the target molecule **4** confirmed the (*E*)-configuration of its imine double bond. A molecular docking study predicted the binding mode of compound **4** into its target protein and hence its usefulness as a potential precursor for new imidazole-bearing antifungal agents featuring both benzodioxole and imidazole pharmacophore moieties.

Supplementary Materials: Supplementary materials are available online.

Acknowledgments: The authors would like to extend their sincere appreciation to the Deanship of Scientific Research at King Saud University for its funding of this research through the Research Group Project No. RGP-196.

**Author Contributions:** R.I.A.-W. and A.R.A.-G. synthesized and characterized the title molecule. H.A.G. carried out the X-ray analysis of the title molecule. M.H.A.-A. performed the in vitro antifungal screening for the title compound. J.C.M. and I.H.J. conducted the computational work. M.I.A. proposed the work, prepared the single crystals of the title compound and prepared the manuscript for publication. All authors discussed the contents of the manuscript.

Conflicts of Interest: The authors have declared no conflict of interest.

#### References

- 1. Caston-Osorio, J.J.; Rivero, A.; Torre-Cisneros, J. Epidemiology of invasive fungal infection. *Int. J. Antimicrob. Agents* **2008**, *32* (Suppl. 2), S103–S109. [CrossRef]
- Cuenca-Estrella, M.; Bernal-Martinez, L.; Buitrago, M.J.; Castelli, M.V.; Gomez-Lopez, A.; Zaragoza, O.; Rodriguez-Tudela, J.L. Update on the epidemiology and diagnosis of invasive fungal infection. *Int. J. Antimicrob. Agents* 2008, 32 (Suppl. 2), S143–S147. [CrossRef]
- 3. Denning, D.W.; Kibbler, C.C.; Barnes, R.A. British Society for Medical Mycology proposed standards of care for patients with invasive fungal infections. *Lancet Infect. Dis.* **2003**, *3*, 230–240. [CrossRef]
- 4. Kathiravan, M.K.; Salake, A.B.; Chothe, A.S.; Dudhe, P.B.; Watode, R.P.; Mukta, M.S.; Gadhwe, S. The biology and chemistry of antifungal agents: A review. *Bioorg. Med. Chem.* **2012**, *20*, 5678–5698. [CrossRef] [PubMed]

- Pfaller, M.A.; Messer, S.A.; Hollis, R.J.; Jones, R.N. Antifungal activities of posaconazole, ravuconazole, and voriconazole compared to those of itraconazole and amphotericin B against 239 clinical isolates of *Aspergillus* spp. and other filamentous fungi: Report from SENTRY Antimicrobial Surveillance Program, 2000. *Antimicrob. Agents Chemother.* 2002, *46*, 1032–1037. [PubMed]
- Spreghini, E.; Maida, C.M.; Tomassetti, S.; Orlando, F.; Giannini, D.; Milici, M.E.; Scalise, G.; Barchiesi, F. Posaconazole against *Candida glabrata* isolates with various susceptibilities to fluconazole. *Antimicrob. Agents Chemother.* 2008, 52, 1929–1933. [CrossRef] [PubMed]
- Aoyama, Y.; Yoshida, Y.; Sato, R. Yeast cytochrome P-450 catalyzing lanosterol 14 α-demethylation. II. Lanosterol metabolism by purified P-450(14)DM and by intact microsomes. *J. Biol. Chem.* 1984, 259, 1661–1666. [PubMed]
- Vanden Bossche, H.; Koymans, L. Cytochromes P450 in fungi. *Mycoses* 1998, 41 (Suppl. 1), 32–38. [CrossRef]
   [PubMed]
- Aboul-Enein, M.N.; El-Azzouny, A.A.; Attia, M.I.; Saleh, O.A.; Kansoh, A.L. Synthesis and anti-*Candida* potential of certain novel 1-[(3-substituted-3-phenyl)propyl]-1*H*-imidazoles. *Arch. Pharm.* 2011, 344, 794–801. [CrossRef] [PubMed]
- 10. Roman, G.; Mares, M.; Nastasa, V. A novel antifungal agent with broad spectrum: 1-(4-biphenylyl)-3-(1*H*-imidazol-1-yl)-1-propanone. *Arch. Pharm.* **2013**, *346*, 110–118. [CrossRef] [PubMed]
- Attia, M.I.; Radwan, A.A.; Zakaria, A.S.; Almutairi, M.S.; Ghoneim, S.W. 1-Aryl-3-(1*H*-imidazol-1-yl) propan-1-ol esters: Synthesis, anti-*Candida* potential and molecular modeling studies. *Chem. Cent. J.* 2013, 7, 1–12. [CrossRef] [PubMed]
- Cotinguiba, F.; Regasini, L.O.; da Silva Bolzani, V.S.; Debonsi, H.; Passerini, G.D.; Cicarelli, R.M.B.; Kato, M.J.; Furlan, M. Piperamides and their derivatives as potential antitrypanosomal agents. *Med. Chem. Res.* 2009, 18, 703–711. [CrossRef]
- 13. Himaja, M.; Vandana, K.; Ranjitha, A.; Ramana, M.; Karigar, A. Synthesis, docking studies and antioxidant activity of 1, 3-benzodioxole-5-carboxyl amino acids and dipeptides. *Int. Res. J. Pharm.* **2011**, *2*, 57–61.
- Leite, A.C.; da Silva, K.P.; de Souza, I.A.; de Araujo, J.M.; Brondani, D.J. Synthesis, antitumour and antimicrobial activities of new peptidyl derivatives containing the 1,3-benzodioxole system. *Eur. J. Med. Chem.* 2004, *39*, 1059–1065. [CrossRef] [PubMed]
- Aboul-Enein, M.N.; El-Azzouny, A.A.; Attia, M.I.; Maklad, Y.A.; Amin, K.M.; Abdel-Rehim, M.; El-Behairy, M.F. Design and synthesis of novel stiripentol analogues as potential anticonvulsants. *Eur. J. Med. Chem.* 2012, 47, 360–369. [CrossRef] [PubMed]
- Attia, M.I.; El-Brollosy, N.R.; Kansoh, A.L.; Ghabbour, H.A.; Al-Wabli, R.I.; Fun, H.-K. Synthesis, single crystal X-ray structure, and antimicrobial activity of 6-(1,3-benzodioxol-5-ylmethyl)-5-ethyl-2-{[2-(morpholin-4-yl)ethyl]sulfanyl}pyrimidin-4(3H)-one. J. Chem. 2014, 2014, 1–8. [CrossRef]
- 17. Fun, H.-K.; Quah, C.K.; Attia, M.I.; Almutairi, M.S.; Ghoneim, S.W. (*E*)-*N*-[3-(imidazol-1-yl)-1-phenyl-propylidene]hydroxylamine. *Acta Cryst. E* 2012, *68*, o627. [CrossRef] [PubMed]
- 18. Sato, H.; Murakami, R.; Noda, I.; Ozaki, Y. Infrared and Raman spectroscopy and quantum chemistry calculation studies of C–H···O hydrogen bondings and thermal behavior of biodegradable polyhydroxyalkanoate. *J. Mol. Struct.* **2005**, *744*, 35–46. [CrossRef]
- 19. Reed, A.E.; Curtiss, L.A.; Weinhold, F. Intermolecular interactions from a natural bond orbital, donor-acceptor viewpoint. *Chem. Rev.* **1988**, *88*, 899–926. [CrossRef]
- Alabugin, I.V.; Manoharan, M.; Peabody, S.; Weinhold, F. Electronic basis of improper hydrogen bonding: A subtle balance of hyperconjugation and rehybridization. *J. Am. Chem. Soc.* 2003, 125, 5973–5987. [CrossRef] [PubMed]
- 21. Rauhut, G.; Pulay, P. Transferable scaling factors for density functional derived vibrational force fields. *J. Phys. Chem.* **1995**, *99*, 3093–3100. [CrossRef]
- 22. Fogarasi, G.; Zhou, X.; Taylor, P.W.; Pulay, P. The calculation of ab initio molecular geometries: Efficient optimization by natural internal coordinates and empirical correction by offset forces. *J. Am. Chem. Soc.* **1992**, *114*, 8191–8201. [CrossRef]
- 23. Billes, F.; Endrédi, H.; Jalsovszky, G. Vibrational spectroscopy of diazoles. *J. Mol. Struct. THEOCHEM* **1999**, 465, 157–172. [CrossRef]
- 24. Loeffen, P.W.; Pettifer, R.F.; Fillaux, F.; Kearley, G.J. Vibrational force field of solid imidazole from inelastic neutron scattering. *J. Chem. Phys.* **1995**, *103*, 8444–8455. [CrossRef]

- 25. Naumov, P.; Ristova, M.; Šoptrajanov, B.; Zugik, M. Vibrational spectra of bis(acetato)tetrakis(imidazole) copper(II). *J. Mol. Struct.* 2001, *598*, 235–243. [CrossRef]
- 26. Smith, B.C. Infrared Spectral Interpretation: A Systematic Approach; CRC Press: New York, NY, USA, 1999.
- 27. Mesu, J.G.; Visser, T.; Soulimani, F.; Weckhuysen, B.M. Infrared and Raman spectroscopic study of pH-induced structural changes of L-histidine in aqueous environment. *Vib. Spectrosc.* **2005**, *39*, 114–125. [CrossRef]
- 28. Ditchfield, R. Molecular orbital theory of magnetic shielding and magnetic susceptibility. *J. Chem. Phys.* **1972**, *56*, 5688–5691. [CrossRef]
- 29. Baxter, C.A.; Murray, C.W.; Waszkowycz, B.; Li, J.; Sykes, R.A.; Bone, R.G.; Perkins, T.D.; Wylie, W. New approach to molecular docking and its application to virtual screening of chemical databases. *J. Chem. Inf. Comput. Sci.* 2000, *40*, 254–262. [CrossRef] [PubMed]
- 30. Frisch, M.J.; Trucks, G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G.A.; et al. *Gaussian-09*; Revision A.02; Gaussian, Inc.: Wallingford, CT, USA, 2009.
- 31. Lagunin, A.; Stepanchikova, A.; Filimonov, D.; Poroikov, V. PASS: Prediction of activity spectra for biologically active substances. *Bioinformatics* **2000**, *16*, 747–748. [CrossRef] [PubMed]
- Podust, L.M.; Poulos, T.L.; Waterman, M.R. Crystal structure of cytochrome P450 14α-sterol demethylase (CYP51) from Mycobacterium tuberculosis in complex with azole inhibitors. *Proc. Nat. Acad. Sci. USA* 2001, 98, 3068–3073. [CrossRef] [PubMed]
- 33. Morris, G.M.; Huey, R.; Lindstrom, W.; Sanner, M.F.; Belew, R.K.; Goodsell, D.S.; Olson, A.J. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J. Comput. Chem.* **2009**, *30*, 2785–2791. [CrossRef] [PubMed]
- 34. Sanner, M.F. Python: A programming language for software integration and development. J. Mol. Graph. Model. 1999, 17, 57–61. [PubMed]
- 35. APEX2, SAINT and SADABS; Brucker AXS: Madison, WI, USA, 2009.
- 36. Sheldrick, G.M. A short history of SHELX. Acta Cryst. A 2008, 64, 112–122. [CrossRef] [PubMed]
- 37. Sundius, T. Molvib-A flexible program for force field calculations. J. Mol. Struct. 1990, 218, 321–326. [CrossRef]
- 38. Sundius, T. Scaling of ab initio force fields by MOLVIB. Vib. Spectrosc. 2002, 29, 89–95. [CrossRef]
- 39. Glendening, E.D.; Reed, A.E.; Carpenter, J.E.; Weinhold, F. *NBO*, version 3.1; Theoretical Chemistry Institute and Department of Chemistry, University of Wisconsin: Madison, WI, USA, 1998.
- 40. Rodriguez-Tudela, J.L.; Arendrup, M.C.; Barchiesi, F.; Bille, J.; Chryssanthou, E.; Cuenca-Estrella, M.; Dannaoui, E.; Denning, D.W.; Donnelly, J.P.; Dromer, F.; et al. EUCAST Definitive Document EDef 7.1: method for the determination of broth dilution MICs of antifungal agents for fermentative yeasts: Subcommittee on Antifungal Susceptibility Testing (AFST) of the ESCMID European Committee for Antimicrobial Susceptibility Testing (EUCAST). *Clin. Microbiol. Infect.* 2008, 14, 398–405.

Sample Availability: Samples of the synthesized compounds are available from the corresponding author.



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).